## Effect of Population Density on Development Time of Tenebrio molitor

Da Yeon Choi<sup>1†</sup>, Ji Yun Yun<sup>1†</sup>, Seo Yun Kim<sup>1</sup>, Ga Eun Lee<sup>1</sup>, Kyra Batarse<sup>2</sup>, Steven  $\mathrm{Kim}^3$ , and Dong Sub  $\mathrm{Kim}^{4\star}$ 

<sup>1</sup>Undergraduate Student, Department of Horticulture, Kongju National University, Yesan 32439, Korea <sup>2</sup>Undergraduate Student, Department of Mathematics and Statistics, California State University, Monterey Bay, Seaside, CA 93955, United States of America <sup>3</sup> Associate Professor, Department of Mathematics and Statistics, California State University, Monterey Bay, Seaside, CA 93955, United States of America

Assistant Professor, Department of Horticulture, Kongju National University, Yesan 32439, Korea

Abstract. Mealworms are used as food, so it is preferable if the larval stage lasts longer. On the other hand, to accelerate the population growth of mealworms, it is preferable if the larvae become adults quickly. The purpose of this study is to explore the effects of population density on development time of mealworms. We used a container size of 7 cm at the top, 5 cm at the bottom, and 3 cm in height. Mealworms lived in the containers at densities of 1, 2, 5, 10, and 20 per container. The containers were bedded with 1 g of wheat bran and formed two groups, fed and not fed, at each density levels. The experiments were performed three times. In all of the experiments, higher population densities resulted in shorter transformation times from larva to pupa, but the time from pupa to imago was not significantly different. In addition, given the same density, the presence of food accelerated the time to transformation to pupa, but not to imago. The data supported that a lower density is needed to prolong the larval stage, and if adults are needed at a faster rate, the density should be higher. Therefore, we conclude that the development time of mealworms can be controlled by the density which is useful information for mealworm farmers.

Additional key words: Cox proportional-hazards model, imago, mealworms, pupa

## Introduction

Global climate change affects not only human life, but also crops and livestock. Lemonnier and Ainsworth (2018) reported that  $CO<sub>2</sub>$  in the air would be changed from the current 420 ppmto 540 ppm in 2050, and the surface temperature would be elevated due to the greenhouse effect. The elevated temperature accelerates evapotranspiration, triggering soil salinization, and water deficit simultaneously which can result in declining crop productivity. The higher  $CO<sub>2</sub>$  can improve C3 crop yield because although it makes photosynthesis more active, it can also reduce protein and minerals by  $5-15%$  and vitamin B by up to 30% (Loladze, 2014; Myers et al., 2014; Zhu et al., 2018). The change in nutritional quality, more carbohydrates, and less minerals can ultimately affect livestock and human health (Loladze,

2002). For instance, C3 grains and tubers, such as rice, mago, mealworms, pupa<br>2002). For instance, C3 grains and tubers, such as rice,<br>wheat, barley, and potatoes, have a  $7-15%$  reduction in protein content (Myers et al., 2014). Moreover, as  $CO<sub>2</sub>$ increases, the zinc content in plants decreases, and the number of zinc-deficient animals is predicted to increase (Myers et al., 2015).

In addition to the potential impact of climate change, the current trend of world population and urbanization hinders food security. Ritchie and Roser (2023) reported that the world population is expected to increase from the current 7.91 billion people to 9.71 billion in 2050 and 10.35 billion in 2100. According to the statistical data of FAO, however, the average protein supply is declining around the world (Miassi and Dossa, 2023). Tilman and Clark (2014) and Godfray et al. (2018) reported that the current animal protein consumption is not sustainable given the food demand and the projected population. Meanwhile, urbanization reduces farms for feed crop production and ranches for livestock production. Even if new farms and ranches are built, the agricultural system actively contributes to deforestation,

<sup>†</sup>These authors contributed equally to this work

<sup>\*</sup>Corresponding author: dongsub@kongju.ac.kr

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water consumption, and  $CO<sub>2</sub>$  emissions (Semba et al., 2021).

