Food Science of Animal Resources

Food Sci. Anim. Resour. 2021 May 41(3):428~439 DOI https://doi.org/10.5851/kosfa.2021.e8





Evaluation of Quality Characteristics of Low-Nitrite Pork Sausages with Paprika Oleoresin Solution during Refrigerated Storage

Geon Ho Kim and Koo Bok Chin*

Department of Animal Science, Chonnam National University, Gwangju 61186, Korea



Received December 10, 2020
Revised February 1, 2021
Accepted February 23, 2021

*Corresponding author: Koo Bok Chin Department of Animal Science, Chonnam National University, Gwangju 61186, Korea Tel: +82,62,530,2121

Tel: +82-62-530-2121 Fax: +82-62-530-2129 E-mail: kbchin@chonnam.ac.kr

*ORCID

Geon Ho Kim https://orcid.org/0000-0002-4700-4485 Koo Bok Chin https://orcid.org/0000-0002-8062-6331 **Abstract** The objective of this study was to evaluate quality characteristics of pork emulsified-sausage (ES) containing paprika oleoresin solution (POS) as a replacement for sodium nitrite (NaNO2) during refrigerated storage. ESs were prepared with four treatments: 1) REF, 150 ppm NaNO₂; 2) CTL, 75 ppm NaNO₂; 3) TRT1, 75 ppm NaNO₂+0.1% POS (1% paprika oleoresin+99% sunflower seed oil); and 4) TRT2, 75 ppm NaNO₂+0.1% POS (5% paprika oleoresin+95% sunflower seed oil). The addition of POS into ES increased redness and yellowness but decreased lightness (p<0.05). TRT1 and TRT2 had higher redness and yellowness than CTL (p<0.05). TRT1 and TRT2 had lower total plate counts (Log CFU/g) than CTL due to antimicrobial activity of POS, regardless of its levels (p<0.05). Residual nitrite decreased with increasing storage time for all treatments. TRT2 had lower residual nitrite due to nitrite scavenging activity of POS (p<0.05). CTL had the highest thiobarbituric acid reactant substances (TBARS) among all treatments during storage. The addition of POS into ES showed nitrite scavenging activity during refrigerated storage. In conclusion, antimicrobial and antioxidant activities of the ES with a combination of POS and 75 ppm NaNO2 were similar to those of REF (150 ppm NaNO2), and improved color development of redness value. Therefore, the addition of POS could decrease the amount of nitrite in ESs, leading to healthier meat products.

Keywords paprika oleoresin, sodium nitrite, low-nitrite, emulsified-sausage

Introduction

Sodium nitrite (NaNO₂), one of important ingredients in the manufacture of meat products, has many functions. It can react with myoglobin in meat products, resulting in the production of nitrosohemochrome, a pink pigment (Pearson and Gillett, 1996). Since meat color is an important factor for consumers to purchase the meat products, the role of sodium nitrite for the development of cured color cannot be ignored. In addition, nitrite can enhance the shelf-life by inhibiting microbial growth and toxin production of *Clostridium botulinum*. It also imparts a unique flavor of meat products (Christiansen et al., 1973; Hung et al., 2016). However, nitrite might not be completely

depleted when it develops color. If it remains in a meat product during storage, N-nitrosamine, a carcinogen that can bind to amino acid and secondary amine during processing, might be generated (Hawksworth and Hill, 1971). Due to the risk of potential production of nitrosoamine from nitrites, typical cured meat products, such as ham and sausage, have been classified as first-class carcinogens designated by the World Health Organization in 2015. Consumers have a negative perception of meat products containing nitrite. Therefore, functional ingredients that can replace nitrite are needed for manufacturing meat products that health-conscious consumers would prefer. There have been a lot of studies on natural pigments, such as cactus pigment (Kang and Lee, 2008), red beet (Jeong et al., 2010), and Hongkuk (Liu et al., 2010) for the development of sausages with low levels of nitrite.

Paprika (*Capsicum annuum* var. *angulosum*) contains a high amount of antioxidant substances such as carotenoid, a natural antioxidant that can prevent cancer, cell damage, and coronary artery disease (Dragsted et al., 1993). Paprika has a natural color (red, purple, or yellow). It contains a large amount of carotenoids that can be used in food, medicine, and food industry (Rascón et al., 2011). It has been reported that capsanthin increased redness value of meat products (Bázan-Lugo et al., 2012). Thus, paprika is considered as a natural coloring agent to replace nitrite because it can increase the redness of meat products and prolong storage of meat products.

Paprika oleoresin that gives pigment to paprika extract is produced to improve the coloring effect of such paprika. It is processed in the form of oleoresin to preserve the color, taste, and flavor of paprika, which can be changed by heat and pH changes, after extracting paprika fruit with an organic solvent. Color stability of paprika oleoresin is higher than that of paprika fruit (Lee et al., 2002). Paprika oleoresin also contains a large amount of carotenoids. It has been widely used in the food industry, especially spices, soups, and meat products (Mínguez-Mosquera and Pérez-Gálvez, 1998; Yusop et al., 2012). The addition of paprika oleoresin to meat products might replace sodium nitrite for color development because it has higher color stability. In addition, paprika pigment is considered to be able to improve shelf-life of meat products through its antioxidant activity. However, the paprika oleoresin might be evaluated to reduce sodium nitrite in pork sausage by improving both redness and shelf-life. Thus, the objective of this study was to determine quality characteristics of pork ES added with paprika oleoresin and propose appropriate levels of paprika oleoresin to be added in pork ES for developing heathier meat products.

