

Application of Cytochrome *b* Gene Sequences for Identification of Parrots from Korean Zoos

Jung-il Kim¹, Think Dinh Do¹, Duri Lee¹, Yonggu Yeo², Chang-Bae Kim^{1,*}

¹Department of Biotechnology, Sangmyung University, Seoul 03016, Korea

²Conservation and Health Center, Seoul Zoo, Gwacheon 13829, Korea

ABSTRACT

Parrots are common targets for illegal trade because of their beauty and high price. Accurate identification is necessary for the prevention of illegal trade and conservation of parrots. In the present study, mitochondrial markers of cytochrome *b* (*CYTb*) gene were used to identify parrot species from Korean zoos. Totally, 27 samples were collected from Seoul Zoo, Cheongju Zoo, and Uchi Zoo. After collection, total DNA of samples was extracted and used for PCR amplification. *CYTb* fragments were sequenced from all samples examined. The obtained sequences were used for GenBank blast, distance estimation, and phylogenetic analysis. All species were identified using *CYTb* sequences that determined 27 samples belong to 13 species in 7 genera, and 3 families. Our finding demonstrated the usefulness of *CYTb* sequences for identifying parrot species in Korean zoos.

Keywords: zoo, illegal trade, DNA barcoding, *CYTb* gene

INTRODUCTION

Natural exploitation together with human activities has resulted in the loss of world biodiversity. Conservation measures are necessary to protect wild species from extinction. Because of threats to biodiversity in nature, conservation of species in its natural habitat may be challenging. In this circumstance, zoos have changed the roles from animal exhibitions to institutions that handle multiple missions such as conservation, scientific research, and education (Witzenberger and Hochkirch, 2011). It is well-known that zoo is important to raise people's awareness on wildlife conservation and protection. In addition, many endangered species are grown and bred in zoos for conservation targets and they could be reintroduced to their natural habitats in the future (Wirtz et al., 2018). The accumulation of specimens may result in difficult tasks for zoo staff in species recognition. A reliable and accurate method for species identification is useful for zoo staff to conduct their role in conservation and management.

High demand for wildlife products from humans has resulted in the illegal wildlife trade. The illegal wildlife trade is a serious problem that threatens the survival of wild organisms and shows negative effects on wildlife conservation. Because

of their colorful plumage and mimicry ability, parrots (the order Psittaciformes) are in high demand as pets and highly prized. It is reported that parrot is the most traded bird among all avian orders (Bush et al., 2014). According to the International Union for Conservation of Nature (IUCN), 37 parrot species are endangered, 18 species are critically endangered and 16 species are already extinct (<https://www.iucnredlist.org>). Most parrots are protected by the Convention on International Trade in Endangered Species of Wild Fauna and Flora (<http://checklist.cites.org/#/en>). Therefore, accurate identification of parrots is necessary to support authorities to manage parrot trade. However, morphological identification of parrots is challenging due to their slight difference in morphology.

DNA barcoding has been proven to be a powerful tool for discrimination of wildlife products that are obtained from illegal trade (Lahaye et al., 2008; Presti et al., 2015). For animals, the mitochondrial protein-coding genes and ribosomal RNA genes are popular markers for DNA barcoding technique. Of mitochondrial markers, in addition to cytochrome *c* oxidase subunit I (*COI*) gene, cytochrome *b* (*CYTb*) gene is also used for molecular analysis (Arif and Khan, 2009). In avian, particularly *CYTb* gene has been used for the studies of phylogenetics, so that is more represented than other genes on

