**Open Access** Anim Biosci Vol. 36, No. 12:1853-1859 December 2023 https://doi.org/10.5713/ab.23.0097 pISSN 2765-0189 eISSN 2765-0235



# Effects of different types of xanthophyll extracted from marigold on pigmentation of yellow-feathered chickens

Yu Wei<sup>1</sup>, Kun Qin<sup>1</sup>, Xu Qin<sup>1</sup>, Fulong Song<sup>1</sup>, and Xiao Xu<sup>1,\*</sup>

\* Corresponding Author: Xiao Xu Tel: +86-027-8395-6175, Fax: +86-027-8395-6175, E-mail: xuxiao@whpu.edu.cn

<sup>1</sup> Hubei Key Laboratory of Animal Nutrition and Feed Science (Wuhan Polytechnic University), School of Animal Science and Nutritional Engineering, Wuhan Polytechnic University, Wuhan 430023, China

#### ORCID

Yu Wei https://orcid.org/0009-0007-2841-2639 Kun Qin https://orcid.org/0009-0001-2740-5839 Xu Qin https://orcid.org/0009-0008-1250-8086 Fulong Song https://orcid.org/0009-0001-5603-722X Xiao Xu https://orcid.org/0000-0001-6559-3749

Submitted Mar 19, 2023; Revised Apr 28, 2023; Accepted May 17, 2023 **Objective:** This study aimed to explore the effects of different types of xanthophyll extracted from marigold on the growth performance, skin color, and carcass pigmentation.

**Methods:** A total of 192 healthy 60-day-old yellow-feathered broilers weighing an average of 1,279±81 g were randomly allocated to 4 groups, each with 6 replicates and 8 broilers. The 4 treatments were as follows: i) CON group, fed with basal diet; ii) LTN group, supplemented with lutein; iii) MDP group, supplemented with monohydroxyl pigment including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin; iv) LTN+MDP group, supplemented with lutein and monohydroxyl pigment in proportion to 1:1. The supplementary content of LTN, MDP, and LTN+MDP was 2 g/kg. Skin color was measured after 7, 14, 21, and 28 days of feeding the dietary treatments. The breast, thigh, and abdominal fat of slaughtered chickens were stored in cold storage at 4°C for 24 hours and then the meat color of lightness (L\*), redness (a\*), and yellowness (b\*) values was determined.

**Results:** The results showed that all treatments enhanced the yellow scores of subwing skin on day 14, 21, and 28 (p<0.05), and the mixture of lutein and monohydroxyl pigment promoted the yellow scores of shanks on day 14, 21, and 28 (p<0.05). The mixture of lutein and monohydroxyl pigment increased the yellow scores of beaks and all treatments enhanced the yellow of shanks on day 28 (p<0.05). In addition, all treatments improved the yellow (b\*) values of breast and thigh muscle, moreover, the monohydroxyl pigment and the mixture of lutein and monohydroxyl pigment enhanced the values of redness (a\*) and yellow (b\*) of abdominal fat (p<0.05).

**Conclusion:** In summary, different types of xanthophyll extracted from marigold significantly increased the yellow scores of skin color and the yellow (b\*) values of carcass pigmentation. Especially, the mixture of lutein and monohydroxyl pigment was more efficient on skin color.

**Keywords:** Marigold; Monohydroxyl Pigment; Pigmentation; Xanthophyll; Yellow-Feathered Chicken

## **INTRODUCTION**

The color of skin and pigmentation of muscle are decisive factors in determining the preference of consumers and evaluating the meat quality of broilers in many cosmopolitan regions [1,2]. However, it cannot be synthesized in animal bodies, so it must be taken from feed [3], but the quantity and types of pigments provided by corn and soybean-based commercial diets don't satisfy the preference of consumers for yellow-feathered broiler products [4]. Therefore, to meet the consumer's favour and market demand, the pigment has been applied as a feed additive for improving the color of broiler and aquaculture animal products which is the result of the pigment deposited into tissues [5]. The pigment usually includes a natural and a synthetic pigment, which is more steady and lower in price compared with natural pigment, nonetheless, the safety and bio-availability of synthetic pigment has been

Copyright © 2023 by Animal Bioscience

## /ID/

challenged [6]. A large number of works have indicated that carotenoid-enriched products have potential to improve the safety and bio-availability than synthetic pigment [7,8].

