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# Identity of Spirometra theileri from a Leopard (Panthera pardus) and Spotted Hyena (Crocuta crocuta) in Tanzania

### Keeseon S. Eom<sup>1</sup>, Hansol Park<sup>1</sup>, Dongmin Lee<sup>1</sup>, Seongjun Choe<sup>1</sup>, Yeseul Kang<sup>1</sup>, Mohammed Mebarek Bia<sup>1</sup>, Barakaeli Abdieli Ndosi<sup>1,4</sup>, Tilak Chandra Nath<sup>1,5</sup>, Chatanun Eamudomkarn<sup>2</sup>, Julius Keyyu<sup>3</sup>, Robert Fyumagwa<sup>3</sup>, Simon Mduma<sup>3</sup>, Hyeong-Kyu Jeon<sup>1,\*</sup>

<sup>1</sup>Department of Parasitology, Parasitology Research Center and Parasite Resource Bank, Chungbuk National University School of Medicine, Cheongju 28644, Korea; <sup>2</sup>Department of Parasitology, Faculty of Medicine, Khon Kaen University, Khon Kaen, Thailand; <sup>3</sup>Tanzania Wildlife Research Institute, Arusha, Tanzania; <sup>4</sup>Tanzania Wildlife Management Authority, P.O. BOX 2658 Morogoro, Tanzania; <sup>5</sup>Department of Parasitology, Sylhet Agricultural University, Bangladesh

**Abstract:** In the present study, a *Spirometra* species of Tanzania origin obtained from an African leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) was identified based on molecular analysis of cytochrome *c* oxidase I (*cox1*) and NADH dehydrogenase subunit I (*nad1*) as well as by morphological observations of an adult tapeworm. One strobila and several segments of a *Spirometra* species were obtained from the intestine of an African male leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) in the Maswa Game Reserve of Tanzania. The morphological characteristics of *S. theileri* observed comprised 3 uterine loops on one side and 4 on the other side of the mid-line, a uterine pore situated posterior to the vagina and alternating irregularly either to the right or left of the latter, and vesicular seminis that were much smaller than other *Spirometra* species. Sequence differences in the *cox1* and *nad1* genes between *S. theileri* (Tanzania origin) and *S. erinaceieuropaei* were 10.1% (*cox1*) and 12.0% (*nad1*), while those of *S. decipiens* and *S. ranarum* were 9.6%, 9.8% (*cox1*) and 13.0%, 12.6% (*nad1*), respectively. The morphological features of the Tanzania-origin *Spirometra* specimens coincided with those of *S. theileri*, and the molecular data was also consistent with that of *S. theileri*, thereby demonstrating the distribution of *S. theileri* in Tanzania. This places the leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) as new definitive hosts of this spirometrid tapeworm.

Key words: Spirometra theileri, cox1, nad1, morphology, Tanzania

#### INTRODUCTION

Cestode tapeworms of the genus *Spirometra* belong to the family Diphyllobothriidae which infect amphibians, birds and mammals. The adult stage of *Spirometra* spp. parasitize the intestine of carnivores, felids and canids including domestic dogs and cats. Spargana are plerocercoid larvae of cestode tapeworms of the genus *Spirometra*, which are found in paratenic hosts of *Spirometra* spp. such as amphibians, reptiles and mammals including humans.

Spirometrid tapeworms previously reported in African mammals were described by Baer (1924) [1]. *S. pretoriensis* was

