ANIMAL

Concentrate supplementation: A way to mitigate enteric methane emissions in pregnant Hanwoo heifers

Md Raihanul Hoque¹, Hyunjin Cho¹, Mingyung Lee¹, Jakeyom Seo², Sangsuk Lee³, Seongwon Seo^{1,*}

1 Division of Animal and Dairy Sciences, College of Agriculture and Life Sciences, Chungnam National University, Daejeon 34134, Korea

²Department of Animal Science, Life and Industry Convergence Research Institute, Pusan National University, Miryang 50463, Korea

 3 Department of Animal Science and Technology, Sunchon National University, Suncheon 57922, Korea

* Corresponding author: swseo@cnu.ac.kr

Abstract

The objective of this study was to investigate the effect of supplementing concentrates in a forage-based diet on methane emissions of pregnant Hanwoo heifers. Twenty-one pregnant Hanwoo heifers (481 \pm 42.4 kg) were divided into two groups: 1) a group receiving forage only (control, CON) and 2) the other group receiving forage with 4 kg of a concentrate mix (treatment, TRT). Methane (CH₄) concentration was measured using a laser methane detector, following an 18-d adaptation period, according to previously established protocols. Feed intake was recorded throughout the experimental period. Ruminal fluid was collected and analyzed for pH, ammonia-nitrogen (NH₃-N), and volatile fatty acid (VFA). The TRT exhibited higher dry matter and neutral detergent fiber intake than CON ($p < 0.05$) with elevated NH₃-N ($p < 0.001$) and total VFA concentrations ($p = 0.013$). The proportions of butyrate, valerate, and iso-valerate were higher in TRT than CON ($p < 0.05$). Notably, CH₄ concentrations per kg dry matter intake was lower in the TRT group, both from respiration and eructation ($p < 0.05$). In conclusion, supplementing concentrates in a low-quality forage-based diet for pregnant Hanwoo heifers fulfills nutrient requirements and reduces CH₄ emissions, suggesting a potential strategy to reduce environmental impact of Hanwoo production.

Keywords: eructation, Hanwoo, laser methane detection, respiration, rumen characteristics

Introduction

Pregnant beef heifers are often fed only forage as their diet (Carlson et al., 2022). Feeding high-quality forage hay should supply sufficient nutrients to meet the nutrient requirements of beef heifers (Adams et al., 1996). Nevertheless, supplementation of corn, by-products, and crop residues can offer economic benefits, given the reduction in perennial grasslands and the high

OPEN ACCESS

Citation: Hoque MR, Cho H, Lee M, Seo J, Lee S, Seo S. 2024. Concentrate supplementation: A way to mitigate enteric methane emissions in pregnant Hanwoo heifers. Korean Journal of Agricultural Science 51:283-294. https://doi. org/10.7744/kjoas.510304

Received: February 14, 2024

Revised: June 28, 2024

Accepted: July 16, 2024

Copyright: © 2024 Korean Journal of Agricultural Science

This is an Open Access article distributed under the terms

of the Creative Commons Attribution Non-Commercial License (https://creativecommons.org/ licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

cost of hay (Carlson et al., 2022). Moreover, in countries like Korea, which heavily rely on imported hay and domestic rice straw for cattle feeding (Chang, 2018), a forage-based diet is often found to be insufficient in providing necessary nutrients for cattle (Aquino et al., 2020). Inadequate nutrition during pregnancy potentially leads to complications such as calf mortalities and respiratory or intestinal dysfunction (Wu et al., 2006). Thus, maintaining body condition during gestation and lactation by providing sufficient nutrients is crucial for the development of the uterus, fetal growth, and the subsequent calf's weight gain (Funston et al., 2010). In this regard, concentrate supplementation is crucial and cost-effective for the productivity of low-quality forage-feeding heifers (Pritchard and Males, 1982).

In addition to its nutritional and economic advantages, concentrate supplementation may also offer environmental benefits. Dietary forage leads to higher production of acetate and hydrogen during anaerobic fermentation, resulting in increased methane (CH4) production (Janssen, 2010). Methane is one of the greenhouse gases produced by livestock farming, which is reported to be responsible for 30% of global greenhouse gas emissions (FAO, 2023). Furthermore, methanogenesis in the rumen also represents an energy loss, ranging from 2 to 12% of the gross energy intake (Johnson and Johnson, 1995). Previous research has demonstrated that concentrate supplementation (forage : concentrate = 50 : 50) in dairy cows reduces CH4 production per unit of milk production (Patel et al., 2011). Bayat et al. (2017) reported that a diet with a forage-to-concentrate ratio of 35 : 65 reduced CH₄ emissions, lowered pH levels, and increased volatile fatty acid (VFA) production by organic matter intake in lactating cows. In Jersey and Holstein cows, concentrate supplementation at levels of 70% and 91% led to a modified rumen VFA profile and reduced CH₄ gas emissions (Olijhoek et al., 2022). Based on these findings, it is hypothesized that concentrate mix supplementation can meet the nutrient requirements of pregnant heifers while simultaneously reducing CH4 emissions. However, this has not yet been studied in pregnant Hanwoo heifers.