Materials and Methods

Materials

Raw pork ham lean and back fat were purchased from a local meat market (Hyundai Retail Meat Market, Gwangju, Korea). After the external fat and connective tissue were removed from pork ham, and it was ground with a meat chopper (M-12S, Korea Fuji Kogyo, Busan, Korea) and stored frozen at –50°C until used for sausage manufacturing. The frozen ground raw meat and back fat were thawed and refrigerated at 4°C for 24 h before sausages manufacturing. Paprika oleoresin was provided by Kalsec (Kalamanzoo, MI, USA). Sunflower seed oil was purchased from Sajo (Seoul, Korea) and used for preparing diluted paprika oleoresin solution (POS; 20 times and 100 times).

Experimental design

Formulation of pork sausages is listed in Table 1. In this experiment, POS was prepared by diluting paprika oleoresin and sunflower seed oil to avoid excessive increase of redness and yellowness from undiluted paprika oleoresin. Sunflower seed oil was extracted using organic solvents similar to paprika oleoresin (Salgin et al., 2006). It had no trans-fatty acids.

Table 1. The formulation for manufacturing pork emulsified-sausages added with different contents of oleoresin paprika solution

In anodianta (0/)	Treatment				
Ingredients (%)	REF	CTL	TRT1	TRT2	
Lean meat	60.0	60.0	60.0	60.0	
Fat	20.0	20.0	20.0	20.0	
Water	18.0	18.0	18.0	18.0	
Non meat ingredients	1.97	1.96	2.06	2.06	
Salt	1.50	1.50	1.50	1.50	
Sodium tripolyphosphate	0.40	0.40	0.40	0.40	
Sodium erythorbate	0.05	0.05	0.05	0.05	
Sodium nitrite	0.015	0.0075	0.0075	0.0075	
Paprika oleoresin solution	0.00	0.00	0.10	0.10	
Paprika oleoresin	0.00	0.00	0.001	0.005	
Sunflower seed oil	0.00	0.00	0.099	0.095	
Total	100.0	100.0	100.1	100.1	

However, it had linolenic acid, linoleic acid, unsaturated fatty acids, vitamin A, and vitamin D (Lee and Park, 2010). Pork ESs were prepared for the following four treatments: 1) REF, 150 ppm NaNO₂; 2) CTL, 75 ppm NaNO₂; 3) TRT1, 75 ppm NaNO₂ and 0.1% POS (1% paprika oleoresin+99% sunflower seed oil); and 4) TRT2, with 75 ppm NaNO₂ and POS 0.1% (5% paprika oleoresin+95% sunflower seed oil). Physicochemical chracteristics, textural properties, antioxidant activities, and antimicrobial activities of pork emulsified-sausages (ESs) at 0, 3, 7, 14, 21, 28, and 35 days after manufacture were then determined.

Manufacture of sausages

Ground raw meat was mixed with an ice water for 30 sec using a hood mixer (Mixer, HMC-401, Hanil Electric, Seoul, Korea). The mixture and curing agents were then mixed with an ice water for 1 min. After the addition of curing agents, back fat and POS were added. The mixture was then emulsified for 1 min. Finally, the meat batter was ground for 1 min. Stuffed meat batter in 50 mL conical tube (SPL Life Science, Pocheon, Korea) was heated in a water bath (WB-22, Daihan Scientific, Seoul, Korea) at 75°C for 30 min, vacuum packaged, and refrigerated stored at 4°C until analyzed.

pH and color determination

pH values were measured five times for the inside of each treatment using a pH meter (Model 340, Mettler-Toledo, Columbus, OH, USA). pH calibration was performed based on optimum slope measuring by pH 4.01 and 7.00 buffer.

Meat color was measured six times. Lightness (CIE L*), redness (CIE a*), and yellowness (CIE b*) of cross-sections of sausages were measured using a Minolta Color Reader (CR-10, Minolta, Tokyo, Japan). White color plate (CIE L*=94.8, CIE a*=1.0, CIE b*=0.1) was used as the standard.

Microbiological analysis

Homogenized sausage sample (10 g) and double-distilled (dd) water (90 mL) were mixed with a stomacher (BagMixer®

400 CC®, Interscience, Saint-Nom-la-Bretèche, France) for microbial counts. Total plate count (TPC) and violet red bile (VRB) agar plates were used for measuring numbers of total bacteria and Enterobacteriaceae, respectively. After spreading samples onto agar plates in petri dishes, plates were incubated at 37°C in an incubator for 48 h.