© 2019, Korean Society for Parasitology and Tropical Medicine This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. assigned the name Lühella pretoriense (Baer, 1924) and became synonymous with Diphyllobothrium pretoriense by Baer (1926), which was found in bat-eared fox (Otocyon megalotis) from South Africa (Baer, 1926) and later in African wild dog (Lycaon pictus) from Congo (Baer and Fain, 1955) [2]. S. theileri has been found in bush cat (Leptailurus serval), tiger cat (Felis lybica) near Pretoria, and lion from Congo (Baer, 1959) [3]. According to some studies, Spirometra and Taenia species were the most prevalent parasites in African lion, spotted hyena and African wild dog [4-8]. The spotted hyenas in Kenya and Zambia were reported to have a high rate of Spirometra infection with 74.3% and 22% in the feces of examined animals, respectively [5]. Some lions in Tanzania and Zambia were noted as having 63% and 87% infection of spirometrid tapeworms, respectively [5,6]. Spirometra species were the most commonly found in lion and second most common parasite in hyenas. The Spirometra species in lions is S. theileri whereas the species found in hyenas is generally accepted to be S. pretoriensis, although both

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\*Corresponding author (jeonhk@chungbuk.ac.kr)

Spirometra species may occur in lions [9]. The plerocercoids of Spirometra have been found in many herbivores including antelope, zebra and warthog in Kenya and Tanzania [8,9]. Baboons and other monkeys have also been infected with Spirometra species in East Africa [10,11]. Host susceptibility of African Spirometra species was reported from experimentally infected dogs and cat. They were able to infect dogs but not cats with spargana recovered from wart hogs in Tanzania [12]. They were also unable to infect various amphibians and reptiles with oral doses of infected copepods [12]. Although 2 Spirometra species have been reported as being S. theileri and S. pretoriensis from South Africa and Congo, it is unknown if these Spirometra species can infect humans. There have been a few reports and sporadic records of human sparganosis in Africa [13,14]. Human infections are unclear and not fully understand in Africa. Water containing procercoid larvae has been suggested as a possible mode of human infection in North and East Africa, and of wild animals in Masailand and Chad [13-15].

Previous studies have dealt with the validity of making *Spirometra* species assignments based on morphological characteristics that has given rise to 6 *Spirometra* species as independent species, 38 *Spirometra* species as the 1 species, and 4 species as valid species [16-18]. To date, the genus *Spirometra* includes *S. erinaceieuropaei*, *S. decipiens*, *S. masonoides* and *S. ranarum* as valid species [19-22]. In the present study, a *Spirometra* species of Tanzania origin obtained from an African leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) was identified based on molecular analysis of cytochrome c oxidase I (*cox1*) and NADH dehydrogenase subunit I (*nad1*) as well as by morphological observations of an adult tapeworm.

#### MATERIALS AND METHODS

#### Specimens and morphological analysis

One strobila and several segments of a *Spirometra* species were obtained in intestine from an African male leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) in the Maswa Game Reserve of Tanzania in February 2012. The Maswa Game Reserve (34°35'E, 03°16'S) was established in 1962, covers an area of 2,200 square kilometers, and is located in the south-western boundary of Serengeti National Park (34°50'E, 02°30'S). Parasite materials used in this study were provided by the Parasite Resource Bank of the Korea National Research Resource Center, Republic of Korea (PRB000720).

The collected worm was fixed in 70% ethanol and some

gravid proglottids were compressed between 2 glass slides prior to fixation in alcohol-formalin-acetic acid (AFA) for carmine staining. Part of the specimen was subjected to hematoxylin and eosin (H-E) staining after longitudinal sections were obtained for observation of the cirrus pouch, cirrus sac, seminal vesicle, uterus and genital pore. The following morphological characteristics of the *Spirometra* species were compared to those of Baer [2] with carmine stained mature and gravid proglottids: width and length of mature and gravid proglottids, number of uterine loops, position of uterine pore, genital pore, vaginal opening, size and number of testes, and size of eggs.