Furthermore, CH4 emissions from cattle in farm settings should be assessed with a large number of animals in practical conditions. The laser methane detection (LMD) method has been proven effective for this purpose (Chagunda et al., 2013). The LMD utilizes infrared absorption spectroscopy and incorporates a semiconductor laser as a focused excitation source. It employs the technique of second harmonic detection in wavelength modulation spectroscopy to determine the concentration of CH4 (Chagunda et al., 2009). Kang et al. (2022) have developed protocols and further validated the LMD's capability to distinguish CH₄ emissions in a large number of cattle under practical conditions. Therefore, this study aimed to assess CH4 emissions using LMD in a large population of pregnant Hanwoo heifers under practical field conditions when a low-quality forage-based diet was supplemented with a concentrate mix to provide more nutrients.

Materials and Methods

This study was conducted at the Center for Animal Science Research, Chungnam National University (CNU), Korea. The use of animals and protocols for this experiment were reviewed and pre-approved by the CNU Animal Research Ethics Committee (202203A-CNU-058).

Animals, experimental design, and diet

A total of twenty-one Hanwoo heifers $(481 \pm 42.4 \text{ kg})$ were used in this experiment. All the Hanwoo heifers were bought when they were of similar age (8 mo) from a local commercial Hanwoo market (Korea). They were vaccinated upon arrival at the CNU Animal Science Research Centre and were raised here until the experiment. Initially, there were 25 heifers, of which 4 were excluded because they were not pregnant. The experiment lasted for 24 d, consisting of 18 d of adaptation and 6 d of measuring periods. Each two heifers of similar BW were housed in a pen (5 m \times 5 m) equipped with a forage feed bin, which enabled us to automatically determine individual forage intake (Dawoon Co., Korea). Heifers were divided into two groups: 1) a group receiving forage only $(n = 11$; control, CON) and 2) the other group receiving forage supplemented with 4 kg as-fed of a concentrate mix ($n = 10$; treatment, TRT). The formulation and chemical composition of the experimental diets are shown in Tables 1 and 2, respectively. The heifers were fed twice daily at 08:00 and 18:00 h. Forage and drinking water were freely accessible to the animals throughout the experiment. For the TRT group, each heifer was individually offered a concentrate mix of 4 kg per day. These pregnant heifers had a 30-min to consume the concentrate mix, after which any remaining amount was measured to determine actual intake. Roughage intake was measured daily using an automatic feed intake measurement device (Dawoon Co., Korea). Consequently, this allowed for the measurement of dry matter intake (DMI) during the experimental period.

^z 33,330,000 IU·kg⁻¹ vitamin A; 40,000,000 IU·kg⁻¹ vitamin D; 20.86 IU·kg⁻¹ vitamin E; 20 mg·kg⁻¹ Cu; 90 mg·kg⁻¹ Mn; 100 mg·kg-1 Zn; 250 mg·kg-1 Fe; 0.4 mg·kg-1 I; and 0.4 mg·kg-1 Se.

| Items | Concentrate mix | Tall fescue |
|---|-----------------|----------------|
| ADICP | 16 | 9 |
| aNDF | 418 | 716 |
| ${\sf ADF}$ | 246 | 426 |
| $\mbox{\rm ADL}$ | 70 | 59 |
| Ether extract | $22\,$ | $11\,$ |
| ${\boldsymbol{\rm A}}{\boldsymbol{\rm sh}}$ | 114 | $70\,$ |
| Ca | 17 | \mathfrak{Z} |
| $\, {\bf p}$ | 6 | $\sqrt{2}$ |
| $\rm K$ | 12 | 23 |
| $\rm Na$ | $\overline{4}$ | $\,1\,$ |
| $\mathop{\rm Cl}\nolimits$ | 9 | $\sqrt{5}$ |
| S | $\overline{4}$ | $\sqrt{2}$ |
| Mg | $\sqrt{4}$ | $\sqrt{2}$ |
| TDN | 604 | 528 |
| $\text{NEm}\left(\text{MJ}\!\cdot\!\text{kg}^{\text{-1}}\,\text{DM}\right)$ | 5.8 | 4.4 |
| $NEg (MJ·kg-1 DM)$ | 3.3 | 2.1 |
| Total carbohydrates | 709 | 847 |
| ${\rm NFC}$ | 309 | 144 |
| Carbohydrate fraction $(g \cdot kg^{-1}$ carbohydrate) | | |
| ${\rm CA}$ | $87\,$ | 63 |
| CB1 | 253 | $\sqrt{2}$ |
| CB2 | 96 | 106 |
| CB ₃ | 299 | 660 |
| CC | 235 | 168 |
| Protein fraction (g·kg ⁻¹ CP) | | |
| $PA + B1$ | 378 | 386 |
| PB ₂ | 368 | 415 |
| PB3 | 149 | 73 |
| ${\rm P}{\bf C}$ | 105 | 126 |

Table 2. Analyzed chemical composition (g·kg⁻¹DM or as stated) of the experimental diets.

