

Wood Chemical Compositions of Raru Species Originating from Central Tapanuli, North Sumatra, Indonesia: Effect of Differences in Wood Species and Log Positions¹

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ABSTRACT

Raru is a lesser-known plant species originating from North Sumatra, Indonesia. Information on the characteristics is still limited, especially its chemical component. Therefore, this study aims to examine the chemical composition information of *Cotylelobium lanceolatum*, *Cotylelobium melanoxyton*, and *Vatica pauciflora* woods based on their axial log positions (bottom, middle, and top). The wood chemical analysis was performed in terms of the Indonesian National Standard (SNI) method. Furthermore, the analysis measured holocellulose, α -cellulose, hemicellulose, lignin content, alcohol benzene extractive content, the extractive substance in hot and water, and solubility in NaOH 1%. The results indicated that the species and their log axial positions affected different chemical components, which included α -cellulose, hemicellulose, and lignin of *C. lanceolatum* amounting to 41.88%, 19.39%, and 28.68% respectively. Meanwhile, for *C. Melanoxyton*, they were 42.01%, 21.11%, and 24.76% respectively; and for *V. pauciflora* wood, they were 42.95%, 23.24%, and 30.11% respectively. The average values of the extractive contents including the solubility in 1: 2 ethanol benzene, NaOH, and hot water for *C. lanceolatum*, *C. melanoxyton*, and *V. pauciflora* wood were (10.58%, 27.62%, 8.13%), (14.54%, 28.22%, 7.82%), and (10.95%, 28.60%, 7.57%) respectively. The wood species had a significant effect on chemical components including lignin, cellulose and hemicellulose, and extractive solubility in cold water. Furthermore, the axial log position had a significant effect on all the parameters of the chemical composition of the wood being tested.

Keywords: raru wood, chemical component, *C. lanceolatum*, *C. melanoxyton*, *V. pauciflora*

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1. INTRODUCTION

Indonesia is well known to have a very high level of biodiversity, especially in terms of its vegetation diversity. One of the provinces with high biodiversity is North Sumatra. It has a tropical forest area with a relatively high level of species diversity and with several local vegetation, which may be studied for their characteristics. The forest area is around 3,010,160.89 ha or 41.25% of the total land area of North Sumatra Province. Several plant species are local commodities of North Sumatra, namely Kemenyan (*Styrax sumatrana*, *Styrax benzoin* Dryan, *Styrax benzoin* var. *Hiliferum*), *Dryobalanop aromatica*, Raru (*Cotylelobium melanoxyton*, *Cotylelobium lanceolatum*, *Vatica pauciflora*), etc. The characteristics of some local species have not been studied.

Several studies were conducted by Iswanto *et al.* (2016) regarding the physical and mechanical properties of three species of *Styrax* wood originating from North Tapanuli, and the chemical composition based on the differences in the growth location of *Styrax sumatrana* was also studied. A study was conducted on the potential of *styrax* resin as an antioxidant and the cinnamic acid content found in the *styrax* resin of North Tapanuli (Hidayat *et al.*, 2018; Kiswandono *et al.*, 2019). In other related studies, Susilowati *et al.* (2018) and Rahmat *et al.* (2017) detailed the genetics of *styrax* grown in North Sumatra, Indonesia. It was reported that the *styrax* plant can be used for other purposes, such as medicine, perfume, cosmetics, etc.

The Raru plants are the other plant species that can be further studied in the forest areas of North Sumatra. Based on the available information, particularly of the Central Tapanuli area, there are three species of Raru producing plants (Susilowati *et al.*, 2020; Elfiati *et al.*, 2019). They include Dahanon (*Cotylelobium lanceolatum*), Songal (*Vatica pauciflora*), and Pulut (*Cotylelobium melanoxyton*). Hildebrand (1954) showed that there

are several species of wood classified as raru producing plants, including *Shorea maxwelliana* King, *Vatica songa* V.Sl. from the Dipterocarpaceae, and *Garcinia sp.* from the Guttifera family.

