## Korean Journal of Environmental Biology

### **Original article**

https://doi.org/10.11626/KJEB.2020.38.3.476

#### Korean J. Environ, Biol.

38(3): 476-480 (2020) ISSN 1226-9999 (print) ISSN 2287-7851 (online)

# Plocamium serrulatum (Plocamiaceae), a red algal species newly recorded in Korea

Pil Joon Kang, Jae Woo An and Ki Wan Nam\*

Department of Marine Biology, Pukyong National University, Busan 48513, Republic of Korea

#### \*Corresponding author

Ki Wan Nam Tel. 051-629-5922 E-mail. kwnam@pknu.ac.kr

Received: 28 August 2020 Revised: 21 September 2020

Revision accepted: 21 September 2020

**Abstract:** A marine red algal species was collected from Uljin, located on the eastern coast of Korea, during a survey of marine algal flora. This alga shares the generic features of *Plocamium*, and is characterized by linear, flat and thin thalli with narrow axes, branches developing in alternating pairs from margins of the axes, two to four times alternately pinnately branching, linear or occasionally adaxially curved lowermost branchlets and distinctly and compactly clustered stichidia. In a phylogenetic tree based on *rbc*L sequences, the Korean alga nests in the same clade as *P. serrulatum*. The genetic distance between both sequences within the clade was calculated as 0.0–0.2%. Based on the morphological and molecular data, this Korean species is identified as *P. serrulatum* described originally from Taiwan. This is the first record of *P. serrulatum* in Korean marine algal flora.

Keywords: Plocamium serrulatum, red alga, morphological and molecular data, Korea

#### INTRODUCTION

Plocamium Lamouroux (1813), which is a red algal genus currently thought to contain 45 species, is widely distributed throughout the world's oceans. However, it is most diverse in the southern hemisphere (Cremades et al. 2011; Guiry and Guiry 2020). Silva et al. (1996) reported 16 species from the Red Sea. Some of these also have distributions more or less restricted to South Africa, whereas several others are known only from western Australia (Wynne 2002). In Korea, six species of Plocamium have been recorded (Kim et al. 2013).

Traditional taxonomy distinguishes between *Plocamium* species largely on the basis of the number of ramuli in alternating series, the morphology of the lower ramuli, the length, width, color and consistency of the thallus, and the morphology and arrangement of tetrasporangial stichidia and cystocarps (Simons 1964; Womersley 1971; South

and Adams 1979; Gabrielson and Scagel 1989; Cremades et al. 2011).

During the indigenous survey of marine algal flora, a species belonging to *Plocamium* was collected from Uljin in Korea. This is identified based on morphological and molecular data, and is newly recorded in Korea herein.

#### MATERIALS AND METHODS

Specimens for this study were collected from Onyang-ri, Uljin located in the eastern coast of Korea. Taxonomic data were obtained from fresh, liquid-preserved and herbarium specimens. Liquid-preserved material was stored in a 10% solution of Formalin/seawater. Blades dissected from the cleared materials were hand sectioned, transferred to a slide with distilled water, and mounted in pure glycerin. For permanent slides, the glycerin was exchanged with 10–20%

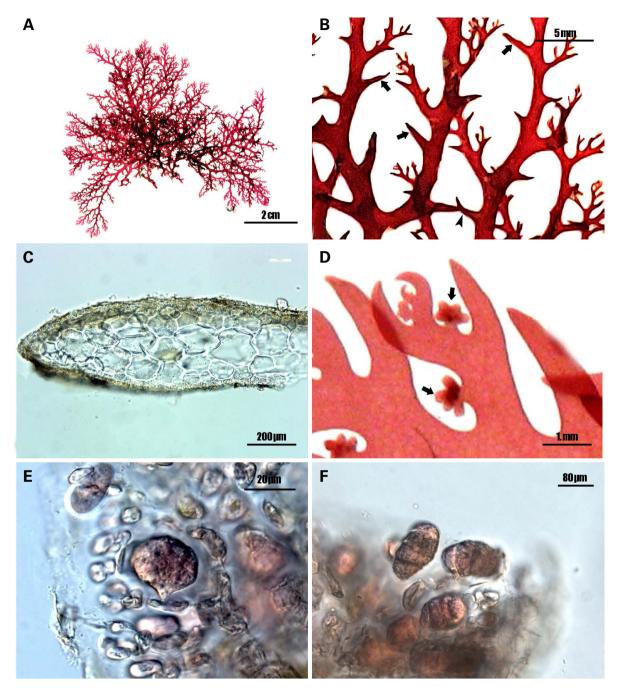


Fig. 1. Plocamium serrulatum Okamura. A. Habit of vegetative plants with narrow axes. B. Details of branches developing in alternating pairs from margins of the axes with linear (arrows) or occasionally adaxial curving lowermost branchlets (arrowhead). C. Transverse section of main branch. D. Distinct and compact clustered stichidia (arrows). E. Young tetrasporangium cut off from cortical cell of stichidium. F. Fully developed tetrasporangia divided zonately.

corn syrup.