#### PCR and DNA sequencing

Total genomic DNA extraction and PCR reactions were employed as previously described by Jeon et al. [21]. Mitochondrial cox1 and nad1 genes were amplified and sequenced by cycle sequencing. Primers used to amplify the mitochondrial cox1 gene (Spi-CO1F: 5'-GAC TAA GTG TTT TCA AAA CAC TAA GTG-3' and Spi-CO1R: 5'CAC CCT ACC CCT GAT TTA CAA AAT-3') and nad1 gene (Spi-ND1F: 5'-GGA GAA TAT TGG TTT GTC TAA CCA-3' and Spi-ND1R: 5'-CCT TCT TAA CGT TAA CAG CAT TAC GAT-3') were designed from the cox1 and nad1 sequences of S. erinaceieuropaei (KJ599680), S. decipiens (KJ599679) and S. ranarum (MH298843) mitochondrial genomes. PCR was performed in a 25 µl reaction volume containing 50 ng of genomic DNA, 10 pmol of each primer, 2.5 µl of 10X buffer, 12.25 µl of 2X buffer (MgCl<sub>2</sub>, dNTP) and 1.25 units Taq polymerase (TAKARA BIO INC, Kusatsu, Shiga, Japan). PCR was employed using a Biometra T-Gradient Thermoblock (Analytikjena, Göttingen, Germany) instrument under conditions comprising: 94°C for 3 min; 35 cycles of 94°C for 30 sec, 46°C for 40 sec, 72°C for 90 sec, and a final step at 72°C for 10 min. DNA sequencing was performed using a Big-Dye terminator kit (version 3.1, Applied Biosystems, Foster City, California, USA) and reaction products were sequenced directly using a DNA sequencer (ABI3730XL, Applied Biosystems).

#### DNA sequence analyses

DNA sequences of the mitochondrial *cox1* gene were assembled using the Geneious 9.1 program (Biometer, Auckland, New Zealand). These sequences were aligned using MAFFT methods in the Geneious 9.1 program by comparison with sequences of *S. erinaceieuropaei*, *S. decipiens* and *S. ranarum* in the GenBank database. Molecular Evolution Genetics Analysis (MEGA) software version 7.0 was employed to analyze nucleo-

tide composition, conserved sites, variable sites, singleton sites and parsimony-informative sites [23]. Evaluation of the number of haplotypes, nucleotide diversity and haplotype diversity were performed using DnaSP software version 6.12 [24].

Phylogenetic analysis was evaluated by employing Bayesian inference (BI) and maximum-likelihood (ML) using the complete mitochondrial cox1 (1,566 bp) and nad1 (891 bp) sequences of S. erinaceieuropaei (KJ599680), S. decipiens (KJ599679), S. ranarum (MH298843, MH298844), Sparganum proliferum (AB015753), Dibothriocephalus nihonkaiense (=Diphyllobothrium nihonkaiense) (EF420138) and D. latus (= Diphyllobothrium latum) (DQ985706). The ML tree was constructed with 7 taxa and the nucleotide dataset of the mitochondrial cox1 gene as implemented in RAxML v. 7.3.1 [25] after TRN+G substitution model sampling was chosen according to the ModelTest using the program Partition Finder [26]. The nodes were assessed by bootstrapping with 1,000 pseudoreplicates. The BI tree was reconstructed with the same dataset using MrBayes 3.2 with a HKY+G substitution model and running 4 simultaneous Monte Carlo Markov chains (MCMC) for 10 million generations, sampling every 1,000 generations and discarding the first 25% of the generations as burnin [27].

#### **RESULTS**

Morphological characteristics

The strobila collected from the intestine of a leopard (Panthera pardus) and spotted hyena (Crocuta crocuta) was 45 cm and 134.5 cm, respectively. The dimensions of immature to gravid proglottids ranged from 0.4 mm to 1.0 mm in width and 0.15 to 0.25 mm in length. The genital pore was situated ventrally on the midline in the anterior 1/5 of the segment. The vaginal pore was situated closely behind the male genital pore. The uterine pores were on the midline behind the anterior margin of the terminal ball 0.03 mm in diameter. The uterus consisted of 3-4.5 loops on each side. The dumbbellshaped ovary was connected to the uterus and situated near the posterior margin of the segment (Fig. 1A, B). The longitudinal section of the cirrus sac showed an oval form, 0.16 mm in length and 0.13 mm in width. The seminal vesicle was elliptical and measured 0.13 in length and 0.14 mm in width (Fig. 1C) (Table 1). The testes measured about 0.06 mm in diameter. The eggs measured 0.054-0.059 mm in length and 0.033-0.035 mm in width.



