Genetic Diversity of Hard Ticks (Acari: Ixodidae) in the South and East Regions of Kazakhstan and Northwestern China

Yicheng Yang^{1,2,†} , Jin Tong^{3,†}, Hongyin Ruan^{4,†}, Meihua Yang⁵, Chunli Sang¹, Gang Liu¹, Wurelihazi Hazihan⁶, Bin Xu⁷, Sándor Hornok⁸, Kadyken Rizabek⁹, Kulmanova Gulzhan⁹, Zhiqiang Liu¹⁰, Yuanzhi Wang^{1,*}

¹Department of Basic Medicine, School of Medicine, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ²Emergency Department, Shihezi City People's Hospital, Shihezi City, Xinjiang Uygur Autonomous Region 832000, People's Republic of China; ³Department of Ultrasound, the First Affiliated Hospital of Medical College, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ⁶Department of Stomatology, School of Medicine, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ⁶Department of Forestry, College of Agriculture, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ⁶School of Animal Science and Technology, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ⁶School of Animal Science and Technology, Shihezi University, Shihezi City, Xinjiang Uygur Autonomous Region 832002, People's Republic of China; ⁷National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention, WHO Collaborating Centre for Tropical Diseases, Key Laboratory of Parasitology and Vector Biology of the Chinese Ministry of Health, Shanghai 200025, China; ⁶Department of Parasitology and Zoology, University of Veterinary Medicine, Budapest 1078, Hungary; ⁹Department of Food Engineering, Kazakh National Agrarian University, Almaty Oblast 050010, Kazakhstar; ¹⁰Institute of Veterinary Medicine, Xinjiang Academy of Animal Science, Urumqi City, Xinjiang Uygur Autonomous Region 830000, People's Republic of China

Abstract: To date, there is no report on the genetic diversity of ticks in these regions. A total of 370 representative ticks from the south and east regions of Kazakhstan (SERK) and Xinjiang Uygur Autonomous Region (XUAR) were selected for molecular comparison. A fragment of the mitochondrial cytochrome *c* oxidase subunit I (*cox1*) gene, ranging from 631 bp to 889 bp, was used to analyze genetic diversity among these ticks. Phylogenetic analyses indicated 7 tick species including *Hyalomma asiaticum*, *Hyalomma detritum*, *Hyalomma anatolicum*, *Dermacentor marginatus*, *Rhipicephalus sanguineus*, *Rhipicephalus turanicus* and *Haemaphysalis erinacei* from the SERK clustered together with conspecific ticks from the XUAR. The network diagram of haplotypes showed that i) *Hy. asiaticum* from Almaty and Kyzylorda Oblasts together with that from Yuli County of XUAR constituted haplogroup H-2, and the lineage from Chimkent City of South Kazakhstan was newly evolved; and ii) the *R. turanicus* ticks sampled in Israel, Almaty, South Kazakhstan, Usu City, Ulugqat and Baicheng Counties of XUAR were derivated from an old lineage in Alataw City of XUAR. These findings indicate that: i) *Hy. asiaticum*, *R. turanicus* and *Ha. erinacei* shared genetic similarities between the SERK and XUAR; and ii) *Hy. marginatum* and *D. reticulatus* show differences in their evolution.

Key words: Genetic diversity, hard tick, Kazakhstan, northwestern China

The Republic of Kazakhstan is located in Central Asia between 39°49°–55°49° N and 46°28°–87°18° E, with its western part extending into Eastern Europe. Kazakhstan is the ninth largest country in the world with a total area of 2,727,300 km², also ranking as the world's largest landlocked country [1,2]. It borders Russia, Kyrgyzstan, Turkmenistan, Uzbekistan and China [3]. Xinjiang Uygur Autonomous Region (XUAR), occupies 1/6 of China and borders 8 countries, covering an

* These authors contributed equally to this work.

© 2021, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (https://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. area of approximately 1,660,000 km² [4]. The borderline between the south and east regions of Kazakhstan (SERK) and XUAR is 1,783 km long [5]. The ecological environment, topography, climate and natural landscape are similar both in the SERK and XUAR [6].