© This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

***To whom correspondence should be addressed**

Tel: 82-2-2287-5288, Fax: 82-2-2287-0070
E-mail: evodevo@smu.ac.kr

Table 1. Parrot samples collected and examined in this study

Family	Species name	Zoos	Form	No. of samples
Cacatuidae	<i>Cacatua alba</i>	Seoul zoo	Blood	1
	<i>Cacatua ducorpsii</i>	Uchi zoo	Feather	1
	<i>Cacatua galerita</i>	Seoul zoo	Blood	1
	<i>Cacatua goffiniana</i>	Seoul zoo	Blood	1
	<i>Cacaua moluccensis</i>	Seoul zoo	Feather	1
		Cheongju Zoo	Feather	1
	<i>Nymphicus hollandicus</i>	Seoul zoo	Feathers	2
		Cheongju Zoo	Feathers	2
Psittacidae	<i>Ara ararauna</i>	Seoul zoo	Feather	1
		Cheongju Zoo	Feather	1
		Uchi zoo	Feather	1
	<i>Ara chloropterus</i>	Seoul zoo	Feather	1
		Uchi zoo	Feather	1
	<i>Ara macao</i>	Seoul zoo	Feather	1
	<i>Myiopsitta monachus</i>	Seoul zoo	Feathers	2
Psittaculidae	<i>Eclactus roratus</i>	Seoul zoo	Feather	1
		Uchi zoo	Feather	1
	<i>Lorius garrulus</i>	Seoul zoo	Feather	1
		Cheongju Zoo	Feather	1
	<i>Melopsittacus undulatus</i>	Seoul zoo	Feathers	2
		Cheongju Zoo	Feathers	2
	Uchi zoo	Feather	1	

Table 2. Cytochrome *b* (*CYTB*) gene primers used in this study

Primer name	Direction	Sequence (5'-3')	Reference
MT-A1	F	CAACATCTCAGCATGATGAAACTTCG	Wink and Sauer-Gürth (2000)
Mte	R	GCAAATAGGAAGTATCATTCTGG	Fritz et al. (2006)
ND5L 14754	F	GGACCAGAAGGACTTGCCGACCTA	Ribas (2004)
H15400	R	AAGAATCGGGTTAGGGTGGGG	Braun (2014)
L15311	F	GTCCTACCATGAGGTCAAATATC	Braun (2014)
HThr 16082	R	TCTTTGGTTACAAGACCAATG	Kornegay et al. (1993)

reference databases such as GenBank (Branicki et al., 2003; Coghlan et al., 2013). Also, *CYTB* gene have been widely applied for avian species identification (Lee et al., 2008; Aliabadian et al., 2009; Boonseub et al., 2009) and estimating interspecific genetic relationship (Moore and DeFilippis, 1997). In this study, partial mitochondrial protein-coding genes (*CYTB* gene) were used to identify parrot species from Korean zoos. For this purpose, the partial *CYTB* gene of all samples was sequenced and analyzed.

MATERIALS AND METHODS

There were 15 feather and blood samples collected from

Seoul Zoo (Seoul). Another set of samples including 12 feathers were collected from Cheongju Zoo (Cheongju), and Uchi Zoo (Gwangju). The sample information used in this study is presented in Table 1. The ethical procedure for using blood and muscle tissue was approved by the Seoul Zoo IACUC (no. 2019-001).

Upon sample collection, total DNA was extracted and measured purity and concentration. The partial mitochondrial *CYTB* gene was amplified for all collected samples. *CYTB* primers used in this study are listed in Table 2. The 20 μ L of PCR reaction mixture contained 10 μ L of 2 \times DyeMix (Enzynomics, Korea) with 1.0 U of Taq polymerase, 1 μ L of each primer (5 pmole/ μ L), 100 ng DNA and distilled water up to 20 μ L. The amplification protocol was as follows: ini-

Table 3. Cytochrome *b* (*CYTB*) gene sequences of parrot species in the present study in comparison to GenBank database

No.	Family	Species name	No. of samples	Accession no.	Identity (%)	E-value
1	Cacatuidae	<i>Cacatua alba</i>	1	MT275977	99.0	0
2		<i>Cacatua ducorpsii</i>	1	MT275978	100	3E-163
3		<i>Cacatua galerita</i>	1	MT275979	100	0
4		<i>Cacatua goffiniana</i>	1	MT275980	99.9	0
5		<i>Cacatua moluccensis</i>	2	MT275981–MT275982	100	0
6		<i>Nymphicus hollandicus</i>	4	MT275983–MT275986	100	0
7	Psittacidae	<i>Ara ararauna</i>	3	MT275987–MT275989	100	0
8		<i>Ara chloropterus</i>	2	MT275990–MT275991	99.7	0
9		<i>Ara macao</i>	1	MT275992	100	0
10		<i>Myiopsitta monachus</i>	2	MT275993–MT275994	99.9–100	0
11	Psittaculidae	<i>Eclectus roratus</i>	2	MT275995–MT275996	100	0
12		<i>Lorius garrulus</i>	2	MT275997–MT275998	99.7	0
13		<i>Melopsittacus undulatus</i>	5	MT275999–MT276003	99.7–100	0