Generally, the bark of raru wood is used to improve the taste and the alcohol content of Batakese traditional toddy beverage (Susilowati *et al.*, 2019), anti-hyperuricemic, and antioxidant (Sinaga *et al.*, 2020). The wood is suitable for heavy material constructions, such as electricity poles, and in ship-building (Susilowati *et al.*, 2020). Furthermore, it can be used as natural food preservatives due to its high tannin content (Pasaribu, 2007). However, there is limited information related to the basic properties of this wood. The data found were still limited in terms of the physical, mechanical, and chemical properties of Raru *C. Melanoxyton* as reported by Pasaribu (2007) and Pasaribu *et al.* (2007).

As an effort to optimize the use of the wood, information related to the characteristics is required. Several basic properties have an important role in the use of wood as raw material. One of which is the chemical component consisting of cellulose, hemicellulose, lignin, and extractives. Henriksson *et al.* (2009) reported that the chemical properties, which include cellulose, extractive, and lignin content, are some of the properties that have an impact on pulp and paper production. Furthermore, the chemical properties are also important and should be considered because they are associated with several uses. In the form of cellulose content, these properties can be a reference for using wood as a bioethanol material (Demirbas, 2005). The chemical properties serve as a reference for the use of wood as a pulp and paper material (Sjöström, 1993). The chemical properties in lignin and cellulose content affect the heating value of wood (Bowyer *et al.*, 2003; Lee *et al.*, 2019). Furthermore, the properties can be considered for any purpose such as the cellulose for capacitor material (Li *et al.*, 2019)

to replace the use of ink on plastics with colored films (Tzeng *et al.*, 2015), and as a raw material for 3D printing ink (Mietner *et al.*, 2021). The lignin can be utilized for raw material in wastewater treatment (Xiao *et al.*, 2019; Maulina *et al.*, 2020; Fatriasari *et al.*, 2020), biomedicine (Siddiqui *et al.*, 2018), biorefinery (Liu *et al.*, 2019; Yang *et al.*, 2019), paint and coating (Zikeli *et al.*, 2019), and packaging (Xing *et al.*, 2019).

This study focused on examining the effect of wood species and the axial position of the stem on its chemical components. These two factors play an important role in determining the diversity of chemical components of wood. The variation of the chemical properties was affected by species differences and log positions. In addition, (González-Rodrigo *et al.*, 2013) reported that phenotypic and genotypic factors cause a high variability in wood properties between different and individuals of the same species. Individual variations are not apparent but resulted from a complex system of various factors involved in modifying the physiological processes of wood formation. Previous studies showed that there are various properties such as a chemical component of lignocellulosic materials including wood. Meanwhile, Lestari *et al.* (2016) reported that the extractive content of platinum teak wood is significantly affected by tree age and axial position in the stem. Pettersen (1984) further claimed that chemical component variations are influenced by wood section (roots, stems, and branches), species, growing location, climate, and soil conditions. The chemical makeup differs from pith to bark, from stump to crown, from earlywood and latewood, and between sapwood and heartwood in each portion of a single tree (Kollmann and Cote, 1984). In addition, genetic factors and tree age also have an impact on the chemical composition of wood (Berrocal *et al.*, 2004). Therefore, an axial study of the chemical properties of Raru wood based on the log positions was conducted. This study aimed to analyze the chemical

composition of three species of Raru wood (*C. lanceolatum*, *C. melanoxylon*, and *V. pauciflora*) originating from Central Tapanuli, North Sumatra, Indonesia based on the axial positions of the logs.

