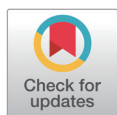


Evaluation of forage production, feed value, and ensilability of proso millet (*Panicum miliaceum* L.)

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Abstract

Whole-plant corn (*Zea mays* L.) and sorghum-sudangrass hybrid [*Sorghum bicolor* (L.) Moench] are major summer crops that can be fed as direct-cut or silage. Proso millet is a short-season growing crop with distinct agronomic characteristics that can be productive in marginal lands. However, information is limited about the potential production, feed value, and ensilability of proso millet forage. We evaluated proso millet as a silage crop in comparison with conventional silage crops. Proso millet was sown on June 8 and harvested on September 5 at soft-dough stage. Corn and sorghum-sudangrass hybrid were planted on May 10 and harvested on September 10 at the half milk-line and soft-dough stages, respectively. The fermentation was evaluated at 1, 2, 3, 5, 10, 15, 20, 30, and 45 days after ensiling. Although forage yield of proso millet was lower than corn and sorghum-sudangrass hybrid, its relative feed value was greater than sorghum-sudangrass hybrid. Concentrations of dry matter (DM), crude protein, and water-soluble carbohydrate decreased commonly in the ensiling forage crops. The DM loss was greater in proso millet than those in corn and sorghum-sudangrass hybrid. The *in vitro* dry matter digestibility declined in the forage crops as fermentation progressed. In the early stages of fermentation, pH dropped rapidly, which was stabilized in the later stages. Compared to corn and sorghum-sudangrass hybrid, the concentration of ammonia-nitrogen was greater in proso millet. The count of lactic acid bacteria reached the maximum level on day 10, with the values of 6.96, 7.77, and 6.95 Log₁₀ CFU/g fresh weight for proso millet, corn, and sorghum-sudangrass hybrid, respectively. As ensiling progressed, the concentrations of lactic acid and acetic acid of the three crops increased and lactic acid proportion became higher in the order of sorghum-sudangrass hybrid, corn, and proso millet. Overall, the shorter, fast-growing proso millet comparing with corn and sorghum-sudangrass hybrid makes this forage crop an alternative option, particularly in areas where agricultural inputs are limited. However, additional research is needed to evaluate the efficacy of viable strategies such as chemical additives or microbial inoculants to minimize ammonia-nitrogen formation and DM loss during ensiling.

Keywords: Proso millet, Corn, Sorghum-sudangrass hybrid, Silage, Conservation

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Availability of data and material

Upon reasonable request, the datasets of this study can be available from the corresponding author.

Authors' contributions

Conceptualization: Wei SN, Kim JG.
Data curation: Wei SN, Ahmadi F, Kim JG.
Formal analysis: Jeong EC, Li YF, Kim HJ.
Methodology: Wei SN, Jeong EC, Kim HJ.
Software: Wei SN, Li YF.
Validation: Ahmadi F, Kim JG.
Investigation: Li YF, Kim HJ.
Writing - original draft: Wei SN, Kim JG.
Writing - review & editing: Wei SN, Jeong EC, Li YF, Kim HJ, Ahmadi F, Kim JG.

Ethics approval and consent to participate

This article does not require IRB/IACUC approval because there are no human and animal participants.

INTRODUCTION

Korea is a country with scarce agricultural resources. About two thirds of its land area is mountains and hills. The cultivated area only accounts for 22% of the total land area. It has one of the lowest per capita cultivated land areas in the world. The livestock industry accounts for almost 40% of total agricultural production in Korea [1]. With the development of the livestock industry, the forage industry has attracted increasing attention. The forage industry is the basis for the survival and development of the livestock industry. However, Korea's current self-produced feed resources are relatively limited, and some feeds must still be imported from overseas. As the most basic production source of animal products, problems with the feed supply will affect the sustainable development of the whole livestock industry. To stabilize the livestock industry and agricultural production, the production of high-quality forage would reduce feed costs and have an import substitution effect.

Corn and sorghum-sudangrass hybrids are the two most common forage crops that are used mainly as summer-season forages in dairy and beef rations. They have low production costs, high yield, and a relatively high nutritional value. Proso millet (*Panicum miliaceum* L.) is a short growing, summer season crop (60 to 100 days) with unique agronomic properties such as high tolerance to heat and drought conditions, and is cultivated in abundance in Asian and African countries [2–4]. Proso millet crop has the potential to remain productive in areas with marginal lands and limited agricultural inputs, where cultivation of major crops such as corn is restricted [5–7]. Proso millet could be a viable alternative to main summer forages in areas where cultivation of corn or sorghum-sudangrass is restricted due to a longer growing season or poor agricultural conditions [8].

Ensiling has long been recognized as a simple and effective method of preserving moist forage, ensuring a continuous supply of forage to animals [9,10]. To our knowledge, few studies have investigated the fermentation dynamics of proso millet forage. The purpose of this research was to provide basic information about the ensiling feasibility of forage from proso millet in comparison to commonly cultivated summer crops (whole-plant corn and sorghum-sudangrass hybrid).

MATERIALS AND METHODS

Crop establishment and management

Establishment of experimental plots was made at the experimental site of Seoul National University, Pyeongchang Campus (located at 37° 32' 40" N, 128° 26' 33" E, average altitude is about 550 m above sea level) during the summer season of 2019. A detailed description of meteorological data including temperature and precipitation throughout the growing season (May to September, 2019) is illustrated in Fig. 1. During the growing season, temperature ranged from 15.9 °C to 26.8 °C (average = 21.5 °C). Soil analysis on the 0–15-cm soil depth of the experimental site showed that it was slightly acidic (pH 6.55; soil:water suspension = 1:5), with 14.1% organic matter, 0.12% total nitrogen, and a cation exchange capacity of 16.5 cmol(+)/kg. Concentration of exchangeable cations including Ca, K, Mg, and Na averaged 1.75, 4.01, 0.92, and 0.10 mg/kg, respectively. For the three crops, nitrogen, phosphorus, and potassium fertilizers were applied at a rate of 200, 150, and 150 kg/ha, respectively. After preparation of seedbed, seeds were sown manually and grown on 3 replicate plots/each crop. Each plot was 3 m × 5 m in size. Proso millet (*Panicum miliaceum* L. var. Geumsilchal) was planted on June 8 at a seeding rate of 20 kg/ha, and harvested on September 5. Sorghum-sudangrass hybrid (*Sorghum bicolor* L. var. Turbo-gold) was sown at a seeding rate of 40 kg/ha on May 10 and harvested on September 10. Corn (*Zea mays* L. var. Gwangpyeongok) was sown on May 10 at a plant-to-plant distance of 20 cm and an inter-row spacing of 75 cm. Whole-