Entomophagy has been used as a nutritional source for both animal feed and food for humans. The edible insects contain protein, essential amino acids, minerals, and vitamins. Not only from the nutritional perspective but also from an environmental perspective, entomophagy might play a major role in protein sources. For instance, insects require less water (Miglietta et al., 2015) and arable land (Oonincx and Boer,  $2012$ ) and emit lower  $CO<sub>2</sub>$  (Oonincx et al.,  $2010$ ) than pigs, cattle, and poultry. Thus, entomophagy is more sustainable than the existing protein production system (Heckmann et al., 2018).

Mealworm (Tenebrio molitor) is well known as an edible insect. In general, the insects react sensitively to air temperature and relative humidity, and the environmental conditions for the mealworm are 20°C and 75% (Punzo and Mutchmor, 1980). To run a mealworm farm effectively, there are two main strategies to consider for farmers. The first strategy is to maintain the edible larval period to the maximum, and the second strategy is to produce as many adults as possible for reproduction. Since these two strategies may be naturally contradictory, it is hard to find environmental conditions that satisfy both strategies, and it may require more cost and space. One simple strategy, if it is effective, is to induce food competition among mealworms by changing the density of their habitat. Therefore, we performed an experimental study by controlling the density of mealworm habitat and the presence of food.

## Materials and Methods

#### 1. Samples

Mealworm larvae (Tenebrio molitor L) with a length of Materials and Methods<br>1. Samples<br>Mealworm larvae (*Tenebrio molitor* L) with a length of<br>2-3 cm were purchased from an insect farm, which specializes in mealworms, in South Korea. The larvae were grown in cylindrical containers with a dimension of 7 cm at the top, 5 cm at the bottom, and 3 cm in height. The temperature was maintained about 25°C, and the photoperiod was kept at 14 Light: 10 Dark (Kim et al., 2015; Punzo and Mutchmor, 1980). The relative humidity was maintained at about 60%. The containers were bedded with 1 g of wheat bran (Deen et al., 2021; Langston et al., 2023) and were labeled "F" if mealworms were fed and "S" if they were not

fed. Mealworms were placed in the containers at densities of 1, 2, 5, 10, and 20 and were provided at 0.2 g of radish per larva in the F-labeled containers (Park et al., 2012; Kim et al., 2014). The mealworms that became pupae were immediately transferred to another container. Three experiments were performed. Experiment 1 was from July 19 to September 16, Experiment 2 was from September 23 to November 20, and Experiment 3 was from November 16 to December 16 in 2023. The treatment groups and number of containers are shown in Table 1.

#### 2. Statistical analysis

The date of starting the experiment, the date of transforming to pupa, and the date of transforming to imago were recorded for each mealworm. Using these records, the time to pupa and the time to imago from pupa were calculated. Some mealworms were censored (i.e., did not become a pupa or an imago) because they died in the middle of the test or did not become a pupa or an imago until the end of the experiment. To account for the censored data, survival analysis was performed (Klein and Moeschberger, 2003). There were two experimental factors of interest, food and density. There were two levels of food, fed or not, and there were five levels of density: 1, 2, 5, 10, or 20 mealworms per container of the

Table 1. The number of containers and the sample size in Experiment 1 (pilot study), Experiment 2 (main study), and Experiment 3 (repeated study).

Experiment	Density	Fed	Not fed
	1 per container	20	20
	2 per container	10	10
	5 per container	$\overline{4}$	4
	10 per container	2	2
	20 per container		
	1 per container	20	20
	2 per container	10	10
$\mathcal{D}_{\mathcal{L}}$	5 per container	4	4
	10 per container	$\mathfrak{D}$	2
	20 per container		
	1 per container	0	36
	2 per container	0	20
3	5 per container	0	7
	10 per container	0	5
	20 per container		2

same size. The time to each phase was visualized using the Kaplan-Meyer estimator, and the effects of the two factors, food and density, were estimated using the Cox proportionalhazards model.