2. MATERIALS and METHODS

2.1. Materials

Raru wood species of *C. lanceolatum*, *V. pauciflora*, and *C. melanoxylon* were collected from Central Tapanuli Regency, North Sumatra Province (1° 11' 00"- 2° 22' 0" north latitude, and 98° 07'- 98° 12' east longitude). They were *C. lanceolatum*, *V. pauciflora*, and *C. melanoxylon* with the criteria of a tree diameter greater than 15 cm. In addition, only one tree was used for each species and was converted into three log sections after been felled. Fig. 1 showed the base, middle, and top of the wood.

2.2. Methods

2.2.1. Samples preparation

The samples were then ground with a ring flaker for over 40 mesh, then oven-dried with a maximum temperature of 40°C. They were then stored in sealed plastic containers for further testing, as shown in Fig. 2. The preparation was based on the Indonesia National

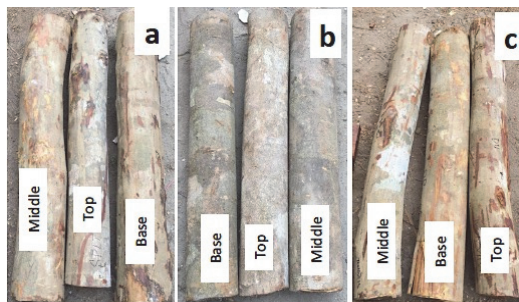


Fig. 1. (a) *C. melanoxylon*, (b) *V. pauciflora*, and (c) *C. lanceolatum* based on log axial direction.



Fig. 2. Samples for chemical analysis of (a) *C. melanoxylon*., (b) *V. pauciflora*, and (c) *C. lanceolatum* based on log axial direction.

Standard (SNI 08-7070: 2005). For chemical component analysis, the moisture content was determined using an oven which was set at a temperature of $105 \pm 3^\circ\text{C}$ for 24 h, and it ranged from 2-4% for lignin analysis, 3-6% for ethanol and benzene, and 1-2% for alpha-cellulose.

2.2.2. Analysis of wood chemical component

The chemical component analysis was performed based on the Indonesian National Standard (SNI). This standard was used to determine the chemical components of wood (Hastuti *et al.*, 2017; Purwita and Sugesty, 2018; Jasni *et al.*, 2016). The analysis was observed following the structural chemical compositions of the cell wall, such as Acid-insoluble lignin (SNI 8429: 2017), alpha-cellulose (SNI 8400: 2017), and holocellulose (ASTM D1104-56). Furthermore, extractives content as a non-structural chemical component was also observed. The extractive in alcohol benzene was performed based on SNI 8401:2017, and its solubility in cold and hot water, as well as 1%

NaOH, was referred to the SNI 01-1305-1989 and SNI 14-1838-1990.

2.2.2.1. Lignin content

In the 50 mL cup, about $1.0 \pm 0.1\text{g}$ free ethanol benzene extracted samples were placed, and 15 mL of 72% sulfuric acid was added and stirred for 2-3 min until fully dispersed. Furthermore, 300 mL of distilled water was added to a 1000 mL Erlenmeyer, and water was applied until the amount reached 575 mL, at which point the sulfuric acid concentration reached 3%. The solution was heated and the temperature was maintained for 4 h with a constant volume. Meanwhile, the deposit was separated and washed with hot water until acid-free, then precipitated in an oven at $105 \pm 3^\circ\text{C}$.

2.2.2.2. Holocellulose content

A 2 g sample of the ethanol-benzene extracted was weighted and washed with hot water before the benzene alcohol solution was removed. It was wetted with cold water (10°C) and placed in a filtering paper containing the sample in the chlorination apparatus and chlorinated for 3 min. The filtering paper was then carefully stirred, and the chlorination was repeated for 2 min before being washed with 95% alcohol to dissolve the excess chlorine gas and HCl. After 1 min, a vacuum pump was used to remove the solution and moistened with cold water. In addition, methanolamine 3% ($\pm 75^\circ\text{C}$) was added and left for 2 min before removing the solution with a vacuum pump. This step was repeated once more, and the residue was washed twice with 95% alcohol and twice with cold water. The solvent was extracted from each washing by using a vacuum pump, and the chlorination was repeated every 2-3 min, followed by two washes with 95% alcohol to extract the alcohol-monoethanol solution. The residue was washed two more times with cold water and 95% alcohol until it was no longer acidic. Furthermore, ether was added to speed up the drying process, and the resi-

due-filled filtering paper was oven dried for 2.5 h at $105 \pm 3^\circ\text{C}$, then cooled in the desiccator and weighed.

