

pISSN 1598-298X • eISSN 2384-0749 J Vet Clin 2023;40:44-49 https://doi.org/10.17555/jvc.2023.40.1.44

Check for updates

The Effect of Water-Filtered Infrared-A (wIRA) on Body Core and Body Surface Temperatures in **Anesthetized Rabbits Maintained with Isoflurane**

Geonho Choi Abstract The purpose of this study was to evaluate body temperature changes **Dongseok Kim** in rabbits anesthetized using water-filtered infrared-A (wIRA). Ten rabbits were **Eungmo Tae** used for this study. For the experimental group (wIRA group; wG, n = 5), the Ilgwon Jung experimental equipment was used and irradiated using wIRA. The control group Sang-Kwon Lee (CG, n = 5) did not have any warming device. There were no significant differ-Won-Jae Lee ences in heart rate, respiration rate, and end tidal CO₂ (EtCO₂) between wG and Sung-Ho Yun CG. After 80 min, the core body temperature of wG rabbits was significantly Young-Sam Kwon higher than that of CG rabbits. The surface body temperature was significantly Min Jang* higher while receiving wIRA support at all time points after 5 min. In conclusion, in rabbits under inhalation anesthesia, the surface body temperature was better Department of Veterinary Surgery, maintained than the core body temperature when using wIRA. College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea **Key words** wIRA, rabbit, body temperature, surface, core. *Correspondence: jangmin@knu.ac.kr ORCID Geonho Choi: https://orcid.org/0000-0001-9127-8064 Dongseok Kim: https://orcid.org/0000-0002-7353-9731 Eungmo Tae: https://orcid.org/0000-0003-4381-7806 Ilgwon Jung: https://orcid.org/0000-0002-7958-0604 Sang-Kwon Lee: https://orcid.org/0000-0002-3097-0345 Won-Jae Lee: https://orcid.org/0000-0003-1462-7798 Sung-Ho Yun: https://orcid.org/0000-0002-9027-3859 Young-Sam Kwon: https://orcid.org/0000-0002-6489-0327 Min Jang: https://orcid.org/0000-0002-2188-1906 Received September 30, 2022 / Revised January 9, 2023 / Accepted January 16, 2023 Copyright © The Korean Society of Veterinary Clinics

This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/ \odot 0 \odot by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited

Introduction

In dogs and cats, hypothermia occurs when the body temperature is lower than 37°C (19); this is a major perioperative complication in veterinary medicine (1). In rabbits, a body temperature lower than 38.5°C indicates hypothermia (29). Hypothermia often occurs during anesthesia because of decreased metabolic rate, drug-related vasodilation, and muscle inactivity (27). Intraoperatively, hypothermia may occur during thoracotomy or lavage using unheated fluid during laparotomy (2). The side effects of hypothermia during anesthesia may include decreased cardiovascular function, decreased metabolism and excretion of anesthetic drugs, and increased vulnerability to infections (10). Moreover, recovery may be delayed and shivering may occur after awakening (26).

Methods for raising body temperature are divided into two categories, i.e., warming of the body surface and warming of the core parts (1). The traditional method to prevent hypothermia is to raise the body surface temperature, i.e., active surface rewarming (1,22), using circulating warm water blankets, warm packs, and forced air-warming blankets (1,3,22). This method can effectively raise the surface body temperature but does not adequately increase the core temperature. In addition, water blankets and warm packs can cause low-temperature burns (4). The process of raising the core body temperature is called active core rewarming and involves warmed and humidified inhaled air and warm intravenous fluid therapy during surgery (1,2).

Water-filtered infrared-A (wIRA) has the characteristic of raising whole-body temperature without raising surface body temperature, and in human medicine, wIRA is also used for wound healing (7). Certain wavelengths (780-1400 nm) pass through water and reach the skin, and water absorbs or reduces unwanted wavelengths in the infrared region (12). The radiation through the water filter has a similar effect as the sun's heat radiation; therefore, the body surface is not burned, and high temperatures (38-42°C) can be delivered to the deep tissues (20). wIRA has two effects, thermal and non-thermal. In the thermal effect, heat energy is transferred to the tissue to increase body temperature, blood flow to the surrounding tissues, and metabolism (13). The non-thermal effect stimulates wound healing and affects cytochrome C oxidase and skin growth factors to activate cellular responses (12). Owing to these properties, wIRA is used to treat burns, skin cancer, and even breast cancer (21).