DM, dry matter; OM, organic matter; CP, crude protein; SOLP, soluble CP; NDICP, neutral detergent insoluble CP; ADICP, acid detergent insoluble CP; aNDF, neutral detergent fiber analyzed using a heat stable amylase and expressed inclusive of residual ash; ADF, acid detergent fiber; ADL, acid detergent lignin; TDN, total digestible nutrients; NEm, net energy for maintenance; NEg, net energy for growth; NFC, non-fiber carbohydrate; CA, carbohydrate A fraction (ethanol soluble carbohydrates); CB1, carbohydrate B1 fraction (starch); CB2, carbohydrate B2 fraction (soluble fiber); CB3, carbohydrate B3 fraction (available insoluble fiber); CC, carbohydrate C fraction (unavailable carbohydrate); PA + B1, protein A and B1 fractions (soluble CP); PB2, protein B2 fraction (intermediate degradable CP); PB3, protein B3 fraction (slowly degradable fiber-bound CP); PC, protein C fraction (unavailable CP).

Measuring CH4 emissions using LMD

The CH4 concentration in the exhaled gas of the animals was measured using the LMD according to Kang et al. (2022). Briefly, the LMD was installed on a tripod, aiming at the animal's nostril from a distance of 1 m. The CH₄ concentrations were measured every 0.5 s for 6 min. The measurements were performed four times $(-2, -1, \text{ and } +1, +2)$ h after the morning feed) daily for all heifers, which were duplicated over 2 d. For data analysis, the peaks of the CH₄ concentration measured by the LMD were detected using the automatic multiscale-based peak detection (AMPD) R package. The peaks were divided into two pathways (respiration and eructation) by fitting a double normal distribution using the mixdist R package. A larger number of peaks belonged to respiration, but they were of lower value. The mean of the normal distribution was assumed to be the representative CH_4 concentration of the exhaled gas from the pathway for the hour. The four-time values of a day were averaged to represent the mean CH4 concentration of the day, and the 2 d' mean values were averaged.

Feed analysis

The diet samples were dried at 60℃ for 96 h and ground through a cyclone mill (Foss, Denmark) and screened through a 1 mm screen before chemical analysis. The nutrient composition of the samples was analyzed at Cumberland Valley Analytical Services Inc. (USA). Feed analysis procedures are described in detail in another study (Jeon et al., 2016). The contents of DM (#934.15), CP (#990.03), ether extract (#920.39), acid detergent fiber (#973.18), and ash (#942.05) were determined according to AOAC International (2006). The total nitrogen was measured by the Dumas method using a Leco FP-528 Nitrogen Combustion Analyzer (Leco Inc., USA). CP was calculated as 6.25 times the nitrogen content. A heat-stable amylase was used to assess the acid detergent lignin (ADL) content and the neutral detergent fiber (NDF) content, which was then expressed inclusive of residual ash (aNDF). The soluble protein, neutral detergent insoluble crude protein (NDICP), and acid detergent insoluble crude protein (ADICP) contents were also determined. The contents of ethanol soluble carbohydrates (ESC), starch, and macro and micro-mineral content were determined.

The content of total digestible nutrient (TDN), net energy for maintenance, and net energy for growth were estimated using NRC (2001) equations. The dietary carbohydrate and protein fractions were estimated according to the Cornell Net Carbohydrate and Protein System (Fox, 2003) with the following modifications. The different carbohydrate fractions were classified as follows: Carbohydrate A fraction (CA) represented sugars and organic acids and was assumed to be equivalent to ESC. Carbohydrate B1 fraction (CB1) referred to starch. Carbohydrate B2 fraction (CB2) was calculated as the soluble fiber, obtained by subtracting CA and CB1 from NFC (non fiber carbohydrates). Carbohydrate B3 fraction (CB3) represented the available NDF, estimated by subtracting 2.4 times ADL from aNDF. Carbohydrate C fraction (CC) indicated the unavailable carbohydrate, estimated as 2.4 times ADL. Concerning the protein fractions, $PA + B1$ (protein A and B1 fractions) denoted the soluble protein, which was identical to soluble CP (crude protein) or soluble CP (SOLP). PB2 referred to the intermediate degradable CP, estimated as 100-NDICP-SOLP. Protein B3 fraction (PB3) represented the slowly degradable fiber-bound CP, estimated as NDICP-ADICP. Lastly, protein C fraction (PC) indicated the unavailable CP, which was equal to ADICP. All the carbohydrate and protein fractions were expressed as grams per kilogram of total carbohydrate or CP, respectively. Minerals were analyzed according to AOAC International (2006) method.

Rumen sampling and analysis

Rumen samples were collected from all heifers over three consecutive days to ensure the acquisition of representative samples for each feeding cycle: at 14:00 (6 h post-feeding) on day 1, 11:00 (3 h post-feeding) on day 2, and 07:00 (before feeding) on day 3. The collection was performed using esophageal stomach tubing, following the method described by Lee et al. (2019). Briefly, after discarding the initially obtained ruminal fluid (approximately 200 mL), 500 mL of ruminal fluid was collected in a glass flask. The rumen contents were instantly subjected to pH measurement (EcoMet P25, Istek Inc., Korea), and the remaining samples were stored at -20℃.