Total genomic DNA was extracted from silica-gel-preserved sample using the DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. Before extraction, dried material was crushed with liquid nitrogen using a mortar and pestle. Extracted DNA was used for amplification of ribulose-1, 5-bisphosphate carboxylase large subunit (*rbc*L) regions. For *rbc*L, the gene

was amplified in three overlapping parts with the primer pairs FrbcL start (5'-TGTGTTGTCGACATGTCTA-ACTCTGTAGAAG-3') - R753 (5'-GCTCTTTCATA-CATATCTTCC-3'), F492 (5'-CGTATGGATAAATTTG-GTCG-3') - R1150 (5'-GCATTTGTCCGCAGT-GAATACC-3'), and F993 (5'-GGTACTGTTGTAGG-TAAATTAGAAGG-3') - RrbcS (5'-TGTGTTGCGGC-CGCCCTTGTGTT AGTCTCAC-3') (Freshwater and Rueness 1994). PCR amplifications were performed in a TaKaRa PCR Thermal Cycler Dice (TaKaRa Bio Inc., Otsu, Japan). PCR was performed with an initial denaturation step at 94°C for 4 min, followed by 35 cycles of 1 min at 94°C, 1 min at 50°C, and 2 min at 72°C, with a final 7-min extension at 72°C. The PCR products were moved to Macrogen Sequencing Service for sequencing (Macrogen, Seoul, Korea). Sequences for the rbcL region were aligned using BioEdit (Hall 1999). Phylogenetic analyses were performed using maximum-likelihood methods. Bootstrap values were calculated with 1,000 replications. *RbcL* sequences of other species were obtained from Gen-Bank. *Sarcodia montagneana* (J.D. Hooker & Harvey) J. Agardh was used as an outgroup.

#### RESULTS AND DISCUSSION

#### Plocamium serrulatum Okamura 1932

Korean name: Yal-beun-gop-seu-ri nom. nov.

(신칭:얇은곱슬이)

Type locality: Kotosho, Taiwan (Aoki, Segawa) (Guiry and Guiry 2020).

Specimens examined: NIBRRD0000004368-NIBRRD

0000004370 (Onyang-ri, Uljin: 26.viii.2014).

Habitat: Epilithic near upper to lower intertidal.

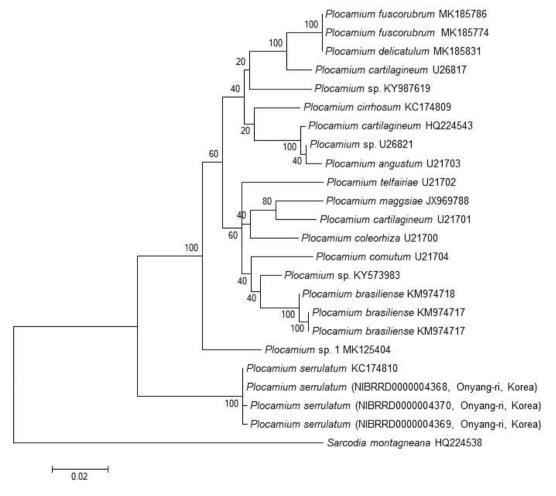


Fig. 2. Phylogenetic tree of *Plocamium* species obtained from maximum-likelihood method based on *rbc*L sequences. Bootstrap proportion values (1,000 replicates samples) are shown above branches. Scale bar = 0.02 substitutions/site.

Morphology: Thalli erect, 5–10 cm in height, linear, flat, bright red in color, attached by a conspicuous holdfast; main axes narrow, 1–2 mm wide, narrowing to 0.5 mm distally, thin membranous, 800–900 µm thick; branches developing in alternating pairs from margins of the axes, with 2–4 times alternately pinnately branched; proliferous branchlets formed mainly in the lower part of lateral branches; lowermost branchlet linear, occasionally adaxially curved; tetrasporangial initial cut off from cortical cell of stichidium; tetrasporangial stichidia distinct and compact clustered, stellate, cylindrical or branched divaricately, scattered among the branches of the proliferations; tetrasporangia divided zonately. Sexual thalli were not collected.

The first dichotomy patterns, such as the ramuli in alternating pairs or alternating series of three to four (or more), are considered to be significant in most keys to species of Plocamium (Simons 1964; Womersley 1971, 1994; South and Adams 1979; Yoshida 1998). In addition, the width of the main axes is also be useful (Wynne 2002; Yano et al. 2004). According to the original description from Kotosho, Taiwan (Okamura 1932), *Plocamium serrulatum* appears to fall into the former branching type with narrow axes rather than the latter dichotomy pattern with broad axes. These features are basically found in Korean alga collected from Uljin in the present study (Fig. 1A, B). In gross morphology together with the branching pattern, the Korean entity is similar to Plocamium serrulatum. It has linear, flat and thin thalli with narrow axes and two to four times pinnate alternating pairs in branching (Fig. 1A, B) as reported by Okamura (1932).

