

Effects of phytogetic feed additives in growing and finishing pigs under different stocking density

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Abstract

This study was to investigate effects of different phytogetic feed additives (PFA) in grower finishing pigs with stressed by high stocking density. A total of 84 growing pigs ([Landrace × Yorkshire] × Duroc) with initial body weight (BW) of 28.23 ± 0.21 kg were used for 10 weeks (4 replicate pens with 3 pigs per pen). The dietary treatment consisted of basal diets in animal welfare density (positive control [PC]), basal diet in high stocking density (negative control [NC]), NC + 0.04% bitter citrus extract (PT1), NC + 0.01% microencapsulated blend of thymol & carvacrol (PT2), NC + 0.10% mixture of 40% bitter citrus extract and 10% microencapsulated blend of thymol and carvacrol (PT3), NC + 0.04% premixture of grape seed and grape marc extract, green tea and hops (PT4), and NC + 0.10% fenugreek seed powder (PT5). The reduction of space allowance significantly decreased ($p < 0.05$) growth performance (average daily gain, average daily feed intake, feed efficiency) and nutrient digestibility (dry matter, crude protein). Also, the fecal score of NC group increased ($p < 0.05$) compared with other groups. In blood profiles, lymphocyte decreased ($p < 0.05$), and neutrophil, cortisol, TNF- α increased ($p < 0.05$) when pigs were in high stocking density. Basic behaviors (feed intake, standing, lying) were inactive ($p < 0.05$) and singularity behavior (biting) were increased ($p < 0.05$) under high stocking density. However, PFA groups alleviated the negative effects such as reducing growth performance, nutrient digestibility, increasing stress indicators in blood and animal behavior. In conclusion, PFA groups improved the health of pigs with stressed by high stocking density and PT3 is the most effective.

Keywords: Pig, Robustness, Additive, Stress, Plant extract, High stocking density

INTRODUCTION

Recently, there has been increased interest in using natural and safe feed additives to enhance robustness for pigs [1,2]. Phytogetic feed additives (PFA) are plant-derived compounds such as leaves, bark, seeds, roots, flowers, twigs, tree herbs, and fruits [3]. According to the European Council, PFAs

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Competing interests

No potential conflict of interest relevant to this article was reported.

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Availability of data and material

Upon reasonable request, the datasets of this study can be available from the corresponding author.

Authors' contributions

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Ethics approval and consent to participate

The experimental protocol for this study was reviewed and approved by the Institutional Animal Care and Use Committee of Chungbuk National University, Cheongju, Korea (approval CBNUA-1530-21-01).

can be categorized as sensory and flavoring compounds and generally feels safe as substitutes for antibiotics [3]. PFAs have been recognized as the latest feed additives and antibiotics alternatives for livestock [4,5]. Previous studies have reported that PFA complex including sunflower, thyme, and garlic can improve growth performance in monogastric animals [6–8]. Mahima et al. [9] have also reported that dietary PFA supplementation has immunomodulatory effects such as immunoglobulin secretion, cytokine, lymphocyte expression, phagocytosis, and histamine release. Essential oils such as thymol, cavacrol, cymene, terpinene reduce the pathogenic microbial load, but also promote digestive enzymes thereby affecting nutrient digestibility [10–12]. Other studies have shown dietary herbs (i.e., onion, fenugreek seed, and anise seed) enhanced economical efficiency to farms by improving the growth and health of mono-gastric animals [7,13,14]. High stocking density is the most significant caused by inducing stress during growing-finishing periods. Stress caused high stocking density can reduce feed intake, thereby causing low body weight (BW) gain [15–17]. Also, this stress can increase aggressive and negative social behavior such as fighting, feeder occupying, tail biting [18,19] and the incidence of body lesions [20–22]. Supplementaion of *Scutellaria baicalensis* L. roots mitigated negative behavior caused by heat stress in mono-gastric animals [23,24]. However, studies on the relationship between high stocking density and PFA have not been reported. In addition, there are few studies searching for effective PFA against stress derived from high stocking density. Therefore, the objective of this study was to explore effective PFA against environmental stress and the exact mechanism alleviated by PFA in a stress situation for grower-finishing pigs.

MATERIAL AND METHODS

The experimental protocol for this study was reviewed and approved by the Institutional Animal Care and Use Committee of Chungbuk National University, Cheongju, Korea (approval CBNUA-1530-21-01).

Preparation of phytogetic feed additives

PFA1 is a bitter citrus extract (BioFlavex® GC, HTBA, Beniel, Spain) that is rich in 25%–27% naringin and 11%–15% neohesperidin. PFA2 is a microencapsulated blend of thymol and carvacrol (AviPower® 2, VetAgro SpA, Reggio, Emilia, Italy) that contains 7% of thymol and 7% carvacrol. PFA3 is a mixture of PFA1, PFA2 and excipient in ratio of 4:1:5. It contains 0.7% thymol, 0.7% carvacrol, 10%–10.8% naringin and 4.4%–6% neohesperidin. PFA4 is a premixture of grape seed & grape marc extract, green tea and hops (AntaOx® Flavosyn, DR. Eckel GmbH, Niederzissen, Germany) containing more than 10% of flavonoids. PFA5 is fenugreek seed powder containing 12% saponin (Fenugreek Seed Powder, P&D Export, Jaguar, India). All PFAs materials were provided by EUGENE BIO (Suwon, Korea).

Animals, housing, and experimental design

A total of 84 crossbred LYD ([Landrace × Yorkshire] × Duroc) mixed-sex growing pigs at 10 weeks of age (average BW 28.23 ± 2.89kg) were used in a 10-week feeding trial. Pigs were allotted to one of seven treatments in a completely randomized block design based on initial BW. Treatments were as follow: positive control-(PC; basal diet in animal welfare density), negative control (NC; basal diet in high stocking density), PT1 (basal diet with 0.05% PFA1 in high stocking density), PT2 (basal diet with 0.04% PFA2 in high stocking density), PT3 (basal diet with 0.10% PFA3 in high stocking density), PT4 (basal diet with 0.04% PFA4 in high stocking density), PT5 (basal diet with 0.05% PFA5 in high stocking density). All pigs were housed in an environmentally controlled

room. There are two types of room area. In growing pig periods, animal welfare stocking density is 0.55m²/pig, high stocking density is 0.40 m²/pig and in finishing pig periods, animal welfare stocking density is 1.00 m²/pig, high stocking density is 0.60 m²/pig. There are 4 replicate pens with 3 pigs per pen during the experiment period. Basal diet was mostly consisted with corn and soybean meal and were formulated to meet or exceed National Research Council [25] recommendations (Table 1). During the experimental period, each pen was equipped with a self-feeder and nipple drinker to allow *ad libitum* access to feed and water.

Sampling and measurements

Growth performance

To calculate average daily gain (ADG), pig's BW was individually measured at the 09:00 on an empty stomach at start of grower (0 weeks), end of grower and start of finisher (4 weeks), end of the finisher (10 weeks). Feed intake and wasted feed were recorded daily to calculate average daily intake (ADFI). Feed efficiency (G:F) was calculated by ratio of body weight gain and feed intake.

Nutrient digestibility

Apparent total tract digestibility (ATTD) of dry matter (DM) and nitrogen (N) were estimated using 0.2% of chromic oxide as an inert indicator [26]. Crude proteins (CP) were measured from the N. Pigs were fed diets mixed with chromic oxide on 4th week and 10th week. Fresh fecal grab

Table 1. Ingredients and chemical composition of the basal experimental diets (as fed basis)

Items	Grower (0–4 w)	Finisher (4–10 w)
Ingredients (%)		
Corn	65.10	72.38
Soybean meal	23.90	17.40
Wheat bran	7.00	6.00
Soybean oil	1.00	1.00
L-Lysine	0.10	0.28
DL-Methionine	0.04	0.04
L-Threonine	0.03	0.03
Dicalcium phosphate	1.00	1.00
Limestone	1.20	1.25
Salt	0.50	0.50
Vitamin premix ¹	0.08	0.08
Mineral premix ²	0.05	0.05
Calculated composition		
ME (kcal/kg)	3,276	3,284
Crude protein (%)	18.00	15.50
Lysine (%)	1.01	0.97
Methionine (%)	0.33	0.29
Calcium (%)	0.78	0.76
Phosphorus (%)	0.62	0.58

¹Provided per kilogram of complete diet: 20,000 IU of vitamin A, 4,000 IU of vitamin D₃, 80 IU of vitamin E, 16mg of vitamin K₃, 4 mg of thiamine, 20mg of riboflavin, 6 mg of pyridoxine, 0.08 mg of vitamin B₁₂, 120 mg of niacin, 50 mg of Ca-Pantothenate, 2 mg of folic acid, 0.08 mg of biotin.