During the past several decades the premier natural pigment additive sources of carotenoids in poultry feed included yellow corn, dehydrated alfalfa meal, and corn gluten meal [9,10]. In recent years, curcumin and lutein gradually replaced yellow corn and corn gluten meal becoming the most common natural dietary carotenoids in commercial poultry feed [8]. Lutein, 3 '3 dihydroxy-a-carotene, is extracted from an abundant diversity of species of marigold [11]. And many monohydroxyl pigments occur in the metabolism and transformation of lutein including dehydrated lutein,  $\beta$ -cryptoxanthin,  $\alpha$ -cryptoxanthin, and so on [12]. A large amount of research has resulted in lutein substituted synthetic pigment becoming a universal additive, which can enhance egg yolk color more effectively [13,14], and increase the yellow scores of skin and muscle of broilers [4,8]. However, there are few reports on the effects of monohydroxyl pigments, such as dehydrated lutein, β-cryptoxanthin, and a-cryptoxanthin on pigmentation in broilers.

Hence, this study was conducted to explore the differences in pigmentation between lutein and monohydroxyl pigments (dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin) extracted from marigold, in addition, we mixed lutein and monohydroxyl pigments in proportion in 1:1 to discover the function.

### MATERIALS AND METHODS

#### Experimental animals and design

All experiments and procedures in this study were conducted according to the Animal Scientific Procedures Act 1986 (Home Office Code of Practice. HMSO: London January 1997) and EU regulations (Directive 2010/63/EU) and have been approved by the Institutional Animal Care and Use Committee of Wuhan Polytechnic University (No. WPU2022 11059). A total of 192 healthy 60-day-old broilers with an average body weight (BW) of 1,279±81 g were randomly divided into 4 groups with 6 replicates per group and 8 broilers per replicate. The 4 treatments were as follows: i) CON group, fed with basal diet; ii) LTN group, supplemented with lutein extracted from marigold; iii) MDP group, supplemented with monohydroxyl pigment extracted from marigold including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin; iv) LTN+MDP group, supplemented with lutein and monohydroxyl pigment in proportion to 1:1. All treatments were supplemented with different types xanthophyll according to the ratio of 0.2%. The yellow scores of skin color was evaluated on day 7, 14, 21, and 28, and breast muscle, thigh muscle, and abdominal fat of slaughtered chickens were stored in cold storage at 4°C for 24 hours and taken out to determine the meat color of lightness (L\*), redness (a\*), and yellowness (b\*) values. All chicks had free access to feed and water and were exposed to natural light and ventilation. The ambient temperature was maintained at 22 to 23 degrees Celsius with relative humidity controlled at 55% to 65%. The experiment lasted for 32 days, including 4 days of adaptation and 28 days of the experiment. The corn soybean meal-based basal diet formulated to meet the nutrition requirement for broilers is shown in Table 1, formulated according to the Poultry Nutrient Requirements [15]. The LTN, MDP, and the mixture of LTN+MDP were commercial products provided by Chen Guang Bio Co., Ltd, Hebei, China. The extraction of lutein from marigold is usually done with organic solvent extraction. According to the principle of similar phase dissolution, the solubility of lutein in different solvents is different, and the solvent with relatively high solubility is selected to dissolve lutein. Acid-catalyzed dehydration of lutein in a homogenous phase was carried out in a variety of solvents such as ethers, chlorinated solvents, acetone, and toluene at ambient temperature. Under reaction conditions, the monohydroxyl pigment including dehydrated lutein, β-cryptoxanthin, and α-cryptoxanthin was produced. The efficient content of LTN, MDP, and LTN+MDP is 1,662.8 ppm, 1,259 ppm, 841.4+629.2