For the statistical model, we let  $X = 1$  if a mealworm was fed and  $X = 0$  if not fed (the reference level of the food group). To indicate the five density levels, four dummy variables were defined as follows:  $D_2 = 1$  if two mealworms were in the container ( $D_2 = 0$  otherwise);  $D_5 = 1$  if five mealworms were in the container ( $D_5 = 0$  otherwise);  $D_{10} = 1$  if ten mealworms were in the container ( $D_{10} = 0$  otherwise); and  $D_{20} =$ 1 if twenty mealworms were in the container  $(D_{20} = 0$ otherwise). That is,  $D_2 = D_5 = D_{10} = D_{20} = 0$  if only one mealworm was in the container alone (the reference level of the density group). Given these notations, the Cox proportional-hazards model was specified as follows:

$$
\lambda(t|X,D)=\lambda_0(t)\exp(\beta_1X+\beta_2D_2+\beta_3D_5+\beta_4D_{10}+\beta_5D_{20})
$$

The parameters of interest  $(\beta_1, \beta_2, \beta_3, \beta_4, \text{ and } \beta_5)$  were estimated using the survival package in R (Therneau and Grambsch, 2000; Therneau, 2023). Under the model assumption,  $exp(\beta_1)$  is the ratio of the likelihood of the transformation to the next phase (to pupa or from pupa to imago) at any time when the F group is compared to the S group of the same density;  $exp(\beta_2)$  is the ratio when the group of 2 per container and the group of 1 per container (the reference group) are compared given the same food con-

dition;  $\exp(\beta_3)$  is the ratio when the group of 5 per container and the reference group are compared given the same food condition; and so on. The model assumption of proportionalhazards was tested using the Schoenfeld residuals (Grambsch and Therneau, 1994). All statistical analyses were performed in R version 4.3.0 (R Core Team, 2023).

## Results

An expected power outage occurred during Experiment 1, and it was regarded as a pilot experiment in this study. The main experiment, referred to as Experiment 2, allowed us to estimate both the effect of food and density. The results of Experiment 2 associated with the transformation to pupa from larva are reported in Section 3.1, and the results of Experiment 2 associated with the transformation to image from pupa are reported in Section 3.2. Given the remaining resources, we could repeat the experiment without food, referred to as Experiment 3, and the results of Experiment 3 are reported in Section 3.3. All combined results are summarized in Section 4.

#### 1. Experiment 2 (time to pupa from larva)

Among the 200 mealworms, 152 mealworms transformed to pupa, and the remaining 48 mealworms did not transform to pupa until the end of observation or died before the end of the test. As shown in the left panel of Fig. 1, the median survival time was 19 days for the group with food and 28



Fig. 1. The Kaplan-Meyer estimators by food group (A) and by density group (B) for transformation to pupa (Experiment 2).

Table 2. Results of the Cox proportional-hazards model (Experiment 2; transformation to pupa).

	Estimate	95% CI	$p$ -value
$exp(\beta_1)$	1.558	(1.129, 2.152)	0.007
$exp(\beta_2)$	1.414	(0.811, 2.467)	0.222
$exp(\beta_3)$	2.126	(1.242, 3.639)	0.005
$exp(\beta_4)$	2.471	(1.456, 4.193)	${}< 0.001$
$exp(\beta_5)$	2.798	(1.642, 4.769)	${}< 0.001$



Fig. 2. The Kaplan-Meyer estimators by food group (A) and density group (B) for transformation to imago (Experiment 2).

days for the group without food. Under the Cox proportionalhazards model, at any time point before transforming to pupa, a mealworm with food is 1.56 times more likely to transform to pupa than a mealworm without food given the same density ( $p = 0.007$ ). The right panel of Fig. 1 shows that the group of mealworms in the container of a higher density transformed to pupa faster when compared to those of a lower density. Under the Cox model, when the group of 20 mealworms per container is compared to the group of one mealworm per container (the reference group) given the same food condition, the group of 20 mealworms per container is 2.80 times more likely to transform to pupa ( $p \leq$ 0.001). Relative to the reference, given the same food condition, the group of 10 mealworms per container was statistically significant (2.47 times;  $p \le 0.001$ ), the group of 5 mealworms per container was statistically significant as well (2.13 times;  $p = 0.005$ ), and the group of 2 mealworms per container was not statistically significant (1.41 times;  $p =$ 0.222) as reported in Table 2. The Cox proportional-hazards assumption was assessed, and the data supported the model

assumption ( $p = 0.75$ ).