Thiobarbituric acid reactive substances (TBARS)

Lipid oxiatation products were measured using the method of Sinnhuber and Yu (1977). Briefly, each sample was prepared by adding 1% of thiobarbituric acid solution (3 mL) to ground sausage (2 g). Then 2.5% of trichloroacetic acid solution (17 mL) was blended with the mixture in a glass tube. The tube was heated at 100°C in a water bath (WB-22, Daihan Scientific, Seoul, Korea) for 30 min. After heating, the supernatant of the heated mixture (5 mL) and 5 mL of chloroform were mixed together for 1 min and centrifuged at 1,660×g (Model VS-5500, Vision Science, Korea) for about 5 min. The supernatant of the sample (3 mL) was blended with petroleum ether (3 mL) for 1 min. The mixture was then centrifuged at 1,660×g for 10 min. The absorbance of the reaction product was determined using a spectrophotometer (Model UV-1601, Shimadzu, Kyoto, Japan) at a wavelength of 532 nm. TBARS value was derived by calculating the measured absorbance with the following equation:

TBARS value (mg of MDA/of sample) =
$$\frac{0. \text{ D. value} \times 9.48}{\text{Sample weight (g)}}$$

The constant 9.48 was obatained from the reaction product of red thiobarbituric acids's sample dilution factor and its absorption coefficient (152,000 M⁻¹ cm⁻¹).

Expressible moisture (EM, %)

For EM, each sample (1.5 g) was prepared by shaping into a rectangular shape. The sample was covered with three pieces of filter paper (Whatman #3, GE Healthcare, Little Chalfont, UK), placed in the bottom of a conical tube, and centrifuged at 1,660×g (VS-5500, Vision Science, Gyeongsan, Korea) for 15 min. After the centrifugation was completed, EM was determined based on changed weight of the sample and the filter paper using the following formula:

Expressible moisture (%) =
$$\frac{\text{Expressible water weight of filter paper (g)}}{\text{Sample weight (g)}} \times 100$$

Proximate composition

Proximate composition was performed according to methods of the Association of Offical Analytical Chemists (AOAC, 2000). Moisture content (%) was measured with the dry oven method. Briefly, 2 g of each sample in a paper thimble was dried for 18 h in a dry oven at 102°C and the weight of the samples was measured after drying. Crude fat content (%) was measured with the Soxhlet extraction. Briefly, the paper thimble after moisture drying was placed in Soxhlet's extractor, and fat in the samples was extracted with petroleum ether to measure the weight loss. Crude protein content (%) was determined based on the Kjeldahl method.

Cooking loss

Cooking loss (CL) was expressed in percentage (%) by calculating the weight of sausage before and after heating with the

following formula.

Cooking loss (%) =
$$100 - \left(\frac{\text{Sample weight after heating (g)}}{\text{Sample weight before heating (g)}} \times 100\right)$$

Texture profile analysis

For texture profile analysis, sausage samples with 1.3 cm in height and 1.25 cm in diameter were prepared. Texture properties such as hardness (gf), springiness (mm), gumminess, chewiness, and cohesiveness of each sample were determined with a bellow cross head compression probe (cross speed of 300 mm/min and load cell of 50 kg) using an Instron Universal Machine (Model 3344, Instron, Canton, MA, USA).

Residual nitrite

Residual nitrite content was measured by a modified method of AOAC (2000). Briefly, homogenized sausage sample (5 g) was blended with double-distilled water (300 mL), and heated in a water bath (WB-22, Daihan Scientific, Seoul, Korea) for 1 h at 100°C. After heating, filteration was performed using a funnel and a filter paper (Whatman #2, GE Healthcare, Little Chalfont, UK) to remove sausage grounds. Filtrated sample was diluted with distilled water to make a mixture of 500 mL. Sulfanilamide solution (2.5 mL) was added to 25 mL of this mixture, blended, and incubated at room temperature for 5 min for reactions to occur. Then, N-(1-naphthyl)ethylene dihydrochloride solution (2.5 mL) was added to the reactant, blended, and incubated at room temperature for 15 min. The absorbance of the final reaction product was measured with a spectrophotometer (Model UV-1601, Shimadzu, Kyoto, Japan) at wavelength of 540 nm. A standard curve was obtained by measuring the absorbance of nitrite solution to derive residual amount of nitrite. It was used to determine residual amount of nitrite based on the measured absorbance of the sample.

Statistical analysis

All experiments for this study were carried out in triplicate for each sample of treatments. Data were represented as mean and standard deviation and were analyzed using two-way analysis of variance (ANOVA) and IBM SPSS Statistics 23 (SPSS, Chicago, IL, USA) with Duncan's multiple range test at a significance level of 5% (p<0.05). If a significant interaction between treatments and storage time was observed, means and standard deviations were separated by treatments within each storage time and by storage time within each treatment. When the interaction was not significant (p>0.05), data were pooled to test the primary effect by each main factor.