2.2.2.3. Alpha-Cellulose Content

The Alpha-Cellulose content was determined by weighing 1.5 ± 0.1 g and placing it in a 300 mL cup glass with 75 mL of 17.5% NaOH solution, which was heated in a $25 \pm 0.2^\circ\text{C}$ until the sample was dispersed. To make a 100 mL complete solution, 25 mL of 17.5% NaOH solution was added. After 30 min of the first NaOH solution addition, 100 mL of aquades was added and left in the hot plate stirrer for 30 min until the total extraction time is approximately 60 min. After that, 25 mL of the filtrate and 10 mL of 0.5 N potassium dichromate solution were put into a 250 mL flask, then 50 mL of concentrated H_2SO_4 was added. The solution was left for 15 min in the hot plate stirrer. Furthermore, 50 mL of aquades was added and left to cool, and 2-4 drops of pheromoin indicator were titrated with 0.1 N ferro ammonium sulfate solution until purple. The filtrate was replaced with 12.5 mL of 17.5% NaOH solution and 12.5 mL of aquades for the blank titration.

2.2.2.4. Extractives content in ethanol-benzene of 1:2

About 1 ± 0.1 g of sample powder (40-60 mesh) was weighed in a petri dish, and then samples were put in the filtering paper, covered and tied. The soluble fraction in ethanol benzene of samples was then calculated. Following that, the samples were put in a 250 mL soxhlet apparatus (the initial weight was determined), and 150-200 mL of a 1:2 alcohol-benzene mixture was applied. This apparatus was connected to the cooler and extracted in a water bath for 6 h, with five circulations per hour. Furthermore, the filtering paper containing samples was removed from the apparatus, and the extraction solution was steamed until nearly dry. The residual evaporation was heated in pumpkin extraction at 105°C for 3 h or more.

2.2.2.5. Hot water extractive content

A total of 2.0 ± 0.1 g of oven-dried samples were inserted into the 200 mL Erlenmeyer flask and 100 mL water was added before placing the cooler upright. In addition, the sample-containing Erlenmeyer was placed in a water bath containing boiling water for 3 h, with the surface in the bath set to be higher than that in the Erlenmeyer. The sample was filtered with a mouthpiece filtering paper considered to weigh after Erlenmeyer was removed from the water bath. Furthermore, the filtrate was then washed with hot water until it was clear, and the residue was then dried in a $105 \pm 3^\circ\text{C}$ oven for 4 h before being cooled in a desiccator and measured.

2.2.2.6. Cold Water extractive content

A total of 2.0 ± 0.1 g of oven-dried samples was inserted into a 400 mL cup glass and added 300 mL of water. The solution was stirred and placed at a room temperature of $23 \pm 2^\circ\text{C}$ for 48 h. Also, the sample was filtered with a masir funnel and washed with cold water until the filtrate was clear. The funnel containing the sample residue was oven-dried at a temperature of $105 \pm 3^\circ\text{C}$ for 4 h then cooled in a desiccator and weighed.