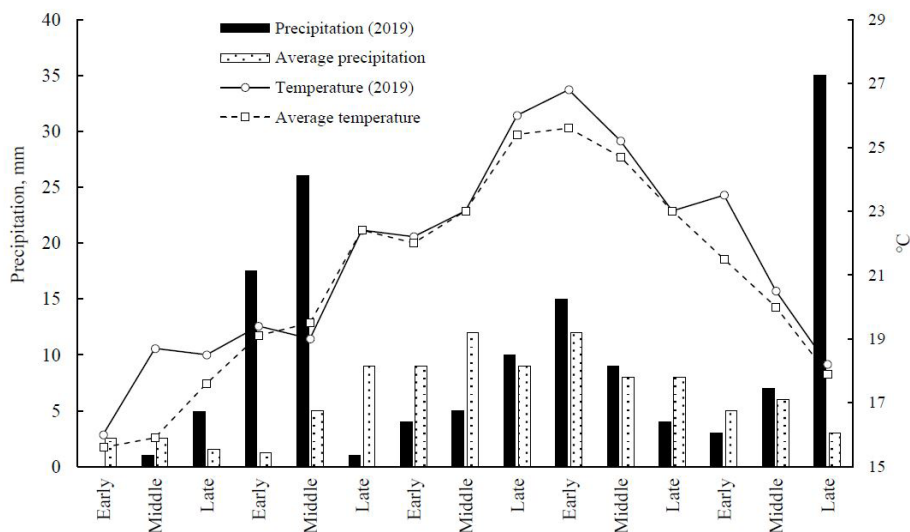


Fig. 1. Temperature and precipitation during the growing season (May to September, 2019) and comparison with the average climatic normal. The data were obtained from the Korean Meteorological Administration.

crop corn was harvested on September 10. Sorghum-sudangrass hybrid and proso millet were harvested when they reached soft-dough stage of the seedhead. Whole-crop corn was harvested at about the half milk-line stage, which is a reliable visionary criterion indicating the optimum time to harvest whole plant for silage making [11]. This was accomplished by splitting the corn ear in the center and visually inspecting the kernel milkline. Whole-crop corn was fractionated into cob (containing kernel and rachis) and stover component that was consisted of the remaining components of the plant after cob removal [12]. These fractions were separately weighed and approximately 1-kg representative subsamples were collected for dry matter (DM) determination. The proportion of these fractions in the whole plant was then calculated. Forage yield was determined by manually harvesting the forage material in the whole plot and calculating the fresh forage yield, which was then converted to units of fresh and DM/hectare.

Silage preparation

At harvest, four whole-crop plants from center rows in each plot were randomly selected and chopped into approximately 2–3 cm long pieces using a chopper (Richi Machinery, Henan, China). The chopped crops were grouped into separate piles per each plot for silage experiment. The representative allotments were also collected for quality assessment of fresh biomass before ensiling. Ensiling was made by packing approximately 600 g chopped material into plastic film bags (28 cm × 36 cm). The bags were vacuum-sealed (Zhejiang Hongzhan Packing Machinery, Wenzhou, China) and stored in a dark and dry condition at room temperature (about 22 °C). Bags were randomly opened on days 1, 2, 3, 5, 10, 15, 20, 30, and 45 of ensiling for quality assessment of silage fermentation. Silos were weighed at designated openings for DM loss determination [13]. Number of replicate silos for each crop at each opening was 3. Therefore, the design arrangement for the three forage types in the silage trial was as follows: 3 forage types × 9 silo openings × 3 replications, resulting in formation of a total of 81 silos. At each silo opening, the ensiled material inside each silo was emptied, mixed thoroughly and divided into 3 representative portions. The first portion was dried (65 °C) to a constant weight and used for the chemical composition analysis. The second portion was stored in a freezer at –80 °C (TSE400D, Thermo Fisher Scientific, Waltham, MA,

USA) for quantification of organic acids and ammonia nitrogen ($\text{NH}_3\text{-N}$). The third subsample was used for enumeration of microbial population in ensiled biomass.

Analytical analyses

A 10-g fresh silage sample was placed into a 250 mL conical flask and covered with 100 mL distilled water. The flasks were shaken for 1 h on a mechanical shaker (Green Sseriker, Vision Scientific, Daejeon, Korea) and stored in refrigerator for 24 h. The conical flasks were shaken by hand every 2 hours during refrigeration. The mixture was filtered through a filter paper (Whatman No. 6, Advantech, Zurich, Switzerland). Silage pH was determined in the filtrate with a pH meter (AB 150, Fisher Scientific International, Pittsburgh, PA, US). A 1.5 mL portion of the filtrate was used for analysis of the organic acid concentration using high performance liquid chromatography (HPLC, Agilent Technologies, Santa Clara, CA, US) equipped with a refractive index detector [8]. $\text{NH}_3\text{-N}$ was analyzed via the method described by Broderick and Kang [14]. The spread-plate method [15] was used to enumerate the population of microorganisms. In brief, a 10-g sample was diluted with 90 mL sterilized saline solution (8.50 g/L NaCl) and shaken for 1 h. Lactic acid bacteria (LAB), molds, and total microorganisms were enumerated on Rogosa, and Sharpe agar medium, potato dextrose agar, and plate count agar media, respectively. The limit of detection was $2 \text{ Log}_{10} \text{ CFU/g}$ fresh mass.

DM concentration in ensiled material was determined in triplicate at 65°C in a forced drying oven for 72 h. The dried samples were ground to pass through a 1 mm screen (Thomas Scientific, Swedesboro, NJ, USA) for nutrient composition analysis. Total nitrogen was quantified via the Dumas method [16], and crude protein (CP) was calculated as nitrogen \times 6.25. Acid detergent fiber (ADF) and neutral detergent fiber (NDF) were measured following the method of Van Soest et al. [17]. Water-soluble carbohydrate (WSC) was analyzed via a modification of the anthrone method proposed by Yemm and Willis [18].

In vitro dry matter digestibility

In vitro DM digestibility (IVDMD) was performed in triplicate using an Ankom Daisy^{II} incubator (ANKOM Technologies, Fairport, NY, USA) [19], as described by Goering and Van Soest [20]. Ground samples (0.5–0.6 g) were weighed into F57 filter bags and sealed using a heat sealer. Samples were evenly distributed on both sides of the digestion jars. Then, 1,330 mL buffer solution A and 266 mL buffer solution B were added to each jar. Two ruminally cannulated Holstein steers were selected and their rumen fluid was collected before the morning feed and passed through four layers of cheesecloth. Then, 400 mL rumen fluid was added to the buffer solution and samples. The digestion jar was purged with CO_2 gas for 30 s and then closed with a lid. The jars were incubated at 39°C for 48 h. Undigested NDF residues in original bags were extracted using an ANKOM²⁰⁰⁰ fiber analyzer.

Statistical analysis

Field experiment was arranged in a completely randomized block design with three replications. Data were subjected to analysis of variance (ANOVA) using the general linear model (GLM) in SPSS (IBM SPSS Statistics, Version 24.0, Armonk, NY, USA). Individual plot was regarded as the experimental unit in the model for analysis of data from the field experiment (Table 1). Individual silo served as the experimental unit in the model for analysis of data from silage experiment. Prior to statistical analysis, microbial data (Table 4) were logarithmically transformed. Mean treatment differences were obtained by Duncan's multiple range tests, with a statistical significance level of 5%.

Table 1. Forage yield and forage quality of proso millet, corn, and sorghum-sudangrass hybrid

Items	Forage type			SEM	p-value
	Proso millet	Corn	Sorghum-sudangrass hybrid		
Dry matter (g/kg)	303 ^a	277 ^b	193 ^c	11.7	< 0.01
TDN (g/kg DM)	631 ^b	677 ^a	541 ^c	16.2	< 0.01
RFV	97 ^b	117 ^a	77 ^c	7.63	< 0.01
Yield (tons/ha)					< 0.01
Fresh matter	25.4 ^c	67.6 ^b	121.7 ^a	8.97	< 0.01
Dry matter	7.69 ^c	18.7 ^b	23.5 ^a	1.41	< 0.01

^{a-c}Means with different letter within each row differ ($p < 0.05$).