To our knowledge, there are no reports of clinical studies on anesthesia on changes in body temperature using wIRA. The objective of this study was to evaluate the thermal effect of core body temperature compared with surface body temperature during anesthesia using wIRA. The purpose of this study was to investigate the effect of wIRA on hypothermia induced by inhalation anesthesia. We hypothesized that the surface body temperature and core body temperature will be higher in the wIRA group compared to the control group.

Materials and Methods

Experimental animals

Ten New Zealand White male rabbits (weight, 2.5-3.0 kg; mean \pm SD age, 1.2 \pm 0.2 years, n = 10) were used after approval by the Institutional Animal Care and Use Committee of Kyungpook National University (approval number: KNU2022-0006). The following conditions were used: room temperature, 20-25°C; air humidity, 50-60%; light/dark cycle, 12 h; stabilization period, seven days. Water and food were supplied during the experimental periods.

Preparation of irradiation with wIRA

All rabbits were fasted a day before anesthesia. For the experimental group (wIRA group; wG, n = 5), an experimental equipment (RBp 3000; Ray-bio, Korea) with a wavelength of 400-1,100 nm was used and irradiation was done from 30 cm distance. The control group (CG, n = 5) was not exposed to any warming device. Alfaxalone (3 mg kg⁻¹, IV Alfaxan[®]; Jurox, Australia) was administered after a 24 gauge catheter was inserted into the marginal vein of each rabbit's ear, for induction. After induction, V-gel (size R3, V-gel[®]; Docsinnovent, UK) was intubated, and respiratory anesthesia was induced. Then, after intubation, the rabbit was placed in the ventral-dorsal position (VD) using a W-plate, and an eye lubricant was applied. Isoflurane concentration was maintained at 2%, and anesthesia was induced for 2 h. N/S solution was administered intravenously at a rate of 5 mL kg⁻¹ h⁻¹. After anesthesia, the abdominal hair was clipped from the sternum to the umbilicus using a clipper. The room temperature was maintained at 25°C.

Physical evaluation and data collection

For physical evaluation, core body temperature, surface body temperature, heart rate (HR), respiration rate (RR), end tidal CO₂ (EtCO₂), and hemoglobin oxygen saturation (SpO₂) were measured. The core temperature was measured using an esophageal thermometer (M1024251, GE Healthcare, US) and the surface temperature was measured using a skin infrared thermometer (BNT 400; Braun, Germany). The remaining indicators were measured by monitoring the patients (CARESCAPETM B650; GE Healthcare, US). Data were collected at five-minute (min) intervals for 20 min, and continued at ten-minute intervals for 2 h.

Statistical analysis

After normality testing using the Shapiro-Wilk test, data for the control and wIRA groups were compared using a t-test. Core and superficial temperatures were compared using two-way ANOVA. Statistical analyses were performed using SPSS 25.0 (IBM SPSS Inc., Armonk, New York, US). p < 0.05 was considered significant.

Results

There were no significant differences in HR, RR, and $EtCO_2$ between animals of wG and CG (Table 1). Core body temperature was not significantly different between animals of wG and CG (Fig. 1) until 120 min (p = 0.05).

There was no significant difference in the surface body temperature at base (0 min) (Fig. 2). However, after 5 min, the surface body temperature was significantly higher while receiving wIRA support at all times (5, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120 min; p < 0.05).

Discussion

The main finding of this study was that maintaining the surface body temperature was a much better strategy than maintaining the core body temperature. In other study showed that wIRA raises the core body temperature without affecting the surface body temperature (12); however, the opposite was found in this study. According to recent research results, it has been reported that changes in skin and subcutis temperature are affected by irradiance, exposure time, and tissue depth, and temperature maxima were between 4 and 7 mm (30). According to another study, superficial temperature was significantly increased from 39°C to 43°C when wIRA was used, and core body temperature increased with time (21). An interesting fact is that the fat composition of pigs (saturated fats: 69%, unsaturated fats: 31%) and rabbits (saturated fats: 30%, unsaturated fats: 70%) differs (7). Hence, using wIRA, the core body temperature would increase in pigs owing to their lower heat distribution and heat loss (21,30). Although there have been no studies on body temperature maintenance in rabbits, differences in fat composition between the two animals may have affected the results; therefore, further studies are required.