#### 2. Experiment 2 (time to imago from pupa)

Among the 152 mealworms that had transformed into a pupa, 140 pupae had then metamorphosed into imago. The remaining 12 pupae either died or did not transform into imago before the end of the experiment. Overall, the effect of food and density on the transformation to imago was not as drastic as on the transformation to pupa. Using the Cox proportional hazard model, there was no statistically significant effect of food or density. When the group of pupae fed was compared to the group of pupae not fed, given the same density of container, the likelihood of transforming into imago was 0.8 times at any time of the imago stage, and it was not statistically significant ( $p = 0.508$ ). On the left panel of Fig. 2, the estimated median transformation time was 8 and 9 days for the group fed and the group not fed. Likewise, the estimated median transformation time was 8 and 9 days for all five density groups (1, 2, 5, 10, and 20 pupae per container) as shown in the right panel of Fig. 2.

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Table 3. Results of the Cox proportional-hazards model (Experiment 2; transformation to imago).

	Estimate	95% CI	$p$ -value
$exp(\beta_1)$	0.890	(0.631, 1.256)	0.508
$exp(\beta_2)$	0.932	(0.516, 1.683)	0.814
$exp(\beta_3)$	1.104	(0.628, 1.941)	0.732
$exp(\beta_4)$	1.257	(0.727, 2.176)	0.413
$exp(\beta_5)$	1.250	(0.719, 2.170)	0.429

Table 4. Results of the Cox proportional-hazards model (Experiment 3; all mealworms not fed).





Fig. 3. The Kaplan-Meyer estimators by density group (1, 2, 5, 10, and 20 mealworms per container) for transformation to pupa (A) and transformation to imago (B) where all mealworms were not fed (Experiment 3).

When compared to the reference density group (1 per container), the effect of higher density was not statistically significant as reported in Table 3. There was no strong evidence of violating the proportional-hazards assumption  $(p = 0.073)$ .

## 3. Experiment 3

The experiment was repeated to replicate the observed pattern. In this experiment, where all mealworms were not fed, 201 mealworms that were not fed, 123 out of the 201

mealworms transformed to pupa before the end of observation, and 106 out of the 123 pupae transformed to imago before the end of observation. Table 4 and Fig. 3 show that the time to transformation to pupa was significantly different with respect to the density group (Fig. 3A), but the time to transformation to imago was not (Fig. 3B). The time to transformation to pupa observed in this repeated experiment was consistent with the previous experiment. Especially, the two highest density groups (20 and 10 per container) had similar time to transformation to pupa.

#### 4. Combined results

Fig. 4 shows the estimated relative likelihood of transformation to pupa from Experiment 2 (the main study) and Experiment 3 (the experiment for replication). The observed trends are similar between the two experiments that the transformation to pupa tends to be faster at high density groups. Given the density group, in Experiment 2, we could observe that food accelerated the transformation to pupa. Fig. 5 shows the estimated relative likelihood of transformation to imago from Experiment 2 and Experiment 3. The trend with respect to density is constant (i.e., no effect of density), and food does not seem to be effective on the time to the transformation to imago.

## Discussion

High population density can be a stressor for animals. Higher population density of chickens caused pecking themselves, resulting in poor coat condition (Hofmann et al., 2021), and higher population density of pigs increased the tendency to attackother pigs (Hwang et al., 2019). Lee et al. (2023) found that flounder had an increased deformity rate at higher population density. Our study also showed that rapid pupation occurs as larval population density increases. This means that as the population density increases, the larvae may also experience stress which results in a decrease in development time.