Table 4. Intraspecific distance and interspecific distance (%) of congenetic species based cytochrome *b* (*CYTB*) sequences

No.	Family	Species name	Intraspecific distance (%)	Interspecific distance (%)
1	Cacatuidae	<i>Cacatua alba</i>	0.00–0.33	5.97–11.30
2		<i>Cacatua ducorpsii</i>	0.00	2.03–11.82
3		<i>Cacatua galerita</i>	0.00–0.67	3.07–8.60
4		<i>Cacatua goffiniana</i>	0.00–0.33	1.34–11.00
5		<i>Cacatua moluccensis</i>	0.00–1.68	5.23–13.52
6	Psittacidae	<i>Ara ararauna</i>	0.00–0.52	6.00–13.39
7		<i>Ara chloropterus</i>	0.00–0.52	4.32–10.24
8		<i>Ara macao</i>	0.00–0.78	3.76–11.16
9	Psittaculidae	<i>Lorius garrulus</i>	0.16	2.30–3.48

Distances were estimated based sequences in this study and GenBank sequences.

tial denaturation at 95°C for 2 min followed by 35 cycles of denaturation at 95°C for 45 s, annealing at 52°C for 1 min, extension at 72°C for 1 min and final elongation at 72°C for 5 minutes. The PCR products were analyzed by electrophoresis in 1% (w/v) agarose gels in 1% tris-acetate buffer. The PCR products were sequenced directly by Sanger sequencing.

DNA sequence data sets were verified quality, and the consensus sequence was extracted from forward and reverse sequences using Geneious 9.1 software (Kearse et al., 2012). The consensus sequence was blasted on GenBank database and identified species with the highest similarity. For the genus with multiple species and sequences available on Genbank, additional sequences were retrieved from GenBank for further analyses. Sequence distances were estimated using Kimura-2-parameter (K2P) distance model (Kimura, 1980) in MEGA X software (Kumar et al., 2018). For reconstruction of the phylogenetic tree, the Maximum likelihood method with 1,000 bootstrap replicates in MEGA X software was used

(Kumar et al., 2018).

RESULTS AND DISCUSSION

Totally, 27 sequences of *CYTB* gene were generated. The blast result of *CYTB* sequences showed identity >99.0% with sequences available on Genbank for all samples (Table 3). Intraspecific distance and interspecific distance for examined species are presented in Table 4. Among five *Cacatua* species collected in this study, maximum intraspecific distance varied from 0% (*C. ducorpsii*) to 1.68% (*C. moluccensis*). These values are lower than minimum interspecific distance of the same species. The similar pattern was also found in *Ara* spp. and *Lorius garrulus* (Table 4). Combination of blast result and comparison of sequence distances, there were 13 parrot species identified from three Korean zoos. The examined parrots belonged to 7 genera and 3 families of the order Psittaciformes. The finding based on *CYTB* sequences is consistent