The mitochondrial cytochrome *c* oxidase subunit I (*cox1*) gene, also regarded as DNA barcoding, was informative in resolving recent diverging events between closely related species and at the intraspecific level [7]. Recently, 14 tick species, including *Dermacentor nuttalli*, *Dermacentor niveus*, *Dermacentor marginatus*, *Dermacentor silvaru*, *Hyalomma asiaticum*, *Hyalomma detritum*, *Haemaphysalis punctata*, *Haemaphysalis concinna*, *Haemaphysalis erinacei*, *Rhipicephalus turanicus*, *Rhipicephalus sanguineus*, *Argas vespertilionis*, *Argas persicus*, and Ixodes kaiseri,

Received 25 August 2020, revised 17 January 2021, accepted 19 January 2021.
*Corresponding author (wangyuanzhi621@126.com)

have been molecularly characterized in the border regions of XUAR [8]. At the same time, while at least 13 tick species have been identified in Kazakhstan, including *Dermacentor daghestanicus*, *D. marginatus*, *D. niveus*, *Dermacentor reticulatus*, *Hy. asiaticum*, *Hyalomma anatolicum*, *Hyalomma scupense*, *Hyalomma marginatum*, *Haemaphysalis punctata*, *Haemaphysalis sulcata*, *Rhipicephalus turanicus*, *Boophilus calcaratus* and *Ixodes persulcatus* [9, 10], and their molecular characteristics are still unknown. Here we report the genetic diversity of ticks in the SERK, in comparison with ticks from the XUAR.

During 2015-2019, a total of 4,392 hard ticks were collected from 605 domestic animals (287 sheep, 210 cattle, 101 camels, 7 dogs) and 16 hedgehogs at 24 sampling sites belonging to 15 districts of 5 Oblasts (East Kazakhstan, Almaty, Jambyl, South Kazakhstan and Kyzylorda) in the SERK. After morphological identification, 213 representative ticks were selected for further molecular analysis. Meanwhile, 157 representative ticks collected from marbled polecats, sheep and camels between 2015 and 2018 in 8 districts of the XUAR were used for comparison.

The genomic DNA was extracted from the selected 370 ticks individually using TIANamp Genomic DNA Kit (TIANGEN, Beijing, China) according to the manufacturer's instructions. In this study, we methodologically omitted shorter than 450 bp *cox*1 sequences, and chose 631-889 bp fragment to analyze inter- and intra-species genetic diversity. The primers used for PCR were LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO2198 (5'-TAAACITCAGGGTGACCAAAAAATCA-3') [11]. The cycling conditions consisted of an initial 5-min denaturation at 95°C, followed by 37 cycles at 94°C for 40 sec, 55°C for 40 sec, and 72°C for 1 min, with a final extension at 72°C for 10 min.

Sequences of the *cox*1 were compared with GenBank data using BLAST program (http://www.ncbi.nlm.nih.gov/BLAST/) after manual editing. This dataset was resampled 1,000 times to generate bootstrap values. Phylogenetic relationships were inferred using the Maximum Likelihood (ML) method. The best-fitting substitution model was determined with the Akaike Information Criterion using the ML model test implemented in MEGA 7.0 software [12]. Genetic diversity was estimated as haplotype (h) and nucleotide diversity (π) indices with the programme DNAsp v5.10.1 [13]. Median-joining (MJ) networks were generated with the software Network v5.10.1 [14] to display the configuration of haplotypes [15].

Morphological and molecular identification revealed the

presence of 9 tick species in the 5 evaluated southeastern border oblasts of Kazakhstan, including *Hy. asiaticum, Hy. detritum, Hy. anatolicum, Hy. marginatum, D. marginatus, D. reticulatus, R. sanguineus, R. turanicus* and *Ha. erinacei*. Sequence identities ranged from 96.27% to 100% using BLAST analysis of the *cox*1 in intra-species. The sequences in this study were deposited in GenBank (Accession No.: MF002579-MF002581, MK213071, MK307807, MK610453, MN817302, MN841460-MN841462, MN841464, MN853165-MN853167, MN862754, MN865123, MN868560, MN868592, MN892553, MN961479, MT506455, MT664833, KU364300-KU364301, KU364303, KU364307, KX882100, KY996841).