Samples were then transferred to the laboratory, where they were thawed and centrifuged at $14,000 \times g$ for 10 min at 4℃ for VFA and ammonia-N analysis, as described in detail by Lee et al. (2019). Briefly, for the VFA analysis, ruminal fluid supernatant (1 mL) was mixed with 0.2 mL of metaphosphoric acid (250 g·L⁻¹) and kept at 4[°]C for 30 min. Following centrifugation of the mixture at $14,000 \times g$ for 10 min at room temperature, the supernatant was injected into a gas chromatograph (HP 6890, Hewlett-Packard Co., USA) equipped with a flame ionization detector and capillary column (Nukol Fused silica capillary column 30 m \times 0.25 mm \times 0.2 µm, Supelco Inc., USA). The temperature of the oven, injector, and detector was 90℃ to 180℃, 185℃, and 210℃, respectively. Nitrogen was used as the carrier gas at a flow rate of 40 mL·min⁻¹. For the NH₃-N analysis, the stored ruminal fluid underwent re-centrifugation at 21,000 \times g for 15 min, and 20 µL of the supernatant was mixed with 1 mL of phenol color reagent and 1 mL of alkali-hypochlorite reagent. The mixture was then incubated in a water bath for 15 min at 37℃. After being mixed with 8 mL of distilled water, the optical density of the mixture was measured at 630 nm, using a spectrophotometer (UV-1800, Shimadzu Corp., Japan).

Statistical analysis

The animals were used as unit per treatment where animal number was within power range. The data were checked for normality and homoscedasticity before further analysis. All statistical analysis was performed using SAS software (SAS Institute Inc., USA). The data was analyzed using PROC TTEST. Significance was declared at p < 0.05, and a trend was discussed at $0.05 \le p < 0.1$.

Results and Discussion

The metabolic demands for energy and protein escalate during the latter stage of gestation in pregnant heifers, a critical period for sustaining both maternal health and fetal growth (Patterson et al., 2003; Gionbelli et al., 2016). Considering the nutritional requirement, it might be necessary to supplement energy and protein, especially when the diet is based on low-quality forage (Schillo et al., 1992).

The results of concentrate supplementation on feed intake are shown in Table 3. In the TRT group, DMI increased by 2.4 kg per day compared to CON ($p < 0.001$). This rise was mainly due to the voluntary intake of concentrate mix at the rate of 3.5 kg DM. Furthermore, concentrate supplementation resulted in a 24% decrease in forage intake in the TRT group ($p = 0.002$). However, despite this reduction in forage consumption, there was a 38% increase in total NDF intake (NDFI) due to the concentrate supplementation ($p = 0.014$).

In cattle, when a nutrient-deficient diet is supplemented with concentrate, DMI increases, which aligns with our findings (DelCurto et al., 2000). Stafford et al. (1996) indicated a 16% increase in total DMI when poor quality tallgrass was supplemented with a concentrate mix (CP 17.5%) at 0.15% of BW in growing beef steers. In their study, the concentrate supplementation did not alter forage intake. However, Loy et al. (2007) reported decreased forage DMI in beef heifers when forage was supplemented with concentrates. In their study (Loy et al., 2007), concentrate supplementation at 0.40% of BW increased total DMI by 12%, while forage feed intake was reduced by 10% in heifers. Conversely, protein supplementation could increase forage intake (DelCurto et al., 2000). The impact of concentrate supplementation in forage and total DMI appears to depend on nutritional balance, frequency of feeding, and the types of forage and concentrates in the diet (Huston et al., 1999; Loy et al., 2007).

| Items (kg) | Treatment | | | |
|-------------------|------------|------------|------------|--------------|
| | CON | TRT | SEM | p-value |
| Initial BW | 480 | 483 | 19.2 | 0.851 |
| fDMI | 4.2 | 3.2 | 0.36 | 0.002 |
| cDMI | ٠ | 3.5 | ٠ | - |
| Total DMI | 4.2 | 6.8 | 0.44 | ${}_{0.001}$ |
| Total NDFI | 2.9 | 3.8 | 0.30 | 0.014 |

Table 3. Effect of different forage to concentrate ratio diet on feed intake of Hanwoo heifers.

CON (control), receiving forage only; TRT (treatment), receiving forage with 4 kg concentrate mix; SEM, standard error of the mean; BW, body weight; fDMI, forage dry matter intake; cDMI, concentrate dry matter intake; DMI, dry matter intake; NDFI, neutral detergent fiber intake.

Supplementation of concentrate slightly reduced ($p < 0.05$) the pH of ruminal fluid in the TRT group (Table 4). The concentrate supplementation significantly elevated rumen NH₃-N concentration ($p \le 0.001$), maintaining it above 5 $mg \cdot L^{-1}$, which is the minimum concentration for optimal rumen microbial protein synthesis (Firkins et al., 2007). There was also a 12% increase in the total VFA concentration in the rumen of TRT group animals ($p = 0.013$). In the TRT group, rumen acetate concentration was decreased compared to CON ($p < 0.001$). On the other hand, concentrate supplementation increased the concentrations of butyrate ($p < 0.001$), valerate ($p < 0.001$), and iso-valerate ($p = 0.041$) in the rumen. Propionate concentration did not differ between the groups $(p > 0.05)$. Finally, concentrate supplementation tended to decrease the acetate : propionate ratio ($p = 0.074$).