TDN, total digestible nutrients. For proso millet and sorghum-sudangrass hybrid, TDN was calculated according to the following equation: $[889 - (0.79 \times \text{ADF, g/kg DM})]$. For corn plant, TDN was calculated using the following equation: $[878.4 - (0.70 \times \text{ADF, g/kg DM})]$ [46]; RFV, relative feed value calculated according to the following equation: $[(\text{dry matter intake} \times \text{digestible dry matter})/1.29]$, where dry matter intake = $120/(\text{NDF}\%)$ and digestible dry matter = $88.9 - (0.779 \times \text{ADF}\%)$ [47].

RESULTS AND DISCUSSION

Forage quality and yield

Yield and forage quality of experimental forage crops are presented in Table 1. Forage DM concentration was greatest in proso millet (303 g/kg), intermediate in corn (277 g/kg), and lowest in sorghum-sudangrass hybrid (193 g/kg). Whole-plant corn had the highest relative feed value (RFV) of 117, which was 20 and 40 units higher on average than proso millet and sorghum-sudangrass hybrid, respectively. A forage crop with an RFV between 103 and 124 is considered a high-quality forage [21], indicating the superiority of corn over sorghum-sudangrass hybrid and proso millet forage. Similar to our observations, Jahansouz et al. [22] also reported a similar trend in fresh forage yield. Concentration of total digestible nutrients was highest in corn (667 g/kg DM), intermediate with proso millet (631 g/kg DM), and lowest with sorghum-sudangrass hybrid (541 g/kg DM). In general, the forage nutritive value of proso millet is comparable to the value reported by Kim et al. [23] harvesting “Geumsilchal” variety in reclaimed lands located in Sihwa (Korea).

Forage yield was significantly different by forage types, with proso millet producing the least DM. The forage DM yield was greater in the order of sorghum-sudangrass hybrid (23.5 t/ha), corn (18.7 t/ha), and proso millet (7.68 t/ha). Forage yield of proso millet (fresh or DM basis) agrees with the values reported by Shin et al. [24]. Calamai et al. [4] also reported that total dry biomass in proso millet averaged 6.43 t/ha. Data of NDF and CP concentration of these forage crops is previously reported [8]. NDF was highest in sorghum-sudangrass hybrid, intermediate in proso millet, and lowest in corn. No difference existed in CP concentration among crops, averaging 58 g/kg DM.

Chemical composition during ensiling

Changes in DM loss and chemical composition of the three forage crops during ensiling are reported in Table 2. As ensiling progressed, DM loss occurred in all crops, with proso millet losing the most DM than corn or sorghum-sudangrass hybrid, most likely because a higher number of epiphytic molds existed on proso millet biomass. Loss of DM was faster in proso millet during the first day of fermentation, which may be justified by the significantly greater population of total microorganisms in fresh mass of proso millet than in corn or sorghum-sudangrass hybrid (Table 4). Microbial degradation of nutrients into carbon dioxide and water could possibly explain loss of DM with ensiling [25,26]. CP concentration displayed a downward trend during the ensiling process, which is suggestive of protein degradation with ensiling. A downward trend was also observed in

Table 2. Dry matter (DM) concentration, DM loss and chemical composition during ensiling

Items	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
DM (g/kg)	Proso millet	284.4 ^{aA}	284.1 ^{aA}	278.6 ^{abA}	278.9 ^{abA}	272.2 ^{bcA}	270.2 ^{cA}	269.6 ^{cA}	266.8 ^{cA}	266.4 ^{cA}	3.22
	Corn	275.2 ^{abB}	274.6 ^{abB}	274.7 ^{aA}	272.5 ^{aA}	272.1 ^{aA}	271.0 ^{aA}	268.4 ^{abA}	264.1 ^{bcA}	260.7 ^{cA}	2.82
	Sorghum-sudan-grass hybrid	190.3 ^{aC}	187.5 ^{abC}	183.1 ^{abcB}	180.0 ^{bcB}	174.3 ^{cb}	177.7 ^{cb}	176.7 ^{cb}	178.4 ^{bcB}	174.7 ^{cb}	3.82
DM loss	Proso millet	19.0 ^{cA}	19.30 ^{cA}	24.8 ^{bcA}	24.50 ^{bcA}	31.2 ^{abA}	33.20 ^{aA}	33.8 ^{aA}	36.6 ^{aA}	37.0 ^{aA}	2.95
	Corn	2.14 ^{dB}	2.73 ^{dB}	2.63 ^{dB}	4.78 ^{cdC}	5.20 ^{cdC}	6.28 ^{cdC}	8.94 ^{bcC}	13.2 ^{abB}	16.6 ^{abB}	2.11
	Sorghum-sudan-grass hybrid	2.50 ^{dB}	5.30 ^{dcB}	9.70 ^{bcC}	12.8 ^{abB}	18.5 ^{ab}	15.1 ^{ab}	16.1 ^{ab}	15.6 ^{ab}	18.1 ^{ab}	2.23
Crude protein	Proso millet	62.3 ^{aA}	61.0 ^{abA}	59.8 ^{abA}	58.3 ^{abA}	60.3 ^{abA}	59.9 ^{abA}	57.9 ^{ba}	59.6 ^{abA}	57.1 ^{ba}	1.46
	Corn	57.4 ^{ab}	56.5 ^{ab}	58.4 ^{aA}	54.6 ^{abB}	54.5 ^{abB}	53.40 ^{abB}	52.7 ^{abB}	53.0 ^{abB}	50.8 ^{bb}	2.12
	Sorghum-sudan-grass hybrid	53.4 ^{aC}	48.9 ^{abC}	49.5 ^{abB}	49.6 ^{abBC}	48.4 ^{abC}	46.2 ^{bc}	46.6 ^{bc}	46.6 ^{bc}	46.2 ^{bc}	1.99
ADF	Proso millet	324.9 ^{bb}	327.3 ^{bb}	324.3 ^{bb}	342.4 ^{ab}	344.4 ^{ab}	340.1 ^{abB}	347.1 ^{ab}	330.1 ^{bb}	345.8 ^{ab}	4.76
	Corn	260.5 ^{aC}	256.2 ^{abC}	251.5 ^{bc}	243.4 ^{dcC}	248.9 ^{bcC}	249.2 ^{bcC}	246.1 ^{cdC}	241.5 ^{dc}	252.0 ^{bc}	3.21
	Sorghum-sudan-grass hybrid	419.1 ^A	419.7 ^A	420.4 ^A	414.9 ^A	420.5 ^A	422.1 ^A	427.6 ^A	413.2 ^A	415.1 ^A	4.98
NDF	Proso millet	608.5 ^{ab}	610.8 ^{ab}	604.3 ^{ab}	606.6 ^{ab}	601.8 ^{abB}	610.5 ^{ab}	602.5 ^{abB}	586.0 ^{bb}	590.5 ^{bb}	6.01
	Corn	496.1 ^{aC}	491.0 ^{aC}	467.6 ^{bc}	445.3 ^{cc}	454.4 ^{cc}	455.8 ^{cc}	455.6 ^{cc}	445.5 ^{cc}	449.2 ^{cc}	4.37
	Sorghum-sudan-grass hybrid	674.7 ^{aA}	673.1 ^{aA}	668.0 ^{aA}	665.0 ^{aA}	666.2 ^{aA}	669.8 ^{aA}	671.7 ^{aA}	635.0 ^{ba}	640.1 ^{ba}	5.45

Values were expressed as g/kg DM, unless otherwise stated.