Phylogenetic analysis based on the cox1 (Fig. 1) revealed that: i) Hy. detritum ticks from East Kazakhstan Oblast (MN841460) clustered together with conspecific ticks from Turpan Region of XUAR (KF583581) and Inner Mongolia Province of China (JQ737068); ii) Ha. erinacei ticks from Almaty Oblast, Kazakhstan (MN841464), and Altaw City of XUAR (KU364301) belonged to a different phylogenetic group than ticks from European countries; iii) Hy. anatolicum ticks from Jambyl Oblast (MN853167), Karshgar City of XUAR (KF583576) and Iran (KT920180) represented an ancestral clade to ticks from Iraq, India and Pakistan; iv) D. marginatus ticks from Qapqal County of XUAR (JX051151), East Kazakhstan Oblast (MN868592), and South Kazakhstan Oblast (MN868560) showed a separated branch with ticks from D. marginatus in Alataw City of XUAR (KU364300) with high (100%) bootstrap support; and v) R. sanguineus sensu lato from Alamaty Oblast (MN862754) was more closely related to a specimen from Iran than to ticks from Altaw City of XUAR (KU364307).

Haplotypes is defined as specific combinations of alleles on an individual chromosome [16]. The network diagram of *Hy. asiaticum* showed the haplotypes in the SERK and XUAR based on the *cox*1. The analyses are as follows: i) the H-2 haplotype was the most dominant and the widespread in SERK and XUAR, such as Almaty Oblast (MN892553), Kyzylorda Oblast (MN961479) and Yuli County of XUAR (KF527400); ii) the H-5, H-6, and H-7 haplotypes were originated from the H-2 haplotype; and iii) the H-3 and H-4 haplotypes were furtherly originated from the H-6 haplotype. Interestingly, the H-1 haplotype, being distributed in South Kazakhstan, was derivated from the H-3 haplotype, which was distributed in Shihezi City, Fuhai County and Alataw City of XUAR. Meanwhile, 3 lineages of *R. turanicus* from Almaty and South Kazakhstan Oblast



Fig. 1. Maximum-likelihood (ML; 1,000 bootstrap replicates) phylogenetic tree of the cox1 constructed with MEGA 7.0 software, using the sequences of tick species from SERK (\blacktriangle) and XUAR (\triangle) in this study and the sequences available in the GenBank. The scale bar represented the inferred substitutions per nucleotide site.

(MN853166 and MN841462, H-1 haplotype), XUAR (MF002579-MF002581, H-3 haplotype), and Israel (KF219748, H-4 haplotype), originated from an ancestral lineage from Alataw City of XUAR (KU364303, H-2 haplotype) (Fig. 2). In addition, our study revealed many highly divergent haplotypes for *Hy. asiaticum*, e.g. H-1, H-4, H-7, and H-8, while many highly divergent haplotypes for *R. turanicus*, e.g. H-1, H-3, and H-4, this phenomenon maybe be related to a long and complex evolutionary history of *Hy. asiaticum* and *R. turanicus* in the XUAR and SERK.

Based on ecological feature, *Hy. scupense* is a one-host tick while *Hy. detritum* is a 2-host species, although they are identical in morphological characteristics [17]. Here *Hy. detritum* ticks were all sampled from herbivorous livestock, and phylo-

genetic analyses showed that *Hy. detritum* in East Kazakhstan Oblast (MN841460) shared a common branch with these in Turpan Region (the north of XUAR, KF583581) and Inner Mongolia Province of China (JQ737068), and that *Hy. detritum* ticks in Almaty Oblast (MT664833) were their ancestral lineages.

The brown dog tick *R. sanguineus* (Latreille, 1806), which is generally accepted as the taxonomic baseline for the *R. sanguineus* group, is the most widely distributed tick species globally [18]. At least 2 lineages of *R. sanguineus* seno lato (s.l), namely temperate and tropical lineages, were presented [19]. Here phylogenetic analysis indicated *R. sanguineus* both Almaty Oblast in Kazakhstan (MN862754) and Alataw City of XUAR (KU364307) belonged to *R. sanguineus* s.l temperate