Fig. 4. Estimated relative likelihood of transformation to pupa by density group (A) and food group (B). The statistical significance is marked (\*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001) relative to each reference group (one mealworm per container and not fed).



Fig. 5. Estimated relative likelihood of transformation to imago by density group (A) and food group (B). The effects of density and food are not statistically significant.

In terms of insect development, larvae of some fly species, Calliphora vicina, have reduced development times at high population density (Saunders and Bee, 1995). Previous studies reported that the geometric moth (Välimäki et al., (2013) and the tropical butterfly Bicyclus anynana (Bauerfeind and Fischer, 2005) also had a shortened developmental time by high density. However, in the case of mosquitoes (Agnew et al., 2002; Gimnig et al., 2002; Muriu et al., 2013) and flies (Diamantidis et al., 2020; Henry et al., 2018), there was a significant tendency for development times to be delayed as the larval population density increased.

Coleoptera and Lepidoptera had inconsistent development times across population density. For the Zophobas atratus Fab species, higher density has been delayed development times (Quennedey et al., 1995). Gibbs et al. (2004) found the same developmental delay response among Lepidoptera species. The same response was found in the studies using the Tenebrio molitor (Connat et al., 1991). However, the environmental conditions of the Connat's study were different from our experiments. Connat conducted the study using 9.9 cm<sup>2</sup> sized containers, which is approximately 10 times greater than the  $0.95 \text{ cm}^2$  containers in our experiments. On the other hands, in the other Tenebrio molitor studies, population density did not result in significant differences in pupal stage times (Weaver and McFarlane, 1990). Therefore, the materials (container size) could be an effect modifier in the relationship between larval population density and development stage. In the scope of our experimental settings, however, we have observed the consistent pattern that higher density accelerated the transformation to pupa but not to imago.

At high population density, feeding efficiency might have been reduced, leading to lack of nutrients. Morales-Ramos and Rojas (2015) fed a constant amount of food per Tenebrio molitor larva, but as the population density increased, food consumption and individual larval weight decreased. It may be due to 20-hydroxyendisone (20E), which induces lipolysis and pupalization (Riddiford, 2020; Liu et al., 2015). Terashima et al. (2005) reported that starvation in Drosophila melanogaster increases 20E concentration due to lack of nutrients. It is potentially associated with the time of development stage, and the potential causal paths needs to be proven in future study.

## Literature Cited

- Agnew P., M. Hide, C. Sidobre, and Y. Michalakis 2002, A minimalist approach to the effects of density-dependent competition on insect life-history traits. Ecol Entomol 27: 396-402. doi:10.1046/j.1365-2311.2002.00430.x
- Bauerfeind S.S., and K. Fischer 2005, Effects of food stress and density in different life stages on reproduction in a butterfly. Oikos 111:514-524. doi:10.1111/j.0030-1299.2005.13888.x
- Connat L., J.P. Delbecque, I. Glitho, and J. Delachambre 1991, The onset of metamorphosis in Tenebrio molitor larvae (Insecta, Coleoptera) under grouped, isolated and starved conditions. J Insect Physiol 37:653-662. doi:10.1016/0022- 1910(91)90042-X
- Deen N.E., S. Lamaj, F. Verrastro, V. Al Bitar, and L.F. Baldacchino 2021, Effects of two diets on adults' survival and productivity in mass-rearing of Tenebrio molitor (Coleaoptera: Tenebriodae). J Insects Food Feed 7:1149-1157. doi:10.3920/JIFF2020.0129
- Diamantidis A.D., C.S. Ioannou, C.T. Nakas, J.R. Carey, and N.T. Papadopoulos 2020, Differential response to larval crowding of a long‐ and a short‐lived medfly biotype. J Evol Biol 33:329-341. doi:10.1111/jeb.13569
- Gibbs M., L.A. Lace, M.J. Jones, and A.J. Moore 2004, Intraspecific competition in the speckled wood butterfly Pararge aegeria: Effect of rearing density and gender on larval life history. J Insect Sci 4:16. doi:10.1093/jis/4.1.16
- Gimnig J.E., M. Ombok, S. Otieno, M.G. Kaufman, J.M. Vulule, and E.D. Walker 2002, Density-dependent development of Anopheles gambiae (Diptera: Culicidae) larvae in artificial habitats. J Med Entomol 39:162-172. doi:10.1603/0022- 2585-39.1.162
- Godfray H.C.J., P. Aveyard, T. Garnett, J.W. Hall, T.J. Key, et al. 2018, Meat consumption, health, and the environment. Science 361:eaam5324. doi:10.1126/science.aam5324
- Grambsch P., and T. Therneau 1994, Proportional hazards tests and diagnostics based on weighted residuals. Biometrika 81:515-526. doi:10.1093/biomet/81.3.515
- Heckmann L.H., J.L. Andersen, N. Gianotten, M. Calis, C.H. Fischer, and H. Calis 2018, Sustainable mealworm production for feed and food. In A Halloran, R Flore, P Vantomme, N Roos, eds, Edible Insects in Sustainable Food Systems. Springer, New York, NY, USA, pp 321-328. doi:10.1007/ 978-3-319-74011-9\_19
- Henry Y., D. Renault, and H. Colinet 2018, Hormesis-like effect of mild larval crowding on thermotolerance in Drosophila flies. J Exp Biol 221:jeb169342. doi:10.1242/ jeb.169342
- Hofmann T., S. Schmucker, M. Grashorn, and V. Stefanski 2021, Short- and long-term consequences of stocking