Fig. 1. Gravid proglottids of *Spirometra theileri* (Tanzania origin) collected from a leopard (*Panthera pardus*) (A) and spotted hyena (*Cro-cuta crocuta*) (B). (A, B) Whole mounted specimens of proglottids showing the genital pore (GP), vaginal pore (VP), and uterus (U) (×40), (C) Longitudinal section of a gravid proglottid showing the cirrus sac (CS), seminal vesicle (SV), and uterus (U) (H&E stain).

Organs	Morphological factures	S. theireli <sup>1</sup>	Spirometra sp.2	Spirometra sp.3	Othoro
	Morphological leatures		Others		
Scolex	Spatulate	0.25-0.3	-	-	Diameter
		0.53-0.86	-	-	Length
Proglottids <sup>4</sup> (n = 10)	Trapezoid	0.4-2.4	0.60-0.65	0.4-1.0	Width
		0.02-1.9	0.15-0.25	0.15-0.25	Length
Uterus (n = 10)	Coiling	3-4.5	3-4.5	3-4.5	Loops
Eggs (n=10)	Operculate	0.057	0.054-0.059	0.054-0.059	Length
		0.034	0.033-0.035	0.033-0.035	Width
Cirrus pouch (n = 10)	Oval	0.16	0.16	0.16	Length
		0.13	0.13	0.13	Width
Cirrus (n = 10)	Cylinderical	0.3	0.3	0.3	Length
		0.13	0.13	0.13	Diameter
Seminal vesicle (n = 10)	Elliptical	-	0.13	0.13	Length
			0.14	0.14	Width
Vaginal opening (n = 10)	Crescentic	-	0.25	0.25	Width
Testes (n = 10)	Spherical	0.11-0.15:0.06-0.08	0.11-0.12:0.04-0.06	0.11-0.12:0.04-0.06	Diameter

#### Table 1. Morphological features of Spirometra theileri and Spirometra sp.

1Baer (1926).

<sup>2</sup>This study (Tanzania origin collected from a leopard).

<sup>3</sup>This study (Tanzania origin collected from a spotted hyena).

<sup>4</sup>Range from the first segment to the end of the segment of the strobila.

Table 2. Pairwise sequence differences of mitochondrial cox1 and nad1 genes of Spirometra species of Tanzania origin and various Spirometra species, Dibothriocephalus latus and D. nihonkaiense

	-1	-2	-3	-4	-5	-6
(1) Spirometra species (Tanzania)*						
(2) <i>S. decipiens</i> (KJ599679)	9.6/13.0					
(3) S. ranarum (MH298843/MH298844)	9.8/12.6	2.1/2.1				
(4) S. erinaceieuropaei (KJ599680)	10.1/12.0	9.4/8.8	8.8/10.0			
(5) Sparganum proliferum (AB015753)	12.2/-	12.1/-	12.2/-	12.6/-		
(6) <i>D. latus</i> (DQ985706)	16.7/20.3	17.4/22.0	17.4/21.7	16.9/21.7	16.9/-	
(7) D. nihonkaiense (EF420138)	17.8/21.8	17.5/22.3	17.8/22.0	17.6/21.7	17.6/7.2	7.2/7.2

\*S. theileri (GenBank No. MK955901 for cox1 and MN244299 for nad1).

Table 3. Genetic diversity of mitochondrial DNA sequences of Spirometra species including S. erinaceieuropaei, S. decipiens, S. ranarum and S. theileri

	V-S	C-S	Sin-S	Parsi-S	Syn-S	NonSyn-S	Sq-D	Hd±S.D.	Pi±S.D.
<i>cox1</i> (1,566 bp)	227	1,339	169	59	374.75	1188.25	0-0.0101	1.000±0.177	0.08280±0.01946
<i>nad1</i> (891 bp)	157	734	116	41	209.00	682.00	0-0.0130	$1.000 \pm 0.177$	$0.09914 \pm 0.02443$

V-S: variable site, C-S: conserved site, Sin-S: singleton site, Parsi-S: parsimony informative site, Syn-S: synonymous site, NonSyn-S: nonsynonymous site, Sq-D: sequence divergence, Hd: haplotype diversity, Pi: nucleotide diversity.