^{a-c}Values with different lowercase letters within each row show significant difference among ensiling days with the same forage type.

^{A-C}Values with different capital letters within each column show significant differences among forage types in the same ensiling day ($p < 0.05$).

ADF, acid detergent fiber; NDF, neutral detergent fiber.

NDF concentration of all forage crops with ensiling. From day 0 to 45, NDF concentration of proso millet decreased from 607 to 591 g/kg DM, which is less than the corresponding values in corn and sorghum-sudangrass hybrid. Chen et al. [26] suggested that hemicellulose degradation during the ensiling process is mainly responsible for NDF reduction with ensiling. This loss could be due to a combination of enzymatic and acid hydrolysis of the more digestible cell-wall fractions during the fermentation [10,27]. After 45 days of ensiling, ADF concentration of proso millet silage declined by about 20 g/kg DM. Similar decreases also occurred for corn and sorghum-sudangrass hybrid.

Fermentation quality during ensiling

Changes in silage pH as a function of fermentation time are illustrated in Fig. 2. The day-0 pH of corn crop (5.80) was generally lower than proso millet or sorghum-sudangrass hybrid (mean 6.05), which is in agreement with the mean values (5.50 to 6.0) reported for the different forages after chopping [28,29]. Silage pH of corn and sorghum-sudangrass hybrid fell rapidly to below 5 within 24 hrs of ensiling, but it took 3 days for proso millet pH to decline below this value. During the late phase of ensiling, silage pH remained stable and was significantly lower in corn than in proso millet or sorghum-sudangrass hybrid ($p < 0.05$), possibly due to the higher population of LAB in corn silage biomass (Table 4). During the 45-day ensiling period, silage pH of corn, proso millet, and sorghum-sudangrass hybrid decreased by 1.94, 1.65, and 2.04 units, respectively. Buffering capacity, WSC concentration, and moisture level have been identified as critical parameters influencing the ensilability of forages if epiphytic LAB exist in sufficient numbers [30]. Buffering capacity was lowest in corn (24.2 mEq/kg DM), intermediate in proso millet (32 mEq/kg DM), and highest in

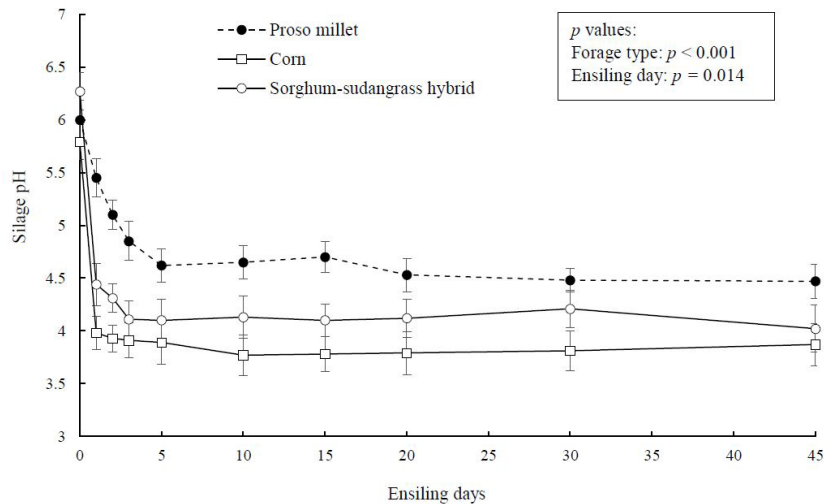


Fig. 2. The pH value of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling days. Bars indicate standard error.

sorghum-sudangrass hybrid (55.5 mEq/kg DM) [8]. Forages with higher buffering capacity require more acids for pH reduction. This supports the faster pH reduction in corn plant at the initial phase of ensiling than proso millet or sorghum-sudangrass hybrid.

Time-course of silage ammonia-nitrogen development, expressed as a proportion of total N is illustrated in Fig. 3. Initial NH₃-N (g/kg total N) level before ensiling was highest in corn (35), intermediate in proso millet (30), and lowest in sorghum-sudangrass hybrid (14.4). Ammonia-N concentration increased in three forage crops as ensiling progressed, with proso millet exhibiting the highest rise. This indicates that protein fractions in proso millet were degraded to a greater extent during ensiling, perhaps because of accelerated rate of proteolysis and deamination [31]. The NH₃-N concentration of less than 70 g/kg total N indicates successful silage fermentation, whereas amounts greater than 100 g/kg total N have been linked to poor silage fermentation [32]. This

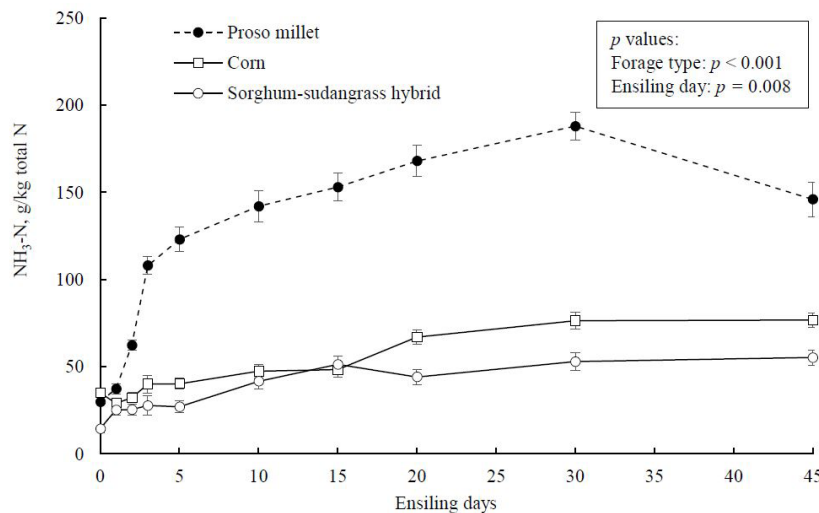


Fig. 3. Ammonia-nitrogen concentration of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling days. Bars indicate standard error.

criterion indicates more degradation of protein in proso millet than corn and sorghum-sudangrass hybrid. The rapid acidification of silage mass is known to inhibit growth and activity of undesirable microorganisms as well as proteolytic activity [10,33]. The higher $\text{NH}_3\text{-N}$ concentration in proso millet silage could be attributed to its higher pH during ensiling, which was likely insufficient to effectively suppress enzymes and microorganisms involved in protein degradation during fermentation.