2.2.2.7. Extractives content in the 1% NaOH

2.0 ± 0.1 g of oven-dried sample was transferred into a 200 mL cup glass, and 100 mL of 1% NaOH solution was added and stirred in a closed glass cup. For 60 min, it was put in a water bath containing boiling water and positioned where water in the handler was higher than the surface of the NaOH solution in the cup glass. The sample was filtered with a mouthpiece filtering paper that was known to weigh after the cup glass containing the sample was removed from the water bath. Furthermore, it was washed with hot water until the filtrate was colorless. 25 mL of 10% acetic acid was applied and allowed to stand for 1 min before re-filtration, and the items used were washed

in hot water until they were acid-free. The glass containing the sample was removed from the water bath, and the sample was filtered with a mouthpiece filtering paper used for weighing. The funnel containing the sample residue was dried at $105 \pm 3^\circ\text{C}$ for 4 h before being cooled in a desiccator and weighed.

2.3. Statistical Analysis

A completely factorial randomized design consisting of factors A and B was used. Factor A includes the three species of wood (*C. lanceolatum*, *V. pauciflora*, and *C. melanoxylon*), and factor B includes the three positions of the stem in the axial direction (base, middle, and top). The sample analysis for the chemical component was carried out in duplo for each treatment. Therefore, the total sample analyzed was 18 units. Analysis of Variance (ANOVA) test at a 95% confidence interval was conducted to determine whether this treatment provided a real difference or not to the chemical component parameters tested.

3. Results and Discussion

3.1. Structural Chemical Composition

The average value of structural chemical composition was presented in Table 1.

3.1.1. Lignin

The lignin content based on the axial direction ranges from 22.77 (**1.09**) - 31.25 (**0.21**)%, and the highest was located in the middle part of *V. pauciflora* wood, while the lowest was found in the top part of *C. melanoxylon* wood. The highest and lowest lignin content of all wood species was located in the middle and top part of the log. This result was contrary to a previous study conducted by Rahman *et al.* (2018) concerning lignin content in Kelampayan wood. It was reported that the highest lignin content was located in the top position because it was dominated by juvenile wood. Furthermore, raru wood was not significantly different concerning specific gravity at the base (1.07), middle (0.97), and top (0.93). The three positions are included in the category of high specific gravity since the phenomenon of the influence of juvenile wood on kelampayan as proposed by Rahman *et al.* (2018) does not occur in this raru wood.

Pasaribu (2007) showed that the specific gravity of *C. melanoxylon* wood at the three log's position of the base, middle, and top trunk ranged from 1.02 to 1.09. The highest and lowest value was located in the base and then the top respectively.

All parts of *V. pauciflora* contain lignin, which showed that the wood has better strength than the other species. Lignin contributes to the stiffness of cell walls (Gindl *et al.*, 2002), and Fagerstedt *et al.* (2015) stated

Table 1. Structural chemical composition of Raru wood

Wood Species	Log Position	Lignin	Holocellulose	Alpha Cellulose	Hemicellulose
<i>C. lanceolatum</i>	Base	28.78 (0.68)	61.78 (1.10)	42.44 (0.08)	19.34 (0.48)
	Middle	29.62 (1.73)	61.03 (0.04)	41.6 (0.85)	19.43 (0.47)
	Top	27.66 (0.65)	61.03 (0.04)	41.62 (0.74)	19.62 (0.45)
<i>V. pauciflora</i>	Base	31.13 (0.47)	62.24 (0.34)	41.3 (0.68)	20.94 (0.62)
	Middle	31.25 (0.21)	67.34 (0.48)	44.26 (0.51)	23.08 (0.37)
	Top	27.97 (0.81)	69.00 (0.85)	43.29 (0.41)	25.71 (0.44)
<i>C. melanoxylon</i>	Base	25.23 (0.33)	62.60 (0.85)	40.60 (1.56)	22.00 (0.28)
	Middle	26.30 (0.42)	67.20 (0.28)	44.99 (1.12)	22.21 (0.30)
	Top	22.77 (1.09)	59.59 (0.83)	40.46 (1.50)	19.13 (0.18)

that it provides resistance to the comprehensive force of structures.

