

Parvidepsidone, a Novel Depsidone from the Barks of Garcinia parvifolia Miq.

Sri Hartati, Megawati Megawati, and Lucia Dwi Antika*

Research Center for Chemistry, National Research and Innovation Agency of Indonesia, Banten 15314, Indonesia

Abstract – *Garcinia parvifolia* Miq., belongs to *Garcinia* genus and Clusiaceae family, is one of the well-known species from *Garcinia* genus which widely found in tropical and subtropical countries, and has been reported to contain natural bioactive compounds. A novel depsidone, parvidepsidone (2), was isolated from the barks of *G parvifolia* along with two known compounds: stigmasterol (1) and rubraxhantone (3). Based on information of LC-MS, 1D and 2D NMR spectra, the structure of parvidepsidone (2) was fully assigned. Keywords – *Garcinia parvifolia* Miq., Parvidepsidone, Stigmasterol, Rubraxhantone

Introduction

Garcinia parvifolia Mig., belongs to Garcinia genus and Clusiaceae family, is one of the well-known species from Garcinia genus which is widely distributed in tropical and subtropical countries including Indonesia, Malaysia, Thailand and Brunei. G. parvifolia Miq., locally known as "asam kandis", "asam kundong" or cherry mangosteen, has been reported to has several natural bioactive compounds.1 The genus of Garcia are rich of secondary metabolites such as polyphenols, flavonoids, xanthones, and polyisoprenylated benzophenones.²⁻⁵ Those metabolites were reported to be the ingredients that contribute to the biological capabilities of G. parvifolia Miq. including antioxidant,⁶ antiviral⁷ and antidiabetes.⁸ The crude extracts and pure compounds from this plant also were found to be cytotoxic against several cancer cell line, including p388 leukemia cell line,⁹ L1219 murine leukemia cell line¹⁰ and human breast cancer cell,¹¹ thus possessing its potential as anti-carcinogenic agent.

This current study aimed to discover a new compound from the barks of *G parvifolia* Miq. Column chromatography separation of the *G parvifolia* crude methanolic backs extract was performed. Additionally, the isolation, structure elucidation, and chemical constituents were also conducted by LC-MS/MS, 1D and 2D NMR analysis. A new depsidone, namely parvidepsidone (2), as well as two well-known compounds, stigmasterol (1) and rubraxanthone (3) were successfully isolated from *G parvifolia* Miq. barks. (Fig. 1).

Experimental

General experimental procedures - Melting points were obtained with an electrothermal digital melting point apparatus and was uncorrected. UV spectra were measured on HP 8453A UV-visible spectroscopy systems (Hewlett-Packard, CA, USA), while IR spectra were recorded on a diode array spectrophotometer FTIR Prestige-21 (Shimadzu, Kyoto, Japan). LC-MS analysis to measure molecular weight was performed by using a Hitachi L-6200 Intelligent Pump coupled to a MarinerTM API-TOF Biospectrometry Workstation (Applied Biosystem, CA, USA). Mass spectroscopy measurements were performed on a time-of-flight (TOF). Samples were introduced to MS via electrospray ionization (ESI system) in the positive ion mode. Thin layer chromatography (TLC) was performed using silica gel GF₂₅₄ (TLC sheets, silica-gelprecoated) (Merck, Darmstadt, Germany). Spots were visualized on TLC under UV light at 254 nm using TLC visualizer (CAMAG Scientific Inc., Wilmington, NC). Column chromatography was performed by using silica gel 100 (70-230 Mesh ASTM) (Merck, Darmstadt, Germany) with hexane, EtOAc and MeOH as organic solvents. Samples were subjected to Sephadex LH-20 column and eluted with CH_2Cl_2 and MeOH (1:1, v/v). ¹H NMR, ¹³C NMR, and correlation NMR spectra including correlation microscopy (COSY), heteronuclear multiple bond correlation (HMBC) and heteronuclear single quantum coherence (HSQC) of compounds in CDCl₃ examined

^{*}Author for correspondence

Lucia Dwi Antika, Research Center for Chemistry, National Research and Innovation Agency of Indonesia, Banten 15314, Indonesia

Tel: +62-21-756-0929; E-mail: lucia.dwi.antika@brin.go.id

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Fig. 1. The structures of compounds (1 - 3) isolated from *G parvifolia* Miq. barks.