Concentration of WSC in silage mass over the course of the 45-d fermentation is presented in Fig. 4. Initial WSC concentration (before ensiling) was higher in proso millet than in corn or sorghum-sudangrass hybrid (170 vs. mean 141 g/kg DM). An initial WSC concentration between 60 and 80 g/kg DM has been suggested as an adequate amount to promote an efficient silage fermentation [34]. This indicates that the forage crops evaluated in this study contained sufficient WSC to promote a good-quality silage fermentation. The exhaustion of WSC was faster in corn plant as ensiling progressed, reaching a minimum of 6.70 g/kg DM after 3 days of ensiling, after which WSC concentration decreased slightly until day 45 of ensiling (5.20 g/kg DM). Proso millet experienced a comparatively slower rate of decline in WSC during ensiling, decreasing to 18.2 g/kg DM on day 15 of ensiling and reaching a mean value of 5.9 g/kg DM after 45 days of ensiling.

During the ensiling fermentation, LAB consume WSC as a readily available source of energy and primarily convert it to lactic acid, which is associated with silage mass acidification and inhibition of the activities of undesirable microorganisms [26]. Variations in WSC consumption rates amongst forage crops during the early phase of ensiling might be ascribed to differences in microbial activity and plant enzymes in the crops prior to ensiling. In general, WSC supplies the energy required to drive silage fermentation [35]. A sufficient quantity of WSC has been identified as an important factor in fast acidification during the initial phase of ensiling, which is associated with DM loss reduction and improvement of silage quality [10]. In our experiment, the faster reduction of WSC in corn compared to proso millet forage represented a faster decline in silage pH, which was associated with less DM loss and $\text{NH}_3\text{-N}$ production during ensiling.

The IVDMD of the experimental forage crops as a function of ensiling duration are illustrated in Fig. 5. Before ensiling, IVDMD of proso millet and sorghum-sudangrass hybrid was not different, averaging 643 g/kg DM, which was approximately 16% less than corn (746 g/kg DM).

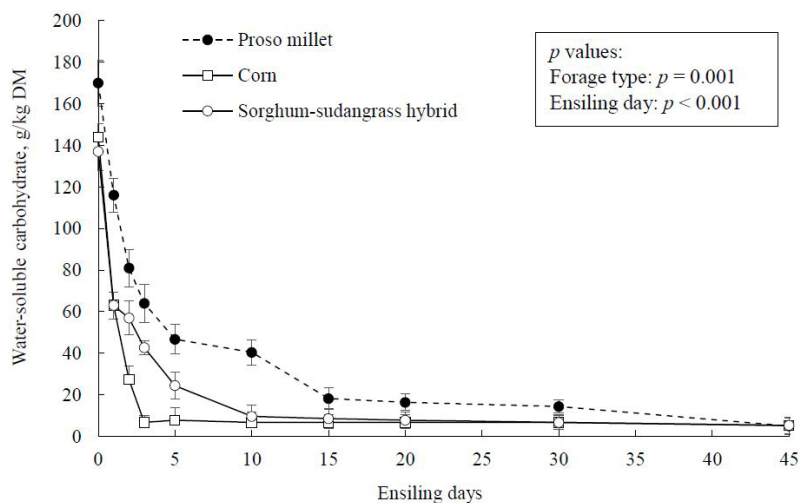


Fig. 4. Water-soluble carbohydrate concentration of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling days. Bars indicate standard error. DM, dry matter.

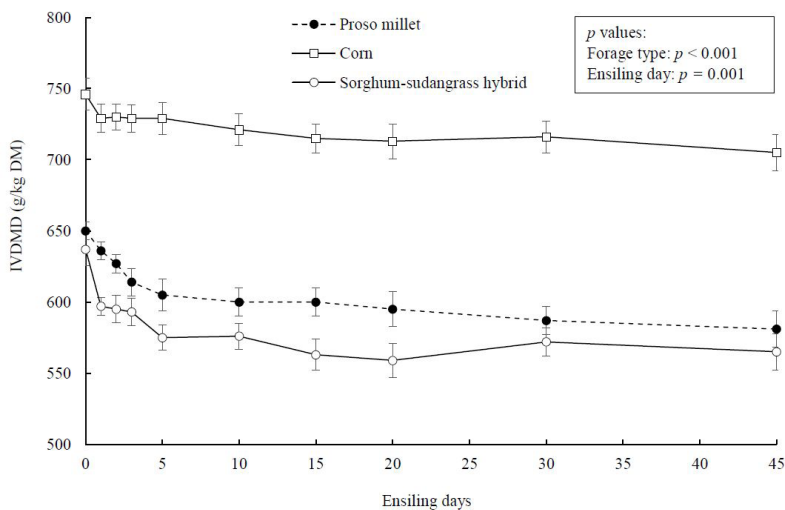


Fig. 5. *In vitro* dry matter digestibility (IVDMD) of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling days. Bars indicate standard error. DM, dry matter.

All crops experienced a decline in IVDMD with ensiling. Previous studies have identified that ADF and NDF concentrations correlate negatively with IVDMD [36]. This supports findings of the current study because corn had less NDF and ADF fractions than proso millet or sorghum-sudangrass hybrid, resulting in the higher digestibility of corn than the other two crops.

Organic acids formation during ensiling

Formation of lactic acid and acetic acid as a function of ensiling duration is illustrated in Table 3. Butyric acid was undetectable during the 45-day ensiling period, which indicates a well-fermented silage and a lack of clostridial activity during ensiling process [10,26,29]. High silage pH, typically greater than 4.5, low DM concentration, and high buffering capacity have been identified as probable factors which contribute to clostridia growth and proliferation during ensiling [32,37]. This suggests that among the forage types evaluated in this experiment, proso millet had a greater susceptibility to clostridial activity and, thus butyric acid production. However, such an effect was not observed in this experiment and the absence of butyric acid detection during silage fermentation of proso millet indicates its low susceptibility to putrefaction by clostridial fermentation.

Lactic acid formation increased as silage fermentation progressed, and the magnitude of this increase was generally greater in sorghum-sudangrass hybrid, intermediate in corn, and lowest in proso millet. During the 45-day ensiling period, lactic acid concentration displayed an upward trend and reached a maximum on day 45, with values of 42.5 g/kg DM for proso millet, 67.7 g/kg DM for corn, and 127 g/kg DM for sorghum-sudangrass hybrid. Lactic acid is typically found in concentrations ranging from 20 to 40 g/kg DM in commonly used silages [29], which indicates that all forages in the present experiment underwent an adequate lactic acid fermentation. Similar to lactic acid production, acetic acid also increased with ensiling, the rate of its production was generally larger in the earlier phase of silage fermentation. During ensiling process, acetic acid was usually lower in proso millet than in corn and sorghum-sudangrass hybrid. Acetic acid concentration in sorghum-sudangrass hybrid reached a maximum concentration of 100 g/kg DM on day 45 of silage fermentation. The higher lactic acid and acetic acid production in sorghum-sudangrass hybrid during silage fermentation could be explained by its higher moisture concentration than the other two crops, which accelerates microbial activity and acid production during the ensiling process.