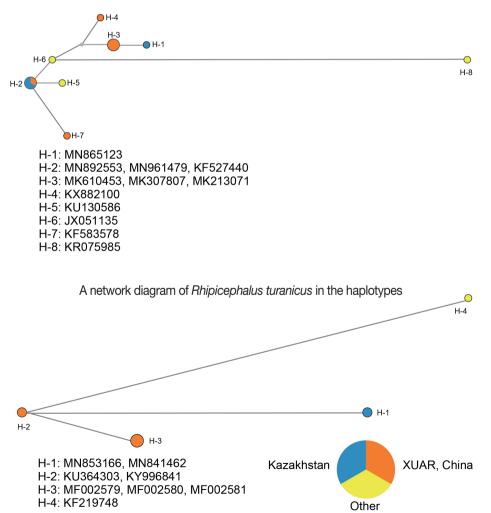


Fig. 2. Network diagrams of *Hyalomma asiaticum* and *Rhipicephalus turanicus* showing the haplotypes in the SERK and XUAR based on the mitochondrial *cox*1 gene.

lineages, which was a separated clade with *R. sanguineus* tropical lineage from India, Australia, Brazil and South Africa.

The mitochondrial cox_1 , as the standard DNA barcode, is the best choice for identification of tick species [7]. In the present study, 7 tick species from the SERK, including Hy. asiaticum, Hy. detritum, Hy. anatolicum, D. marginatus, R. sanguineus, R. turanicus and Ha. erinacei, were clustered together with conspecific ticks from the XUAR. Nevertheless, the mitochondrial lineages were unknown in Hy. marginatus and D. reticulatus ticks from the SERK and XUAR, as for the cox1 data of Hy. marginatum and D. reticulatus from the XUAR were absent. Animal transportation likely contributed to the dispersal of different haplotypes. Movements of wildlife and migratory birds are well-documented in these regions [20], which might contribute to common genetic lineages of tick species. In the future, it is vital to sample more tick species both in the SERK and XUAR, which is helpful to explore their commonness and difference in tick's evolution.

ACKNOWLEDGMENTS

This research was supported in part by the National Key Research & Development Program of China (2018ZX10101002-007 and 2018ZX10101002-003), National Natural Science Foundation of China (81960379 and 31960709), Humanities and Social Sciences Research Projects (18YJCZH220), Highlevel Talent Project of Shihezi University (RCZK2018C04), and the International Cooperation Projects of Xinjiang Uygur Autonomous Region (2020E01008).

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

- Cherednichenko A. Estimating agricultural adaptation to climate change through cloud activation for northern Kazakhstan. In Qi J, Evered KT eds, Environmental Problems of Central Asia and Their Economic, Social and Security Impacts. Tashkent, Uzbekistan. Springer. 2008, pp. 183-190. https://doi.org/10.1007/978-1-4020-8960-2
- Yamanaka H, Murata K, Tabata R, Kawaguchi F, Sasazaki S, Yamamoto Y, Bakhtin M, Kazymbet P, Meldevekob A, Suleimenov MZ, Nishibori M, Mannen H. Kazakhstani Native Cattle Reveal Highly Divergent mtDNA From Bos Taurus and Bos Indicus Lin-

eages With an Absence of Bos Indicus Y Chromosome. Anim Sci J 2019; 90: 29-34. https://doi.org/10.1111/asj.13128