There were side effects such as burning sensation, hyperpigmentation, and blister formation when applying local hyperthermia (44°C for 30 min) (14). However, no skin side effects were observed when wIRA was applied in another study (8). In addition, wIRA is well accepted by patients without side effects even when used chronically and acutely (33). In addition, wIRA was well accepted by patient with no side effects when used chronically and acutely (33). In this study, skin was irradiated with wIRA for 2 h, and there were no significant side effects.

In this study, anesthesia was induced with alfaxalone and maintained with isoflurane. Alfaxalone is a steroid anesthetic

Table 1. Results of changes in HR, RR, EtCO₂ during 120 min of anesthesia

	5	. 2 3				
Time (min)	Control group (CG)			wIRA group (wG)		
	HR (beats/min)	RR (breaths/min)	EtCO₂ (mmHg)	HR (beats/min)	RR (breaths/min)	EtCO₂ (mmHg)
0 (base)	214.4 ± 54.6	17.8 ± 10.0	42.0 ± 5.6	222.2 ± 7.3	26.6 ± 12.3	42.8 ± 12.2
5	238.4 ± 17.7	27.0 ± 7.1	40.0 ± 5.3	227.0 ± 13.7	27.4 ± 10.3	42.4 ± 10.9
10	230.4 ± 21.3	24.0 ± 9.4	40.8 ± 4.6	221.2 ± 9.5	26.6 ± 11.1	42.6 ± 9.5
15	219.2 ± 22.9	24.0 ± 9.4	41.4 ± 4.6	221.8 ± 6.4	29.0 ± 7.4	43.0 ± 9.5
20	225.0 ± 22.0	28.2 ± 9.2	41.6 ± 4.5	213.6 ± 10.9	29.8 ± 6.2	43.4 ± 9.6
30	218.2 ± 27.9	24.8 ± 5.9	43.2 ± 6.8	216.4 ± 16.3	27.8 ± 8.7	43.0 ± 9.0
40	210.6 ± 29.2	27.4 ± 3.6	44.2 ± 9.7	204.8 ± 15.9	30.6 ± 4.6	43.8 ± 8.5
50	193.4 ± 25.7	24.8 ± 9.0	46.2 ± 11.3	189.0 ± 10.1	29.2 ± 4.6	45.6 ± 8.5
60	154.5 ± 70.6	29.4 ± 3.9	46.0 ± 10.4	192.4 ± 13.5	30.0 ± 3.3	45.0 ± 8.4
70	182.4 ± 24.3	28.8 ± 4.6	45.6 ± 7.9	185.4 ± 17.7	30.2 ± 2.5	43.6 ± 7.6
80	175.4 ± 22.0	29.0 ± 4.8	46.8 ± 8.8	179.8 ± 18.5	28.8 ± 4.3	44.4 ± 8.9
90	166.4 ± 20.4	26.4 ± 4.3	47.8 ± 9.7	173.0 ± 26.1	30.0 ± 4.4	44.0 ± 6.7
100	158.4 ± 24.2	26.0 ± 3.9	50.0 ± 12.1	162.0 ± 25.2	29.0 ± 5.1	45.2 ± 6.7
110	161.2 ± 26.6	22.0 ± 5.2	48.0 ± 16.8	167.2 ± 18.7	29.0 ± 4.7	44.6 ± 7.0
120	160.6 ± 26.1	37.0 ± 15.7	45.6 ± 14.3	170.2 ± 21.4	27.6 ± 3.2	44.6 ± 6.8

Data are presented as mean \pm standard deviation.

HR, heart rate; RR, respiratory rate; EtCO₂, end-tidal CO₂.