Results and Discussion

pH and color determination

Table 2 shows results of pH of pork ESs added with POS during refrigerated storage. Since no interaction between treatment and storage time was found (p>0.05), data were pooled by treatment within storage time and storage time within treatment. For pork ESs, their pH values ranged from 6.06 to 6.09, showing no difference among ESs (p>0.05). In a study of Eskandari et al. (2013), pH values of frankfurters added with 0.1% or 1.5% of paprika extract were not different from the pH values of frunfurters without adding paprika extract (p>0.05). Similar to results of their study, our results also showed that the

Table 2. pH and color values of pork emulsified-sausages added with paprika oleoresin solution

Treatments ¹⁾	рН	CIE L*	CIE a*	CIE b*
REF	$6.07{\pm}0.05^{a}$	73.6±0.64°	11.8±0.25 ^b	6.10±0.29°
CTL	$6.06{\pm}0.07^a$	75.8 ± 0.65^a	9.90 ± 0.50^{d}	5.53 ± 0.35^{d}
TRT1	$6.09{\pm}0.03^a$	$74.7{\pm}1.19^b$	$11.1 \pm 0.60^{\circ}$	6.37 ± 0.34^{b}
TRT2	$6.08{\pm}0.05^a$	$73.9 \pm 1.06^{\circ}$	12.1 ± 0.06^a	$6.75{\pm}0.34^a$
Storage time (d)				
0	6.08 ± 0.03^a	74.7 ± 0.89^a	11.1±1.11 ^a	6.19 ± 0.55^{abc}
3	6.10 ± 0.03^a	74.6 ± 1.30^a	11.6 ± 1.08^a	$6.00 \pm 0.50^{\circ}$
7	$6.08{\pm}0.02^a$	$74.6{\pm}1.50^a$	11.6 ± 1.16^a	6.14 ± 0.64^{bc}
14	$6.09{\pm}0.03^a$	74.6 ± 1.43^a	$11.3{\pm}1.30^a$	6.10 ± 0.50^{bc}
21	$6.08{\pm}0.04^a$	74.3 ± 1.43^a	11.2±1.04 ^a	6.15 ± 0.62^{bc}
28	$6.04{\pm}0.08^a$	74.2 ± 1.33^a	11.4 ± 0.87^{a}	$6.32{\pm}0.54^{ab}$
35	$6.05{\pm}0.08^a$	74.5 ± 1.19^a	11.2±0.88a	$6.40{\pm}0.50^a$

¹⁾ Treatments: REF, emulsified-sausage (ES) added with 150 ppm sodium nitrite (NaNO₂); CTL, ES added with 75 ppm NaNO₂; TRT1, ES added with 75 ppm NaNO₂+0.1% paprika oleoresin solution (POS, 1% paprika oleoresin+99% sunflower seed oil); TRT2, ES added with 75 ppm NaNO₂+0.1% POS (5% paprika oleoresin+95% sunflower seed oil).

addition of paprika extract did not affect pH values of meat products (p>0.05).

As shown in Table 2, results of color determination were pooled by treatment within storage time and storage time within treatment since no interaction between treatment and storage time was found (p>0.05). Lightness (L*) values of sausages were higher for the CTL than those of the treatments added with 75 ppm of nitrite alone (p<0.05). However, lightness (L*) values of sausages in REF and TRT2 were not different from each other (p>0.05), although they were lower than those of other treatments (p<0.05). The reason for the difference in L* values among CTL, TRT1, and TRT2 with the same amount of nitrite added might be partially due to a decrease in lightness with the addition of diluted POS. A previous study has reported that 15 or 30 g/kg of paprika powder decreased L* values of dry-cured sausages at 20 mm of meat mincing level (Fernández-López et al., 2002). Redness (a*) values of sausages in the TRT2 added with 75 ppm of nitrite and 0.1% of POS (paprika oleoresin 5% and sunflower seed oil 95%) were higher than those of the REF added with 150 ppm of nitrite alone (p<0.05). The addition of paprika oleoresin resulted in higher redness values (p<0.05). Thus, the addition of paprika oleoresin could increase the redness of pork ES, however, redness values showed no change during the storage time (p>0.05). The addition of paprika powder (0.1%, 0.5%, or 2%) into fresh pork sausages also increased a* values in the study of Martínez et al. (2006). Paprika has a high amount of ketocarotenoids such as capsorubin and capsanthin (Levy et al., 1995). These carotenoids, called red xanthophylls (Minguez-Mosquera et al., 1992), are specific red compounds that can increase the redenss of foods including meat products. Yellowness (b*) values of TRT2 were the highest in the TRT2 among all treatments (p<0.05). The addition of diluted paprika oleoresin also increased yellowness (p<0.05), which showed no change during the storage time (p>0.05). According to a study of Jokanović et al. (2011), the addition of 1% of paprika oleoresin increased the yellowness of marinated chicken breast. Thus, meat products with paprika oleoresin increased their yellowness values, as well.

Microbiological anaylsis

As shown in Table 3, there was no interaction between treatments and storage time in results of microbiological analysis

a-d Means having the same superscripts in the same column are not significantly different (p>0.05).