Item	Ratio
Ingredients (%)	
Corn	60.58
Soybean meal, 44% crude protein	32.00
Soybean oil	3.00
Calcium hydrophosphate	1.20
Limestone	1.20
Salt	0.30
L-Lysine	0.35
DL-Methionine	0.22
L-Threonine	0.15
Premix <sup>1)</sup>	1.00
Total	100.00
Nutrient levels <sup>2)</sup>	
ME (kcal/kg)	3,100
Crude protein (%)	16.4
Calcium (%)	0.81
Available phosphorus (%)	0.35
Lysine (%)	0.72
Methionine + cystine (%)	0.49
Threonine (%)	0.54
Tryptophan (%)	0.17

ME, metabolizable energy

<sup>1)</sup> Premix supplied per kg diet: vitamin A, 12,500 IU; vitamin D<sub>3</sub>, 2,600 IU; vitamin E, 24 mg; vitamin K<sub>3</sub>, 4 mg; riboflavin, 5 mg; D-calcium pantothenate, 24 mg; nicotinic acid, 55 mg; cobalamin, 0.03 mg; manganese (MnSO<sub>4</sub>·H<sub>2</sub>O), 55 mg; rion (FeSO<sub>4</sub>·H<sub>2</sub>O), 75 mg; zinc (ZnSO<sub>4</sub>·7H<sub>2</sub>O), 36 mg; copper (CuSO<sub>4</sub>·5H<sub>2</sub>O), 8 mg; selenium (Na<sub>2</sub>SeO<sub>3</sub>), 0.15 mg. <sup>2)</sup> The nutrients contents of metabolizable energy and available phospho-

rus were calculated values, and lysine, methionine + cystine, threonine, crude protein and calcium content were analytical value. ppm, respectively.

#### Growth measurements

The health and death conditions of broilers were recorded every day during the trial period, and the BW and feed intake of broilers were determined on day 1 and day 28, respectively, and average daily gain (ADG), average daily feed intake (ADFI), and feed to gain ratio (F/G) were calculated.

#### Skin color

Four chickens per replicate were randomly selected on day 7, 14, 21, and 28 during the trial period, and the yellow scores of shanks and beaks were evaluated by using the Kemin color fan obeying color scores from 1 to 14, according to previous research [2]. Skin color of the lateral subwing area of the chickens, including lightness (L\*), redness (a\*), and yellowness (b\*) values, were measured using a Chromameter (CR-410, Konica Minolta, Tokyo, Japan).

#### **Carcass pigmentation**

After sampling on the 28th day of the trial period, three chickens were randomly selected from each replicate for slaughtering, and breast muscle, thigh muscle, and abdominal fat were collected. Samples of slaughtered chickens were stored in cold storage at 4°C for 24 hours and taken out to determine the meat color of lightness (L\*), redness (a\*), and yellowness (b\*) values of breast, thigh, and abdominal fat with a Chromameter calibrated with a whiteboard before measured. Each sample was measured three times at different sites and then its average value was taken.

### Statistical analysis

All data were analyzed by one-analysis of variance of SAS 9.1 software (SAS Inst. Inc., Cary, NC, USA) and presented as means and standard error of the mean. The pen was considered as the experimental unit for growth performance, and selected chickens that were slaughtered for sample collection were regarded as the experimental unit for skin color and carcass pigmentation. The significant level of data was de-

clared at p<0.05. If significant effects were found, individual means were compared using Duncan's multiple comparison tests.

## **RESULTS**

#### **Growth performance**

As shown in Table 2, the BW on day 28 of the broilers supplemented MDP and LTN+MDP diets were not significantly different from the CON group (p>0.05), however, dietary supplementation LTN significantly decreased the BW compared to the CON group (p<0.05). Moreover, a similar consequence was shown in ADG. There was no significant difference in ADFI and F/G of treatment groups compared with the CON group (p>0.05).