## Sequence divergence of mitochondrial *cox1* and *nad1* genes

The divergence of the mitochondrial *cox1* and *nad1* sequences of the *Spirometra* species collected from a leopard and spotted hyena (Tanzania origin) was 9.6%/13.0% (*S. decipiens*, KJ599670), 9.8%/12.6% (*S. ranarum*, MH298843), 10.1%/ 12.0% (*S. erinaceieuropaei*, KJ599680), 12.2%/- (Sparganum

proliferum, AB015753), 16.7%/20.3% (*D. latus*, DQ985706) and 17.8%/21.8% (*D. nihonkaiense*, EF420138) (Table 2). The *cox1* (1,566 bp) and *nad1* (891) sequences of the *Spirometra* species including *S. erinaceieuropaei*, *S. decipiens*, *S. ranarum* and *S. theileri* revealed 227 and 157 polymorphic sites, 59 and 41 parsimony informative sites, and 169 and 116 singleton variable sites, respectively. There were 374.75 (*cox1*) and



Fig. 2. Phylogenetic relationship of *Spirometra* species based on *cox1* and *nad1* sequences. Numbers on branches represent bootstrap values for maximum likelihood (ML) and the support value of Bayesian inference (BI). GenBank No. MK955901 and MN244299 are registered as *S. theileri* by complete mitochondrial *cox1* and *nad1* sequences.

209.00 (*nad1*) synonymous substitutions, and 1,188.25 (*cox1*) and 682.00 (*nad1*) non-synonymous substitutions. The genetic divergence of the *cox1* and *nad1* sequences of the *Spirometra* species ranged from 0 to 10.1% and 0 to 13.0%, respectively. These *cox1* and *nad1* genes showed high haplotype diversity (Hd) and low nucleotide diversity (Pi) (Table 3).

Phylogenetic analyses of the *Spirometra* species was conducted using 2 analytic methods (ML and BI) based on the *cox1* genes of *S. erinaceieuropaei*, *S. decipiens*, *S. ranarum*, *Sparganum proliferum*, *D. latus and D. nihonkaiense*. The phylogenetic tree showed high level confidence values for 4 branches of the 4 *Spirometra* species that divided into 3 clades containing *S. theileri*, *S. erinaceieuropaei* and *S. decipiens*, and *S. ranarum* subclades (Fig. 2).

#### DISCUSSION

Spirometrid tapeworms found in African mammals were described as *S. pretoriensis* and *S. theileri* (under the name *Diphyllobothrium theileri*) by Baer (1924). The morphological characteristics of *S. theileri* described by Baer were as follows: the average length was 350 to 400 mm, and the greatest width was 3.3 mm. The length of the scolex ranged from 0.53 to 0.86 mm, and the width 0.25 to 0.3 mm. The first segments were 0.4 mm wide and 0.02 mm long, becoming 1.9 mm in length and 2.4 mm in width. The testes measured 0.11 to 0.15 vs 0.06 to 0.08 mm in spherical shape and occupied nearly all of the lateral fields of the medullary parenchyma. The cirrus measured 0.16 mm in length and 0.06 mm in diameter. The cirrus pouch measured 0.3 mm in length and 0.13 mm in diameter. The vaginal opening was situated behind the cirrus pouch. The ovary was situated on the dorsal side of uterus in the posterior region of the segment. The uterus was characterized by the great diameter of its coils, of which there were 3 on one side and 4 on the other side of the mid-line. The uterine pore was situated posterior to the vagina and usually alternating irregularly either to the right or left of the latter. The ova were thick-shelled and operculated, and measured  $57 \times 34 \mu m$  [2] (Baer, 1924).