Table 3. Microbial counts, TBARS, and expressible moisture of emulsified-sausages added with different contents of paprika oleoresin solution

Treatments ¹⁾	Total plate counts	Enterobacteriaceae counts	TBARS ²⁾	EM ³⁾
REF	<2.00 ^b	<2.00 ^a	0.19 ± 0.08^{b}	$20.0{\pm}0.14^a$
CTL	$2.07{\pm}1.18^a$	<2.00 ^a	0.24 ± 0.12^a	$20.3{\pm}0.35^a$
TRT1	<2.00 ^b	<2.00 ^a	0.18 ± 0.07^{b}	18.9 ± 0.57^{ab}
TRT2	<2.00 ^b	<2.00a	0.16 ± 0.06^{b}	18.2 ± 1.06^{b}
Storage time (d)				
0	<2.00°	<2.00°	0.09 ± 0.01^{e}	$17.3 \pm 0.48^{\circ}$
3	<2.00°	<2.00°	0.13 ± 0.02^{e}	18.0 ± 0.89^{c}
7	<2.00°	<2.00°	0.16 ± 0.03^{de}	18.5±2.31°
14	<2.00°	<2.00°	0.20 ± 0.02^{cd}	18.1 ± 1.65^{bc}
21	2.31 ± 1.27^{c}	<2.00°	$0.21 \pm 0.03^{\circ}$	19.7 ± 2.46^{b}
28	$3.53{\pm}0.74^{b}$	2.80 ± 0.86^{b}	0.27 ± 0.07^{b}	21.5 ± 1.84^a
35	4.18 ± 0.81^{a}	$3.72{\pm}0.86^{a}$	$0.31{\pm}0.10^{a}$	22.5±1.05 ^a

¹⁾ Treatments: REF, Emulsified-sausage (ES) added with 150 ppm sodium nitrite (NaNO₂); CTL, ES added with 75 ppm NaNO₂; TRT1, ES added with 75 ppm NaNO₂+0.1% paprika oleoresin solution (POS, 1% paprika oleoresin+99% sunflower seed oil); TRT2, ES added with 75 ppm NaNO₂+0.1% POS (5% paprika oleoresin+95% sunflower seed oil).

(p>0.05). TPC in the CTL were higher than those in other treatment groups (p<0.05). However, TRT1 and TRT2 were lower than those with TPC, indicating that paprika oleoresin might have an antibacterial activity. Salih (2006) reported that oil extract from paprika possessed antimicrobial activity against *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* due to the presence of capsanthin from paprika. In addition, oleoresins including paprika extract at concentration above 5,000 ppm had antimicrobial activity by fracturing bacterial membrane. This mechanism can inhibit the growth of foodborne pathogens such as *Listeria monocytogens*, *Staphylococcus aureus*, *Bacillus cerus*, *Salmonella*, and *Pseudomosnas aeruginosas* (Dussault et al., 2014). However, microbial counts of *Enterobacteriaceae* (VRB) were not changed during storage for any treatment (p>0.05). Both TPC and VRB anlaysis results increased with increasing storage time (p<0.05).

TBARS

Table 3 shows results of TBARS of pork ESs added with POS during storage time. Data were pooled by treatment within storage time and storage time within treatment since no interaction between treatment and storage time was found (p>0.05). Although no difference in the TBARS values were observed among REF, TRT1, and TRT2 (p>0.05), TBARS values of the CTL were the highest among treatments (p<0.05). These results indicated that the addition of paprika oleoresin could inhibit lipid oxidation during storage. Antioxidant activity and lipid oxidation inhibited the ability of paprika as reported by previous studies. *Capsicum annuum* group including paprika contained a great antioxidant activity from rich polyphenol pattern (Marín et al., 2004). When paprika extract is added to meat products, it can be expected to inhibit lipid oxidation. In fact, Shim and Chin (2013) reported that pork patties containing oven-dried paprika powder (0.5% and 1.0%) had lower TBARS values than control without paprika powder. Kim et al. (2013) have also reported that the lipid oxidation of pork

²⁾ TBARS, thiobarbituric acid reactive substances (mg/100 kg).

³⁾ EM, expressible moisture.

a-c Means having the same superscripts in the same column are not significantly different (p>0.05).

patties added with 0.323% of paprika oleoresin for nitrite replacement was lower than those of the control without adding paprika oleoresin during refrigerated storage. Carotenoids, pigments present in paprika, have a radical scavenging ability. They can function as antioxidants by physically trapping singlet oxygen (Kiritsakis and Dugan, 1985). Carotenoids in liposome systems reported to increase antioxidant activity with a decrease of oxygen content (Kiokias and Gordon, 2004). TBARS value started to increase rapdily from day 14 of storage (p<0.05). They continued to increase and tended to be the highest at day 35 of storage (p<0.05).

Expressible moisture (EM, %)

Results of EM are shown in Table 3. There was no interaction between treatments and storage time in results of EM (p>0.05). EMs of sausages in REF and CTL were not significantly different(p>0.05). However, EM values in the TRT2 were higher than those of REF and CTL (p<0.05). During storage, EM (%) showed an increase from day 21 (p<0.05). It continued to increase up to day 35 during storage. Bázan-Lugo et al. (2012) have reported that sausage batter added with more than 1% of paprika powder have higher EM values than those without adding paprika powder. These results were different from our results due to the differences in the structure of paprika powder and oleoresin type.