| Items | Treatment | | SEM | p-value |
|---|------------|------------|------------|--------------|
| | CON | TRT | | |
| pH | 6.90 | 6.72 | 0.084 | 0.032 |
| NH_3-N (mg·dL ⁻¹) | 3.12 | 6.79 | 0.789 | ${}_{0.001}$ |
| Total VFA (mM) | 65.7 | 73.9 | 2.98 | 0.013 |
| Acetate (mmol·mol ⁻¹) | 691 | 656 | 4.9 | ${}_{0.001}$ |
| Propionate ($mmol·mol·$) | 187 | 185 | 3.1 | 0.517 |
| Butyrate (mmol·mol ⁻¹) | 73 | 105 | 2.9 | ${}_{0.001}$ |
| Iso-butyrate (mmol·mol ⁻¹) | 19 | 19 | 0.9 | 0.615 |
| Valerate (mmol·mol ⁻¹) | 14 | 18 | 0.6 | ${}_{0.001}$ |
| $Iso\text{-}valerate \text{ (mmol·}mol^{-1})$ | 16 | 18 | 0.8 | 0.041 |
| Acetate: Propionate | 3.69 | 3.55 | 0.072 | 0.074 |

Table 4. Effect of different forage to concentrate ratio diet on ruminal fluid characteristics of Hanwoo heifers.

CON (control), receiving forage only; TRT (treatment), receiving forage with 4 kg concentrate mix; SEM, standard error of the mean; NH3-N, ammonia nitrogen; VFA, volatile fatty acid.

In our study, ruminal NH₃-N concentration in the CON group was only 3.12 mg·dL⁻¹, which might limit the metabolizable protein (MP) supply to the animals. A ruminal NH₃-N concentration below 5 mg·dL⁻¹ is generally considered insufficient for adequate ruminal microbial protein synthesis (Firkins et al., 2007), which is crucial for meeting the MP requirements of ruminants. This lower $NH₃-N$ concentration in the CON group might be an indicator of nutrient deficiency of a forage-based diet. With high-quality forage, this might not be an issue, as it can provide sufficient NH3-N for effective microbial protein synthesis (Broderick, 1995). However, low-quality forage often requires concentrate supplementation, recognizing the importance of nitrogen availability.

Concentrate supplementation altered the rumen ecosystem as expected. It increased total VFA production and reduced pH in the rumen, which is consistent with previous studies (Sun et al., 2010; Jiang et al., 2022). Highly fermentable carbohydrate supply due to concentrate supplementation can reduce rumen pH by increasing total VFA and other organic acid production (Stone, 2004). Furthermore, consistent with existing research, we observed a reduction in acetate production. Concentrate supplementation reduces the proportion of insoluble carbohydrates and NDF, which are primary producers of acetate (Sairanen et al., 2005). Typically, a high level of concentrate supplementation (around 70% of dietary DM) increases propionate production in the rumen (Olijhoek et al., 2018). However, our study diverged from this trend, showing no difference in propionate concentration. The reason may be attributed to the limited quantity of concentrate supplementation, constituting approximately 50 - 60% of the total diet. The non-fiber carbohydrate (NFC) of the concentrate might not have been sufficiently high enough to stimulate propionate production (Sairanen et al., 2005; Moorby et al., 2006). Furthermore, our study corroborated previous findings indicating an increase in butyrate concentration following concentrate supplementation (Sairanen et al., 2005; Miguel et al., 2019). Butyrate production, however, is not solely contingent on the diet, as acetate and butyrate are interconvertible between each other in the rumen. A portion of acetate may be converted to butyrate before absorption (Kristensen, 2001). Supplementation of concentrate can act as a stimulant for butyrate-producing microorganisms and promote the conversion of acetate to butyrate (Hackmann and Firkins, 2015). Similarly, the observed increase in valerate concentration in our study was aligned with previous study (Bayat et al., 2017). Within the rumen, acetate and propionate can be elongated to valerate and iso-valerate (Chen et al., 2011).

Concentrate supplementation reduced CH4 concentration per kg DMI in pregnant Hanwoo heifers (Table 5). In the TRT group, the CH₄ concentration per kg DMI during respiration was 32% lower than CON ($p = 0.002$). Similarly,

| Items | Treatment | | | |
|------------------------|------------|------------|------------|---------|
| | CON | TRT | SEM | p-value |
| $CH4$ from respiration | | | | |
| ppm | 7.8 | 8.6 | 0.48 | 0.108 |
| ppm per kg DMI | 1.9 | 1.3 | 0.20 | 0.002 |
| ppm per kg NDFI | 2.8 | 2.3 | 0.30 | 0.139 |
| $CH4$ from eructation | | | | |
| ppm | 48.1 | 54.2 | 4.49 | 0.196 |
| ppm per kg DMI | 12.2 | 8.1 | 1.45 | 0.011 |
| ppm per kg NDFI | 17.1 | 14.6 | 2.20 | 0.272 |

Table 5. Effect of different forage to concentrate ratio diet on methane emissions of Hanwoo heifers.

