Development and Validation of a Canopy Photosynthetic Rate Model of Lettuce Using Light Intensity, CO₂ Concentration, and Day after Transplanting in a Plant Factory

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Abstract. The photosynthetic rate is an indicator of the growth state and growth rate of crops and is an important factor in constructing efficient production systems. The objective of this study was to develop a canopy photosynthetic rate model of romaine lettuce using the three variables of CO₂ concentration, light intensity, and growth stage. The canopy photosynthetic rates of the lettuce were measured at five different CO₂ concentrations (600-2,200µmol·mol⁻¹), five light intensities (60-340µmol·m⁻²·s⁻¹), and four growth stages (5-20 days after transplanting) in three closed acrylic chambers ($1.0 \times 0.8 \times 0.5$ m). A simple multiplication model expressed by multiplying three single-variable models and the modified rectangular hyperbola model including photochemical efficiency, carboxyl-ation conductance, and dark respiration, which vary with growth stage, were also considered. In validation, the R² value was 0.923 in the simple multiplication model, while it was 0.941 in the modified rectangular hyperbola model. The modified rectangular hyperbola model appeared to be more appropriate than the simple multiplication model in expressing canopy photosynthetic rates. The model developed in this study will contribute to the determination of an optimal CO₂ concentration and light intensity with the growth stage of lettuce in plant factories.

Additional key words : growth stage, photosynthesis model, rectangular hyperbola model, regression model

Introduction

Plant factories using artificial lights are highly energyintensive production systems (Mills, 2012). These production systems require efficient production management (Li et al., 2016). Because photosynthetic rate is an indicator of the growth state and growth rate of crops, it plays an important role in constructing an efficient production system. Several models have already been established for photosynthesis and growth in various plant cultivation systems (Caporn 1989; Shimizu et al., 2008). Lettuce is one of the most suitable model crops for cultivation in plant factories, and the photosynthesis of lettuce has often been studied (Jung et al., 2016).

Existing photosynthesis models are shown as simple saturation curves using the two variables of light intensity and CO_2 concentration. The models based on light intensity and CO_2 concentration have been studied as rectangular hyperbolas (Acock et al., 1971; Thornley, 1976; Acock and Allen, 1985; Goudriaan et al., 1985). Recently, there have been studies on harmonizing the canopy level from the leaf photosynthesis level in crop modeling (Hikosaka et al., 2016). Since the light intensity depends on the position of leaves in the canopy, measuring the photosynthetic rate of a single leaf has limitations in representing the entire crop photosynthesis. The canopy photosynthetic rate is more closely related to crop yield than to the leaf photosynthetic rate. Therefore, when using photosynthesis models as an indicator of crop growth, it is necessary to measure the canopy photosynthetic rate in plant factories.

Most of the current photosynthesis models have rarely reflected time even though the crop response to growth stages is different (Perez-Peña and Tarara, 2015) and the efficiency of light utilization decreases as leaf area increases (Green, 1987; Leadly et al., 1990; Jung et al., 2016). In particular, it is important to maximize the resource utilization efficiency of plant factories, so the photosynthetic rate of cultivated crops should be known (Kozai, 2013). Also, understanding the change in light use efficiency of crops over time helps to establish the cultivation strategy of plant factories. Niinemets (2016) reported

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photo-plasticity, in which photosynthetic capacity varies with leaf age. Therefore, relative factors for photosynthetic reaction over time should be applied to models.

To express the photosynthetic rate for light intensity and CO_2 concentration, rectangular hyperbola equations and negative exponential equations have been used. These two equations are suitable for representing situations in which the rate of photosynthesis gradually increases to a specific saturation point. Thornley (1974) and Kaitala et al. (1982) suggested a simple model for the chemical reactions that take place in light and dark reactions of photosynthesis. This model is known to be suitable for expressing the photosynthetic rate with changes in light intensity and CO_2 concentration, but there are limitations to tracking changes in other environmental factors, such as temperature and growth stage.

Modifying the existing rectangular hyperbola model would be suitable for expressing the effects of other environmental factors. Although the widely used Farquhar, von Caemmerer, and Berry (FvCB) photosynthetic model expresses the change in photosynthetic rate according to various environmental factors (Farquhar et al., 1980), it does not provide a simple form that can include the three variables of light intensity, CO₂ concentration, and growth stage. Jung et al. (2017) established a photosynthetic rate model with light intensity, temperature, and growth period, but it is still necessary to construct a model including CO₂ concentrations, which are more important for photosynthesis. The objectives of this study were to analyze canopy photosynthetic rates at various combinations of CO₂ concentration, light intensity, and growth stage and to develop a canopy photosynthetic rate model based on a rectangular hyperbola equation.