Proximate composition

Table 4 shows proximate composition. Moisture, fat and protein contents (%) were not different among treatments (p>0.05). The moisture content of ESs ranged from 62.2% to 62.9%, fat content ranged from 18.1% to 18.8%, and the protein content ranged from 13.9% to 14.0%. These results indicated that lower level (0.1%) of POS might not affect the proximate composition of ES. Yusop et al. (2012) have reported that the addition of 1% to 3% of paprika oleoresin did not affect moisture or fat contents of marinated chicken breasts. Therefore, the addition levels of paprika oleoresin at 0.1% might not be enough to change proximate composition.

Table 4. Proximate composition and functionality of pork emulsified-sausages added with paprika oleoresin solution

		Treatments ¹⁾				
	REF	CTL	TRT1	TRT2		
Moisture (%)	62.2±0.66a	62.2±1.10 ^a	62.9±1.45 ^a	62.8±1.27 ^a		
Fat (%)	18.1 ± 0.67^{a}	$18.8{\pm}0.84^{a}$	18.2 ± 0.66^a	18.5 ± 0.79^a		
Protein (%)	$13.9{\pm}0.10^{a}$	13.9 ± 0.17^a	14.0 ± 0.13^a	13.9 ± 0.10^a		
Cooking loss (%)	$1.95{\pm}1.14^{a}$	$1.93{\pm}0.59^a$	$2.13{\pm}1.45^a$	1.90 ± 0.77^{a}		
Hardness (gf)	2,861±733a	$2,666\pm602^{a}$	2,833±632a	$3,030\pm574^{a}$		
Springiness (mm)	5.38 ± 0.90^{a}	5.55 ± 0.88^a	5.34 ± 0.13^a	5.15 ± 0.20^{a}		
Gumminess	24.4 ± 5.56^{a}	21.0±6.01 ^a	19.8±2.75 ^a	22.8±3.11 ^a		
Chewiness	133±9.50 ^a	105 ± 19.8^{a}	108 ± 19.4^{a}	119±19.1a		
Cohesiveness	$0.01{\pm}0.00^{a}$	$0.01{\pm}0.00^a$	$0.01{\pm}0.00^a$	0.01 ± 0.00^a		

¹⁾ Treatments: REF, Emulsified-sausage (ES) added with 150 ppm sodium nitrite (NaNO₂); CTL, ES added with 75 ppm NaNO₂; TRT1, ES added with 75 ppm NaNO₂+0.1% paprika oleoresin solution (POS, 1% paprika oleoresin+99% sunflower seed oil); TRT2, ES added with 75 ppm NaNO₂+0.1% POS (5% paprika oleoresin+95% sunflower seed oil).

^a Means having the same superscripts in the same column are not significantly different (p>0.05).

Cooking loss (CL, %)

CL values of pork ESs added with POS are shown in Table 4. There was no differences in CL among the treatments (p>0.05). Kim and Chin (2018) have also reported that CLs of pork low-fat sausages added with paprika powder (0.05% or 0.1%) were not different from those of the control without the addition of paprika powder. These results indicate that paprika pigment does not affect CL of pork ESs.

Texture profile analaysis

Results of texture profile analysis of pork ESs added with POS are shown in Table 4. There were no differences in all textural properties (hardness, springiness, gumminess, chewiness and cohesiveness) among treatments (p>0.05). Bázan-Lugo et al. (2012) have also detected that there were no differences in hardness value between sausage added with 1% of paprika powder and control (0% of paprika powder). However, treatment added with 2% of paprika powder had a higher hardness than control and 1% treatment. It indicates that the ingredients of paprika changed texture of sausage due to retain the water levels during the heating. Thus, the addition of 0.1% of POS into ESs might not be enough to change texture properties of those in our study.

Residual nitrite

Table 5 shows residual nitrite levels in pork ESs added with POS. As expected, residual nitrite levels (ppm) in the REF were the highest among all treatments (p<0.05), and those in the CTL, TRT1, and TRT2 showed no differences until 21 days (p>0.05). Residual nitrite levels in the TRT2 at 28 and 35 days were lower than those of the CTL (p<0.05), indicating that the paprika oleoresin could reduce residual nitrite levels in the TRT2. Jeong et al. (2006) have reported that paprika with higher antioxidant capacity possess higher nitrite scavenge activity. Paprika contains phenolic compounds, ascorbic acid, flavonoid, and carotenoid pigments including capsanthin, capsorubin, and cryptocapsin (Baliga et al., 2003; Zhang and Hamauzu, 2003). Since they are effective in scavengering free radicals, they could inhibit oxidation during storage due to their antioxidant activity. These results indicate that antioxidants in paprika oleoresin can scavenge residual nitrite, thus decreasing residual nitrite levels.