²Provided per kilogram of complete diet: 12.5 mg of manganese, 179 mg of zinc, 140 mg of copper, 0.5 mg of iodine, 0.4 mg of selenium.

ME, metabolizable energy.

samples collected via rectal massage from each pig, and these samples were stored in a freezer at -20°C until analyzed. All feed and fecal samples were analyzed for DM and N following the procedures outlined by the AOAC [27] methods. N was determined with a Kjeltac 2300 nitrogen analyzer (Foss Tecator AB, Hoeganaes Sweden) and Chromium was analyzed via UV absorption spectrophotometry (Shimadzu UV-1201, Shimadzu, Kyoto, Japan) following the method described by Williams et al. [28]. The ATTD of DM and N were calculated with indirect ratio methods using the following formula: Coefficient of ATTD = $[1 - \{(N_f \times C_d)/(N_d \times C_f)\}] \times 100$. Where: N_f = nutrient concentration in faeces (% DM), N_d = nutrient concentration in diet (% DM), C_f = chromium concentration in faeces (% DM), C_d = chromium concentration in diets (% DM).

Fecal score

During experiment, each pig fecal score was measured by same person before daily feeding. The fecal was scored according to its moisture content and shape. Normal feces are 0-point, soft feces are 1-point, mild diarrhea are 2-point and severe diarrhea are 3-point [29]. The score was calculated by averaging each group with the average value of the daily fecal score of each pig.

Blood sample

For the serum profile, at each pen, one pig was randomly selected to collect blood samples through venipuncture at the end of 4th week, and 10th week. At the time of collection, blood samples were collected both whole blood and serum in nonheparinized tubes and vacuum tubes containing K_3EDTA (Becton Dickinson Vacutainer systems, Franklin Lake, NJ, USA), respectively. White blood cells (WBC) and WBC including lymphocyte, neutrophil, basophil concentration in whole blood were measured using an automatic blood analyzer (ADVIA 120, Bayer, Whippany, NJ, USA). After collection, serum samples were centrifuged $12,500 \times g$ for 15 min at 4°C . Samples were stored at -20°C in the refrigerator until analysis. Serum cortisol levels were assessed using enzyme-linked immunosorbent assay kits (LDN GmbH & Co., Nordhorn, Germany) following to the manufacturer's protocol. Tumor necrosis factor-alpha (TNF- α) and interleukine-6 (IL-6) concentration was analyzed with ELISA kit (Quantikine, R&D systems, Minneapolis, MN, USA) and they were measured at 450 nm.

Pig behavior

Collection of each pig image data was recorded by using six-day/night infrared cameras (QNB-7080 RH, Hanwha, Seoul, Korea) installed 3m above each pen. A total of 28 pig behaviors were analyzed by randomly selecting one pig from each pen. Observers collected data based on results of Yang et al. [30], and only one person made all observations and video analysis to see consistent results. The pig behavior analysis was classified for the following criteria (A) Feed intake: the act of eating with the head in the feed bin, or similar behavior. (B) Standing: the act of standing still with the forelimbs and hindlimbs extended perpendicular to the floor, or similar behavior. (C) Lying: the act of lying with the whole body on the floor, lying with the head, front legs, hind legs and abdomen all touching the floor. (D) Sitting: Two front legs are spread vertically to the floor, two rear legs and two hips are sitting on the floor, like a dog sitting on the floor, or something like that. (E) Drinking water: the act of drinking water for 10 seconds by putting your mouth in a drinking nipple. (F) Posture transition (lying \rightarrow standing) A behavior that changes from lying down to standing, in which the two front legs are stretched first, and the hind legs are naturally stretched out. (G): Posture transition (standing \rightarrow lying): A behavior that changes from a standing behavior to a lying behavior, in which the two front legs are bent to the floor first, and then the two hind legs are naturally folded and lying down. (H) Rooting: the act of repeating similar behaviors, such as

scratches, itching, or something on the nose and front legs. (I) Biting: The act of biting another pig's ears, mouth, and tail with teeth and then biting again or doing similar things.

Statistical analysis

All data were analyzed by one-way ANOVA using SPSS software (ver. 20.0, IBM, Armonk, NY, USA), and the differences among treatments were examined by Tukey's multiple range test, which were considered to be significant at $p < 0.05$, unless otherwise stated.

RESULTS

Growth performance

There was no difference among treatment groups in the initial BW of pigs (Table 2). During the growing period (0–4 weeks), PT3 group significantly increased ($p < 0.05$) ADG and G:F ratio than NC group. During the finishing period (4–10 weeks), NC group significantly decreased ($p < 0.05$) ADG and ADFI than PC group. PFA groups ADG significantly higher ($p < 0.05$) than NC group. The PT3-PT4 groups ADFI significantly higher ($p < 0.05$) than NC group. During entire experimental period (0–10 weeks), NC group significantly decreased ($p < 0.05$) ADG, ADFI and G:F ratio than PC group. PFA groups significantly higher ($p < 0.05$) ADFI than NC group. The PT3-PT4 groups significantly increased ($p < 0.05$) ADG and G:F ratio than NC group.

Nutrient digestibility

During the growing period (0–4 weeks), the ATTD of DM significantly increased ($p < 0.05$) in PT1-PT3 groups compared PC group (Table 3). The ATTD of CP significantly decreased ($p < 0.05$) in NC group compared to PC group. However, PFA groups significantly increased ($p < 0.05$)

Table 2. Effects of different phytogetic feed additives on growth performance in growing-finishing pigs with stressed by stocking density¹⁾

Items	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
BW (kg)									
Initial	28.00	27.53	27.64	28.97	28.62	28.48	27.88	0.309	0.868
4 w	56.30 ^{ab}	53.23 ^b	53.77 ^b	57.43 ^{ab}	59.35 ^a	57.04 ^{ab}	56.15 ^{ab}	0.592	0.083
Final	110.63 ^a	96.72 ^c	97.23 ^c	101.37 ^{bc}	109.43 ^a	106.41 ^{ab}	102.39 ^{bc}	0.767	< 0.001
0–4 w									
ADG (kg)	0.98 ^{ab}	0.89 ^b	0.90 ^b	0.98 ^{ab}	1.06 ^a	0.98 ^{ab}	0.98 ^{ab}	0.014	< 0.001
ADFI (kg)	1.98 ^{bc}	2.03 ^{abc}	1.94 ^c	2.01 ^{abc}	2.06 ^{ab}	2.08 ^a	2.03 ^{ab}	0.012	< 0.001
G:F	0.49 ^{ab}	0.44 ^b	0.46 ^{ab}	0.49 ^{ab}	0.51 ^a	0.47 ^{ab}	0.48 ^{ab}	0.006	< 0.001
4–10 w									
ADG (kg)	1.26 ^a	1.01 ^c	1.01 ^c	1.02 ^c	1.16 ^{ab}	1.15 ^b	1.08 ^{bc}	0.013	< 0.001
ADFI (kg)	2.92 ^a	2.54 ^c	2.82 ^{ab}	2.82 ^{ab}	2.88 ^{ab}	2.79 ^b	2.77 ^b	0.016	< 0.001
G:F	0.43 ^a	0.40 ^{abc}	0.36 ^c	0.36 ^c	0.41 ^{ab}	0.41 ^{ab}	0.39 ^{bc}	0.004	< 0.001
Overall period									
ADG (kg)	1.16 ^a	0.97 ^e	0.98 ^e	1.02 ^{de}	1.14 ^{ab}	1.1 ^{bc}	1.05 ^{cd}	0.009	< 0.001
ADFI (kg)	2.58 ^a	2.29 ^c	2.41 ^b	2.43 ^b	2.46 ^b	2.43 ^b	2.41 ^b	0.012	< 0.001
G:F	0.45 ^{ab}	0.43 ^{cd}	0.41 ^d	0.42 ^{cd}	0.46 ^a	0.45 ^{ab}	0.44 ^{bc}	0.004	< 0.001

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

^{a-e}Means within column with different superscripts differ significantly ($p < 0.05$).