Materials and Methods

1. Plant and Cultivation Conditions

Romain lettuce (*Lactuca sativa* L. cv. Asia Heuk Romaine) was hydroponically grown with deep-flow technique systems in plant factory modules at Seoul National University. Yamazaki nutrient solutions with an electrical conductivity of $1.2 \text{ dS} \cdot \text{m}^{-1}$ were supplied to the plants. Light-emitting diodes with a red : blue : white = 8 : 1 : 1 light spectrum were used as light sources (FGL-B1200; FC Poibe Co., Ltd., Seoul, Korea). Light intensity and air temperature in the growth chamber were maintained at 150 ± 20μ mol·m⁻²·s⁻¹ (light : dark = 16 h : 8 h) and $21 \pm 1^{\circ}$ C, respectively. The CO₂ concentration were maintained at 1,000 μ mol·mol⁻¹ for inducing more photosynthesis and relative humidity were maintained at 70 ± 5%. One hundred lettuce plants were grown for 20 days after transplanting (DAT) at a planting density of 16 plants·m⁻².

2. Measurement of Canopy Photosynthetic Rates

Lettuce plants were placed in three identical closed acrylic chambers $(1.0 \times 0.8 \times 0.5m)$ to measure the canopy photosynthetic rate. Four growth stages of lettuce were used for the measurements: 5, 10, 15, and 20 DAT. For calculating the canopy photosynthetic rate, changes in the CO₂ concentration inside the chamber were measured with a combination of light intensity and CO₂ concentration: five light intensities (60, 130, 200, 270, and 340µmol·m⁻²·s⁻¹) and five CO₂ concentrations (600, 1,000, 1,400, 1,800, and 2,200µmol·mol⁻¹). In order to obtain the canopy photosynthetic rate, the CO₂ consumption amount during the measurement period, excluding time lag, was multiplied by the chamber volume and divided by leaf area (Takahashi et al., 2008). Reductions in the internal CO₂ concentration due to the leakage of the chamber were calibrated by using the measured air exchange rate $(0.0077h^{-1})$. The CO₂ emission due to root respiration was considered negligible because it was around 0.69% of the total CO₂ consumption. The temperature and relative humidity were measured by using a sensor (S-VT200B; Soha Tech, Seoul, Korea) and collected with a datalogger (Campbell Scientific, Inc., Logan, UT, USA) every 5s. The air pressure inside the chambers was 1,013hPa. The temperature was controlled by Peltier devices, which were attached to the side wall of the chamber. Increases in relative humidity due to transpiration were controlled by silica gel installed in the chamber. Wind speeds around the plants were maintained at 0.3-0.5m·s⁻¹. The change in CO₂ concentration inside each chamber was measured by using a CO₂ gas analyzer (LI-820; LI-COR, Lincoln, NE, USA) every 5 seconds.

3. Establishment of a Three-Variable Simple Multiplication Model

One-variable models for the photosynthetic rate obtained by measuring the light intensity, temperature, or growth stage are known as a rectangular hyperbola, quadratic, and exponential model, respectively. In a previous study, a three-variable simple multiplication model was constructed to express the lettuce canopy photosynthetic rate (Jung et al., 2017). Since the temperature inside the chamber was kept constant in previous study, the simple multiplication model can be summarized as follows:

$$P = a * \frac{1 * C}{1 + b * C} * e^{c * t} + R \tag{1}$$

where *P* is canopy photosynthetic rate (μ molCO₂·m⁻²·s⁻¹), *I* is light intensity (μ mol·m⁻²·s⁻¹), *C* is CO₂ concentration (μ mol·mol⁻¹), *t* is DAT (d), and *R* is dark respiration (μ mol-CO₂·m⁻²·s⁻¹); *a*, *b*, and *c* are regression parameters. Nonlinear regression analysis was performed in the SPSS (IBM, New York, NY, USA) statistical program using the measured canopy photosynthetic rate according to light intensity, CO₂ concentration, and growth stage.

4. Establishment of a Three-Variable Modified Rectangular Hyperbola Model

A rectangular hyperbola model constructed by Acock et al. (1976) and supplemented by Kaitala et al. (1982) is expressed as follows:

$$P = \frac{\alpha * I * \beta * C}{\alpha * I + \beta * C} + R \tag{2}$$

where α and β are photochemical efficiency (µmolCO₂·mol⁻¹) and carboxylation conductance (s⁻¹), respectively.