Conclusion

The addition of 0.1% POS into ES can increase the redness and yellowness values, but decrease the lightness. TPCs in

Table 5. Residual nitrite of pork emulsified-sausages added with paprika oleoresin solution

Treatments ¹⁾ –	Storage time (d)						
	0	3	7	14	21	28	35
REF	22.1±2.09 ^{aA}	19.4±2.31 ^{bA}	16.7±2.10 ^{cA}	$9.88{\pm}0.82^{\rm dA}$	6.23 ± 2.48^{eA}	$4.64{\pm}0.48^{eA}$	4.25 ± 0.46^{eA}
CTL	$10.8{\pm}1.37^{aB}$	$10.4{\pm}1.10^{abB}$	$8.70{\pm}1.78^{bB}$	$4.91{\pm}1.27^{cB}$	$4.16{\pm}1.54^{cdAB}$	$2.29{\pm}0.51^{\rm dB}$	2.17 ± 0.43^{dB}
TRT1	$9.84{\pm}0.96^{aB}$	$8.89{\pm}0.65^{abB}$	$7.77{\pm}1.08^{bB}$	$4.29{\pm}0.99^{cB}$	$4.04{\pm}1.61^{cAB}$	$2.22{\pm}0.52^{dBC}$	$2.12{\pm}0.55^{\mathrm{dBC}}$
TRT2	$8.76{\pm}0.46^{aB}$	$8.24{\pm}1.09^{bB}$	7.26 ± 1.16^{bB}	$3.60{\pm}0.98^{cB}$	$2.40{\pm}0.82^{\rm dB}$	$1.84{\pm}0.58^{dC}$	1.75 ± 0.44^{dC}

¹⁾ Treatments: REF, Emulsified-sausage (ES) added with 150 ppm sodium nitrite (NaNO₂); CTL, ES added with 75 ppm NaNO₂; TRT1, ES added with 75 ppm NaNO₂+0.1% paprika oleoresin solution (POS, 1% paprika oleoresin+99% sunflower seed oil); TRT2, ES added with 75 ppm NaNO₂+0.1% POS (5% paprika oleoresin+95% sunflower seed oil).

A-C Means having same superscripts in a same column are not different (p>0.05).

^{a-e} Means having same superscripts in a same row are not different (p>0.05).

TRT1 and TRT2 were lower than those in the CTL. TBARS and residual nitrite values for TRT1 and TRT2 were lower than those of the CTL. Nitrite scavenging activity and antioxidant activity were observed in treatments added with POS, regardless of the concentration of paprika oleoresin. These results indicate that the addition of POS into ES can increase redness and yellowness values, inhibit lipid oxidation and growth of microorganisms, and accelerate nitrite scavenging. Therefore, combination of 75 ppm nitrite level and 0.1% POS could be used to manufacture the healthier sausage than those added with nitrite at 150 ppm.

Conflicts of Interest

The authors declare no potential conflicts of interest.

Author Contributions

Conceptualization: Kim GH, Chin KB. Data curation: Kim GH. Formal analysis: Kim GH, Chin KB. Investigation: Kim GH. Writing - original draft: Kim GH. Writing - review & editing: Kim GH, Chin KB.

Ethics Approval

This article does not require IRB/IACUC approval because there are no human and animal participants.

References

- AOAC. 2000. Official methods of analysis of AOAC International. Vol. 2. 17th ed. AOAC International, Gaithersburg, MD, USA. p 1, 3, 7-8.
- Bázan-Lugo E, García-Martínez I, Alfaro-Rodríguez RH, Totosaus A. 2012. Color compensation in nitrite-reduced meat batters incorporating paprika or tomato paste. J Sci Food Agric 92:1627-1632.
- Baliga MS, Jagetia GC, Rao SK, Kiran Babu K. 2003. Evaluation of nitric oxide scavenging activity of certain spices *in vitro*: A preliminary study. Food Nahrung 47:261-264.
- Christiansen LN, Johnston RW, Kautter DA, Howard JW, Aunan WJ. 1973. Effect of nitrite and nitrate on toxin production by *Clostridium botulinum* and on nitrosamine formation in perishable canned comminuted cured meat. Appl Microbiol 25:357-362.
- Dragsted LO, Strube M, Larsen JC. 1993. Cancer-protective factors in fruits and vegetables: Biochemical and biological background. Pharmacol Toxicol 72:116-135.
- Dussault D, Vu KD, Lacroix M. 2014. *In vitro* evaluation of antimicrobial activities of various commercial essential oils, oleoresin and pure compounds against food pathogens and application in ham. Meat Sci 96:514-520.
- Eskandari MH, Hosseinpour S, Mesbahi G, Shekarforoush S. 2013. New composite nitrite-free and low-nitrite meat-curing systems using natural colorants. Food Sci Nutr 1:392-401.
- Fernández-López J, Pérez-Alvarez JA, Sayas-Barberá E, López-Santoveña F. 2002. Effect of paprika (*Capsicum annum*) on color of Spanish-type sausages during the resting stage. J Food Sci 67:2410-2414.
- Hawksworth GM, Hill MJ. 1971. Bacteria and the N-nitrosation of secondary amines. Br J Cancer 25:520-526.