BW, body weight; ADG, average daily gain; ADFI, average daily feed intake; G:F, feed efficiency; PFA, phytogetic feed additives.

Table 3. Effects of different phytogetic feed additives on nutrient digestibility in growing-finishing pigs with stressed by stocking density¹⁾

Items	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
4 week									
DM (%)	85.55 ^a	84.02 ^b	85.72 ^a	85.71 ^a	86.14 ^a	85.43 ^{ab}	85.46 ^{ab}	0.143	0.03
CP (%)	73.35 ^a	69.45 ^b	73.35 ^a	73.58 ^a	74.47 ^a	73.16 ^a	72.88 ^a	0.271	0.01
10 week									
DM (%)	85.85 ^a	83.17 ^b	85.84 ^a	85.75 ^a	86.93 ^a	86.15 ^a	86.11 ^a	0.179	0.01
CP (%)	67.98 ^{bc}	64.75 ^c	70.58 ^{ab}	70.65 ^{ab}	72.87 ^a	71.37 ^a	71.22 ^{ab}	0.398	0.01

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

^{a-c)}Means within column with different superscripts differ significantly ($p < 0.05$).

DM, dry matter; CP, crude protein; PFA, phytogetic feed additives.

CP digestibility than NC group. During the finishing period (4–10 weeks), PFA groups decreased ($p < 0.05$) ATTD of DM and CP compared to NC group. The PT3-PT4 groups CP digestibility numerically increased ($p < 0.05$) than other PFA groups.

Fecal score

During the growing period (0–4 weeks), NC group showed significantly higher ($p < 0.05$) fecal score than PC group (Table 4). However, PFA groups significantly decreased ($p < 0.05$) fecal score compared to NC group. During finishing period (4–10 weeks), the difference of diarrhea incidence was not observed among all treatment groups.

Blood profile

During the growing period (0–4 weeks), there were no significant differences ($p > 0.05$) on WBC, Basophil, and IL-6 among treatment groups (Table 5). The NC group significantly decreased ($p < 0.05$) lymphocyte and increased ($p < 0.05$) neutrophil, cortisol, and TNF- α level in blood compared with PC group. However, PFA groups significantly alleviated ($p < 0.05$) these negative effects by stress with stocking density and was similar with the level of PC group. During the finishing period (4–10 weeks), there were no significant differences ($p > 0.05$) on WBC among treatment groups. NC group significantly decreased ($p < 0.05$) lymphocyte and significantly increased ($p < 0.05$) neutrophil, cortisol, and TNF- α level in blood compared to PC group. However, PFA groups significantly increased ($p < 0.05$) lymphocyte and significantly decreased ($p < 0.05$) neutrophil, cortisol, and TNF- α compared with NC group. PT3 group showed ($p < 0.05$) the lowest results in neutrophil, cortisol, and IL-6 among PFA groups.

Table 4. Effects of different phytogetic feed additives on fecal score in growing pigs with stressed by stocking density¹⁾

Items	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
4 week									
Fecal score ³⁾	0.26 ^b	0.76 ^a	0.33 ^b	0.31 ^b	0.30 ^b	0.29 ^b	0.28 ^b	0.017	0.02

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

³⁾Fecal score was determined as follow : 0, normal feces; 1, soft feces; 2, mild diarrhea; 3, severe diarrhea.

^{a,b)}Means within column with different superscripts differ significantly ($p < 0.05$).

PFA, phytogetic feed additives.

Table 5. Effects of different phytogetic feed additives on blood profile in growing-finishing pigs with stressed by stocking density¹⁾

Items	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
4 week									
WBC (10 ³ /μL)	23.03	23.47	23.18	23.21	22.72	22.99	23.04	0.155	0.93
Lymphocyte (%)	45.31 ^a	36.9 ^b	46.77 ^b	47.47 ^{ab}	46.37 ^b	41.70 ^a	47.80 ^a	0.506	0.01
Neutrophil (%)	39.43 ^{bc}	48.07 ^a	39.63 ^{bc}	38.53 ^{bc}	36.97 ^c	39.83 ^b	40.03 ^b	0.436	0.03
Basophil (%)	0.65	0.70	0.67	0.70	0.63	0.70	0.63	0.027	0.98
Cortisol (μg/dL)	1.82 ^c	3.47 ^a	2.60 ^b	2.19 ^{bc}	1.92 ^c	2.74 ^b	2.78 ^b	0.069	0.01
TNF-α (pg/mL)	61.90 ^b	73.13 ^a	62.63 ^b	62.77 ^b	61.93 ^b	62.67 ^b	62.40 ^b	0.506	0.01
IL-6 (pg/mL)	72.58	72.20	72.50	72.00	72.37	72.13	72.33	0.254	0.99
10 week									
WBC (10 ³ /μL)	17.74	17.76	17.85	17.64	17.76	17.63	17.72	0.148	0.99
Lymphocyte (%)	43.40 ^a	35.90 ^b	45.40 ^a	46.57 ^a	44.37 ^a	45.63 ^a	46.87 ^a	0.492	0.01
Neutrophil (%)	42.08 ^b	52.37 ^a	41.20 ^b	40.83 ^b	42.03 ^b	42.20 ^b	44.13 ^b	0.499	0.01
Basophil (%)	0.68 ^{ab}	0.70 ^a	0.70 ^a	0.63 ^{abc}	0.50 ^{bc}	0.47 ^c	0.47 ^c	0.019	0.01
Cortisol (μg/dL)	0.72 ^b	2.40 ^a	0.61 ^{bc}	0.64 ^{bc}	0.51 ^c	0.67 ^b	0.61 ^{bc}	0.070	0.01
TNF-α (pg/mL)	86.70 ^b	99.83 ^a	85.73 ^b	85.80 ^b	84.90 ^b	85.70 ^b	85.83 ^b	0.596	0.01
IL-6 (pg/mL)	80.80 ^{ab}	81.77 ^a	80.37 ^{ab}	80.70 ^{ab}	80.30 ^b	80.43 ^{ab}	80.53 ^{ab}	0.132	0.04

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

^{a-c)}Means within column with different superscripts differ significantly ($p < 0.05$).

WBC, white blood cell; TNF- α, tumor necrosis factor-α; IL-6, Interleukin-6; PFA, phytogetic feed additives.

Table 6. Effects of different phytogetic feed additives on behavior changes in growing pigs with stressed by stocking density¹⁾

	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
Basic behavior (min/hour)									
Feed intake	4.03	4.01	4.03	4.02	4.10	4.10	4.05	0.009	0.02
Standing	7.05	7.11	6.99	7.12	7.01	6.98	7.13	0.030	0.74
Lying	44.58	44.16	44.39	44.51	44.09	44.66	44.29	0.106	0.78
Sitting	4.34	4.72	4.59	4.35	4.80	4.26	4.53	0.087	0.64
Singularity behavior (count/hour)									
Drink water	5.04	5.19	5.15	5.10	5.11	5.14	5.12	0.015	0.23
Rooting	1.08	1.10	1.11	1.04	1.12	1.06	1.03	0.014	0.46
Posture transition (lying-sitting)	3.54	3.49	3.50	3.48	3.44	3.44	3.51	0.020	0.86
Posture transition (sitting-lying)	3.53	3.48	3.50	3.48	3.43	3.45	3.50	0.015	0.64
Biting	0.18 ^b	0.23 ^a	0.21 ^{ab}	0.18 ^b	0.15 ^c	0.17 ^b	0.18 ^b	0.05	< 0.001

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

^{a,b)}Means within column with different superscripts differ significantly ($p < 0.05$).

PFA, phytogetic feed additives.