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density during rearing on the immune system and welfare of laying hens. Poult Sci 100:101243. doi:10.1016/j.psj.2021. 101243

- Hwang H.S., J.K. Lee, T.K. Eom, T.J. Choi, and S.J. Rhim 2019, Influence of mixing density on behaviors of growing pigs. Ann Anim Resour Sci 30:18-24. doi:10.12718/AARS. 2019.30.1.18
- Kim S.Y., J.B. Park, Y.B. Lee, H.J. Yoon, K.Y. Lee, and N.J. Kim 2015, Growth characteristics of mealworm Tenebrio molitor. J Seric Entomol Sci 53(1):1-5. doi:10.7852/jses. 2015.53.1.1
- Kim S.Y., T.H. Chung, S.H. Kim, S. Song, and N.J. Kim 2014, Recycling agricultural wastes as feed for mealworm (Tenebrio molitor). Korean J Appl Entomol 53:367-373. doi:10.5656/ KSAE.2014.10.0.043
- Klein J.P., and M.L. Moeschberger 2003, Survival analysis: Techniques for censored and truncated data, 2nd Ed. Springer, New York, USA. doi:10.1007/b97377
- Langston K., L. Selaledi, and A. Yusuf 2023, Evaluation of alternative substrates for rearing the yellow mealworm Tenebrio molitor (L). Int J Trop Insect Sci 43:1523-1530. doi:10.1007/s42690-023-01061-z
- Lee S., J.W. Park, M. Jeong, H.S. Jung, J. Kim, W.J. Kim, et al. 2023, Effects of rearing density stress on malformation and stress and immune related gene expression of juvenile olive flounder Paralichthys olivaceus. Korean J Fish Aquat Sci 56:380-387. doi:10.5657/KFAS.2023.0380
- Lemonnier P., and E.A. Ainsworth 2018, Crop responses to rising atmospheric  $[CO_2]$  and global climate change. In SS Redden, R Yadav, eds, Food Security and Climate Change. John Wiley & Sons Ltd., Chichester, England, UK, pp 51-69. doi:10.1002/9781119180661.ch3
- Liu X., F. Dai, E. Guo, K. Li, L. Ma, L. Tian, et al. 2015, 20-Hydroxyecdysone (20E) primary response gene E93 modulates 20E signaling to promote Bombyx larval-pupal metamorphosis. J Biol Chem 290:27370-27383. doi:10.1074/ jbc.m115.687293
- Loladze I. 2002, Rising atmospheric  $CO<sub>2</sub>$  and human nutrition: toward globally imbalanced plant stoichiometry? Trends Ecol Evol 17:457-461. doi:10.1016/S0169-5347(02)02587-9
- Loladze I. 2014, Hidden shift of the ionome of plants exposed to elevated CO<sub>2</sub> depletes minerals at the base of human nutrition. eLife 3:e02245. doi:10.7554/elife.02245
- Miassi Y.E. and K.F. Dossa 2023, Forecasting animal protein supply in Asia and Europe in light of climate change, population growth and land pressure. Trop Plant 2:22. doi: 10.48130/TP-2023-0022
- Miglietta P., F. De Leo, M. Ruberti, and S. Massari 2015, Mealworms for food: A water footprint perspective. Water 2015, 7:6190-6203. doi:10.3390/w7116190
- Morales-Ramos J.A., and M.G. Rojas 2015, Effect of larval density on food utilization efficiency of Tenebrio molitor