Table 3. Concentrations of lactic acid and acetic acid as a function of ensiling days

Organic acids	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid (LA) (g/kg DM)	Proso millet	10.1 ^{ab}	14.1 ^{deB}	23.2 ^{cb}	21.6 ^{cdB}	29.6 ^{bcC}	21.8 ^{cdC}	40.0 ^{ab}	37.0 ^{abC}	42.5 ^{ac}	3.64
	Corn	17.6 ^{ab}	28.6 ^{da}	33.6 ^{daB}	42.9 ^{ca}	44.3 ^{cb}	48.0 ^{cb}	57.5 ^{ba}	62.0 ^{abB}	66.7 ^{ab}	2.98
	Sorghum-sudan-grass hybrid	31.3 ^{aA}	36.1 ^{deA}	39.7 ^{deA}	45.2 ^{da}	67.9 ^{ca}	71.0 ^{ca}	69.3 ^{ca}	98.4 ^{ba}	126.6 ^{aA}	5.49
Acetic acid (AA) (g/kg DM)	Proso millet	5.67 ^{ac}	11.1 ^{deB}	14.7 ^{dc}	14.1 ^{dc}	26.3 ^{cc}	25.1 ^{cc}	62.7 ^{aA}	49.0 ^{bb}	41.7 ^{bb}	3.37
	Corn	10.3 ^{db}	14.8 ^{db}	22.6 ^{cb}	27.1 ^{cb}	37.6 ^{bb}	34.5 ^{bb}	26.0 ^{cb}	48.4 ^{ab}	38.2 ^{bb}	2.24
	Sorghum-sudan-grass hybrid	15.9 ^{aA}	57.6 ^{cdA}	49.6 ^{da}	63.3 ^{ca}	54.8 ^{cdA}	77.5 ^{ba}	61.6 ^{ca}	83.2 ^{ba}	100.3 ^{aA}	4.63
LA/AA	Proso millet	1.78 ^a	1.27 ^{bb}	1.58 ^{abA}	1.53 ^{abA}	1.13 ^b	0.87 ^{cdB}	0.64 ^{dc}	0.76 ^{db}	1.02 ^{bcB}	0.15
	Corn	1.71 ^{bc}	1.93 ^{abA}	1.48 ^{cdA}	1.58 ^{bcdA}	1.18 ^d	1.39 ^{cdA}	2.21 ^{aA}	1.28 ^{da}	1.75 ^{bcA}	0.21
	Sorghum-sudan-grass hybrid	1.97 ^a	0.62 ^{bc}	0.80 ^{bb}	0.71 ^{bb}	1.24 ^{ab}	0.92 ^{bb}	1.13 ^{abB}	1.18 ^{abA}	1.26 ^{abB}	0.45

^{a-c}Values with different lowercase letters within each row show significant difference among ensiling days with the same forage type.

^{A-C}Values with different capital letters within each column show significant differences among forage types in the same ensiling day ($p < 0.05$).

This explanation is supported by the findings of a previous study identifying that a lower moisture level limits silage fermentation [38]. Although no consistent trend was seen in lactic acid: acetic acid ratio, there was a general downward trend for each crop, which is likely indicative of a shift from homo- to hetero-fermentative pattern. This observation is consistent with results reported by Shao et al. [39,40]. The higher ratio of lactic acid: acetic acid in corn is most likely suggestive of the dominance of homofermentative LAB during the ensiling process.

Microbial composition during ensiling

Changes in microbial population as a function of ensiling duration are shown in Table 4. The pre-ensiling population of LAB, mold, and total microorganisms is presented in our companion paper [8]. Briefly, the highest LAB count was detected in corn (6.15 Log₁₀ CFU/g), followed by proso millet (5.91 Log₁₀ CFU/g), and sorghum-sudangrass hybrid (5.88 Log₁₀ CFU/g). An LAB count of 5.0 Log₁₀ CFU/g biomass has been suggested as a minimum number to enable the dominance of the epiphytic LAB during ensiling [41,42]. This suggests that the forage crops had sufficient epiphytic LAB population to initiate an efficient silage fermentation. Number of mold was highest on proso millet biomass (4.53 Log₁₀ CFU/g fresh mass), which was 0.23 and 1.23 Log₁₀ CFU/g fresh mass greater than corn and sorghum-sudangrass hybrid, respectively. Forage species, maturity stage, weather, and field wilting have all been identified as factors causing differences in the population of epiphytic microorganisms in forage crops [43]. During the 45-day of fermentation, LAB count was generally lower in proso millet than corn or sorghum-sudangrass hybrid. Number of LAB increased during the early ensiling period and peaked on day 10 of ensiling. Low pH and the exhaustion of fermentable substrates have been identified as the primary factors contributing to the decline of LAB population as ensiling proceeds [44].

Mold was always present in each crop during fermentation, with a lower number existing on corn than proso millet or sorghum-sudangrass hybrid. The lower mold population in corn biomass is likely related to the rapid acidification (lower pH) of corn silage, inhibiting the growth of undesirable microorganisms [26,42]. Another factor inhibiting mold growth during ensiling is a high acetic acid concentration [45]. Less formation of acetic acid and lactic acid (higher pH) during ensiling fermentation of proso millet could possibly explain the higher mold number in proso millet biomass during ensiling. The count of total microorganisms was generally higher in corn than in

Table 4. Number of lactic acid bacteria, mold and total microorganisms as a function of ensiling days

Microbial count ¹⁾	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid bacteria	Proso millet	6.48 ^{bB}	6.88 ^{aA}	6.94 ^{aB}	6.93 ^{aB}	6.96 ^{aB}	6.48 ^{bB}	5.78 ^{cC}	5.78 ^{cB}	5.34 ^{dB}	0.11
	Corn	6.84 ^{fA}	6.85 ^{fA}	7.23 ^{cdA}	7.30 ^{ca}	7.77 ^{aA}	7.61 ^{abA}	7.04 ^{eA}	6.60 ^{gA}	6.08 ^{hA}	0.08
	Sorghum-sudan-grass hybrid	5.08 ^{dC}	5.68 ^{cB}	6.60 ^{bC}	6.53 ^{bC}	6.95 ^{aB}	6.60 ^{bB}	6.95 ^{aB}	6.62 ^{bA}	5.89 ^{ca}	0.12
Molds	Proso millet	3.49 ^{dA}	4.21 ^{ca}	4.30 ^{bcA}	4.20 ^{ca}	5.00 ^{abA}	5.38 ^{aA}	4.34 ^{bcA}	4.04 ^{cB}	4.80 ^{ba}	0.30
	Corn	3.18 ^{cB}	3.00 ^{cC}	4.00 ^{bb}	3.00 ^{cC}	4.05 ^{bB}	4.00 ^{bC}	3.85 ^{bB}	4.67 ^{aA}	4.18 ^{abB}	0.41
	Sorghum-sudan-grass hybrid	3.48 ^{cdA}	3.30 ^{dB}	3.85 ^{bB}	3.70 ^{bcB}	3.29 ^{dC}	5.11 ^{ab}	3.60 ^{bcC}	3.31 ^{dC}	3.00 ^{ec}	0.13
Total microorganisms	Proso millet	7.43 ^{bA}	7.51 ^{ba}	7.44 ^{ba}	7.79 ^{ab}	7.86 ^{ab}	7.26 ^{bcA}	7.04 ^{cB}	6.30 ^{dC}	6.60 ^{eb}	0.10
	Corn	7.05 ^{cB}	7.29 ^{ca}	7.18 ^{cB}	8.10 ^{ba}	8.85 ^{aA}	7.11 ^{caB}	7.85 ^{ba}	7.04 ^{cB}	7.12 ^{ca}	0.19
	Sorghum-sudan-grass hybrid	6.57 ^{cC}	6.88 ^{bb}	6.48 ^{cc}	7.01 ^{bc}	7.40 ^{ac}	6.95 ^{bb}	7.04 ^{bb}	7.32 ^{aA}	6.51 ^{cb}	0.08

¹⁾Microbial count was expressed as the logarithmic number of colony-forming units per gram fresh mass.