Table 1. ¹H and ¹³C NMR data of **2** in CDCl₃ (δ in ppm, 500 MHz for ¹H and 125 MHz for ¹³C)

No. C	δ 'Η	δ ¹³ C	δ_{C} HMBC
1	13.80 (OH)	161.91	C-1, C-2
2	-	108.57	
3	6.35 (OH)	160.76	
4	6.32 (3H, <i>s</i>)	93.41	C-1; C-2; C4a
4a	-	155.30	
5a	-	152.73	
6	-	135,73	
7	-	156.61	
8	-	147.96	
9	-	114.96	
9a	-	131.77	
11	-	182.87	
11a	-	103.87	
12	3.46 (2H, <i>d</i> , <i>J</i> = 7.35 Hz)	21.61	C-1; C-2: C13
13	5.30 (1H, <i>t</i> , <i>J</i> = 7.35 Hz)	121.67	C-16
14	-	122.6	
15	1.77 (3H, <i>s</i>)	26.09	C-2; C-13; C-16
16	1.85 (3H, <i>s</i>)	18.10	C-2; C-13
17	3.56 (2H, <i>d</i> , <i>J</i> = 7.35 Hz)	23.20	C-5a; C-7; C-18; C-19
18	5.20 (1H, t , J = 5.30 Hz)	122.03	C-20; C-21
19	-	132.35	
20	1.85 (3H, <i>s</i>)	18.17	C-21
21	1.68 (3H, <i>s</i>)	25.95	C-18; C-19; C-21
22	4.11 (2H, <i>d</i> , <i>J</i> = 6.7)	26.16	C-9; C-9a; C-23; C-24
23	5.24 (1H, <i>t</i> , <i>J</i> = 5.50 Hz)	123.72	C-22
24	-	135.82	
25	1.87 (3H, <i>s</i>)	18.34	-
26	1.69 (3H, <i>s</i>)	26.54	-
27	3.96 (3H, <i>s</i>)	60.99	C-7
28	3.79 (3H, <i>s</i>)	60.81	C-8

Values are in parentheses and reported in Hz; the assignments were based on ¹H-¹H COSY and HMBC experiments.

using an NMR Spectrometer ECA-500 (Jeol, Tokyo, Japan) operating at 500 MHz (¹H) and 125 MHz (¹³C) with chemical shifts given in ppm (δ).

were collected in Bulungan Forest, North Kalimantan, Indonesia and identified by Dr. Ismail R. A voucher specimen (#625) of the plant was deposited at the Herbarium Bogoriense, Cibinong, Indonesia.

Plant materials – The barks of G parvifolia (3 kg)

Extraction and isolation – The air-dried G. parvifolia Miq. barks (3.5 kg) were pulverized and extracted with hexane (6 L) quadruplicates at room temperature. The solvent was removed to yield 35.7 g of crude extract. The residue was continued to extract with methanol to yield 374.7 g of methanol crude extract. Separation of the hexane extract (30 g) on a silica gel column chromatography eluted with the mixture of hexane and ethyl acetate (EtOAc) (from 10:0 to 0:10 v/v) obtained 15 fractions (1-15). Fraction 3 (0.89 g) was separated using Sephadex LH-20 column chromatography eluted with hexane-EtOAc to provide compound 1 (600 mg). Fraction 5 (1.333 g) was subjected to Sephadex LH-20 column and eluted with CH_2Cl_2 and methanol (1:1, v/v) to give four sub fractions (F5.1-F5.4). Sub fraction F5.3 (340 mg) was separated by using chromatotron (centrifugal chromatography) with hexane-EtOAc to afford compound 2 (120 mg). The crude of methanol extract (100 g) was partitioned with EtOAc-water (1:1, v/v) quadruplicates to yield 64.83 g of EtOAc extract. The EtOAc extract (32 g) was fractionated by silica gel column (200 g) and eluted with hexane-EtOAc (from 10:0 to 0:10 v/v), followed by EtOAC-MeOH (from 8:2 to 0:10 v/v) to obtain 18 fractions (1-18). After purification by silica gel radial chromatography eluted with acetone-hexane, compound 3 (64 mg) was obtained from fraction F4 (150 mg).