Animal behavior

The effects of different PFA on animal behavior were shown in Tables 6, 7, and Fig. 1. During the growing period (0–4 weeks), there are no significant differences ($p > 0.05$) in basic behavior and most of singularity behavior. The NC group had significantly higher ($p < 0.05$) biting frequency

Table 7. Effects of different phytogetic feed additives on behavior changes in finishing pigs with stressed by stocking density¹⁾

	PC ²⁾	NC	PT1	PT2	PT3	PT4	PT5	SEM	p-value
Basic behavior (min/hour)									
Feed intake	4.63 ^a	4.37 ^b	4.67 ^a	4.59 ^a	4.61 ^a	4.67 ^a	4.61 ^a	0.020	< 0.001
Standing	6.77 ^a	6.32 ^c	6.38 ^c	6.44 ^{bc}	6.71 ^a	6.56 ^{ab}	6.61 ^{ab}	0.031	< 0.001
Lying	44.88 ^c	45.51 ^a	45.21 ^b	45.18 ^b	44.98 ^c	44.99 ^c	44.96 ^{bc}	0.041	< 0.001
Sitting	3.72	3.80	3.74	3.79	3.70	3.78	3.82	0.012	0.75
Singularity behavior (count/hour)									
Drink water	5.34	5.28	5.27	5.30	5.44	5.38	5.29	0.026	0.59
Rooting	1.12	1.08	1.11	1.09	1.21	1.19	1.15	0.021	0.58
Posture transition (lying-sitting)	3.78	3.43	3.49	3.58	3.71	3.68	3.69	0.035	0.60
Posture transition (sitting-lying)	3.77	3.41	3.50	3.60	3.70	3.67	3.68	0.045	0.37
Biting	0.16	0.23	0.18	0.15	0.17	0.16	0.18	0.008	0.20

¹⁾Each value is the mean value of 4 replicates.

²⁾PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density.

^{a-c)}Means within column with different superscripts differ significantly ($p < 0.05$).

PFA, phytogetic feed additives.

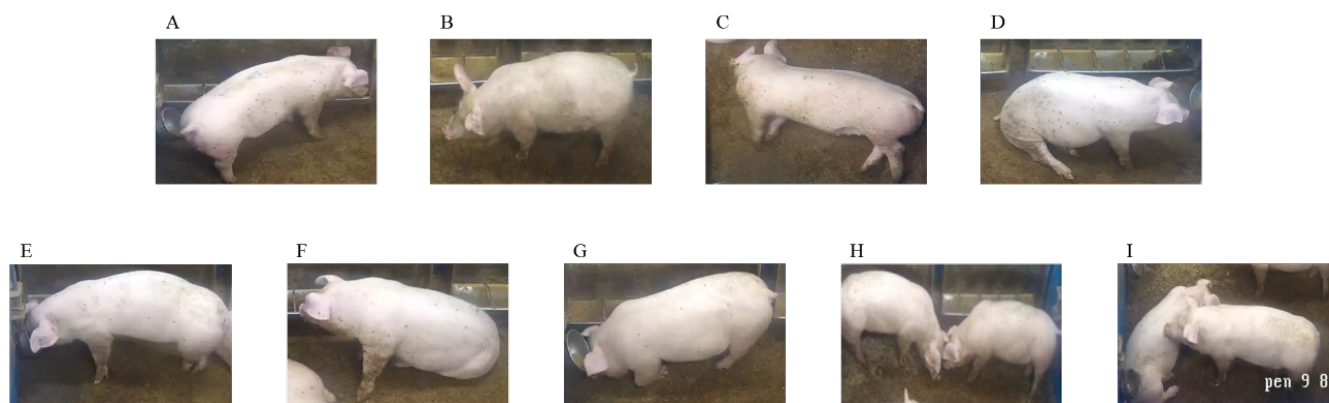


Fig. 1. Classification of pig behavior changes. (A) Feed intake, (B) standing, (C) lying, (D) sitting, (E) drink water, (F) posture transition (lying → standing), (G) posture transition (standing → lying), (H) rooting, (I) biting.

than PC group. However, PFA groups had significantly alleviated ($p < 0.05$) biting frequency compared with NC group. Among PFA groups, PT3 group showed the lowest biting frequency. During the finishing period (4–10 weeks), NC group showed ($p < 0.05$) more lying time and less feed intake and standing time than PC group. Feed intake time significantly increased ($p < 0.05$) in PFA groups than NC group. Standing time significantly increased ($p < 0.05$) in PT2-PT5 groups than NC group. Lying time significantly decreased ($p < 0.05$) in PFA groups than NC group. Especially, PT3-PT5 groups showed similar result with PC group. In singularity behavior, there are no significant difference ($p > 0.05$) in treatment groups. But NC group showed numerically high number of biting than other treatment groups.

DISCUSSION

Growth performance

High stocking density can disturb the movement of animals due to limited feeding environment (space, feeders, and drinkers). Moreover, high stocking density can interfere with airflow and generate heat energy [12]. It can result in difficulty in evacuating body temperature, poor air quality, reduced access to feed and water, and poor performance of animals due to increased ammonia levels [31–33]. High heat energy and poor air quality are known to cause heat stress and adverse effects on growth rate, feed consumption, mortality, and health [34–36]. Similarly, our study showed that pigs under high stocking density (i.e., 0.40 m²/growing pig, 0.60 m²/finishing pig) had reduced ADG and ADFI by 16.38% and 11.24%, respectively, than those under welfare density (i.e., 0.55 m²/growing pig, 1.0 m²/finishing pig) during the whole period (grower: 28–56 kg, finisher: 56–103 kg). Spicer and Aherne [37] have also reported that daily gain and daily feed are reduced 8.47% and 13.15%, respectively, when group size is decreased from 0.72 m²/pig to 0.35 m²/pig. Stress-induced heat and high stocking densities can reduce growth performance by damaging cellular structure, increasing intracellular water imbalance, and increasing free radical concentration [38]. However, our study revealed that pigs under high stocking density with supplementation of PFA showed improvement (i.e., BW decreased 11.61%) in growth performance compared to those in the unsupplemented group. Many researchers reported that dietary supplementation of PFA such as Korean pine extract, cinnamon, turmeric, essential oils, and rosemary can improve growth performance with reducing stress response [38–41]. In our study, PFA supplementation under our high stocking density showed no difference in ADFI between treatments in the growing period, but significantly increased with PFA supplementation in the finishing period. This is consistent with previous studies suggesting that PFA is effective for intake when supplied long-term [42]. Moreover, PFA3 could improve the flavor of feed and increase the palatability of feed intake in pigs [43–46]. Therefore, using natural products with polyphenols (suitable structure for free radical scavenging activity) can effectively alleviate stress caused by low space allowance and heat through their antioxidant activity with improved low feed intake, thereby increasing growth performance.

Nutrient digestibility

High stocking density can negatively affect nutrient digestibility and growth performance. During the whole experiment periods, nutrient digestibility (DM, CP) showed improvement in the treatment group added with PFA than that in the control group without PFA under high stocking density. PFA can also enhance nutrient digestibility and absorption [47,48]. It has been reported that the addition of essential oils to monogastric animals can enhance the activity of trypsin, maltase and pancreatic amylase and increases glucose absorption in the small intestine [49]. Therefore, the addition of PFA can stimulate the secretion of mucus in the intestine, thereby reducing the adhesion of pathogens and stabilizing intestinal microbial symbiosis [50]. It can be seen that improved digestive tract function is associated with increased nutrient digestibility. It can also be said that the antibacterial action of PFA contributes to the increase of nutrient digestibility. PFAs such as carvacrol, thymol, anetol, oregano, anise, and citrus essential oil have antibacterial activity against intestinal microbes when ingested. Among them, phenolic substances are the most active compounds [51,52]. PT2 and PT3 have a phenolic structure in our experiment. It was shown that the digestibility of DM and CP was higher than the high stocking density throughout the experiment period. Fiesel et al. [53] reported an increase in nutrient digestibility due to the antioxidant effect of polyphenols and an increased absorbable surface of the intestine. As the experiment progressed, the digestibility deviation of DM and CP increased according to

the presence or absence of PFA in the feed under high stocking density. In this experiment, it was confirmed that the digestibility was gradually improved when PFA was used, leading to improved performance of pigs. In particular, it was found that the digestibility was significantly improved by flavonoids, a common component of PT3-PT5 additives. A previous study has shown that flavonoids have DM and CP synergistic effects [54]. Therefore, it can be concluded that the use of flavonoid additives can increase the digestibility of nutrients, as it can improve nutrient availability by boosting immunity and antibacterial action in pigs.