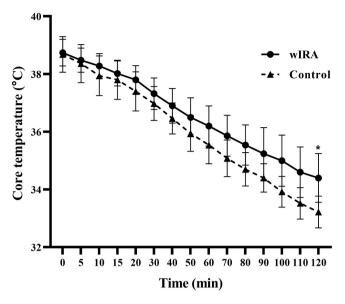


Fig. 1. Change in core temperature in rabbits during anesthesia. Data are presented as mean \pm standard deviation. *p < 0.05.

agent with dose-dependent effects on the cardiovascular and respiratory systems (27). In the cardiovascular system, it decreases blood pressure and increases HR. In the respiratory system, it decreases the respiratory rate, tidal volume, and PaO_2 (18). Isoflurane causes severe respiratory depression and decreases arterial blood pressure, HR, and systemic vascular resistance in a dose-dependent manner (27,34). Inhalation anesthetic agents change body temperature by decreasing thermogenesis or increasing heat loss. In some studies, owing to decreased vascular resistance, the muscle blood flow increased from the deep to the skin, resulting in increased heat loss (23). These results indicate that alfaxalone and isoflurane may have had an effect on body temperature changes. In our study, we investigated whether wIRA was effective at maintaining the surface body temperature following anesthesia-induced hypothermia without any effect on the maintenance of core temperature.

The normal body temperature of rabbits fluctuates between 38.5 and 39.5°C (29). In this study, body temperature decreased significantly with time after anesthesia in rabbits. These results were similar to those obtained in dogs and cats, with hypothermia occurring during surgical anesthesia in 83.6% of the dogs and moderate hypothermia in 66% of the cats (24,25). As a physiological effect of hypothermia, central nervous system depression (reduction of intracranial pressure), cardiovascular abnormalities (decreased cardiac output and blood pressure, bradycardia), delay in metabolic rate, respiratory depression, and immune system depression may occur as complications (5,19). In this study, wIRA was

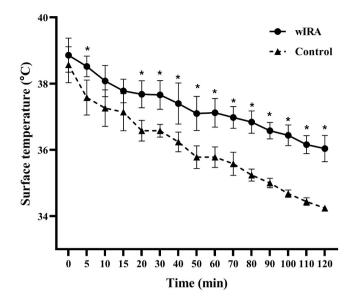


Fig. 2. Change in surface temperature in rabbits during anesthesia. Data are presented as mean \pm standard deviation. *p < 0.05.

used to maintain the core body temperature, but no significant results were obtained. There are several reasons for this. In small animals, the smaller the body weight, the larger the body surface area, and these factors tend to induce hypothermia during anesthesia (28). During general anesthesia, environmental factors also include cold O_2 from ventilation, intravenous fluid supply, ambient temperature, and heat loss through the shaved body area, which may lower the body temperature (4). Due to the above factors, it is assumed that it is difficult to maintain body temperature in rabbits when wIRA is applied under general anesthesia.

The rabbit's normal HR was 198-330 bpm (16), and the vital results showed that it gradually decreased over time and became lower than the normal range after 50 min for both CG and wG. In some studies, mild hypothermia in humans and pigs decreased the HR but increased the overall hemodynamic parameters of myocardial contractility (9,31). In addition, a study in rabbits also reported that hypothermia significantly reduced the HR but resulted in good left ventricular pressure (17). In our study, as the anesthesia time increased, body temperature and HR decreased.

Furthermore, blood pressure was measured using oscillometric techniques, but continuous and accurate results were not obtained. Oscillometric techniques are methods of measuring blood pressure by detecting pulse beats (32). If the pulse rate is measured incorrectly, the blood pressure result is insignificant (6). Further, when the size of the peripheral artery is small, the pulse may not be sufficiently detected (11). In this study, blood pressure using the oscillometric method were measured and excluded because rabbits had a fast HR. small peripheral arteries, and hair in the measurement area, so accurate results could not be obtained. When hypothermia occurs, norepinephrine is attached to the α 1-receptor for compensation. However, if hypothermia is severe or prolonged, this response is reduced, and as a result, the responsiveness to catecholamines and vasoconstriction is decreased. resulting in hypotension (19). Moreover, baroreceptor function decreases in hypothermia, which leads to abnormalities in blood pressure control and HR control in response to volume changes (15). If the cardiovascular system function declines, tissue perfusion will be reduced, which can lead to hypoperfusion or organ dysfunction after anesthesia. In this study, blood pressure measurement was attempted; however, continuous and accurate results were not obtained, and the correlation between hypothermia and the cardiovascular system could not be determined.