Referring to the classification introduced by the Ministry of Environment and Forestry (2020), the lignin content was classified into the moderate category. Therefore, the three wood species can still be used as raw materials for pulp and paper. According to Martawijaya *et al.* (2005), the lignin content in wood commonly ranges from 18-33%, and a high content is not desirable in wood utilization for pulp and paper and bioenergy. This is because it needs to increase the pulp quality or accessibility of enzymes into cellulose. Lignin is the second-largest substance and one of the main chemical components in plants (Casey, 1980). It functions as a binding agent between cells and provides rigidity Ray *et al.* (2009). According to Terashima *et al.* (2009), lignin plays an essential role in the structural assembly of a cell wall.

The lignin content of these three wood species is almost the same as that of *Acacia mangium*, which ranges from 28-32% (Siagian *et al.*, 2009), *Shorea retusa*, *Shorea macroptera*, and *Eucalyptus pelita* contents range from 26-31%, 24-31% (Yunanta *et al.*, 2014; Lukmandaru *et al.*, 2016), and 22-36% (Fatimah *et al.*, 2013) respectively. They have a higher content when compared to *Styrax sumatrana*, *Ceiba pentandra*, and *Ochroma pyramidale* (Iswanto *et al.*, 2019; Purnamawati *et al.*, 2018). When compared with several subtropical wood species such as Larch, Red pine, Korean pine, Cypress, and Cedar as reported by Park *et al.* (2017), the lignin content of the three species of Raru wood is almost similar.

The analysis of variance (ANOVA) showed that the interaction between wood species and log position does not significantly affect the 95% confidence interval on lignin content. However, for a single wood species factor, it has a significantly different effect.

3.1.2. Holocellulose

The holocellulose content of the three observed wood species with an axial direction of log ranged from 59.59 (**0.83**) to 69.00 (**0.85**)%. Meanwhile, the highest holocellulose content was found in the top part of *V. pauciflora* wood, and the lowest was seen in the bottom part of *C. melanoxyton*. According to Fengel and Wegener (1984), a small percentage of lignin remains in holocellulose. This is because there is an ultrastructural and chemical relationship between cell wall macromolecules, such as holocellulose and lignin.

This is slightly different from the result obtained from the study conducted by Rahman *et al.* (2018). It was reported that kelampayan wood, one of the fastest-growing species, had a holocellulose content ranging from 68 to 70%. Most of the cells are dominated by juvenile wood, which has a higher cellulose content. Furthermore, high cellulose affects the high holocellulose content, and the three Raru wood species were not classified as fast-growing wood due to their high specific gravity value.

Based on the species variety, *C. melanoxyton* wood has the highest holocellulose content in all positions. Therefore, it is predicted to have a higher pulp yield than *V. pauciflora* and *C. lanceolatum*. Most of the three Raru wood species contain high holocellulose content (> 65%), which produces good pulp characteristics and a high yield pulp (Fatriasari and Hermiati, 2008). A high proportion of polysaccharides showed a high holocellulose content, which will increase the pulp yield (MacLeod, 2007).

The analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on holocellulose content.

3.1.3. Alpha-cellulose

Material with a higher alpha-cellulose content has high-quality raw material (Sumada *et al.*, 2011). The

cellulose content of Raru wood ranges from 40.46 (1.50) - 44.99 (1.12)%. This is consistent with Pettersen (1984), stating that the cellulose content of dry wood weight ranges from 40-50%. Generally, the wood growth environment contributes to variation of wood's chemical composition, including cellulose (Fatriasari *et al.*, 2019).

The higher content of cellulose is found in the middle position. The highest and the lowest alpha-cellulose contents are located in the middle and the top position of *C. melanoxyton* wood. Meanwhile, the purity is often expressed in the alpha-cellulose content. A high cellulose content showed strong fiber, relatively resistant in separation and purification to chemicals. It is also insoluble in neutral organic solvents and water (Casey, 1980). High alpha-cellulose such as cellulose nitrite, carboxyl methylcellulose, and cellulose xanthate is needed in making filter paper (Fengel and Wegener, 1984).