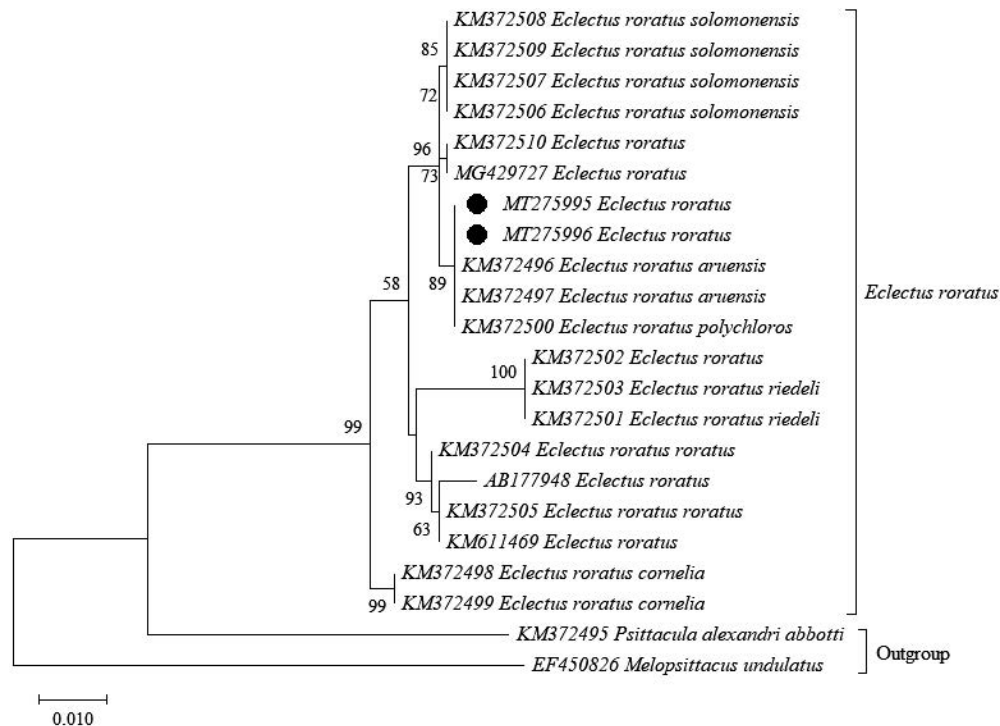


Fig. 1. Phylogenetic analysis of *Eclectus roratus* complex based on cytochrome *b* (*CYTb*) sequences. GeneBank accession numbers of sequences are next to species. The sequences in this study were labelled with black dots. Bootstrap values >50 are shown at nodes.

to species labelled by zoos (Table 1).

Further analyses of *Eclectus roratus* sequences indicated that *E. roratus* is divided into 4 clusters and 2 samples in this study belonged to a cluster that includes 3 subspecies: *E. r. aruensis*, *E. r. polychloros* and *E. r. solomonensis* (Fig. 1). Our finding is congruent with the results reported by Braun et al. (2016). According to NCBI Taxonomy (<http://www.ncbi.nlm.nih.gov/taxonomy>), *E. roratus* is a single species of the genus *Eclectus* and includes six subspecies: *E. r. aruensis*, *E. r. cornelia*, *E. r. polychloros*, *E. r. riedeli*, *E. r. roratus*, and *E. r. solomonensis*. However, based on analyses of *CYTb* sequences together with morphological and geographical data, Braun et al. (2016) revealed that *Eclectus roratus* is species complex and its 6 subspecies could be divided into 4 independent species: *E. roratus*, *E. cornelia*, *E. riedeli*, and *E. polychloros*. Accordingly, *E. polychloros* consisted of 3 subspecies: *E. r. aruensis*, *E. r. polychloros*, and *E. r. solomonensis* while each remaining species consisted of 1 subspecies. This demonstrated that *CYTb* marker is a useful tool for discrimination of species complex in parrot.

Due to their beauty and ability, parrots are one of the most common captive birds (Frynta et al., 2010). The illegal trade that meets human's high demand is a serious threat to parrots. Under biodiversity threats zoo has emerged its role in species conservation. The accurate identification of parrots particular-

ly which taxonomy is under discussion or recently identified in zoo is necessary for management and protection of parrots. In this study, DNA barcoding was applied to identify parrots from different Korean zoos. With *CYTb* marker, all collected species were identified based on blast results and the comparison between intraspecific and interspecific variations. Especially, *E. roratus* in this study might be identified to *E. polychloros* following Braun et al. (2016). It could be helpful to for management and protection of parrots in Korean zoos by correct identification. Since the first study of Hebert et al. (2004), numerous studies have used DNA barcoding to identify bird species, including parrots. Together with other mitochondrial markers, more *CYTb* sequences have been generated and accumulated on databases. With the increasing accumulation of *CYTb* sequences from different localities in the world, *CYTb* gene becomes a powerful approach for avian. Therefore, the gene has been applied to study avian identification and phylogeny (Astuti et al., 2006; Lee et al., 2008). This step is necessary to protect endanger bird species like many parrots from extinction.