^{a-h}Values with different lowercase letters within each row show significant difference among ensiling days with the same forage type.

^{A-C}Values with different capital letters within each column show significant differences among forage types in the same ensiling day ($p < 0.05$).

sorghum-sudangrass hybrid or proso millet. Total microorganisms reached the maximum number on day 10 of ensiling, and then followed a downward trend, which could be explained by pH reduction at this time point, limiting the growth of microorganisms.

CONCLUSION

Silage fermentation of proso millet forage resulted in a significant increase in NH₃-N generation and a larger loss of DM when compared to corn or sorghum-sudangrass hybrid, perhaps because of its higher buffering capacity and silage pH. However, butyrate was undetectable during its ensiling fermentation. Further research is needed to optimize the fermentation quality of proso millet forage, possibly by using the appropriate silage additives to minimize ammonia-nitrogen formation during fermentation, as well as to promote greater lactate production, which is associated with a further decline in silage pH and mold growth inhibition, and thus with a reduction in DM loss. Despite the lower productivity (less forage production per unit of cultivated land) than corn and sorghum-sudangrass hybrid, nutrient value of proso millet was comparable to sorghum-sudangrass hybrid. Proso millet could be harvested in a shorter period of time, making it a potential summer crop in situations where cultivation of other major summer crops is limited.

REFERENCES

1. KOSIS [Korean Statistical Information Service]. Farm households by size of raising Korean beef cattle/total head [Internet]. 2017 [cited 2021 September 3]. https://kosis.kr/statHtml/statHtml.do?orgId=101&tblId=DT_1EO221&conn_path=I2
2. McCartney D, Fraser J, Ohama A. Potential of warm-season annual forages and Brassica crops for grazing: a Canadian review. *Can J Anim Sci.* 2009;89:431-40. <https://doi.org/10.4141/CJAS09002>
3. Habiyaremye C, Matanguihan JB, D'Alpoim Guedes J, Ganjyal GM, Whiteman MR, Kidwell KK, et al. Proso millet (*Panicum miliaceum* L.) and its potential for cultivation in the Pacific Northwest, U.S.: a review. *Front Plant Sci.* 2017;7:1961. <https://doi.org/10.3389/>

- fpls.2016.01961
4. Calamai A, Masoni A, Marini L, Dell'acqua M, Ganugi P, Boukail S, et al. Evaluation of the agronomic traits of 80 accessions of proso millet (*Panicum miliaceum* L.) under Mediterranean pedoclimatic conditions. *Agriculture*. 2020;10:578. <https://doi.org/10.3390/agriculture10120578>
 5. Amadou I, Gounga ME, Le GW. Millets: nutritional composition, some health benefits and processing—a review. *Emir J Food Agric*. 2013;501-8. <https://doi.org/10.9755/efja.v25i7.12045>
 6. Nematpour A, Eshghizadeh HR, Zahedi M. Comparing the corn, millet and sorghum as silage crops under different irrigation regime and nitrogen fertilizer levels. *Int J Plant Prod*. 2021;15:351-61. <https://doi.org/10.1007/s42106-021-00142-8>
 7. Lyon DJ, Burgener PA, DeBoer KL, Harveson RM, Hein GL, Hergert GW, et al. Producing and marketing proso millet in the Great Plains. Lincoln, NE: University of Nebraska; 2008. Extension Circular #EC 137.
 8. Wei SN, Li YF, Jeong EC, Kim HJ, Kim JG. Effects of formic acid and lactic acid bacteria inoculant on main summer crop silages in Korea. *J Anim Sci Technol*. 2021;63:91-103. <https://doi.org/10.5187/jast.2021.e7>
 9. Wang M, Franco M, Cai Y, Yu Z. Dynamics of fermentation profile and bacterial community of silage prepared with alfalfa, whole-plant corn and their mixture. *Anim Feed Sci Technol*. 2020;270:114702. <https://doi.org/10.1016/j.anifeedsci.2020.114702>
 10. Weinberg ZG, Muck RE. New trends and opportunities in the development and use of inoculants for silage. *FEMS Microbiol Rev*. 1996;19:53-68. <https://doi.org/10.1111/j.1574-6976.1996.tb00253.x>
 11. Wiersma DW, Carter PR, Albrecht KA, Coors JG. Kernel milkline stage and corn forage yield, quality, and dry matter content. *J Prod Agric*. 1993;6:94-9. <https://doi.org/10.2134/jpa1993.0094>
 12. Lynch JP, O'Kiely P, Doyle EM. Yield, quality and ensilage characteristics of whole-crop maize and of the cob and stover components: harvest date and hybrid effects. *Grass Forage Sci*. 2012;67:472-87. <https://doi.org/10.1111/j.1365-2494.2012.00868.x>
 13. Ahmadi F, Lee YH, Lee WH, Oh YK, Park K, Kwak WS. Long-term anaerobic conservation of fruit and vegetable discards without or with moisture adjustment after aerobic preservation with sodium metabisulfite. *Waste Manag*. 2019;87:258-67. <https://doi.org/10.1016/j.wasman.2019.02.010>
 14. Broderick GA, Kang JH. Automated simultaneous determination of ammonia and total amino acids in ruminal fluid and in vitro media. *J Dairy Sci*. 1980;63:64-75. [https://doi.org/10.3168/jds.S0022-0302\(80\)82888-8](https://doi.org/10.3168/jds.S0022-0302(80)82888-8)
 15. Madigan MT, Martinko JM, Parker J, Brock TD. Brock biology of microorganisms. Upper Saddle River, NJ: PrenticeHall; 2003.
 16. Jean-Baptiste-André Dumas. *Science*. 1884;3:750-2. <https://doi.org/10.1126/science.ns-3.72.750>
 17. Van Soest PJ, Robertson JB, Lewis BA. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J Dairy Sci*. 1991;74:3583-97. [https://doi.org/10.3168/jds.S0022-0302\(91\)78551-2](https://doi.org/10.3168/jds.S0022-0302(91)78551-2)
 18. Yemm EW, Willis AJ. The estimation of carbohydrates in plant extracts by anthrone. *Biochem J*. 1954;57:508-14. <https://doi.org/10.1042/bj0570508>
 19. ANKOM Technology. ANKOM Technology method 3: in vitro true digestibility using the DAISYII incubator [Internet]. ANKOM Technology. 2017 [2021 May 10]. http://www.ankom.com/media/documents/TVDMMD_0805_D200.pdf