The three Raru wood species have almost similar cellulose contents and are presumably affected by the high specific gravity category (> 0.8). They have almost the same proportion of mature wood since the cellulose content does not differ. Furthermore, Ona *et al.* (1998) reported that the specific gravity of wood is positively correlated with cellulose content. The Indonesian wood classification assessment for pulp and paper material Ministry of Environment and Forestry (2020) showed that the cellulose content of Raru wood can be categorized into medium class.

The analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on alpha-cellulose content.

3.1.4. Hemicellulose

The hemicellulose content of the three Raru wood species ranges from 19.13 (0.18) - 25.71 (0.44)%. Meanwhile, the highest and the lowest hemicellulose

contents were found on the top log of *V. pauciflora* and *C. melanoxyton* respectively. This value was slightly lower than the hemicellulose content of *Styrax sumatrana* wood. Iswanto *et al.* (2019) also reported that *Styrax sumatrana* wood has a hemicellulose content ranging from 26-29%. Syafii and Siregar (2006) reported that a hemicellulose content with good criteria for pulp and paper raw materials ranges from 15 - 25%. A high hemicellulose content has an essential role in milling the pulp (Stephenson, 1951). It creates more flexible fiber, and it is critical to the fibrillation process. Besides, the fiber would be more plastic because of higher water absorption. This condition causes a high surface area formation during pulp formation Fatriasari and Hermiati (2008). This is due to the gelatin properties of hemicellulose, which facilitates the hydrophilic character of the pulp and facilitates the formation of bonds between fibers.

The analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on hemicellulose content.

3.2. Non-Structural Chemical Composition

The average value of non-structural chemical composition was presented in Table 2.

3.2.1. Extractive Substances in Alcohol Benzene 1:2

The extractive substances are dissolved in both polar and non-polar solvents such as an alcohol-benzene solution of 1: 2. *C. melanoxyton* had the highest value in alcohol-benzene of 1:2 on the base, middle, and top positions with 14.18 (0.25)%, 14.92 (0.88)%, and 14.51 (0.58)% respectively. These values increased up to 5%, which can be included in high solubilization. In addition, it is less suitable to form good pulp properties (Fatriasari *et al.*, 2015) based on the assessment

Table 2. Non-structural chemical composition of Raru wood

Wood Species	Log Position	Alcohol Benzene	1%-NaOH	Hot Water	Cold Water
<i>C. lanceolatum</i>	Base	10.07 (0.10)	28.16 (0.23)	6.57 (0.66)	3.13 (0.18)
	Middle	9.84 (0.91)	26.87 (0.95)	7.57 (0.66)	2.52 (0.45)
	Top	11.84 (0.91)	27.84 (0.91)	8.13 (0.18)	4.39 (0.41)
<i>V. pauciflora</i>	Base	9.67 (0.81)	28.33 (0.47)	7.12 (0.17)	4.00 (0.28)
	Middle	9.40 (0.57)	29.71 (0.58)	7.87 (0.52)	4.12 (0.17)
	Top	13.79 (0.98)	27.76 (0.65)	7.72 (0.45)	3.39 (0.41)
<i>C. melanoxylon</i>	Base	14.18 (0.25)	29.76 (0.65)	7.62 (0.45)	2.48 (0.54)
	Middle	14.92 (0.88)	27.67 (0.66)	8.15 (0.21)	5.27 (0.38)
	Top	14.51 (0.58)	27.22 (0.31)	7.70 (0.28)	6.05 (0.11)

of Indonesian wood classification for pulp and paper material (Ministry of Environment and Forestry, 2020). The extractive solubility in alcohol-benzene of 1:2 also significantly influences the wood color. Lukmandaru (2016) showed a higher extractive content in alcohol-benzene produces lower brightness of wood, which means the heartwood will be darker. *C. melanoxylon* wood has a higher extractive content and produces the darkest color compared to others. On the contrary, this high value in *C. melanoxylon* wood showed it is more difficult to be converted for pulp. This value needs a high amount of chemicals for the cooking process in the pulping process (Fatriasari and Hermiati, 2008).

The analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on alcohol benzene 1:2 content.

3.2.2. Extractive Substances in Hot and Cold Water

Pari *et al.* (2006); Fengel and Wegener (1984) reported that tannins, gums, sugar, and coloring agents can be dissolved in the extraction using hot and cold water, while starches can only be solubilized by hot water.

Extractive solubility in hot and cold water ranged

from 6.57 (**0.66**) to 8.15 (**0.21**)% and 2.48 (**0.54**) to 6.05 (**0.11**)% respectively. The highest and the lowest values in hot water were found in *C. melanoxylon* and *C. lanceolatum* wood respectively. Furthermore, the axial direction showed that the top and the base positions of *C. melanoxylon* Raru wood log have the highest and the lowest extractive content values dissolved in cold water. Generally, there was no difference in extractive content values based on the axial position of the log since hot water is a more solubilize extractive substance than cold water. The partial hydrolysis of hemicellulose that involves the development of an organic acid solution into the water may cause it (Dockzekalska *et al.*, 2010). Also, starch solubilization in hot water extraction contributes to the higher values of the content. This is caused by partial hydrolysis of hemicellulose, which involves developing an organic acid solution into water.

The analysis of variance (ANOVA) showed that the interaction between wood species and log position does not significantly affect the 95% confidence interval. However, the log position has a significantly different effect on the extractive content that dissolves in hot water. For the cold water, the analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on extractive con-

tent that dissolves in cold water.

3.2.3. Extractive Substances in NaOH 1%

The extractives solubility in 1%-NaOH ranged from 26.87 (**0.95**) to 29.76 (**0.65**)%. In the base position of *C. melanoxyton* wood, the solubility was higher than the others. Furthermore, the 1%-NaOH solubility showed the dissolving of wood components in low molecular weight of carbohydrates, tannin, and dyes. The high extractive substances in 1%-NaOH solvent indicated that the wood was dominated by inorganic minerals, carbohydrates, fats, oils, dyes, and aromatic compounds (Pari *et al.*, 2006). In pulping process, the extractive substances are expected to be low due to decreased chemical consumption. A high extractive content should be able to produce pitch in the paper sheet.

The analysis of variance (ANOVA) showed that the interaction of wood species with log position has a significantly different effect at a 95% confidence interval on extractive content that dissolves in NaOH.

4. CONCLUSION

- The mean values of chemical components (α -cellulose, hemicellulose, and lignin) for *C. lanceolatum* species were 41.88%, 19.39%, and 28.68%, respectively. For *C. Melanoxyton* and *V. pauciflora* the values were (42.01%, 21.11%, 24.76%) and (42.95%, 23.24%, 30.11%) respectively. Furthermore, the average values of the extractive contents (the solubility in 1:2 ethanol benzene, NaOH, and hot water) for *C. lanceolatum*, *C. melanoxyton*, and *V. pauciflora* wood were (10.58%, 27.62%, 8.13%), (14.54%, 28.22%, 7.82%), and (10.95%, 28.60%, 7.57%) respectively.
- Wood species significantly affect the chemical components such as lignin, cellulose, and hemicellulose, and extractive solubility in cold water. Furthermore,

the axial log position has a significant effect on the chemical component tested.

- *V. pauciflora* wood had the highest cellulose, hemicellulose, and lignin content compared to the other species. The highest extractives solubility in alcohol-benzene, cold and hot water were found in *C. melanoxyton* wood. Meanwhile, the fractionation of structural component of high chemical in raru woods brings more wide range future application such as biorefinery products.

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