In this study, *CYTb* gene sequences were applied to identify parrot species from three Korea zoos. The result showed that the use of *CYTb* sequences was effective for parrot identification. In addition, the marker is able to separate independent species from species complex group as the case of *E.*

eclectus. More species should be included in the future study to show the effectiveness of *CYTB* gene sequences and resolve better phylogeny of parrots.

ORCID

Jung-il Kim: <https://orcid.org/0000-0001-7851-2493>

Thinh Dinh Do: <https://orcid.org/0000-0001-8945-1518>

Duri Lee: <https://orcid.org/0000-0002-5784-3653>

Yonggu Yeo: <https://orcid.org/0000-0002-5578-2877>

Chang-Bae Kim: <https://orcid.org/0000-0002-1040-7600>

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We thank Seoul Zoo, Cheongju Zoo and Uchi Zoo for sample supplement. Samples were collected with the assistance of So Young Jung, Jung Yeol An, In Hui Park, Han Sol Kim, Su Yeon Seo, Mihyun Yoo, and Hany Lee (Seoul Zoo), Jeong Ho Kim (Cheongju Zoo) and Jong Woog Choi (Uchi Zoo). This work was supported by Korea Environment Industry & Technology Institute (KEITI) through Public Technology Program based on Environmental Policy, funded by Korea Ministry of Environment (MOE) (2018000210004).

REFERENCES

- Aliabadian M, Kaboli M, Nijman V, Vences M, 2009. Molecular identification of birds: performance of distance-based DNA barcoding in three genes to delimit parapatric species. *PLoS ONE*, 4:e4119. <https://doi.org/10.1371/journal.pone.0004119>
- Arif IA, Khan HA, 2009. Molecular markers for biodiversity analysis of wildlife animals: a brief review. *Animal Biodiversity and Conservation*, 32:9-17.
- Astuti D, Azuma N, Suzuki H, Higashi S, 2006. Phylogenetic relationships within parrots (Psittacidae) inferred from mitochondrial cytochrome-b gene sequences. *Zoological Science*, 23:191-198. <https://doi.org/10.2108/zsj.23.191>
- Boonseub S, Tobe SS, Linacre AMT, 2009. The use of mitochondrial DNA genes to identify closely related avian species. *Forensic Science International: Genetics Supplement Series*, 2:275-277. <https://doi.org/10.1016/j.fsigss.2009.08.050>
- Branicki W, Kupiec T, Pawlowski R, 2003. Validation of cytochrome *b* sequence analysis as a method of species identification. *Journal of Forensic Science*, 48:83-87. <https://doi.org/10.1520/JFS2002128>
- Braun MP, 2014. Parrots (Aves: Psittaciformes): evolutionary history, phylogeography, and breeding biology. PhD dissertation, Heidelberg University, Heidelberg, Germany.
- Braun MP, Reinschmidt M, Datzmann T, Waugh D, Zamora R, Häbich A, Neves L, Gerlach H, Arndt T, Mettke-Hofmann C, Wink M, 2016. Influences of oceanic islands and the Pleistocene on the biogeography and evolution of two groups of Australasian parrots (Aves: Psittaciformes: *Eclectus roratus*, *Trichoglossus haematodus* complex). Rapid evolution and implications for taxonomy and conservation. *European Journal of Ecology*, 3:47-66. <https://doi.org/10.1515/eje-2017-0014>
- Bush ER, Baker SE, Macdonald DW, 2014. Global trade in exotic pets 2006-2012. *Conservation Biology*, 28:663-676. <https://doi.org/10.1111/cobi.12240>
- Coghlan ML, White NE, Murray DC, Houston J, Rutherford W, Bellgard MI, Haile J, Bunce M, 2013. Metabarcoding avian diets at airports: implications for birdstrike hazard management planning. *Investigative Genetics*, 4:27. <https://doi.org/10.1186/2041-2223-4-27>
- Fritz U, Auer M, Bertolero A, Cheylan M, Fattizzo T, Hunds-dörfer AK, Sampayo MM, Pretus JL, Široký P, Wink M, 2006. A rangewide phylogeography of Hermann's tortoise, *Testudo hermanni* (Reptilia: Testudines: Testudinidae): implications for taxonomy. *Zoologica Scripta*, 35:531-543. <https://doi.org/10.1111/j.1463-6409.2006.00242.x>
- Frynta D, Lišková S, Bültmann S, Burda H, 2010. Being attractive brings advantages: the case of parrot species in captivity. *PLoS ONE*, 5:e12568. <https://doi.org/10.1371/journal.pone.0012568>
- Hebert PDN, Stoeckle MY, Zemlak TS, Francis CM, 2004. Identification of birds through DNA barcodes. *PLoS Biology*, 2:e312. <https://doi.org/10.1371/journal.pbio.0020312>
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A, 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics*, 28:1647-1649. <https://doi.org/10.1093/bioinformatics/bts199>
- Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16:111-120. <https://doi.org/10.1007/BF01731581>
- Kornegay JR, Kocher TD, Williams LA, Wilson AC, 1993. Pathways of lysozyme evolution inferred from the sequences of cytochrome *b* in birds. *Journal of Molecular Evolution*, 37:367-379. <https://doi.org/10.1007/BF00178867>
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K, 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, 35:1547-