Stigmasterol (1) – white needles (600 mg). m.p. 138 -140 °C; UV (MeOH) λ_{max} nm: 229; IR (KBr) $_{max}$ cm⁻¹: 3419 – 3294, 2937-2864. ¹H NMR (CDCl₃, 500 MHz): $\delta_{\rm H}$ 1.01 (3H, *s*), 1.03 (3H, *s*), 0.84 (3H *d*, *j* = 3.15 Hz), 0.79 (3H, *d*, *j* = 5.9 Hz), 0.83 (3H, *d*, *j* = 5.6 Hz), 0.91 (3H, *dd*), 5.35 (1H, *d*. *J* = 5.0 Hz, H-6), 5.16 (1 H, *t*, *J* = 8.4 Hz, H-22), 5.01 (1 H, *t*, *J* = 8,4 Hz, H-23), 3.52 ppm (1H, *m*); ¹³C NMR (CDCl₃, 125 MHz): $\delta_{\rm C}$ 121.92 (C-6), 138.53 (C-22), 129.43 (C-23), 140.92 (C-5). The comparison of NMR spectra of **1** has similarities to the literature data.¹²

Parvidepsidone (2) – yellow powder (120 mg). UV (MeOH) λ_{max} nm: 216.0, 244.5, 259.5 and 314.0; IR (KBr) $_{max}$ cm⁻¹: 3415 – 1644. Table 1 displays the NMR spectral data of **2**. ESI-MS *m*/*z* [M+H⁺] and [2M +Na⁺] calculated for C₃₀H₃₆O₇ found 509.12 (cacld 509.25338) and 1039.21 (cacld 1039.48143), respectively.

Rubraxhantone (3) – yellow powder (64 mg). m.p 201-203 °C; UV (MeOH) λ_{max} nm: 217, 240, 310 and 344; IR (KBr) _{max} cm⁻¹: 3424 – 1643. ¹H NMR (CDCl₃, 500 MHz): $\delta_{\rm H}$ 13.50 (1H,s, OH), 6.83 (1H, s, H-5), 6.19 (1H, *d*, *J* = 1.85 Hz, H-2), 6.30 (1H, *d*, *J* = 1.85 Hz, H-4), 3.79 (3H, s, 11'-OCH₃), 5.27 (1H, *t*, *J* = 5.5, 1.3 Hz, H-1'), 5.03 (1H, *t*, *J* = 5.5, 1.3 z, H-6), 4.11 (2H, *d*, *J* = 6.5 Hz,

H-2'), 2.05 (2H, m, H-4'), 1.97 (2H, *t*, J = 7.9 Hz, H-5'), 1.52 (3H,s, H-8'), 1.55 (3H, s, H-9'), 1.82 (3H, s, H-10'); ¹³C NMR (CDCl₃, 125 MHz): $\delta_{\rm C}$ 164.07 (C-3), 26.46 (C-4'), ESI-MS: m/z [M+H⁺] = 411.08 (calcd 411.18022). The comparison of NMR spectra of **3** has similarities to the literature data.¹³

Result and Discussion

Compounds 1 and 3 were determined by comparing ¹H NMR, ¹³C NMR and MS spectral data with those in the literature and identified as stigmasterol¹² and rubraxanton,¹³ respectively. Compound 2 (parvidepsidone) was isolated as a yellow powder with molecular ion peaks at m/z509.12 (calcd 509.25338) and 1039.21 (cacld 1039.48143), which were confirmed by high-resolution mass spectrometry to be compatible with the chemical formula $C_{30}H_{36}O_7$. The UV-VIS spectrum showed maximum absorption (λ_{max}) at 216.0, 244.5, 259.5 and 314.0, which indicate the presence of an aromatic moiety and a typical UV patter of the depsidone chromophore.¹⁴ The IR v_{max} (KBr) spectrum of compound 2 indicated the presence of hydroxyl (3415 cm⁻¹) and lactone carbonyl groups (1644 cm⁻¹), indicating the presence of a depsidone chromophore.¹⁵ The ¹H NMR spectrum of **2** (Table 1) reveals protons for one aromatic unit and one hydroxyl group at $_{\rm H}$ 6.32 (1H, s, H-4) and $\delta_{\rm H}$ 6.35 (1H, brs, 3-OH), respectively.