- Bismil'din FB, Shapieva ZZ, Anpilova EN. Current malaria situation in the Republic of Kazakhstan. Med Parazitol (Mosk) 2001; 181: 24-33 (in Russian).
- Zhou M, Cao S, Sevinc, F, Sevinc M, Ceylan O, Moumouni PF, Jirapattharasate C, Liu M, Wang G, Iguchi A, Vudriko P, Suzuki H, Xuan X. Molecular detection and genetic identification of *Babesia bigemina, Theileria annulata, Theileria orientalis* and *Anaplasma marginale* in Turkey. Ticks Tick Borne Dis 2016; 7: 126-134. https://doi.org/10.1016/j.ttbdis.2015.09.008
- Mihailovich SS, Isaevna MG. Land border delimitation of Republic of Kazakhstan: practical knowledge of border policy formation. Eur Sci Rev 2018; 2018: 202-204. https://doi.org/10.29013/ESR-18-7.8-202-204
- Hanapia S, Peng D, Junqi Y, Li C. Species diversity characteristics and conservation status in Xinjiang and Kazakhstan, China. Anhui Agronomy Bull 2017; 23: 46-48 (in China). https://doi. org/10.16377/j.cnki.issn1007-7731.2017.16.014
- Kushimo, Omobolanle. The Tick Genus Amblyomma in Africa: Phylogeny and Mutilocus DNA Barcoding. Electronic Theses and Dissertations. 2013, p 835. https://digitalcommons.georgiasouthern.edu/etd/835
- Sheng J, Jiang M, Yang M, Bo X, Zhao S, Zhang Y, Wureli H, Wang B, Tu C, Wang Y. Tick distribution in border Oblasts of Northwestern China. Ticks Tick Borne Dis 2017; 10: 665-669. https://doi.org/10.1016/j.ttbdis.2019.02.011
- Nurmakhanov T, Sansyzbaev Y, Atshabar B, Deryabin P, Kazakov S, Zholshorinov A, Matzhanova A, Sadvakassova A, Saylaubekuly R, Kyraubaev K, Hay J, Atkinson B, Hewson R. Crimean-Congo haemorrhagic fever virus in Kazakhstan (1948-2013). Int J Infect Dis 2015; 38: 19-23. https://doi.org/10.1016/j.ijid.2015.07.007
- Turebekov N, Abdiyeva K, Yegemberdiyeva R, Dmitrovsky A, Yeraliyeva L, Shapiyeva Z, Amirbekov A, Oradova A, Kachiyeva Z, Ziyadina L, Hoelscher M, Froeschl G, Dobler, G, Zinner J, Frey S, Essbauer S. Prevalence of *Rickettsia* species in ticks including identification of unknown species in two regions in Kazakhstan. Parasit Vectors. 2019; 12: 197. https://doi.org/10.1186/s13071-019-3440-9
- 11. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotechnol 1994; 3: 294-299.
- Tamura K, Nei M. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. Mol Biol Evol 1993; 10: 512-526. https://doi.org/10.1093/oxfordjournals.molbev.a040023
- Librado P, Rozas J. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. Bioinformatics. 2009; 25: 1451-1452. https://doi.org/10.1093/bioinformatics/btp187
- Bandelt HJ, Forster P, Röhl A. Median-joining networks for inferring intraspecific phylogenies. Mol Biol Evol 1999; 16: 37-48. https://doi.org/10.1093/oxfordjournals.molbev.a026036

- Wetjen M, Schmidt T, Schrimpf A, Schulz R. Genetic diversity and population structure of burbot *Lota lota* in Germany: Implications for conservation and management. Fish Manag Ecol 2020; 27: 170-184. https://doi.org/10.1111/fme.12396
- Puffenberger EG, Hosoda K, Washington SS, Nakao K, deWit D, Yanagisawa M, Chakravart A. A missense mutation of the endothelin-B receptor gene in multigenic Hirschsprung's disease. Cell 1994; 79: 1257-1266. https://doi.org/10.1016/0092-8674(94)90016-7
- Apanaskevich DA, Filippova NA, Horak IG. The genus *Hyalomma* Koch, 1844. X. Redescription of all parasitic stages of *H*. (*Euhyalomma*) scupense Schulze, 1919 (=*H. detritum Schulze*) (Acari: Ixodidae) and notes on its biology. Folia Parasitol (Praha) 2010; 57: 69-78. https://doi.org/10.14411/fp.2010.009
- 18. Chandra S, Ma GC, Burleigh A, Brown G, Norrisa JM, Ward MP,

Emerya D, Šlapetaa J. The brown dog tick *Rhipicephalus sanguine-us* sensu Roberts, 1965 across Australia: Morphological and molecular identification of *R. sanguineus* s.l. tropical lineage. Ticks Tick Borne Dis 2020; 11: 101305. https://doi.org/10.1016/j.ttbdis.2019.101305

- Zemtsova GE, Apanaskevich DA, Reeves WK, Hahn M, Snellgrove A, Levin ML Phylogeography of *Rhipicephalus sanguineus* sensu lato and its relationships with climatic factors. Exp Appl Acarol 2016; 69: 191-203. https://doi.org/10.1007/s10493-016-0035-4
- 20. Guo LP, Mu LM, Xu J, Jiang SH, Wang AD, Chen CF, Guo G, Zhang WJ, Wang YZ. *Rickettsia raoultii* in *Haemaphysalis erinacei* from marbled polecats, China-Kazakhstan border. Parasit Vectors 2015; 8: 461. https://doi.org/10.1186/s13071-015-1065-1