CON (control), receiving forage only; TRT (treatment), receiving forage with 4 kg concentrate mix; SEM, standard error of the mean; DMI, dry matter intake; NDFI, neutral detergent fiber intake.

concentrate supplementation reduced the CH4 concentration per kg DMI during eructation by 34% in the TRT group compared to CON ($p = 0.011$). Methane concentration in the exhaled gas from respiration and eructation per kg of NDFI were not different between the groups ($p > 0.05$).

Laser Methane Detector does not give a total CH4 measurement for a day. However, short time measurements by LMD as ppm and ppm per kg DMI can correlate positively to the 24-h measurements of CH₄ (Kobayashi et al., 2021; Kang et al., 2022). Increasing concentrates in the diet at a level of 40% or more is widely acknowledged as a way to mitigate CH4 production (Knapp et al., 2014). In our study, the concentration of CH4 per kg DMI, was reduced through a diet supplemented with 50 - 60% concentrate. This aligns with the results from Bayat et al. (2017), where a 35 : 65 forage-to-concentrate ratio diet in cows resulted in a similar outcome. Similarly, a diet comprising 70% concentrate demonstrated a reduction in CH4 per kg of DMI in beef cattle (Doreau et al., 2011). Beef heifers on a 90% concentrate diet and cows on a 61% concentrate diet also exhibited a comparable decrease in CH4 per kg of DMI (Lovett et al., 2003; Olijhoek et al., 2018). Aguerre et al. (2011) supplemented a 47 : 53 forage-to-concentrate ratio diet and found a linear reduction of CH4 per kg of DMI emissions in cows. The decrease in CH4 production appears to be associated with a shift in rumen fermentation from acetate to other VFA like propionate, butyrate and valerate, along with a reduction of rumen pH (Martin et al., 2010). Propionate, butyrate and valerate production, as opposed to acetate production, serves as an alternative H₂ sink, diminishing its availability for methanogenesis (Moss et al., 2000). Additionally, high-concentrate diets, which are characterized by elevated starch levels, are linked to a decline in the population of ciliated protozoa (Morgavi et al., 2010). This reduction in protozoa, combined with a high starch diet and reduced pH, can contribute to decreased CH4 production, especially since certain methanogens harboring ciliated protozoa are pH sensitive (Newbold et al., 1995). However, contrasting results exist. A low-level increase in the concentrate content in the diet does not consistently lead to a reduction in CH4. Lovett et al. (2003) observed an increase in CH4 g per kg DMI when feeding heifers a diet with a 40 : 60 forage-to-concentrate ratio. Similarly, an 8 kg·d⁻¹ concentrate supplementation did not impact CH₄ emissions from Jersey cows (Van Wyngaard et al., 2018). Another study comparing 1 kg·d⁻¹ vs. 5 kg·d⁻¹ concentrate supplementation in cows revealed no difference in CH₄ per kg DMI but did show an increase in daily CH4 production (Muñoz et al., 2015). The observed variations in CH⁴ production could be attributed to factors such as forage quality, the *ad libitum* supply of forage, DMI from forage and concentrate and variations in CH4 measuring methods (Muñoz et al., 2015). In our study, concentrate supplementation led to an increase in soluble carbohydrates in the diet, a reduction in rumen pH, and the promotion of alternative H_2 sinks by diverting VFA production.

Conclusion

Supplementing pregnant Hanwoo heifers with 4 kg of as-fed concentrate where average 3.5 kg of DMI from concentrate mix per animal effectively increased their DMI and ensured the provision of essential nutrients necessary for maintaining optimal rumen conditions. Additionally, this supplementation contributed to a notable reduction in CH4 yield. Therefore, concentrate supplementation for pregnant heifers offers economic benefits, fulfills nutritional requirements, and presents environmental advantages. These collective benefits highlight its potential as a suitable strategy in Hanwoo cattle production.

Data Availability Statement

The data presented in this study are available on request from the corresponding author.

Conflict of Interests

No potential conflict of interest relevant to this article was reported.

Acknowledgements

This work was supported by Korea Institute of Planning and Evaluation for Technology in Food, Agriculture and Forestry (IPET) through Livestock Industrialization Technology Development Program, funded by Ministry of Agriculture, Food and Rural Affairs (321083-5).