- Hung Y, de Kok TM, Verbeke W. 2016. Consumer attitude and purchase intention towards processed meat products with natural compounds and a reduced level of nitrite. Meat Sci 121:119-126.
- Jeong CH, Ko WH, Cho JR, Ahn CG, Shim KH. 2006. Antioxidative and nitrite scavenging activities of Korean paprika according to cultivars. J Agric Life Sci 40:27-34.
- Jeong HJ, Lee HC, Chin KB. 2010. Effect of red beet on quality and color stability of low-fat sausages during refrigerated storage. Korean J Food Sci Anim Resour 30:1014-1023.
- Jokanović MR, Džinić NR, Tomović VM, Savatić SB, Tasić TA, Ikonić PM, Šojić BV. 2011. Effect of ground paprika and its oleoresin on marinated chicken breast meat quality. Acta Period Technol 42:55-62.
- Kang JO, Lee SG. 2008. Effects of *Opuntia ficus-indica* pigment and sodium lactate on nitrite-reduced sausages. J Anim Sci Technol 50:551-560.
- Kim GH, Chin KB. 2018. Physicochemical and textural properties of low-fat pork sausages with paprika powder. J Korean Soc Food Sci Nutr 47:917-925.
- Kim HS, Seong PN, Chang MJ, Kim MH. 2013. Quality characteristics of cold storage pork patty by micro-encapsulation of natural additives. Food Eng Prog 17:355-361.
- Kiokias S, Gordon MH. 2004. Antioxidant properties of carotenoids in vitro and in vivo. Food Rev Int 20:99-121.
- Kiritsakis A, Dugan LR. 1985. Studies in photooxidation of olive oil. J Am Oil Chem Soc 62:892-896.
- Lee JW, Park JW. 2010. Changes of fatty acid composition and oxidation stability of edible oils with frying number of French fried potatoes. J Korean Soc Food Sci Nutr 39:1011-1017.
- Lee SO, Kyung SH, Park KD, Kang HG, Park JS, Lee SK. 2002. A study on detection of residual solvent, ethoxyquin and color stability in oleoresin paprika extracts. App Biol Chem 45:77-83.
- Levy A, Harel S, Palevitch D, Akiri B, Menagem E, Kanner J. 1995. Carotenoid pigments and beta-carotene in paprika fruits (*Capsicum* spp.) with different genotypes. J Agric Food Chem 43:362-366.
- Liu DC, Wu SW, Tan FJ. 2010. Effects of addition of anka rice on the qualities of low-nitrite Chinese sausages. Food Chem 118:245-250.
- Marín A, Ferreres F, Tomás-Barberán FA, Gil MI. 2004. Characterization and quantitation of antioxidant constituents of sweet pepper (*Capsicum annuum* L.). J Agric Food Chem 52:3861-3869.
- Martínez L, Cilla I, Antonio Beltrán J, Roncalés P. 2006. Effect of *Capsicum annuum* (red sweet and cayenne) and *Piper nigrum* (black and white) pepper powders on the shelf life of fresh pork sausages packaged in modified atmosphere. J Food Sci 71:S48-S53.
- Minguez-Mosquera MI, Jaren-Galan M, Garrido-Fernandez J. 1992. Color quality in paprika. J Agric Food Chem 40:2384-2388.
- Mínguez-Mosquera MI, Pérez-Gálvez A. 1998. Color quality in paprika oleoresins. J Agric Food Chem 46:5124-5127.
- Pearson AM, Gillett TA. 1996. Processed meats. Aspen, New York, NY, USA. p 69.
- Rascón MP, Beristain CI, García HS, Salgado MA. 2011. Carotenoid retention and storage stability of spray-dried encapsulated paprika oleoresin using gum Arabic and soy protein isolate as wall materials. LWT-Food Sci Technol 44:549-557.
- Salgın U, Döker O, Çalımlı A. 2006. Extraction of sunflower oil with supercritical CO₂: Experiments and modeling. J Supercrit Fluid 38:326-331.
- Salih AA. 2006. Extraction and identification of oil extract from Capsicum annuum L. fruits and study of its antimicrobial

- activity. J Bas Res (Sci) 32:80-87.
- Sinnhuber RO, Yu TC. 1977. The 2-thiobarbituric acid reaction, an objective measure of the oxidative deterioration occurring in fats and oils. J Oleo Sci 26:259-267.
- Shim YW, Chin KB. 2013. Antioxidant activity of the oven-dried paprika powders with various colors and phycochemical properties and antioxidant activity of pork patty containing various paprika powder. Korean J Food Sci Anim Resour 33:626-632.
- Yusop SM, O'Sullivan MG, Preuß M, Weber H, Kerry JF, Kerry JP. 2012. Assessment of nanoparticle paprika oleoresin on marinating performance and sensory acceptance of poultry meat. LWT-Food Sci Technol 46:349-355.
- Zhang D, Hamauzu Y. 2003. Phenolic compounds, ascorbic acid, carotenoids and antioxidant properties of green, red and yellow bell peppers. J Food Agric Environ 1:22-27.