(Coleoptera: Tenebrionidae). J Econ Entomol 108:2259-2267. doi:10.1093/jee/tov208

- Muriu S.M., T. Coulson, C.M. Mbogo, and H.C. Godfray 2013, Larval density dependence in Anopheles gambiae s.s., the major African vector of malaria. J Anim Ecol 82:166-174. doi:10.1111/1365-2656.12002
- Myers S.S., A. Zanobetti, I. Kloog, et al. 2014, Increasing  $CO<sub>2</sub>$ threatens human nutrition. Nature 510:139-142. doi:10.1038/ nature13179
- Myers S.S., K.R. Wessells, I. Kloog, A. Zanobetti, and J. Schwartz 2015, Effect of increased concentrations of atmospheric carbon dioxide on the global threat of zinc deficiency: a modelling study. Lancet Global Health 3:e639-e645. doi: 10.1016/s2214-109x(15)00093-5
- Oonincx D.G. and I.J. De Boer 2012, Environmental impact of the production of mealworms as a protein source for humans – a life cycle assessment. PLoS ONE 7:e51145. doi:10.1371/ journal.pone.0051145
- Oonincx D.G., J. Van Itterbeeck, M.J. Heetkamp, H. Van Den Brand, J.J. Van Loon, and A. Van Huis 2010, An exploration on greenhouse gas and ammonia production by insect species suitable for animal or human consumption. PLoS ONE 5:e14445. doi:10.1371/journal.pone.0014445
- Park Y.K., Y.C. Choe, Y.B. Lee, S.H. Lee, J.S. Lee, and S.H. Kang 2012, Fecundity, life span, developmental periods and pupal weight of Tenebrio molitor L. (Coleoptera: Tenebrionidae). J Seric Entomol Sci 50:126-132. doi:10.7852/jses. 2012.50.2.126
- Punzo F., and J.A. Mutchmor 1980, Effects of temperature, relative humidity and period of exposure on the survival capacity of Tenebrio molitor (Coleoptera: Tenebrionidae). J Kans Entomol Soc 53(2):260-270.
- Quennedey A., N. Aribi, C. Everaerts, and J.P. Delbecque 1995, Postembryonic development of Zophobas atratus Fab. (Coleoptera: Tenebrionidae) under crowded or isolated conditions and effects of juvenile hormone analogue applications. J Insect Physiol 41:143-152. doi:10.1016/0022-1910(94) 00091-T
- R Core Team 2023, R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at: https://www.R-project.org/.
- Riddiford L.M. 2020, Rhodnius, golden oil, and Met: A history of juvenile hormone research. Front Cell Dev Biol 8:679. doi:10.3389/fcell.2020.00679
- Ritchie H. and M. Roser 2023, The world population is changing: For the first time there are more people over 64 than children younger than 5. Our World in Data.
- Saunders D.S., and A. Bee 1995, Effects of larval crowding on size and fecundity of the blow fly, Calliphora vicina (Diptera: Calliphoridae). Eur J Entomol 92:615-622.
- Semba R.D., R. Ramsing, N. Rahman, K. Kraemer, and M.W. Bloem 2021, Legumes as a sustainable source of protein in