Author Information

Seongwon Seo, https://orcid.org/0000-0002-4131-0545

References

- Adams DC, Clark RT, Klopfenstein TJ, Volesky JD. 1996. Matching the cow with forage resources. Rangelands 18: 57-62.
- Aguerre MJ, Wattiaux MA, Powell J, Broderick GA, Arndt C. 2011. Effect of forage-to-concentrate ratio in dairy cow diets on emission of methane, carbon dioxide, and ammonia, lactation performance, and manure excretion. Journal of Dairy Science 94:3081-3093.
- AOAC (Association of Official Analytical Chemists) International. 2006. Official Methods of Analysis of AOAC International (18th). AOAC International, Gaithersburg, MD, USA.
- Aquino D, Del Barrio A, Trach NX, Hai NT, Khang DN, Toan NT, Van Hung N. 2020. Rice straw-based fodder for ruminants. In Sustainable Rice Straw Management edited by Gummert M, Hung N, Chivenge P, Douthwaite B. pp. 111-129. Springer, Cham, Switzerland.
- Bayat A, Ventto L, Kairenius P, Stefański T, Leskinen H, Tapio I, Shingfield K. 2017. Dietary forage to concentrate ratio and sunflower oil supplement alter rumen fermentation, ruminal methane emissions, and nutrient utilization in lactating cows. Translational Animal Science 1:277-286.
- Broderick GA. 1995. Desirable characteristics of forage legumes for improving protein utilization in ruminants. Journal of Animal Science 73:2760-2773.
- Carlson ZE, McPhillips LJ, Erickson GE, Drewnoski ME, MacDonald JC. 2022. Production cow-calf responses from perennial forage-based and integrated beef-cropping systems. Translational Animal Science 6:txac090.
- Chagunda M, Ross D, Roberts D. 2009. On the use of a laser methane detector in dairy cows. Computers and Electronics in Agriculture 68:157-160.
- Chagunda M, Ross D, Rooke J, Yan T, Douglas JL, Poret L, Roberts D. 2013. Measurement of enteric methane from ruminants using a hand-held laser methane detector. Acta Agriculturae Scandinavica, Section A — Animal Science 63:68-75.
- Chang JB. 2018. The effects of forage policy on feed costs in Korea. Agriculture 8:72.
- Chen Y, Penner GB, Li M, Oba M, Guan LL. 2011. Changes in bacterial diversity associated with epithelial tissue in the beef cow rumen during the transition to a high-grain diet. Applied and Environmental Microbiology 77:5770-5781.
- DelCurto T, Hess B, Huston J, Olson K. 2000. Optimum supplementation strategies for beef cattle consuming lowquality roughages. Journal of Animal Science 77:1-16.
- Doreau M, Van Der Werf H, Micol D, Dubroeucq H, Agabriel J, Rochette Y, Martin C. 2011. Enteric methane production and greenhouse gases balance of diets differing in concentrate in the fattening phase of a beef production system. Journal of Animal Science 89:2518-2528.
- FAO (Food and Agriculture Organization of the United Nations). 2023. Reducing enteric methane for improving food security and livelihoods. Accessed in https://www.ccacoalition.org/resources/reducing-enteric-methane-impro ving-food-security-and-livelihoods on 11 November, 2023.
- Firkins J, Yu Z, Morrison M. 2007. Ruminal nitrogen metabolism: Perspectives for integration of microbiology and nutrition for dairy. Journal of Dairy Science 90:E1–E16.
- Fox D. 2003. The Net Carbohydrate and Protein System for evaluating herd nutrition and nutrient excretion: Model documentation. Animal Science Mimeo of Cornell University 213:1-292.
- Funston RN, Larson DM, Vonnahme K. 2010. Effects of maternal nutrition on conceptus growth and offspring performance: Implications for beef cattle production. Journal of Animal Science 88:E205–E215.
- Gionbelli MP, Valadares Filho SC, Duarte MS. 2016. Nutritional requirements for pregnant and non-pregnant beef cows. In Nutrient Requirements of Zebu and Crossbred Cattle (3rd) edited by Valadares Filho SC, Costa e Silva LFC, Gionbelli MP, Rotta PP, Marcondes MI, Chizzotti ML, Prados LF. pp. 251-272. Federal University of Viçosa, Viçosa, Brazil.
- Hackmann TJ, Firkins JL. 2015. Maximizing efficiency of rumen microbial protein production. Frontiers in Microbiology 6:465.
- Huston J, Lippke H, Forbes T, Holloway J, Machen R. 1999. Effects of supplemental feeding interval on adult cows in western Texas. Journal of Animal Science 77:3057-3067.
- Janssen PH. 2010. Influence of hydrogen on rumen methane formation and fermentation balances through microbial growth kinetics and fermentation thermodynamics. Animal Feed Science and Technology 160:1-22.
- Jeon S, Sohn KN, Seo S. 2016. Evaluation of feed value of a by-product of pickled radish for ruminants: Analyses of nutrient composition, storage stability, and in vitro ruminal fermentation. Journal of Animal Science and Technology 58:34.
- Jiang Y, Dai P, Dai Q, Ma J, Wang Z, Hu R, Xue B. 2022. Effects of the higher concentrate ratio on the production performance, ruminal fermentation, and morphological structure in male cattle-yaks. Veterinary Medicine and Science 8:771-780.
- Johnson KA, Johnson DE. 1995. Methane emissions from cattle. Journal of Animal Science 73:2483-2492.
- Kang K, Cho H, Jeong S, Jeon S, Lee M, Lee S, Seo S. 2022. Application of a hand-held laser methane detector for measuring enteric methane emissions from cattle in intensive farming. Journal of Animal Science 100:skac211.
- Knapp JR, Laur G, Vadas PA, Weiss WP, Tricarico JM. 2014. *Invited review*: Enteric methane in dairy cattle production: Quantifying the opportunities and impact of reducing emissions. Journal of Dairy Science 97:3231-3261.
- Kobayashi N, Hou F, Tsunekawa A, Yan T, Tegegne F, Tassew A, Mekonnen W. 2021. Laser methane detector-based quantification of methane emissions from indoor-fed Fogera dairy cows. Animal Bioscience 34:1415-1424.
- Kristensen NB. 2001. Rumen microbial sequestration of [2-¹³C] acetate in cattle. Journal of Animal Science 79:2491-2498.
- Lee M, Jeong S, Seo J, Seo S. 2019. Changes in the ruminal fermentation and bacterial community structure by a sudden change to a high-concentrate diet in Korean domestic ruminants. Asian-Australasian Journal of Animal Sciences 32:92-102.
- Lovett D, Lovell S, Stack L, Callan J, Finlay M, Conolly J, O'Mara F. 2003. Effect of forage/concentrate ratio and dietary coconut oil level on methane output and performance of finishing beef heifers. Livestock Production Science 84:135-146.
- Loy TW, MacDonald JC, Klopfenstein TJ, Erickson GE. 2007. Effect of distillers grains or corn supplementation fre-

quency on forage intake and digestibility. Journal of Animal Science 85:2625-2630.