This study has several limitations. First, blood pressure was measured using the oscillometric method at the beginning of the study, but blood pressure was discontinued because continuous and accurate results were not obtained. Therefore, a decrease in the core body temperature due to hypoperfusion cannot be ignored. Finally, the number of rabbits (10 rabbits) used in the study was small, which is an obvious statistical limitation.

In conclusion, in rabbits under inhalation anesthesia, although wIRA had a mild effect on maintaining body temperature, the surface body temperature was maintained better than the core body temperature. As these results may differ depending on the effect of anesthetic drugs or inhalation anesthesia, the effectiveness of wIRA can be evaluated later by conducting the same study using conscious rabbits.

Acknowledgements

This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (Ministry of Science and ICT) (No. 2021R1G1A1013034).

Conflicts of Interest

The authors have no conflicting interests.

References

 Aarnes TK, Bednarski RM, Lerche P, Hubbell JA. Effect of pre-warming on perioperative hypothermia and anesthetic recovery in small breed dogs undergoing ovariohysterectomy. Can Vet J 2017; 58: 175-179.

- Armstrong SR, Roberts BK, Aronsohn M. Perioperative hypothermia. J Vet Emerg Crit Care 2005; 15: 32-37.
- 3. Cabell LW, Perkowski SZ, Gregor T, Smith GK. The effects of active peripheral skin warming on perioperative hypothermia in dogs. Vet Surg 1997; 26: 79-85.
- 4. Clarke KW, Trim CM, Hall LW. Veterinary anaesthesia. 11th ed. Edinburgh: Elsevier Health Sciences. 2013: 56.
- Congdon JM. Cardiovascular disease. In: Johnson RA, Snyder LBC, Schroeder CA, editors. Canine and feline anesthesia and co-existing disease. Hoboken: John Wiley & Sons. 2022: 1-85.
- Cooper E. Hypotension. In: Silverstein DC, Hopper K, editors. Small animal critical care medicine. 2nd ed. St. Louis: Elsevier Health Sciences. 2015: 46-50.
- Dean HK, Hilditch TP. The body fats of the pig: the influence of body temperature on the composition of depot fats. Biochem J 1933; 27: 1950-1956.
- Fuchs SM, Fluhr JW, Bankova L, Tittelbach J, Hoffmann G, Elsner P. Photodynamic therapy (PDT) and waterfiltered infrared A (wIRA) in patients with recalcitrant common hand and foot warts. Ger Med Sci 2004; 2: 1-16.
- Götberg M, van der Pals J, Olivecrona GK, Götberg M, Koul S, Erlinge D. Mild hypothermia reduces acute mortality and improves hemodynamic outcome in a cardiogenic shock pig model. Resuscitation 2010; 81: 1190-1196.
- Heier T, Caldwell JE, Sessler DI, Miller RD. Mild intraoperative hypothermia increases duration of action and spontaneous recovery of vecuronium blockade during nitrous oxide-isoflurane anesthesia in humans. Anesthesiology 1991; 74: 815-819.
- 11. Henik RA, Dolson MK, Wenholz LJ. How to obtain a blood pressure measurement. Clin Tech Small Anim Pract 2005; 20: 144-150.
- 12. Hoffmann G. Principles and working mechanisms of water-filtered infrared-A (wIRA) in relation to wound healing. GMS Krankenhhyg Interdiszip 2007; (2): 1-16.
- 13. Hoffmann G. Water-filtered infrared-A (wIRA) in acute and chronic wounds. GMS Krankenhhyg Interdiszip 2009; 4: Doc12.
- Hu L, Qi R, Hong Y, Huo W, Chen HD, Gao XH. One stone, two birds: managing multiple common warts on hands and face by local hyperthermia. Dermatol Ther 2015; 28: 32-35.
- 15. Kaul SU, Beard DJ, Millar RA. Preganglionic sympathetic activity and baroreceptor responses during hypothermia. Br J Anaesth 1973; 45: 433-439.
- Lord B, Boswood A, Petrie A. Electrocardiography of the normal domestic pet rabbit. Vet Rec 2010; 167: 961-965.
- Mattheussen M, Mubagwa K, Van Aken H, Wusten R, Boutros A, Flameng W. Interaction of heart rate and hypothermia on global myocardial contraction of the isolated rabbit heart. Anesth Analg 1996; 82: 975-981.
- 18. Muir W, Lerche P, Wiese A, Nelson L, Pasloske K, Whittem T. Car-

diorespiratory and anesthetic effects of clinical and supraclinical doses of alfaxalone in dogs. Vet Anaesth Analg 2008; 35: 451-462.