Lowermost branchlet shape is also important for distinguishing between *Plocamium* species (Yano *et al.* 2004). *P. serrulatum* appears to be an adaxial curving type based on the original description (Okamura 1923, p. 189, pl. 198, f. 1–4, as *P. costatum*). However, both types of adaxial curving and linear shape in the branchlets are observed in Korean alga, even though the former shape only occurs occasionally (Fig. 1B).

The position of tetrasporangial stichidia varies among *Plocamium* species (Womersley 1971; Gabrielson and Scagel 1989). *P. serrulatum* shows distinctly and compactly clustered stichidia of divaricate and stellate shape in the usual form (Okamura 1932). This feature is also found in the Korean alga (Fig. 1D).

In the phylogenetic tree based on *rbc*L sequence, the sequence of the Korean alga nests in the same clade as that of *P. serrulatum* from the type locality (Taiwan) with a genetic distance of 0.0–0.2% (Fig. 2). These morphological

and molecular evidence suggests that this Korean species should be identified as *P. serrulatum*. This is the first record of the *Plocamium serrulatum* in Korea.

#### **ACKNOWLEDGEMENTS**

This work was supported by a Research Grant of Pukyong National University (2019).

#### **REFERENCES**

- Cremades J, R Barreiro, I Maneiro and W Saunders. 2011. A new taxonomic interpretation of the type of *Plocamium cartilagineum* (Plocamiales, Florideophyceae) and its consequences. Eur. J. Phycol. 46:125–142.
- Freshwater DW and J Rueness. 1994. Phylogenetic relationships of some European *Gelidium* (Gelidiales, Rhodophyta) species based upon *rbc*L nucleotide sequence analysis. Phycologia 33:187–194.
- Gabrielson PW and RF Scagel. 1989. The marine algae of British Columbia, northern Washington and Southeast Alaska: division Rhodophyta (red algae), class Rhodophyceae, order Gigartinales, families Caulacanthaceae and Plocamiaceae. Can. J. Bot. 67:1221–1234.
- Guiry MD and GM Guiry. 2020. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. http://www.algaebase.org. Accessed 26 August 2020.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucl. Acids Symp. Ser. 41:95–98.
- Kim HS, SM Boo, IK Lee and CH Sohn. 2013. National List of Species of Korea Marine Algae. Jeonghaengsa, Seoul.
- Lamouroux JVF. 1813. Essai sur les genres de la famille des Thalassiophytes non articulées. Annales du Muséum d'Histoire Naturelle, Paris 20: 21–47, 115–139, 267–293, pls 7–13.
- Okamura K. 1923. Icones of Japanese Algae. Vol. IV(10). Published by the author, Tokyo.
- Okamura K. 1932. Icones of Japanese Algae. Vol. VI(10). Published by the author, Tokyo.
- Silva PC, PW Basson and RL Moe. 1996. Catalogue of the benthic marine algae of the Indian Ocean. Univ. Calif. Publ. Bot. 79:1–1259.
- Simons RH. 1964. Species of *Plocamium* on the South African Coast. Bothalia 8:183–193.
- South GR and NM Adams. 1979. A revision of the genus *Ploca-mium* Lamouroux (Rhodophyta, Gigartinales) in New Zea-

#### Korean J. Environ. Biol. 38(3): 476-480 (2020)

- land. Phycologia 18:120-132.
- Womersley HBS. 1971. The genus *Plocamium* (Rhodophyta) in southern Australia. Trans. R. Soc. S. Aust. 95:9–27.
- Womersley HBS. 1994. The Marine Benthic Flora of Southern Australia Part IIIA Bangiophyceae and Florideophyceae (Acrochaetiales, Nemaliales, Gelidiales, Hildenbrandiales and Gigartinales sensu lato). Australian Biological Resources Study, Canberra.
- Wynne MJ. 2002. A description of Plocamium fimbriatum sp.
- nov. (Plocamiales, Rhodophyta) from the Sultanate of Oman, with a census of currently recognized species in the genus. Nova Hedwigia 75:333–356.
- Yano T, M Kamiya, S Arai and H Kawai. 2004. Morphological homoplasy in Japanese *Plocamium* species (Plocamiales, Rhodophyta) inferred from the Rubisco spacer sequence and intracellular acidity. Phycologia 43:383–393.
- Yoshida T. 1998. Marine Algae of Japan. Uchida Rokakuho Published, Tokyo.