1549. <https://doi.org/10.1093/molbev/msy096>
- Lahaye R, van der Bank M, Bogarin D, Warner J, Pupulin F, Gigot G, Maurin O, Duthoit S, Barraclough TG, Savolainen V, 2008. DNA barcoding the floras of biodiversity hotspots. *Proceedings of the National Academy of Sciences of the United States of America*, 105:2923-2928. <https://doi.org/10.1073/pnas.0709936105>
- Lee JCI, Tsai LC, Huang MT, Jhuang JA, Yao CT, Chin SC, Wang LC, Linacre A, Hsieh HM, 2008. A novel strategy for avian species identification by cytochrome *b* gene. *Electrophoresis*, 29:2413-2418. <https://doi.org/10.1002/elps.200700711>
- Moore WS and DeFilippis VR, 1997. The window of taxonomic resolution for phylogenies based on mitochondrial cytochrome *b*. In: *Avian molecular evolution and systematics* (Ed., de Mindeel DP). Academic Press, New York, pp. 84-119.
- Presti FT, Guedes NMR, Antas PTZ, Miyaki CY, 2015. Population genetic structure in Hyacinth Macaws (*Anodorhynchus hyacinthinus*) and identification of the probable origin of confiscated individuals. *Journal of Heredity*, 106(S1):491-502. <https://doi.org/10.1093/jhered/esv038>
- Ribas CC, 2004. *Filogenias Moleculares e Biogeografia Histórica em Psitacídeos (Aves: Psittacidae): Padrões e Processos de Diversificação no Neotrópico*. Universidade de São Paulo, Instituto de Biociências, São Paulo, pp. 1-151.
- Wink M, Sauer-Gürth H, 2000. Advances in the molecular systematics of African raptors. In: *Raptors at risk* (Eds., Chancellor RD, Meyburg BU). WWGBP/Handcock House, Surrey, pp. 135-147.
- Wirtz S, Böhm C, Fritz J, Kotschal K, Veith M, Hochkirch A, 2018. Optimizing the genetic management of reintroduction projects: genetic population structure of the captive Northern Bald Ibis population. *Conservation Genetics*, 19:853-864. <https://doi.org/10.1007/s10592-018-1059-6>
- Witzenberger KA, Hochkirch A, 2011. *Ex situ* conservation genetics: a review of molecular studies on the genetic consequences of captive breeding programmes for endangered animal species. *Biodiversity and Conservation*, 20:1843-1861. <https://doi.org/10.1007/s10531-011-0074-4>

Received April 7, 2020

Revised July 3, 2020

Accepted July 7, 2020