The photochemical efficiency, carboxylation conductance, and dark respiration obtained by nonlinear regression analysis are shown on a two-dimensional plane with the growth stage as the X axis (Fig. 3). The photochemical efficiency, carboxylation conductance, and dark respiration were determined using empirical equations (Jung et al., 2017):

$$\alpha = a_1 * e^{b_1 * t} \tag{3}$$

$$\beta = a_2 * t^2 + b_2 * t + c_2 \tag{4}$$

$$R = a_3 * t^2 + b_3 * t + c_3 \tag{5}$$

where a_1 , a_2 , b_1 , b_2 , c_1 , and c_2 are regression parameters. The regression coefficients were determined through the nonlinear regression analysis of photochemical efficiency, carboxylation conductance, and dark respiration. The modified rectangular hyperbola model was constructed by substituting Eqs. 3-5 into Eq. 2.

5. Validation of the Canopy Photosynthetic Rate Models

One hundred additional lettuce plants were cultivated under the same environmental conditions as for the measurement of the canopy photosynthetic rate. The R^2 value was obtained through a regression analysis of the measured and estimated canopy photosynthetic rates.

Results

1. Changes in Shoot Fresh Weight and Leaf Area with Growth Stage

As the growth stage progressed, the fresh weight and leaf area of the lettuce exponentially increased and reached 97.3 \pm 2.94g and 872.0 \pm 44.50cm² at 20 DAT, respectively. (Fig. 1). The fresh weight and leaf area were regressed to the following exponential models with days after transplanting as the growth stage:

$$W = 20.485 * e^{0.086 * t} - 16 \tag{6}$$

$$LA = 272.32 * e^{0.071 * t} - 250 \tag{7}$$

where *W* is fresh weight (g) and *LA* is leaf area (cm²). Regression analysis showed that the R^2 values of Eqs. 6 and 7 were 0.991 and 0.984, respectively.

2. Regression Results of Simple Multiplication and Modified Rectangular Hyperbola Models

The canopy photosynthetic rate of the lettuce was plotted on a three-dimensional space using light intensity and CO_2 concentrations as the *X* and *Y* axes (Fig. 2). In the figure, black dots show the canopy photosynthetic rate



Fig. 1. Fresh weight and leaf area of the lettuce at 0 - 20 days after transplanting. Vertical bars represent the mean \pm SD (n = 15).

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Fig. 2. Canopy photosynthetic rates of the lettuce expressed with the simple multiplication model according to light intensity and CO₂ concentration at 5 (A), 10 (B), 15 (C), and 20 (D) days after transplanting.

obtained from actual measurement and the curved surface is the estimated canopy photosynthetic rate acquired from the simple multiplication model. The simple multiplication model (curved surface) was determined by Eq. 8:

$$P = \frac{0.106*I}{I+1.392*C} * e^{-0.024*t} + 0.020*t^2 - 0.581*t + 3.977$$
(8)

Regression analysis of Eq. 8 resulted in an R^2 value of 0.918. As light intensity increased, the canopy photosynthetic rate linearly increased. As the CO₂ concentration increased, the canopy photosynthetic rate increased and finally saturated. As the growth stage progressed, the maximum canopy photosynthetic rate decreased, while the saturation pattern of the canopy photosynthetic rate with light intensity and CO₂ concentration was maintained.

The photochemical efficiency, carboxylation conductance, and dark respiration are shown on a two-dimensional plane by substituting the regression coefficients in Table 1 (Fig. 3). The photochemical efficiency exponentially decreased as the growth stage progressed (Fig. 3A).

Table 1. Regression coefficients and R² values calculated through regression analysis in the modified rectangular hyperbola model according to growth stage.

Parameter -	Regression coefficient			D ²
	а	b	c	ĸ
Photochemical efficiency $(\mu molCO_2 \cdot \mu mol^{-1})$	0.078	-0.025		0.958
Carboxylation conductance (s ⁻¹)	-0.001	-23.220	24.083	0.871
Dark respiration $(\mu molCO_2 \cdot m^{-2} \cdot s^{-1})$	-0.02	-29.030	198.864	0.999

The carboxylation conductance and dark respiration appear as quadratic curves, which increased and decreased as the growth stage progressed (Figs. 3B and 3C). In the canopy photosynthetic rates plotted on the three-dimensional space with light intensity and CO_2 concentration (Fig. 4), the black dots show the canopy photosynthetic rate obtained from actual measurement and the curved surface is the estimated canopy photosynthetic rate acquired from the modified rect-



Fig. 3. Regressed photochemical efficiency (A), carboxylation conductance (B), and dark respiration (C) with the modified rectangular hyperbola model according to growth stage.