Table 1 also exhibits signals for hydrogen-bonded hydroxyl proton at $\delta_{\rm H}$ 13.50 (1H,s,1-OH), three olefinic proton signals at $\delta_{\rm H}$ 5.20 (1H, *t*, *J* = 5.30 Hz), 5.24 (1H, *t*, J = 5.5 Hz), 5.30 (1H, t, J = 7.35 Hz); three methylene proton signals at $\delta_{\rm H}$ 3.46 (2H, *d*, J = 7.35 Hz), 3.56 (2H, d, J = 7.35 Hz), 4.11 (2H, d, J = 6.70 Hz); and six methyl proton signals at δ_H 1.77 (3H, s, H-15), 1.85 (3H, s, H-16), 1.85 (3H,s, H-20), 1.68 (3H,s, H-21), 1.89 (3H,s, H-25), 1.69 (3H, s, H-26) indicating three 3-methyl-2-enyl structure. Compound 2 contains two methoxyl groups at $\delta_{\rm H}$ 3.69 (3H, s) and 3.79 (3H, s). The ¹³C NMR signal assigned the oxygenated quaternary aromatic carbons 4a $(\delta_{\rm C} 155.30)$; 5a $(\delta_{\rm C} 152.73)$; 9a $(\delta_{\rm C} 114.92)$, and substituted aromatic carbon C-11 abandon to the carbonyl carbon of depsidone. In the HMBC spectrum (Table 1), the hydrogenbonded hydroxyl proton signal at $\delta_{\rm H}$ 13.80 (H-1') was correlated with the signal at δ_c 161.91 (C-1) and δ_C 108.57 (C-2) which showed that the hydroxyl group is located at C-1 position. Two methoxyl signals at δ_H 3.96 (3H, s) and $\delta_{\rm H}$ 3.79 (3H, s) were correlated with the signal at δ_c 156.61 (C-7) and δ_c 147.96 (C-8), as well as C-8 (δ_c 147.96). It significantly demonstrated that H-27 attached to C-7 and H-28 attached to C-8, which indicates that

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Fig. 2. HMBC (a) and COSY (b) correlations of 2.

methoxyls are located at C-2 and C3 positions of **2** (Fig. 2). The ¹³C NMR signals at δ_C 182.87 (C=O) and strong absorption at v_{max} 1644 cm⁻¹ in the IR spectrum indicated a characteristic of lactone carbonyl and showed that compound **2** is a depsidone.¹⁵ These data allowed us to assign the structure of **2** as 1,3-dihydroxy-7,8-dimethoxy-2,6,9tris(3-methylbut-2-enyl)-11*H*-dibenzo[*b*,*e*][1-4]

dioxepin -11-one, which was given as the name pervidepsidone.

Previous studies reported some depsidones have been isolated form Garcinia genus, namely schomburgdepsidone B from *Garcinia schomburgkiana*¹⁵ and garcidepsidone B from *G parvifolia*.¹⁶ The NMR spectral data of **2** were similar to those schomburgdepsidone B isolated from *Garcinia schomburgkiana*,¹⁵ except for the replacement of hydroxyl on C-7 [δ_H 3.96 (3H, s); δ_C 60.99] and C-8 [δ_H 3.79 (3H, s); δ_C 60.81] of ring B to methoxyl, as well as substitution of methoxyl on C-3 of ring A to hydroxyl. Compared to study conducted by Rukachaisirikul (2008),¹⁶ parvidepsidone showed different structure in ring A, in which parvidepsidone has one isophrene in C-2 while the previously reported garcidepsidone B has two isophrene in the same position, suggesting this compound is a new depsidone isolated from *G parvifolia* Miq.

Among isolated compounds from this current study, stigmasterol and rubraxanthone have been reported to possess several pharmacological activities including antiinflamatory,^{17,18} antioxidant,^{19,20} and anticarcinogenic.^{21,22} Altogether, this present study reported that three compounds have been successfully isolated from the bark of *G parvifolia* Mig. Based on spectral data, the structures of isolated compounds were determined as stigmasterol (1), parvidepsidone (2) and rubraxhantone (3). Out of those three, compound 2 (parvidepsidone) is considered as a novel depsidone from the barks of *G parvifolia* Miq. However, further studies are necessary to identify the potential of these compounds, particularly parvidepsidone, for bioactivity-related research materials.

Acknowledgments

This research was supported by the Doctoral Scholarship Program funded by the Indonesian Institute of Sciences. Authors are grateful Dr. Ismail R. at the Bogor Botanical Garden for his help in identification and collection of plant material. Authors also thank Nurul Apriani and Renny A. Khairunnisa for their experimental helps during this study.