20. Goering HK, Van Soest PJ. Forage fiber analysis (apparatus, reagents, procedures, and some applications). Washington, DC: US Department Agriculture-Agricultural Research Service (USDA-ARS); 1970. Agricultural Handbook No. 379.
21. Horrocks RD, Valentine JF. Harvested forages. San Diego, CA: Academic Press; 1999.
22. Jahansouz MR, Keshavarz Afshar R, Heidari H, Hashemi M. Evaluation of yield and quality of sorghum and millet as alternative forage crops to corn under normal and deficit irrigation regimes. *Jordan J Agric Sci.* 2014;10:699-715. <https://doi.org/10.12816/0031747>
23. Kim JG, Jeong EC, Kim MJ, Li YF, Kim HJ, Lee SH. Comparison of growth characteristics and productivity of summer forage crops in Sihwa reclaimed land. *J Korean Soc Grassl Forage Sci.* 2021;41:110-8. <https://doi.org/10.5333/KGFS.2021.41.2.110>
24. Shin JS, Kim WH, Lee SH, Shin HY. Comparison of forage yield and feed value of millet varieties in the reclaimed tidelands. *J Korean Soc Grassl Forage Sci.* 2006;26:215-20. <https://doi.org/10.5333/KGFS.2006.26.4.215>
25. Kim JS, Lee YH, Kim YI, Ahmadi F, Oh YK, Park JM, et al. Effect of microbial inoculant or molasses on fermentative quality and aerobic stability of sawdust-based spent mushroom substrate. *Bioresour Technol.* 2016;216:188-95. <https://doi.org/10.1016/j.biortech.2016.05.056>
26. Chen L, Yuan XJ, Li JF, Dong ZH, Wang SR, Guo G, et al. Effects of applying lactic acid bacteria and propionic acid on fermentation quality, aerobic stability and in vitro gas production of forage-based total mixed ration silage in Tibet. *Anim Prod Sci.* 2019;59:376-83. <https://doi.org/10.1071/AN16062>
27. McDonald P, Henderson N, Heron S. The biochemistry of silage. Marlow: Chalcombe Publications; 1991.
28. Diepersloot EC, Pupo MR, Ghizzi LG, Gusmão JO, Heinzen C Jr, McCary CL, et al. Effects of microbial inoculation and storage length on fermentation profile and nutrient composition of whole-plant sorghum silage of different varieties. *Front Microbiol.* 2021;12:660567. <https://doi.org/10.3389/fmicb.2021.660567>
29. Kung L Jr, Shaver RD, Grant RJ, Schmidt RJ. Silage review: interpretation of chemical, microbial, and organoleptic components of silages. *J Dairy Sci.* 2018;101:4020-33. <https://doi.org/10.3168/jds.2017-13909>
30. Martinez-Fernandez A, Soldado C, de la Roza Delgado B, Vicente F, Gonzalez-Arrojo MA, Argamenteria A. Modelling a quantitative ensilability index adapted to forages from wet temperate areas. *Span J Agric Res.* 2013;11:455-62. <https://doi.org/10.5424/sjar/2013112-3219>
31. Arriola KG, Queiroz OCM, Romero JJ, Casper D, Muniz E, Hamie J, et al. Effect of microbial inoculants on the quality and aerobic stability of bermudagrass round-bale haylage. *J Dairy Sci.* 2015;98:478-85. <https://doi.org/10.3168/jds.2014-8411>
32. Lima R, Lourenço M, Díaz RF, Castro A, Fievez V. Effect of combined ensiling of sorghum and soybean with or without molasses and Lactobacilli on silage quality and in vitro rumen fermentation. *Anim Feed Sci Technol.* 2010;155:122-31. <https://doi.org/10.1016/j.anifeedsci.2009.10.008>
33. Pahlow G, Muck RE, Driehuis F, Elferink SJWHO, Spoelstra SF. Microbiology of ensiling. In: Buxton DR, Muck RE, Harrison JH, editors. *Silage Science and Technology*. Madison, Wisconsin: American Society of Agronom; 2003. p. 31-93.
34. Amer S, Hassanat F, Berthiaume R, Seguin P, Mustafa AF. Effects of water soluble carbohydrate content on ensiling characteristics, chemical composition and in vitro gas production of forage millet and forage sorghum silages. *Anim Feed Sci Technol.* 2012;177:23-9. <https://doi.org/10.1016/j.anifeedsci.2012.07.024>

35. Contreras-Govea FE, Muck RE, Broderick GA, Weimer PJ. Lactobacillus plantarum effects on silage fermentation and in vitro microbial yield. *Anim Feed Sci Technol.* 2013;179:61-8. <https://doi.org/10.1016/j.anifeedsci.2012.11.008>
36. Ammar H, López S, González JS, Ranilla MJ. Chemical composition and in vitro digestibility of some Spanish browse plant species. *J Sci Food Agric.* 2004;84:197-204. <https://doi.org/10.1002/jsfa.1635>
37. Queiroz OCM, Ogunade IM, Weinberg Z, Adesogan AT. Silage review: foodborne pathogens in silage and their mitigation by silage additives. *J Dairy Sci.* 2018;101:4132-42. <https://doi.org/10.3168/jds.2017-13901>
38. Kim JG, Chung ES, Seo S, Ham JS, Kang WS, Kim DA. Effects of maturity at harvest and wilting days on quality of round baled rye silage. *Asian-Australas J Anim Sci.* 2001;14:1233-7. <https://doi.org/10.5713/ajas.2001.1233>
39. Shao T, Ohba N, Shimojo M, Masuda Y. Dynamics of early fermentation of Italian ryegrass (*Lolium multiflorum* Lam.) silage. *Asian-Australas J Anim Sci.* 2002;15:1606-10. <https://doi.org/10.5713/ajas.2002.1606>
40. Shao T, Zhang ZX, Shimojo M, Wang T, Masuda Y. Comparison of fermentation characteristics of Italian ryegrass (*Lolium multiflorum* Lam.) and guineagrass (*Panicum maximum* Jacq.) during the early stage of ensiling. *Asian-Australas J Anim Sci.* 2005;18:1727-34. <https://doi.org/10.5713/ajas.2005.1727>
41. Cai Y, Benno Y, Ogawa M, Kumai S. Effect of applying lactic acid bacteria isolated from forage crops on fermentation characteristics and aerobic deterioration of silage. *J Dairy Sci.* 1999;82:520-6. [https://doi.org/10.3168/jds.S0022-0302\(99\)75263-X](https://doi.org/10.3168/jds.S0022-0302(99)75263-X)
42. Muck R, Kung L. Effects of silage additives on ensiling. In: *Proceedings from the Silage: Field to Feedbunk, North American Conference; 1997; Hershey, PA.*
43. Fenton MP. An investigation into the sources of lactic acid bacteria in grass silage. *J Appl Bacteriol.* 1987;62:181-8. <https://doi.org/10.1111/j.1365-2672.1987.tb02397.x>
44. Xu Z, He H, Zhang S, Kong J. Effects of inoculants *Lactobacillus brevis* and *Lactobacillus parafarraginis* on the fermentation characteristics and microbial communities of corn stover silage. *Sci Rep.* 2017;7:13614. <https://doi.org/10.1038/s41598-017-14052-1>
45. Danner H, Holzer M, Mayrhuber E, Braun R. Acetic acid increases stability of silage under aerobic conditions. *Appl Environ Microbiol.* 2003;69:562-7. <https://doi.org/10.1128/AEM.69.1.562-567.2003>
46. Schroeder JW. *Forage nutrition for ruminants.* Fargo, ND: NDSU Extension Service, North Dakota State University; 2004.
47. Rohweder DA, Barnes RF, Jorgensen N. Proposed hay grading standards based on laboratory analyses for evaluating quality. *J Anim Sci.* 1978;47:747-59. <https://doi.org/10.2527/jas1978.473747x>