### Skin color

No significant difference in the yellow scores of beaks and shanks was observed between all treatments and the CON group in the first week (p>0.05, Fugure 1). The analogous results in L\*, a\*, and b\* values of subwing skin are presented in Figure 2 (p>0.05). After two weeks, the notable difference is shown in Figure 1 that the yellow scores of shank supplementing MDP and LTN+MDP diets were significantly enhanced over the CON group (p<0.05). In addition, the b\* values of subwing skin in all treatments were significantly increased (p<0.05), and a better effect was exhibited in LTN and LTN +MDP groups than the MDP group in Figure 2 (p<0.05). As shown in Figure 1, in the third week, the LTN group significantly promoted the yellow scores of beaks, and the LTN+MDP group significantly enhanced the yellow scores of shanks (p<0.05). Moreover, the b\* values of subwing skin in all treatments were significantly increased over the CON group in Figure 2 (p<0.05). Figure 1 and Figure 2 also demonstrates that after four weeks, supplementing with LTN+ MDP significantly increased the yellow scores of beaks, and the yellow scores of shank of all treatment groups (p<0.05). Furthermore, compared with the CON group, all treatment groups dramatically increased the b\* values of subwing skin

Table 2. Effects of different types of xanthophyll extracted from marigold on the growth performance

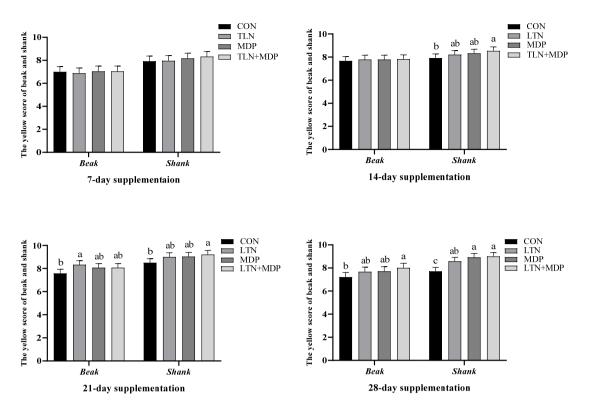
Item	Treatments <sup>1)</sup>				0514	
	CON	LTN	MDP	LTN+MDP	SEM	p-value
D 1 BW (g)	1,261	1,265	1,275	1,281	21	0.856
D 28 BW (g)	1,897 <sup>ab</sup>	1,809°	1,884 <sup>ab</sup>	1,922ª	25	0.025
ADG (g/d)	22.7ª	19.4 <sup>b</sup>	21.8ª	22.9ª	1.1	0.008
ADFI (g/d)	103.4	98.7	104.9	108.8	5.75	0.126
F/G	4.56	5.09	4.81	4.75	0.42	0.652

n = 8 (1 bird per cage)

SEM, standard error of the mean; BW, body weight; ADG, average daily gain; ADFI, average daily feed intake; F/G, feed to gain ratio.

<sup>1)</sup> CON, fed with basal diet; LTN, supplemented with lutein; MDP, supplemented with monohydroxyl pigment including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin; LTN+MDP, supplemented with lutein and monohydroxyl pigment in proportion to 1:1.

 $^{\rm a-c}$  Means with different small letters within a row differ (p<0.05).



**Figure 1.** Effects of different types of xanthophyll extracted from marigold on the yellow scores of beaks and shanks in yellow-feathered chickens. CON, fed with basal diet; LTN, supplemented with lutein extracted from marigold; MDP, supplemented with monohydroxyl pigment extracted from marigold including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin; LTN+MDP, supplemented with lutein and monohydroxyl pigment in proportion to 1:1. The statistics of the Y axis were measured, and the X axis presented the position of survey based on Kemin colour fan. Scores are mean and pooled standard error of the mean, n = 6 (1 bird per cage). <sup>arc</sup> Different letters indicate significant differences between mean values for a given parameter (p<0.05).

(p<0.05).