Since then, Opuni and Muller (1974) reported on S. theileri adult tapeworms recovered from dogs that were fed plerocercoids obtained from a wart hog in Tanzania. The morphology of S. theileri that has been described included the following features: mature proglottids measured 8 to 12 mm in width with a spirally coiled uterus of 4 to 7 loops, the final centrally placed loops were much larger than the others and were constricted and hemispherical in shape [12]. Graber (1981) described this species in detail as possessing the following characteristics: mature or gravid proglottids measured 2.5-3.5 mm, the segments were wider than they were long and measured 0.24-0.36 mm in length in the anterior part and  $2.5-3.5 \times 1.2$ -1.3 mm in the middle and posterior part, with the last segment being a little narrow (2.4×1.95 mm), the scolex measured  $0.6 \times 0.18$  mm and was elongated with a spoon-like shape, and the mean size of the eggs was  $49 \times 32.1 \,\mu\text{m}$  [28].

In the present study, the morphological features of the Spirometra species collected from the intestine of a leopard (Panthera pardus) and spotted hyena (Crocuta crocuta) were examined according to the aforementioned studies. The morphological features of 2 Spirometra species isolated from Tanzania resembled that of S. theileri in a previous study (Baer, 1926), whereby the uterus consisted of 3 loops on one side and 4 loops on the other side of the mid-line, and the aspect of the gravid uterus in the longitudinal section. The morphological characteristics of S. theileri have been described by Baer (1926) [2] as comprising a uterus with 3 uterine loops on one side and 4 on the other side of the mid-line, the uterine pore was situated posterior to the vagina and alternating irregularly either to the right or left of the latter, and the vesicular seminis were much smaller than other Spirometra species. The distinct features of S. theileri relative to other Spirometra species included a uterine morphology comprising uterine coils that were larger than the terminal ball and ellipse shape. The morphological features of S. theileri were distinct from that of S. pretoriensis. The major differentiating features concern the number of uterine loops and shape of the cirrus pouch. For example, the uterus of S. theileri consists of 3-4.5 coils and a cirrus pouch communicated through a short canal with a second, much smaller vesicular seminis, while the uterus of S. pretoriensis forms a single large loop on either side and the cirrus pouch possesses a large vesicular seminis of variable shape and size owing to the presence of very muscular walls [2].

Molecular identification of *S. theileri* was examined by sequence differences in the *cox1* and *nad1* genes with other *Spirometra* species including *S. erinaceieuropaei*, *S. decipiens* and *S. ranarum*. Sequence differences in the *cox1* and *nad1* genes between *S. theileri* (Tanzania origin) and *S. erinaceieuroapei* were 10.1% (*cox1*) and 12.0% (*nad1*), while those of *S. decipiens* and *S. ranarum* were 9.6% and 9.8% (*cox1*) and 13.0% and 12.6% (*nad1*), respectively. The nucleotide diversity (Pi) value of *cox1* and *nad1* was greater than 0.01 in this study, suggesting that the nucleotide sequences of *Spirometra* species such as *S. erinaceieuropaei*, *S. decipiens* and *S. theileri* have large variation. The phylogenetic analyses based on ML and BI methods suggested that the mitochondrial DNA sequences of *S. theileri* were separated from *S. erinaceieuropaei* and *S. decipiens*.

In conclusion, this study identified a *Spirometra* species of Tanzania origin by morphological examination and mitochondrial *cox1* and *nad1* sequences analysis of the adult worms. The morphological features and molecular data of the Tanzania-origin *Spirometra* specimens coincided with those of *S. theileri*, thereby demonstrating the distribution of *S. theileri* in Tanzania. *S. theileri* was found in leopard (*Panthera pardus*) and spotted hyena (*Crocuta crocuta*) as new definitive hosts of this spirometrid tapeworm in Tanzania.

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### **CONFLICT OF INTEREST**

We have no conflict of interest related to this work.

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