References

(1) Kosela, S.; Hu, L.; Yip, S.; Rachmatia, T.; Sukri, T.; Daulay, T. S.; Tan, G; Vittal, J. J.; Sim, K. *Phytochemistry* **1999**, *52*, 1375-1377.

(2) Acuña, U. M.; Dastmalchi, K.; Basile, M. J; Kennelly, E. J. J. Food Compos. Anal. 2012, 25, 215-220.

(3) Rao, A. V. R.; Sarma, M. R.; Venkataraman, K.; Yemul, S. S. *Phytochemistry* **1974**, *13*, 1241-1244.

- (4) Bennett, G. J.; Lee, H. H. Phytochemistry 1989, 28, 967-998.
- (5) Gao, X.-M.; Ji, B.-K.; Li, Y.-K.; Ye, Y.-Q.; Jiang, Z.-Y.; Yang, H.-Y.; Du, G; Zhou, M.; Pan, X.-X.; Liu, W.-X.; Hu, Q. F. *J. Braz. Chem. Soc.* **2016**, *27*, 10-14.

(6) Ali Hassan, S. H.; Fry, J. R.; Abu Bakar, M. F. *Biomed. Res. Int.* **2013**, *2013*, 138950.

(7) Adnan, A.; Allaudin, Z. N.; Hani, H.; Loh, H. S.; Khoo, T. J.; Ting,
K. N.; Abdullah, R. *BMC Complement. Altern. Med.* 2019, *19*, 169.

(8) Phukhatmuen, P.; Raksat, A.; Laphookhieo, S.; Charoensup, R.; Duangyod, T.; Maneerat, W. *Heliyon* **2020**, *6*, e03625.

(9) Xu, Y. J.; Chiang, P. Y.; Lai, Y. H.; Vittal, J. J.; Wu, X. H.; Tan, B. K.; Imiyabir, Z.; Goh, S. H. *J. Nat. Prod.* **2000**, *63*, 1361-1363.

(10) Kardono, L.; Hanafi, M.; Sherley, G.; Kosela, S.; Harrison, L. *Pakistan J. Biol. Sci.* **2006**, *9*, 483-486.

(11) Ahmad, N. E.; Bakar, M. F. A.; Suleiman, M.; Bakar, F. I. A.;

- Sabran, S. F.; Kormin, F. IOP Conf. Ser. Earth Environ. Sci. 2021, 736, 012004.
- (12) Goad, L. J.; Akihisa, T. Analysis of Sterols; Springer; Netherlands, 1997, pp 235-255.
- (13) Ampofo, S. A.; Waterman, P. G. Phytochemistry 1986, 25, 2351-2355.
- (14) Rukachaisirikul, V.; Naklue, W.; Phongpaichit, S.; Hutadilok-Towatana, N.; Maneenoon, K. *Tetrahedron* **2006**, *62*, 8578-8585.
- (15) Sukandar, E. R.; Siripong, P.; Khumkratok, S.; Tip-Pyang, S. *Fitoterapia* **2016**, *111*, 73-77.
- (16) Rukachaisirikul, V.; Trisuwan, K.; Sukpondma, Y.; Phongpaichit, S. Arch. Pharm. Res. 2008, 31, 17-20.
- (17) Wahyuni, F. S.; Ali, D. A. I.; Lajis, N. H.; Dachriyanus. *Pharmacogn. J.* **2017**, *9*, 55-57.
- (18) Gabay, O.; Sanchez, C.; Salvat, C.; Chevy, F.; Breton, M.; Nourissat, G; Wolf, C.; Jacques, C.; Berenbaum, F. Osteoarthritis

Cartilage 2010, 18, 106-116.

- (19) Mohamed, G. A.; Ibrahim, S. R. M.; Shaaban, M. I. A.; Ross, S. A. *Fitoterapia* **2014**, *98*, 215-221.
- (20) Yoshida, Y.; Niki, E. J. Nutr. Sci. Vitaminol (Tokyo). 2003, 49, 277-280.

(21) Kangsamaksin, T.; Chaithongyot, S.; Wootthichairangsan, C.; Hanchaina, R.; Tangshewinsirikul, C.; Svasti, J. *PLoS One* **2017**, *12*, e0189628.

(22) Ee, G. C. L.; Izzaddin, S. A.; Rahmani, M.; Sukari, M. A.; Lee, H. L. *Nat. Prod. Sci.* **2006**, *12*, 138-143.

Received August 12, 2021

Revised November 26, 2021

Accepted November 30, 2021