Fecal score

In high stocking density, the frequency of diarrhea was increased during the growing period, although it showed no significant difference during the finishing period. Many studies have found that diarrhea in pigs is more likely to be induced by stress [55–57]. Actually, the frequency of diarrhea is increased in weaned pigs during stress [58]. Diarrhea has been found intermittently in growing pigs [59]. When pigs get stressed, their immunity is lower and pathogens in the intestine are activated. Intestinal pathogens can suppress unnecessary energy loss such as reduced feed intake and G:F ratio known to interfere with immune system activation. In addition, intestinal pathogens can inhibit homeostasis of the epithelial barrier, causing secretory diarrhea due to intestinal damage through osmotic stress or inflammatory diarrhea by increasing inflammatory cytokines. However, in our study, the frequency of diarrhea was significantly reduced when PFAs were fed to pigs in a stressful situation. These results indicate that PFA can improve fecal status by improving intestinal health, and further studies on fecal microflora should be conducted. When pigs are fed with natural products reduces the frequency of diarrhea due to stress as the natural product's antibacterial action improves intestinal health and increased digestibility [60,61]. Many researchers have checked diarrhea scores of weaning pigs, but not those of growing to finishing pigs. In the present study, complete diarrhea was not found even in the growing period, although a lot of soft feces were observed for pigs under a high stocking density condition. The difference between growing period and finishing period is that as pigs grow, their immune system gets better, and their gut health improves. Therefore, we can confirm a meaningful diarrhea score even in pigs during the growing period. Thus, it is necessary to check the status of feces.

Blood profile

In the present study, there were no significant differences in blood profile between the entire experiment period WBC or growing period basophil and IL-6 of pigs between treatment groups. However, pigs fed with PFAs under a high stocking density condition had better blood results than those without addition of additives under a high stocking density condition. Pigs with high stocking density are subjected to critical psychological, social, and environmental stresses. High stocking density can also cause chronic severe stress that affects immunity and health [62]. Lymphocytes show various immunological responses including modulation of immune defense and immunoglobulin [63]. In our study, lymphocytes were decreased during stress situation, but returned to PC levels when PFAs were added. According to Dhabhar [64], in stressful situations, lymphocyte counts are decreased due to changes induced by trafficking or redistribution of lymphocytes to other body compartments of glucocorticoids. This result was similar to our study. In our study, the number of neutrophils was increased when pigs were stressed. This number was then decreased after supplementation with PFA in our study. It has been reported that stressful situations cause decreasing lymphocytes and increasing neutrophils in the blood [65]. As a result, it was possible to confirm the indirect change caused by supplementation of PFA to relieve stress. Cortisol, a steroid hormone, or glucocorticoid produced by the adrenal gland and released in

response to stress, is often used as a physiological marker to quantify animal stress [66]. It is well known that cortisol can regulate intermediary metabolism, immunity, and growth [67,68]. A poor welfare situation can cause animals to be extremely stressed. In this study, cortisol level was increased under high stocking density compared with animal welfare density (space decreased in growing pig 27.27%, in finishing pig 40%). This result was agreement with the results of Jang et al. [69] that reported decreasing space allowance (decreasing 28.13%) induced increasing cortisol level (2.3 to 4 µg/dL).

However, PFAs supplementation alleviated high cortisol level in blood caused by high stocking density. Li et al. [39] observed that flavonoids, which are physiologically active substances of PFA, down-regulated immune responses by mediated viruses and the T-cell, thereby reducing psychological stress. This observation suggests that PFA mitigates the increased cortisol concentration by high stocking density.

Pro-inflammatory cytokines such as TNF- α and IL-6 are potential outputs of the cellular immune system and can indirectly reflect immune responses due to the activation of T-cells [70,71]. This study showed that high stocking density increased pro-inflammatory cytokine level. These results suggest that the environmental stress caused by a limited space allowance can induce a cellular immune response. When stressed out, pro-inflammatory cytokines are secreted to promote cortisol secretion and suppress growth hormone secretion [72,73]. Excessive pro-inflammatory cytokines can induce fever, inflammation, tissue destruction [74], and in some cases, even shock and death [75]. Thus, the immune system is activated due to high stocking stress, which shifts nutrient distribution priorities from growth to host defense [70,72,76].

In addition, TNF- α and IL-6 content is reduced through improved gut microbiota, antioxidant, and anti-inflammatory effects, due to improved digestibility of nutrients, alleviating stress response, and strengthening immunity [77,78]. However, PFA was effective methods to alleviate negative effects of a high stocking density in our study. Other researchers also reported essential oil and herb extract reduced pro-inflammatory cytokines [79,80].

Therefore, PFA is effective in relieving stress, and PT3 group showed the highest effect among PFAs group. The reason the PT3 group outperformed the others was due to the construction of the PFA group. Flavonoids and terpenoids (carvacrol and thymol) may protect cells from the harmful effects of autoxidation.

Animal behavior

A high stocking density equates to a reduced floor space allowance. Decreasing floor space allowance per pig increases the frequency of contact, social tension, and aggression [81–84]. In addition, when heat production per unit floor area is increased, heat stress will occur and induce oxidative stress [22,85]. If this stress is not well managed in pigs, it can increase their susceptibility to stress and hence reduce their immune and health status. Throughout our study, animal behavior at high stocking density improved when fed with PFA. The biting frequency was increased in NC but decreased after PFA treatment similar to PC. Among all treatment groups, PT3 group showed the lowest biting frequency. Greene et al. [38] has reported that biting as a representative form of aggressive behavior can occur in pigs under chronic stress. This is consistent with our study. When ingesting phenolic compounds as components of PT3 group, it is possible to restore redox homeostasis and prevent oxidative stress by improving the activity of antioxidant enzymes SOD, CAT, GPx, and GR [86]. Therefore, the effect of adding PFA3 not only can help pigs cope with biting behavior caused by stress, but also can overcome it. During the finishing period, basic behaviors (eating, standing, lying down) were more active when fed with PFA added in high stocking density. In addition, the feed intake increased during PFA feeding in growth performance.

Feed intake is an important indicator because it is related to body weight, ADG, ADFI, and G:F ratio. Pigs with a high stocking density face difficulty in feeding due to competition in the feeder. In this study, PC group showed less time than other treatments in feed intake. Therefore, the number of trips to the feeder is directly related to intake and can affect growth performance. Also, standing and lying time were similar to NC group. They were more active than PC group. Especially, PT3-PT5 groups are more activated than others. Pearce and Paterson [87] have reported that observation of the behavior of standing motionless in a narrow space is a behavior that pigs do to cope with stress at a high stocking density. As stress increased, the amount of physical activity decrease. It can be seen that when the standing time decrease, the lying time increases at the same time. This indicates that there is a close relationship between basic behavior and growth performance. Through this experiment, it can be seen that when pigs get stressed, their basic behaviors (standing, lying, and feeding) were affected at the same time.

CONCLUSION

Dietary supplementation of PFA improves the growth performance, nutrient digestibility, immunity, fecal score, and animal behavior in grower-finishing pigs. As a result, lymphocytes, neutrophils, cortisol, IL-6, and TNF- α in the blood, bites, and basic behaviors were improved, indicating that stress was reduced and strengthened. The diarrhea index improved because of getting healthier, which means less damage to the intestines and increased digestibility. Due to these positive effects, growth performance was improved, and it was found that PFA is an effective additive for stress due to high stocking density. Among them, the most effective and additional advantages were found when using PFA3 (mixture of PFA1 40%, PFA2 10% and excipient 50%) rather than using PFA1 (bitter citrus extract) and PFA2 (microencapsulated blend of thymol and carvacrol) separately.