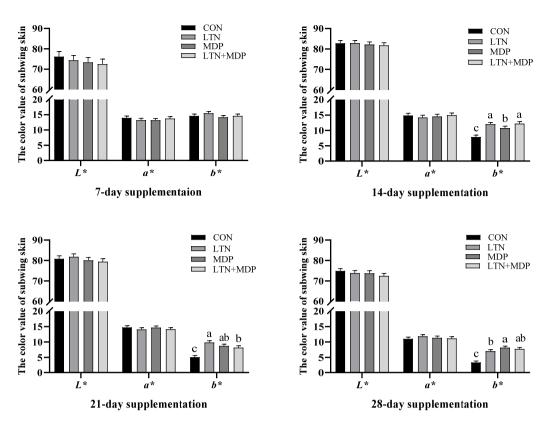
#### **Carcass pigmentation**

As shown in Table 3, all treatment groups of LTN, MDP, and LTN-MDP significantly promoted the b\* values of the breast muscle in comparison to the CON group (p<0.05). The skin results shown in Table 3 indicate that all treatments significantly enhanced the b\* values of the thigh muscle (p<0.05). However, there was no significant difference in the L\* and a\* values of breast and thigh muscle between all treatments and the CON group (p>0.05). Additionally, Table 3 displays that the MDP and LTN+MDP groups dramatically increased the a\* and b\* values of abdominal fat more than the CON group (p<0.05).

## DISCUSSION

Carotenoids have become important pigment additives for improving animal products in poultry production [16,17], from the former yellow corn, dehydrated alfalfa meal, and corn gluten meal to the current common use of plant-derived curcumin, lutein and so on [18,19]. In corn-soybean meal-based diets, yellow corn contains about 17 ppm of needed to meet consumer preferences [20]. Particularly, lutein extracted from marigold is extensively used and there were many monohydroxyl pigments-dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin produced in the metabolic process [12]. A large amount of research has shown that carotenoids have no significant influence on growth performance. The addition of xanthophyll-rich okra meal had no significant difference in daily gain and the feed to gain ratio [2]. And the performance of pullets was not influenced by supplementing 4% marigold [21]. Our results indicated that compared with the CON group, the MDP and LTN+MDP had no significant difference in the BW of day 28 and the ADG, whereas the LTN group dramatically decreased the BW of day 28 and the ADG. Many studies have shown that lutein supplementation has no significant difference in growth performance. However, a few studies have suggested that large amounts of lutein may have negative effects on the liver. Supplementation with 400 mg/kg lutein has been shown to result in a decrease in serum erythrocyte count and hemoglobin content, as well as a decrease the relative organ weight of liver, and the presence of cystic tubular dilatation of kidneys, fatty change

xanthophyll, which is so low that additional colorants are



**Figure 2.** Effects of different types of xanthophyll extracted from marigold on the lightness (L\*), redness (a\*), and yellowness (b\*) values of subwing skin in yellow-feathered chickens. CON, fed with basal diet; LTN, supplemented with lutein extracted from marigold; MDP, supplemented with monohydroxyl pigment extracted from marigold including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin; LTN+MDP, supplemented with lutein and monohydroxyl pigment in proportion to 1:1. Scores are mean and pooled pooled standard error of the mean, n = 6 (1 bird per cage).

and accessory cortical nodules of the adrenal have been observed [22]. Moreover, studies have shown that aspartate aminotransferase and alanine aminotransferase significantly increased in a dose-dependent manner when lutein was added at 100 to 1,000 mg/kg [23]. Based on this consideration, it is speculated that a high dosage of lutein may affect liver health and lead to decrease the BW of day 28 and ADG. The color of animal products, such as eggs, salmon and

ltem	Treatments <sup>1)</sup>				0514	
	CON	LTN	MDP	LTN+MDP	SEM	p-value
Breast						
L*	74.02	74.70	76.11	75.17	2.22	0.269
a*	10.96	9.91	10.74	10.44	1.03	0.335
b*	9.28 <sup>b</sup>	12.48ª	11.74ª	13.74ª	1.21	< 0.001
Thigh						
L*	83.06	79.52	80.81	79.39	2.46	0.762
a*	14.65	15.26	15.20	15.65	0.88	0.726
b*	9.26 <sup>b</sup>	13.96ª	13.76ª	14.30ª	1.12	< 0.001
Abdominal fat						
L*	96.11	96.57	97.22	96.67	0.85	0.885
a*	6.48 <sup>b</sup>	7.72 <sup>ab</sup>	8.80ª	8.69ª	0.62	< 0.001
b*	26.91 <sup>b</sup>	31.04 <sup>ab</sup>	32.41ª	35.46ª	2.31	< 0.001

**Table 3.** Effect of different types of xanthophyll extracted from marigold on the lightness (L\*), redness (a\*), and yellowness (b\*) values of breast, thigh muscle, and abdominal fat in yellow-feathered chickens

Scores are mean and pooled standard error of the mean, n = 6 (1 bird per cage).