human diets. Glob Food Secur 28:100520. doi:10.1016/j.gfs. 2021.100520

- Terashima J., K. Takaki, S. Sakurai, and M. Bownes 2005, Nutritional status affects 20-hydroxyecdysone concentration and progression of oogenesis in Drosophila melanogaster. J Endocrinol 187:69-79. doi:10.1677/joe.1.06220
- Therneau T. 2023, A package for survival analysis in R. R package version 3.5-5, https://CRAN.R-project.org/package= survival
- Therneau T.M. and P.M. Grambsch 2000, Modeling survival data: Extending the Cox model. Springer, New York. ISBN 0-387-98784-3. doi:10.1007/978-1-4757-3294-8
- Tilman D. and M. Clark 2014, Global diets link environmental sustainability and human health. Nature 515:518-522. doi:

10.1038/nature13959

- Välimäki P., S.M. Kivelä, and M.I. Mäenpää 2013, Temperature- and density-dependence of diapause induction and its life history correlates in the geometrid moth Chiasmia clathrata (Lepidoptera: Geometridae). Evol Ecol 27:1217-1233. doi:10.1007/s10682-013-9657-8
- Weaver D.K. and J. McFarlane 1990, The effect of larval density on growth and development of Tenebrio molitor. J Insect Physiol 36:531-536. doi:10.1016/0022-1910(90)90105-O
- Zhu C., K. Kobayashi, I. Loladze, et al. 2018, Carbon dioxide  $(CO<sub>2</sub>)$  levels this century will alter the protein, micronutrients, and vitamin content of rice grains with potential health consequences for the poorest rice-dependent countries. Sci Adv 4:eaaq1012. doi:10.1126/sciadv.aaq1012

# Tenebrio molitor의 개체수 밀도가 발달 시간에 대한 영향

- /라/��//�� //��//�� - / //에 | - 글포기 클클 시단에 베른 OO<br>최다연<sup>1†</sup>ㆍ윤지윤<sup>1†</sup>ㆍ김서윤<sup>1</sup>ㆍ이가은<sup>1</sup>ㆍ키라 바타르세<sup>2</sup>ㆍ스티븐 김<sup>3ㆍ</sup>김동섭<sup>4</sup>

<sup>1</sup>공주대학교 산업과학대학 원예학과 학부생, <sup>2</sup>캘리포니아 주립대학교 몬트레이베이 수학통계학과 학부생, <sup>3</sup>캘리포니아 주립대학교 몬트레이베이 수학통계학과 부교수, <sup>4</sup>공주대학교 산업과학대학 원예학과 조교수

적 요. 거저리는 식품으로 사용되기 때문에 유충기가 오래 지속되면 더 좋다. 반면에 거저리의 개체수 유지를 위 해 성장을 가속화하기 위해서는 유충이 빨리 성충이 되면 더 좋다. 이 연구에서는 개체군 밀도가 거저리의 발달 시 간에 미치는 영향을 구명하였다. 이를 위해 상단 7 cm, 하단 5 cm, 높이 3 cm 크기의 용기를 사용했다. 거저리는 용기 당 1, 2, 5, 10, 20마리의 밀도로 용기에서 서식하였다. 용기에 밀기울 1 g을 넣고 거저리의 먹이 여부에 따라 라벨을 붙였다. 실험은 세 번 반복되었다. 모든 실험에서 개체군 밀도가 높을수록 유충에서 번데기로의 변환 시간이 짧았 지만 번데기에서 성충으로 변환되는 시간은 크게 다르지 않았다. 또한 먹이가 있는 그룹에서 번데기로의 변환 시간 이 단축되었지만, 성충으로 변환되는 시간에는 차이가 없었다. 이 연구 결과는 유충기를 연장하기 위해 더 낮은 밀 도가 필요하고 더 빠른 속도로 성충이 필요하다면 밀도가 더 높아야 한다는 것을 보여주었다. 결론적으로 거저리의 발달 시간은 개체수 밀도에 의해 제어할 수 있을 것이다.