- 19. Oncken AK, Kirby R, Rudloff E. Hypothermia in critically III dogs and cats. Compendium 2001; 23: 506-521.
- Piazena H, Meffert H, Uebelhack R. Spectral remittance and transmittance of visible and infrared- a radiation in human skin- comparison between in vivo measurements and model calculations. Photochem Photobiol 2017; 93: 1449-1461.
- 21. Piazena H, Müller W, Pendl W, von Ah S, Cap VH, Hug PJ, et al. Thermal field formation during wIRA-hyperthermia: temperature measurements in skin and subcutis of piglets as a basis for thermotherapy of superficial tumors and local skin infections caused by thermosensitive microbial pathogens. Int J Hyperthermia 2019; 36: 938-952.
- 22. Potter J, Murrell J, MacFarlane P. Comparison of two passive warming devices for prevention of perioperative hypothermia in dogs. J Small Anim Pract 2015; 56: 560-565.
- 23. Ramachandra V, Moore C, Kaur N, Carli F. Effect of halothane, enflurane and isoflurane on body temperature during and after surgery. Br J Anaesth 1989; 62: 409-414.
- 24. Redondo JI, Suesta P, Gil L, Soler G, Serra I, Soler C. Retrospective study of the prevalence of postanaesthetic hypothermia in cats. Vet Rec 2012; 170: 206.
- 25. Redondo JI, Suesta P, Serra I, Soler C, Soler G, Gil L, et al. Retrospective study of the prevalence of postanaesthetic hypothermia in dogs. Vet Rec 2012; 171: 374.
- Reynolds L, Beckmann J, Kurz A. Perioperative complications of hypothermia. Best Pract Res Clin Anaesthesiol 2008; 22: 645-657.

- Schauvliege S. Patient monitoring and monitoring equipment. In: Duke-Novakovski T, de Vries M, Seymour C, editors. BSAVA manual of canine and feline anaesthesia and analgesia. 3rd ed. Gloucester: British Small Animal Veterinary Association. 2016: 77-96.
- Stepaniuk K, Brock N. Hypothermia and thermoregulation during anesthesia for the dental and oral surgery patient. J Vet Dent 2008; 25: 279-283. Erratum in: J Vet Dent 2009; 26: 8.
- Suckow MA, Brammer DW, Rush HG, Chrisp CE. Biology and diseases of rabbits. In: Fox JG, Anderson LC, Loew FM, Quimby FW, editors. Laboratory animal medicine. 2nd ed. San Diego: Academic Press. 2002: 329-364.
- Wehner H, von Ardenne A, Kaltofen S. Whole-body hyperthermia with water-filtered infrared radiation: technical-physical aspects and clinical experiences. Int J Hyperthermia 2001; 17: 19-30.
- Weisser J, Martin J, Bisping E, Maier LS, Beyersdorf F, Hasenfuss G, et al. Influence of mild hypothermia on myocardial contractility and circulatory function. Basic Res Cardiol 2001; 96: 198-205.
- 32. Williamson JA, Leone S. Noninvasive arterial blood pressure monitoring. In: Creedon JMB, Davis H, editors. Advanced monitoring and procedures for small animal emergency and critical care. Hoboken: John Wiley & Sons. 2012: 134-144.
- Winkel R, Hoffmann G, Hoffmann R. [Water-filtered infrared-A (wIRA) promotes wound healing]. Chirurg 2014; 85: 980-992. German.
- 34. Yang CF, Chen MYC, Chen TI, Cheng CF. Dose-dependent effects of isoflurane on cardiovascular function in rats. Tzu Chi Med J 2014; 26: 119-122.