<sup>1)</sup> CON, fed with basal diet; LTN, supplemented with lutein extracted from marigold; MDP, supplemented with monohydroxyl pigment extracted from marigold including dehydrated lutein, β-cryptoxanthin, and α-cryptoxanthin; LTN+MDP, supplemented with lutein and monohydroxyl pigment in proportion to 1:1. <sup>ab</sup> Different letters indicate significant differences between mean values for a given parameter (p<0.05).

## /ID/

chicken, showed a close relationship with the preference of customers [24-26]. Genetic background, body state, and dietary pigment source and concentration affected the color of poultry to some extent, particularly, dietary pigment source and concentration [24]. It was reported that the gene associated with canthaxanthin and lutein was regarded as a decisive regulatory mechanism of pigmentation in broilers [27]. In this study, compared with the CON group, different lutein enhanced the b\* values of subwing skin during the whole period, the LTN+MDP group increased the yellow scores of shank in the second week, the LTN treatment improved the vellow scores of beaks and the LTN+MDP treatment enhanced the yellow scores of shanks in the third week, and the LTN+ MDP treatment promoted the yellow scores of beaks and all groups boosted the yellow of shanks in the fourth week. Similar to our outcomes, in comparison to a basal diet, a diet supplemented with carotenoid notably boosted the yellow of the skin of broilers [28]. Consistent with our findings, it was demonstrated that supplementing lutein exhibited positive pigmentation efficiency of the skin yellow scores in the broiler [29]. Interestingly, the LTN+MDP group, mixing the lutein and monohydroxyl pigment including dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin displayed a more positive effect on skin pigmentation. The possible reason for this result is that dehydratied lutein, β-cryptoxanthin, and α-cryptoxanthin, which are the metabolizing and transiting products of lutein, are better absorbed and deposited in tissues than lutein.

Various factors determine meat quality such as meat color, drip loss, cooking loss, and shear force, yet color is crucial to the appearance of meat and consumer preference [30,31]. Carotenoids have been used to improve the meat color of poultry, due to the characteristic of amassing in the body [32]. The results of this research showed that dietary supplements different types of xanthophyll enhanced the b\* values of breast and thigh muscle, and the MDP and LTN+MDP groups increased the a\* and b\* values of abdominal fat in comparison to the CON group. Consistent with our results, the addition of xanthophyll-rich okea meal to the diet increased the b\* values of breast and abdomen fat [2]. Many works have indicated xanthophyll extracted from marigold was more efficient deposition in the muscle and skin than synthetic pigment [28,29,33]. Moreover, the dietary supplement of marigold extract enhanced the yellowness values of carcass and meat quality [34]. However, the mechanism is not clear that increasing the a\* value of abdominal fat caused by supplementing with monohydroxyl pigment or lutein and monohydroxyl pigment in proportion to 1:1.

## CONCLUSION

In conclusion, different types of xanthophyll extracted from

marigold including lutein, monohydroxyl pigment (dehydrated lutein,  $\beta$ -cryptoxanthin, and  $\alpha$ -cryptoxanthin), and the mixture of lutein and monohydroxyl pigment can enhance the yellow scores of beaks, shanks and subwing skin, and increase the b\* values of beast, thigh and abdomen fat to improve meat quality. Especially, the mixture of lutein and monohydroxyl pigment is more efficient on skin color.

## **CONFLICT OF INTEREST**

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

## FUNDING

This study was supported by the Dawn Project in Special Project of Knowledge Innovation of Wuhan (No. 202202 0801020396) and the National Natural Science Foundation of China (No. 32272918).