- Martin C, Morgavi DP, Doreau M. 2010. Methane mitigation in ruminants: From microbe to the farm scale. Animal 4: 351-365.
- Miguel MA, Lee SS, Mamuad LL, Choi YJ, Jeong CD, Son A, Lee SS. 2019. Enhancing butyrate production, ruminal fermentation and microbial population through supplementation with Clostridium saccharobutylicum. Journal of Microbiology and Biotechnology 29:1083-1095.
- Moorby JM, Dewhurst RJ, Evans RT, Danelon J. 2006. Effects of dairy cow diet forage proportion on duodenal nutrient supply and urinary purine derivative excretion. Journal of Dairy Science 89:3552-3562.
- Morgavi D, Foran, E, Martin C, Newbold CJ. 2010. Microbial ecosystem and methanogenesis in ruminants. Animal 4:1024-1036.
- Moss AR, Jouany JP, Newbold J. 2000. Methane production by ruminants: Its contribution to global warming. Annales de Zootechnie 49:231-253.
- Muñoz C, Hube S, Morales JM, Yan T, Ungerfeld EM. 2015. Effects of concentrate supplementation on enteric methane emissions and milk production of grazing dairy cows. Livestock Science 175:37-46.
- Newbold C, Lassalas B, Jouany J. 1995. The importance of methanogens associated with ciliate protozoa in ruminal methane production in vitro. Letters in Applied Microbiology 21:230-234.
- NRC (National Research Council). 2001. Nutrient Requirements of Dairy Cattle (7th). The National Academies Press, Washington, D.C., USA.
- Olijhoek D, Hellwing A, Noel S, Lund P, Larsen M, Weisbjerg M, Børsting C. 2022. Feeding up to 91% concentrate to Holstein and Jersey dairy cows: Effects on enteric methane emission, rumen fermentation and bacterial community, digestibility, production, and feeding behavior. Journal of Dairy Science 105:9523-9541.
- Olijhoek D, Løvendahl P, Lassen J, Hellwing A, Höglund J, Weisbjerg M, Lund P. 2018. Methane production, rumen fermentation, and diet digestibility of Holstein and Jersey dairy cows being divergent in residual feed intake and fed at 2 forage-to-concentrate ratios. Journal of Dairy Science 101:9926-9940.
- Patel M, Wredle E, Börjesson G, Danielsson R, Iwaasa A, Spörndly E, Bertilsson J. 2011. Enteric methane emissions from dairy cows fed different proportions of highly digestible grass silage. Acta Agriculturae Scandinavica, Section A — Animal Science 61:128-136.
- Patterson H, Adams DC, Klopfenstein TJ, Clark R, Teichert B. 2003. Supplementation to meet metabolizable protein requirements of primiparous beef heifers: II. Pregnancy and economics. Journal of Animal Science 81:563-570.
- Pritchard RH, Males JR. 1982. Effect of supplementation of wheat straw diets twice a day on rumen ammonia, volatile fatty acids and cow performance. Journal of Animal Science 54:1243-1250.
- Sairanen A, Khalili H, Nousiainen JI, Ahvenjärvi S, Huhtanen P. 2005. The effect of concentrate supplementation on nutrient flow to the omasum in dairy cows receiving freshly cut grass. Journal of Dairy Science 88:1443-1453.
- Schillo KK, Hall JB, Hileman SM. 1992. Effects of nutrition and season on the onset of puberty in the beef heifer. Journal of Animal Science 70:3994-4005.
- Stafford SD, Cochran R, Vanzant E, Fritz J. 1996. Evaluation of the potential of supplements to substitute for lowquality, tallgrass-prairie forage. Journal of Animal Science 74:639-647.
- Stone W. 2004. Nutritional approaches to minimize subacute ruminal acidosis and laminitis in dairy cattle. Journal of Dairy Science 87:E13-E26.
- Sun Y, Mao S, Zhu W. 2010. Rumen chemical and bacterial changes during stepwise adaptation to a high-concentrate diet in goats. Animal 4:210-217.
- Van Wyngaard J, Meeske R, Erasmus LJ. 2018. Effect of concentrate feeding level on methane emissions, production performance and rumen fermentation of Jersey cows grazing ryegrass pasture during spring. Animal Feed Science and Technology 241:121-132.
- Wu G, Bazer F, Wallace J, Spencer T. 2006. Board-Invited Review: Intrauterine growth retardation: Implications for the animal sciences. Journal of Animal Science 84:2316-2337.