REFERENCES

1. Kholif AE, Abdo MM, Anele UY, El-Sayed MM, Morsy TA. *Saccharomyces cerevisiae* does not work synergistically with exogenous enzymes to enhance feed utilization, ruminal fermentation and lactational performance of Nubian goats. *Livest Sci.* 2017;206:17-23. <https://doi.org/10.1016/j.livsci.2017.10.002>
2. Kholif AE, Gouda GA, Galyean ML, Anele UY, Morsy TA. Extract of *Moringa oleifera* leaves increases milk production and enhances milk fatty acid profile of Nubian goats. *Agrofor Syst.* 2019;93:1877-86. <https://doi.org/10.1007/s10457-018-0292-9>
3. European Union. Council directive 2008/120/EC of 18 December 2008 laying down minimum standards for the protection of pigs (codified version). *Off J Eur Union.* 2009;L47:5-13.
4. Ebeid HM, Mengwei L, Kholif AE, Hassan F, Lijuan P, Xin L, et al. *Moringa oleifera* oil modulates rumen microflora to mediate in vitro fermentation kinetics and methanogenesis in total mix rations. *Curr Microbiol.* 2020;77:1271-82. <https://doi.org/10.1007/s00284-020-01935-2>
5. Elghalid OA, Kholif AE, El-Ashry GM, Matloup OH, Olafadehan OA, El-Raffa AM, et al. Oral supplementation of the diet of growing rabbits with a newly developed mixture of herbal plants and spices enriched with special extracts and essential oils affects their productive performance and immune status. *Livest Sci.* 2020;238:104082. <https://doi.org/10.1016/j.livsci.2020.104082>
6. Al-Gharabi HKB, Al-Gharawi JKM, Al-Sahlani AJA. Effect of garlic (*Allium sativum*)

- and onion (*Allium cepa*) water extract on some productive traits of broilers. *Plant Arch.* 2019;19:565-9.
7. Alhaji MS, Alhobaishi M, Ger El Nabi AR, Al-Mufarrej SI. Immune responsiveness and performance of broiler chickens fed a diet supplemented with high levels of Chinese star anise fruit (*Illicium verum* Hook. f). *J Anim Vet Adv.* 2015;14:36-42.
 8. Khan RU, Nikousefat Z, Tufarelli V, Naz S, Javdani M, Laudadio V. Garlic (*Allium sativum*) supplementation in poultry diets: effect on production and physiology. *Worlds Poult Sci J.* 2012;68:417-24. <https://doi.org/10.1017/S0043933912000530>
 9. Mahima, Rahal A, Deb R, Latheef SK, Samad HA, Tiwari R, et al. Immunomodulatory and therapeutic potentials of herbal, traditional/indigenous and ethnoveterinary medicines. *Pak J Biol Sci.* 2012;15:754-74. <https://doi.org/10.3923/pjbs.2012.754.774>
 10. Ahmed ST, Hossain ME, Kim GM, Hwang JA, Ji H, Yang CJ. Effects of resveratrol and essential oils on growth performance, immunity, digestibility and fecal microbial shedding in challenged piglets. *Asian-Australas J Anim Sci.* 2013;26:683-90. <https://doi.org/10.5713/ajas.2012.12683>
 11. Emami NK, Samie A, Rahmani HR, Ruiz-Feria CA. The effect of peppermint essential oil and fructooligosaccharides, as alternatives to virginiamycin, on growth performance, digestibility, gut morphology and immune response of male broilers. *Anim Feed Sci Technol.* 2012;175:57-64. <https://doi.org/10.1016/j.anifeedsci.2012.04.001>
 12. Zhang S, Jung JH, Kim HS, Kim BY, Kim IH. Influences of phytoncide supplementation on growth performance, nutrient digestibility, blood profiles, diarrhea scores and fecal microflora shedding in weaning pigs. *Asian-Australas J Anim Sci.* 2012;25:1309-15. <https://doi.org/10.5713/ajas.2012.12170>
 13. Aji SB, Ignatius K, Ado AY, Nuhu JB, Abdulkarim A, Aliyu U, et al. Effect of feeding onion (*Allium cepa*) and garlic (*Allium sativum*) on some performance characteristics of broiler chickens. *Res J Poult Sci.* 2011;4:22-7. <https://doi.org/10.3923/rjpscience.2011.22.27>
 14. Mamoun T, Mukhtar MA, Tabidi MH. Effect of fenugreek seed powder on the performance, carcass characteristics and some blood serum attributes. *Adv Res Agric Vet Sci.* 2014;1:6-11.
 15. Cho JH, Kim IH. Effect of stocking density on pig production. *Afr J Biotechnol.* 2011;10:13688-92. <https://doi.org/10.5897/AJB11.1691>
 16. Hyun Y, Ellis M, Riskowski G, Johnson RW. Growth performance of pigs subjected to multiple concurrent environmental stressors. *J Anim Sci.* 1998;76:721-7. <https://doi.org/10.2527/1998.763721x>
 17. McGlone JJ, Salak JL, Lumpkin EA, Nicholson RI, Gibson M, Norman RL. Shipping stress and social status effects on pig performance, plasma cortisol, natural killer cell activity, and leukocyte numbers. *J Anim Sci.* 1993;71:888-96. <https://doi.org/10.2527/1993.714888x>
 18. Li Y, Wang C, Huang S, Liu Z, Wang H. Space allowance determination by considering its coefficient with toy provision on production performance, behavior and physiology for grouped growing pigs. *Livest Sci.* 2021;243:104389. <https://doi.org/10.1016/j.livsci.2020.104389>
 19. Cornale P, Macchi E, Miretti S, Renna M, Lussiana C, Perona G, et al. Effects of stocking density and environmental enrichment on behavior and fecal corticosteroid levels of pigs under commercial farm conditions. *J Vet Behav.* 2015;10:569-76. <https://doi.org/10.1016/j.jveb.2015.05.002>
 20. Averós X, Brossard L, Dourmad JY, de Greef KH, Edge HL, Edwards SA, et al. Quantitative assessment of the effects of space allowance, group size and floor characteristics on the lying behaviour of growing-finishing pigs. *Animal.* 2010;4:777-83. <https://doi.org/10.1017/S1751731109991613>

21. Hyun Y, Ellis M, Curtis SE, Johnson RW. Environmental temperature, space allowance, and regrouping: additive effects of multiple concurrent stressors in growing pigs. *J Swine Health Prod.* 2005;13:131-8.
22. Street BR, Gonyou HW. Effects of housing finishing pigs in two group sizes and at two floor space allocations on production, health, behavior, and physiological variables. *J Anim Sci.* 2008;86:982-91. <https://doi.org/10.2527/jas.2007-0449>
23. Casal-Plana N, Manteca X, Dalmau A, Fàbrega E. Influence of enrichment material and herbal compounds in the behaviour and performance of growing pigs. *Appl Anim Behav Sci.* 2017;195:38-43. <https://doi.org/10.1016/j.applanim.2017.06.002>
24. Zmrhal V, Lichovníková M, Hampel D. The effect of phytogenic additive on behavior during mild-moderate heat stress in broilers. *Acta Univ Agric Silvic Mendel Brun.* 2018;66:939-44. <https://doi.org/10.11118/actaun201866040939>
25. NRC [National Research Council]. Nutrient requirements of swine. 11th rev. ed. Washington, DC: National Academies Press; 2012. pp. 208-38.
26. Fenton TW, Fenton M. An improved procedure for the determination of chromic oxide in feed and feces. *Can J Anim Sci.* 1979;59:631-4. <https://doi.org/10.4141/cjas79-081>
27. AOAC [Association of Official Analytical Chemists] International. Official methods of analysis of AOAC International. 21st ed. Gaithersburg, MD: AOAC International; 2019.
28. Williams CH, David DJ, Iismaa O. The determination of chromic oxide in faeces samples by atomic absorption spectrophotometry. *J Agric Sci.* 1962;59:381-5. <https://doi.org/10.1017/S002185960001546X>
29. Marquardt RR, Jin LZ, Kim JW, Fang L, Frohlich AA, Baidoo SK. Passive protective effect of egg-yolk antibodies against enterotoxigenic *Escherichia coli* K88+ infection in neonatal and early-weaned piglets. *FEMS Immunol Med Microbiol.* 1999;23:283-8. <https://doi.org/10.1111/j.1574-695X.1999.tb01249.x>
30. Yang KY, Jeon JH, Kwon KS, Choi HC, Ha JJ, Kim JB, et al. Classification of behavior at the signs of parturition of sows by image information analysis. *J Korea Acad Ind Coop Soc.* 2018;19:607-13. <https://doi.org/10.5762/KAIS.2018.19.12.607>
31. Bilal RM, Hassan F, Farag MR, Nasir TA, Ragni M, Mahgoub HAM, et al. Thermal stress and high stocking densities in poultry farms: potential effects and mitigation strategies. *J Therm Biol.* 2021;99:102944. <https://doi.org/10.1016/j.jtherbio.2021.102944>
32. EFSA [European Food Safety Authority]. Opinion of the Scientific Panel on Animal Health and Welfare (AHAW) on a request from the Commission related to welfare of weaners and rearing pigs: effects of different space allowances and floor. *EFSA J.* 2005;3:268. <https://doi.org/10.2903/j.efsa.2005.268>
33. Feddes JJ, Emmanuel EJ, Zuidhof MJ. Broiler performance, body weight variance, feed and water intake, and carcass quality at different stocking densities. *Poult Sci.* 2002;81:774-9. <https://doi.org/10.1093/ps/81.6.774>
34. Chegini S, Kiani A, Parizadian Kavan B, Rokni H. Effects of propolis and stocking density on growth performance, nutrient digestibility, and immune system of heat-stressed broilers. *Ital J Anim Sci.* 2019;18:868-76. <https://doi.org/10.1080/1828051X.2018.1483750>
35. Estévez M. Oxidative damage to poultry: from farm to fork. *Poult Sci.* 2015;94:1368-78. <https://doi.org/10.3382/ps/pev094>
36. Sohail MU, Ijaz A, Yousaf MS, Ashraf K, Zaneb H, Aleem M, et al. Alleviation of cyclic heat stress in broilers by dietary supplementation of mannan-oligosaccharide and *Lactobacillus*-based probiotic: dynamics of cortisol, thyroid hormones, cholesterol, C-reactive protein, and humoral immunity. *Poult Sci.* 2010;89:1934-8. <https://doi.org/10.3382/ps.2010-00751>