## REFERENCES

- Ouart MD, Bell DE, Janky DM, Dukes MG, Marion JE. Influence of source and physical form of xanthophyll pigment on broiler pigmentation and performance. Poult Sci 1988; 67:544-8. https://doi.org/10.3382/ps.0670544
- Liu GD, Hou GY, Wang DJ, et al. Skin pigmentation evaluation in broilers fed different levels of natural okra and synthetic pigments. J Appl Poult Res 2008;17:498-504. https://doi.org/ 10.3382/japr.2008-00058
- 3. Breithaupt DE. Modern application of xanthophylls in animal feeding—a review. Trends Food Sci Technol 2007;18:501-6. https://doi.org/10.1016/j.tifs.2007.04.009
- Castañeda MP, Hirschler EM, Sams AR. Skin pigmentation evaluation in broilers fed natural and synthetic pigments. Poult Sci 2005;84:143-7. https://doi.org/10.1093/ps/84.1.143
- 5. Pérez-Vendrell AM, Hernández JM, Llauradó L, Schierle J, Brufau J. Influence of source and ratio of xanthophyll pigments on broiler chicken pigmentation and performance. Poult Sci 2001;80:320-6. https://doi.org/10.1093/ps/80.3.320
- Zhu NH, Zhang RJ, Wu H, Zhang B. Effects of lactobacillus cultures on growth performance, xanthophyll deposition, and color of the meat and skin of broilers. J Appl Poult Res 2009;18:570-8. https://doi.org/10.3382/japr.2009-00012
- Karadas F, Grammenidis E, Surai PF, Acamovic T, Sparks NHC. Effects of carotenoids from lucerne, marigold and tomato on egg yolk pigmentation and carotenoid composition. Br Poult Sci 2006;47:561-6. https://doi.org/10.1080/ 00071660600962976
- 8. Rajput N, Naeem M, Ali S, Zhang JF, Zhang L, Wang T. The effect of dietary supplementation with the natural carotenoids

curcumin and lutein on broiler pigmentation and immunity. Poult Sci 2013;92:1177-85. https://doi.org/10.3382/ps.2012-02853

- 9. Quackenbush FW, Kvakovszky S, Hoover T, Rogler JC. Deposition of individual carotenoids in avian skin. J AOAC Int 1965;48;1241-4. https://doi.org/10.1093/jaoac/48.6.1241
- Jonsson G, Mcnab JM. Grass meal as an ingredient in diets for broiler chickens. Br Poult Sci 1983;24:361-9. https://doi. org/10.1080/00071668308416750
- Maoka T. Recent progress in structural studies of carotenoids in animals and plants. Arch Biochem Biophys 2009;483:191-5. https://doi.org/10.1016/j.abb.2008.10.019
- Khachik F. Transformation of naturally occurring (3R, 3'R, 6'R)-lutein and its fatty acid esters to (3R)-β-cryptoxanthin and (3R, 6'R)-α-cryptoxanthin. Arch Biochem Biophys 2018; 653:107-12. https://doi.org/10.1016/j.abb.2018.06.012
- Lokaewmanee K, Yamauchi K, Komori T, Saito K. Enhancement of yolk color in raw and boiled egg yolk with lutein from marigold flower meal and marigold flower extract. J Poult Sci 2011;48:25-32. https://doi.org/10.2141/jpsa.010059
- Chowdhury SD, Hassin BM, Das SC, Rashid MH, Ferdaus AJM. Evaluation of marigold flower and orange skin as sources of xanthophyll pigment for the improvement of egg yolk color. J Poult Sci 2008;45:265-72. https://doi.org/10.2141/ jpsa.45.265
- 15. Ministry of Agriculture of the People's Republic of China. Nutrient requirements of Chinese color-feather chicken. Feeding standard of chicken. 2nd edn. Beijing, China: China Agricultural Press; 2004. (In Chinese)
- Surai PF, Speake BK. Distribution of carotenoids from the yolk to the tissues of the chick embryo. J Nutr Biochem 1998; 9:645-51. https://doi.org/10.1016/S0955-2863(98)00068-0
- Sandeski LM, Ponsano EHG, Neto MG. Optimizing xanthophyll concentrations in diets to obtain well-pigmented yolks. J Appl Poult Res 2014;23:409-17. https://doi.org/10.3382/ japr.2013-00912
- Ortiz D, Lawson T, Jarrett R, et al. Biofortified orange corn increases xanthophyll density and yolk pigmentation in egg yolks from laying hens. Poult Sci 2021;100:101117. https:// doi.org/10.1016/j.psj.2021.101117
- Rajput N, Ali S, Naeem M, Khan MA, Wang T. The effect of dietary supplementation with the natural carotenoids curcumin and lutein on pigmentation, oxidative stability and quality of meat from broiler chickens affected by a coccidiosis challenge. Br Poult Sci 2014;55:501-9. https:// doi.org/10.1080/00071668.2014.925537
- NRC, National Research Council (1994). Nutrient requirements of poultry, 9th revised ed. Washington, DC, USA: National Academy Press; 1994.
- Hasin BM, Ferdaus AJM, Islam MA, Uddin MJ, Islam MS. Marigold and orange skin as egg yolk color promoting agents. Int J Poult Sci 2006;5:979-87. https://doi.org/10.3923/ijps.