37. Spicer HM, Aherne FX. The effects of group size/stocking density on weanling pig performance and behavior. *Appl Anim Behav Sci.* 1987;19:89-98. [https://doi.org/10.1016/0168-1591\(87\)90206-1](https://doi.org/10.1016/0168-1591(87)90206-1)
38. Greene ES, Cauble R, Kadhim H, de Almeida Mallmann B, Gu I, Lee SO, et al. Protective effects of the phytogenic feed additive “comfort” on growth performance via modulation of hypothalamic feeding- and drinking-related neuropeptides in cyclic heat-stressed broilers. *Domest Anim Endocrinol.* 2021;74:106487. <https://doi.org/10.1016/j.domaniend.2020.106487>
39. Li HL, Zhao PY, Lei Y, Hossain MM, Kang J, Kim IH. Dietary phytoncide supplementation improved growth performance and meat quality of finishing pigs. *Asian-Australas J Anim Sci.* 2016;29:1314-21. <https://doi.org/10.5713/ajas.15.0309>
40. Hashemzadeh F, Rafeie F, Hadipour A, Rezadoust MH. Supplementing a phytogenic-rich herbal mixture to heat-stressed lambs: growth performance, carcass yield, and muscle and liver antioxidant status. *Small Rumin Res.* 2022;206:106596. <https://doi.org/10.1016/j.smallrumres.2021.106596>
41. Yan L, Meng QW, Kim IH. Effect of an herb extract mixture on growth performance, nutrient digestibility, blood characteristics, and fecal microbial shedding in weanling pigs. *Livest Sci.* 2012;145:189-95. <https://doi.org/10.1016/j.livsci.2012.02.001>
42. Bartoš P, Dolan A, Smutný L, Šístková M, Celjak I, Šoch M, et al. Effects of phytogenic feed additives on growth performance and on ammonia and greenhouse gases emissions in growing-finishing pigs. *Anim Feed Sci Technol.* 2016;212:143-8. <https://doi.org/10.1016/j.anifeedsci.2015.11.003>
43. Mucha W, Witkowska D. The applicability of essential oils in different stages of production of animal-based foods. *Molecules.* 2021;26:3798. <https://doi.org/10.3390/molecules26133798>
44. Czech A, Kowalczyk E, Grela E. The effect of a herbal extract used in pig fattening on the animals' performance and blood components. *Ann Univ Mariae Curie Sklodowska Sect EE Zootech.* 2009;27:25-33. <https://doi.org/10.2478/v10083-009-0009-7>
45. Wenk C. Herbs and botanicals as feed additives in monogastric animals. *Asian-Australas J Anim Sci.* 2003;16:282-9. <https://doi.org/10.5713/ajas.2003.282>
46. Frankič T, Voljč M, Salobir J, Rezar V. Use of herbs and spices and their extracts in animal nutrition. *Acta Agric Slov.* 2009;94:95-102. <https://doi.org/10.14720/aas.2009.94.2.14834>
47. Amad AA, Männer K, Wendler KR, Neumann K, Zentek J. Effects of a phytogenic feed additive on growth performance and ileal nutrient digestibility in broiler chickens. *Poult Sci.* 2011;90:2811-6. <https://doi.org/10.3382/ps.2011-01515>
48. Hafeez A, Männer K, Schieder C, Zentek J. Effect of supplementation of phytogenic feed additives (powdered vs. encapsulated) on performance and nutrient digestibility in broiler chickens. *Poult Sci.* 2016;95:622-9. <https://doi.org/10.3382/ps/pev368>
49. Lee KW, Everts H, Kappert HJ, Frehner M, Losa R, Beynen AC. Effects of dietary essential oil components on growth performance, digestive enzymes and lipid metabolism in female broiler chickens. *Br Poult Sci.* 2003;44:450-7. <https://doi.org/10.1080/0007166031000085508>
50. Jamroz D, Wartecki T, Houszka M, Kamel C. Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. *J Anim Physiol Anim Nutr.* 2006;90:255-68. <https://doi.org/10.1111/j.1439-0396.2005.00603.x>
51. Giannenas I, Bonos E, Christaki E, Florou-Paneri P. Essential oils and their applications in animal nutrition. *Med Aromat Plants.* 2013;2:140.
52. Panghal M, Kaushal V, Yadav JP. In vitro antimicrobial activity of ten medicinal plants against