2006.979.987

Ravikrishnan R, Rusia S, Ilamurugan G, et al. Safety assessment of lutein and zeaxanthin (Lutemax 2020): subchronic toxicity and mutagenicity studies. Food Chem Toxicol 2011; 49:2841-8. https://doi.org/10.1016/j.fct.2011.08.011

/13/

- 23. Nidhi B, Baskaran V. Acute and subacute toxicity assessment of lutein in lutein-deficient mice. J Food Sci 2013;78:T1636-42. https://doi.org/10.1111/1750-3841.12256
- 24. Hudon J. Biotechnological applications of research on animal pigmentation. Biotechnol Adv 1994;12:49-69. https://doi. org/10.1016/0734-9750(94)90290-9
- 25. Maga JA. Pink discoloration in cooked white meat. Food Rev Int 1994;10:273-86. https://doi.org/10.1080/8755912 9409541003
- Holownia K, Chinnan MS, Reynolds AE, Koehler PE. Evaluation of induced color changes in chicken breast meat during simulation of pink color defect. Poult Sci 2003;82:1049-59. https://doi.org/10.1093/ps/82.6.1049
- Tarique TM, Yang S, Mohsina Z, et al. Identification of genes involved in regulatory mechanism of pigments in broiler chickens. Genet Mol Res 2014;13:7201-16. https://doi.org/ 10.4238/2014.September.5.6
- Sirri F, Petracci M, Bianchi M, Meluzzi A. Survey of skin pigmentation of yellow-skinned broiler chickens. Poult Sci 2010;89:1556-61. https://doi.org/10.3382/ps.2009-00623
- 29. Rajput N, Naeem M, Ali S, Rui Y, Tian W. Effect of dietary supplementation of marigold pigment on immunity, skin and meat color, and growth performance of broiler chickens. Braz J Poult Sci 2012;14:291-5. https://doi.org/10.1590/S1516-635X2012000400009
- 30. Wei Y, Gao QY, Jing XQ, et al. Effect of cardamine violifolia on plasma biochemical parameters, anti-oxidative capacity, intestinal morphology, and meat quality of broilers challenged with lipopolysaccharide. Animals 2022;12:2497. https://doi. org/10.3390/ani12192497
- O'Sullivan MG, Byrne DV, Martens M. Evaluation of pork colour sensory colour assessment using trained and untrained sensory panellists. Meat Sci 2003;63:119-29. https://doi.org/ 10.1016/S0309-1740(02)00065-7
- Hencken H. Chemical and physiological behavior of feed carotenoids and their effects on pigmentation. Poult Sci 1992; 71:711-7. https://doi.org/10.3382/ps.0710711
- 33. Ponsano EHG, Pinto MF, Garcia-Neto M, Lacava PM. Performance and color of broilers fed diets containing Rhodocyclus gelatinosus biomass. Braz J Poult Sci 2004;6:237-42. https://doi.org/10.1590/S1516-635X2004000400008
- 34. Wang SH, Zhang L, Li JL, Cong JH, Gao F, Zhou GH. Effects of dietary marigold extract supplementation on growth performance, pigmentation, antioxidant capacity and meat quality in broiler chickens. Asian-Australas J Anim Sci 2017; 30:71-7. https://doi.org/10.5713/ajas.16.0075