- clinical isolates of oral cancer cases. *Ann Clin Microbiol Antimicrob*. 2011;10:21. <https://doi.org/10.1186/1476-0711-10-21>
53. Fiesel A, Gessner DK, Most E, Eder K. Effects of dietary polyphenol-rich plant products from grape or hop on pro-inflammatory gene expression in the intestine, nutrient digestibility and faecal microbiota of weaned pigs. *BMC Vet Res*. 2014;10:196. <https://doi.org/10.1186/s12917-014-0196-5>
 54. Cui K, Wang Q, Wang S, Diao Q, Zhang N. The facilitating effect of tartary buckwheat flavonoids and *Lactobacillus plantarum* on the growth performance, nutrient digestibility, antioxidant capacity, and fecal microbiota of weaned piglets. *Animals*. 2019;9:986. <https://doi.org/10.3390/ani9110986>
 55. Oh HJ, Park YJ, Cho JH, Song MH, Gu BH, Yun W, et al. Changes in diarrhea score, nutrient digestibility, zinc utilization, intestinal immune profiles, and fecal microbiome in weaned piglets by different forms of zinc. *Animals*. 2021;11:1356. <https://doi.org/10.3390/ani11051356>
 56. Yang KM, Jiang ZY, Zheng CT, Wang L, Yang XF. Effect of *Lactobacillus plantarum* on diarrhea and intestinal barrier function of young piglets challenged with enterotoxigenic *Escherichia coli* K88. *J Anim Sci*. 2014;92:1496-503. <https://doi.org/10.2527/jas.2013-6619>
 57. Zhao Y, Weaver AC, Fellner V, Payne RL, Kim SW. Amino acid fortified diets for weanling pigs replacing fish meal and whey protein concentrate: effects on growth, immune status, and gut health. *J Anim Sci Biotechnol*. 2014;5:57. <https://doi.org/10.1186/2049-1891-5-57>
 58. Moeser AJ, Bliklager AT. Mechanisms of porcine diarrheal disease. *J Am Vet Med Assoc*. 2007;231:56-67. <https://doi.org/10.2460/javma.231.1.56>
 59. Panah FM, Lauridsen C, Højberg O, Nielsen TS. Etiology of colitis-complex diarrhea in growing pigs: a review. *Animals*. 2021;11:2151. <https://doi.org/10.3390/ani11072151>
 60. Cho JH, Chen YJ, Min BJ, Kim HJ, Kwon OS, Shon KS, et al. Effects of essential oils supplementation on growth performance, IgG concentration and fecal noxious gas concentration of weaned pigs. *Asian-Australas J Anim Sci*. 2006;19:80-5. <https://doi.org/10.5713/ajas.2006.80>
 61. Caprarulo V, Turin L, Hejna M, Reggi S, Dell'Anno M, Riccaboni P, et al. Protective effect of phytogenic based additives in enterotoxigenic *Escherichia coli* challenged piglets [Preprint]. 2022 [cited 2023 Jun 12]. <https://doi.org/10.21203/rs.3.rs-1207181/v1>
 62. Griffin JFT. Stress and immunity: a unifying concept. *Vet Immunol Immunopathol*. 1989;20:263-312. [https://doi.org/10.1016/0165-2427\(89\)90005-6](https://doi.org/10.1016/0165-2427(89)90005-6)
 63. Campbell TW. Clinical pathology. In: Mader DR, editor. *Reptile medicine and surgery*. Philadelphia, PA: W.B. Saunders; 1996. p. 248-57.
 64. Dhabhar FS. Stress-induced augmentation of immune function—the role of stress hormones, leukocyte trafficking, and cytokines. *Brain Behav Immun*. 2002;16:785-98. [https://doi.org/10.1016/S0889-1591\(02\)00036-3](https://doi.org/10.1016/S0889-1591(02)00036-3)
 65. Demir S, Atli A, Bulut M, Okan İbiloğlu A, Güneş M, Kaya MC, et al. Neutrophil-lymphocyte ratio in patients with major depressive disorder undergoing no pharmacological therapy. *Neuropsychiatr Dis Treat*. 2015;11:2253. <https://doi.org/10.2147/NDT.S89470>
 66. Warriss PD, Brown SN, Edwards JE, Knowles TG. Effect of lairage time on levels of stress and meat quality in pigs. *Anim Sci*. 1998;66:255-61. <https://doi.org/10.1017/S1357729800009036>
 67. Oyarzún R, Paredes R, Saravia J, Morera FJ, Muñoz JLP, Ruiz-Jarabo I, et al. Stocking density affects the growth performance, intermediary metabolism, osmoregulation, and response to stress in Patagonian blennie *Eleginops maclovinus*. *Aquaculture*. 2020;515:734565. <https://doi.org/10.1016/j.aquaculture.2019.734565>
 68. Chrousos GP, Kino T. Glucocorticoid action networks and complex psychiatric and/or somatic

- disorders. *Stress*. 2007;10:213-9. <https://doi.org/10.1080/10253890701292119>
69. Jang JC, Jin XH, Hong JS, Kim YY. Effects of different space allowances on growth performance, blood profile and pork quality in a grow-to-finish production system. *Asian-Australas J Anim Sci*. 2017;30:1796-802. <https://doi.org/10.5713/ajas.17.0076>
 70. Colditz IG. Effects of the immune system on metabolism: implications for production and disease resistance in livestock. *Livest Prod Sci*. 2002;75:257-68. [https://doi.org/10.1016/S0301-6226\(01\)00320-7](https://doi.org/10.1016/S0301-6226(01)00320-7)
 71. Fossum C. Cytokines as markers for infections and their effect on growth performance and well-being in the pig. *Domest Anim Endocrinol*. 1998;15:439-44. [https://doi.org/10.1016/S0739-7240\(98\)80001-5](https://doi.org/10.1016/S0739-7240(98)80001-5)
 72. Fan J, Molina PE, Gelato MC, Lang CH. Differential tissue regulation of insulin-like growth factor-I content and binding proteins after endotoxin. *Endocrinology*. 1994;134:1685-92. <https://doi.org/10.1210/endo.134.4.7511091>
 73. Johnson OL, Jaworowicz W, Cleland JL, Bailey L, Charnis M, Duenas E, et al. The stabilization and encapsulation of human growth hormone into biodegradable microspheres. *Pharm Res*. 1997;14:730-5. <https://doi.org/10.1023/A:1012142204132>
 74. Murtaugh MP, Baarsch MJ, Zhou Y, Scamurra RW, Lin G. Inflammatory cytokines in animal health and disease. *Vet Immunol Immunopathol*. 1996;54:45-55. [https://doi.org/10.1016/S0165-2427\(96\)05698-x](https://doi.org/10.1016/S0165-2427(96)05698-x)
 75. Dinarello CA. Proinflammatory cytokines. *Chest*. 2000;118:503-8. <https://doi.org/10.1378/chest.118.2.503>
 76. Kim KH, Kim KS, Kim JE, Kim DW, Seol KH, Lee SH, et al. The effect of optimal space allowance on growth performance and physiological responses of pigs at different stages of growth. *Animal*. 2017;11:478-85. <https://doi.org/10.1017/S1751731116001841>
 77. Costa LB, Luciano FB, Miyada VS, Gois FD. Herbal extracts and organic acids as natural feed additives in pig diets. *S Afr J Anim Sci*. 2013;43:181-93. <https://doi.org/10.4314/sajas.v43i2.9>
 78. Dorman HJD, Deans SG. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol*. 2000;88:308-16. <https://doi.org/10.1046/j.1365-2672.2000.00969.x>
 79. Lee DY, Li H, Lim HJ, Lee HJ, Jeon R, Ryu JH. Anti-inflammatory activity of sulfur-containing compounds from garlic. *J Med Food*. 2012;15:992-9. <https://doi.org/10.1089/jmf.2012.2275>
 80. Pirgozliev V, Mansbridge SC, Rose SP, Lillehoj HS, Bravo D. Immune modulation, growth performance, and nutrient retention in broiler chickens fed a blend of phyto-genic feed additives. *Poult Sci*. 2019;98:3443-9. <https://doi.org/10.3382/ps/pey472>
 81. Bryant MJ, Ewbank R. Some effects of stocking rate and group size upon agonistic behaviour in groups of growing pigs. *Br Vet J*. 1972;128:64-70. [https://doi.org/10.1016/S0007-1935\(17\)37133-6](https://doi.org/10.1016/S0007-1935(17)37133-6)
 82. Hemsworth PH, Rice M, Nash J, Giri K, Butler KL, Tilbrook AJ, et al. Effects of group size and floor space allowance on grouped sows: aggression, stress, skin injuries, and reproductive performance. *J Anim Sci*. 2013;91:4953-64. <https://doi.org/10.2527/jas.2012-5807>
 83. Nannoni E, Martelli G, Rubini G, Sardi L. Effects of increased space allowance on animal welfare, meat and ham quality of heavy pigs slaughtered at 160Kg. *PLOS ONE*. 2019;14:e0212417. <https://doi.org/10.1371/journal.pone.0212417>
 84. Turner SP, Ewen M, Rooke JA, Edwards SA. The effect of space allowance on performance, aggression and immune competence of growing pigs housed on straw deep-litter at different group sizes. *Livest Prod Sci*. 2000;66:47-55. [https://doi.org/10.1016/S0301-6226\(00\)00159-7](https://doi.org/10.1016/S0301-6226(00)00159-7)

85. Menchetti L, Nanni Costa L, Zappaterra M, Padalino B. Effects of reduced space allowance and heat stress on behavior and eye temperature in unweaned lambs: a pilot study. *Animals*. 2021;11:3464. <https://doi.org/10.3390/ani11123464>
86. Zhang H, Tsao R. Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. *Curr Opin Food Sci*. 2016;8:33-42. <https://doi.org/10.1016/j.cofs.2016.02.002>
87. Pearce GP, Paterson AM. The effect of space restriction and provision of toys during rearing on the behaviour, productivity and physiology of male pigs. *Appl Anim Behav Sci*. 1993;36:11-28. [https://doi.org/10.1016/0168-1591\(93\)90095-7](https://doi.org/10.1016/0168-1591(93)90095-7)