angular hyperbola model. The modified rectangular hyperbola model is expressed by Eq. 9. As light intensity increased, the canopy photosynthetic rate linearly increased. As the CO_2 concentration increased, the canopy photosynthetic rate increased and gradually saturated. As the growth stage progressed, the maximum canopy photosynthetic rate decreased, while the saturation pattern of the canopy photosynthetic rate with light intensity and CO_2 concentration was maintained.

$$P = \frac{e^{-0.025*t} * (0.001*t^2 - 0.023*t + 0.024)*I*C}{0.013*t^2 - 0.298*t + 0.309 - e^{-0.025*t}*I*C} + 0.020*t^2 - 0.581*t + 3.977$$
(9)

3. Validation of the Canopy Photosynthetic Rate Models For validation, the regression analysis result for the two models was plotted on the 1 : 1 line (Fig. 5). The R^2 value and RMSE of the simple multiplication model was 0.923 and 2.114, and the R^2 value and RMSE of the modified rectangular hyperbola model was 0.941 and 1.622, which was slightly higher than that of the simple multiplication model. The canopy photosynthetic rates ranged from 2.5-23µmol·m⁻²·s⁻¹ for both measured and calculated values.

Discussion

The plant growth stage is considered important when constructing practical photosynthetic models because plants have different responses at different growth stages. Fresh weight and leaf area were used as indicators for classification into germination, vegetation, and reproduction stages (Mokhtarpour et al., 2010), and the photosynthetic rate was also used (Li et al., 2009; Kim et al., 2013). Although the classification of growth stages can vary depending on the characteristics of the plants, Lin (2001) reported that fresh weight was the most economic key element in classifying leaf vegetables. Therefore, it was appropriate to separate the growth stage of leaf vegetables by fresh weight in this study.

The R^2 values of canopy photosynthetic rates obtained by using the developed models varied from 0.95 to 0.78 at each stage with slight overestimations (Fig. 4). This seems to be because the characteristics of the canopy structure were not sufficiently considered in the photosynthesis model (Monsi et al., 1973). Kim et al. (2016) indicated that the photosynthetic rate has a close relationship with the absorption, reflection, and penetration of light in the canopy structure. Therefore, a more precise estimation of light interception by the canopy could increase the R^2 values.

Olessen and Grevesen (1997) also reported that light extinction coefficients differed depending on plant type. Previous research has indicated light extinction coefficients of 0.63-0.86 for certain vegetables, such as tomato, cucumber, and paprika, and 0.40-0.65 for cauliflower. The light extinction coefficients depend on leaf and canopy conditions as well as environmental factors. Particularly in closed plant production systems, light interception significantly differs depending on the levels of light reflection and light diffusion.

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Fig. 4. Canopy photosynthetic rates of the lettuce expressed with the modified rectangular hyperbola model according to light intensity and CO₂ concentration at 5 (A), 10 (B), 15 (C), and 20 (D) days after transplanting.



Fig. 5. Validation of the simple multiplication and modified rectangular hyperbola models by comparing measured and estimated canopy photosynthetic rates.

Among various environmental factors, light is the most influential variable in plant growth and development (Inada and Yabumoto, 1989). Due to the characteristics of the lettuce, the higher the light intensity, the higher the growth

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obtained (Pavlou et al., 2007). The increase in light intensity increased the nitrate reductase in lettuce and it should affect the photosynthesis (Gaudreau et al., 1995). In order to increase light utilization efficiency and improve crop productivity, light sources installed in plant factories should be designed in a form suitable for photosynthesis (Massa et al., 2008).

Some studies have developed the photosynthetic rate models and used the models to predict crop productivity (Kim and Lieth, 2012). In addition, the enrichment of CO_2 concentration promotes the growth of lettuce (Caporn, 1989; Campbell et al., 1990). However, plant cultivation at saturation points of light intensity and CO_2 concentration is economically inefficient. When the photosynthetic rate model of the crop has been established, the economic CO_2 fertilization level can be determined. The modeling of photosynthetic rates using light intensity and CO_2 concentration at each growth stage will be helpful in constructing strategic plant production systems. It is expected that the developed model will contribute to the determination of adequate CO_2 concentration and light intensity conditions with growth stage in plant factories.

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Literature cited

- Acock, B.C., J.H.M. Thornley, and J.W. Wilson. 1971. Photosynthesis and energy conversion. In: Wareing P.F., Cooper, J.P. (Eds.), Potential crop production. Heinemmann Educational Publishers, London, United Kingdom. pp. 43-75.
- Acock, B.C. and L.H. Allen Jr. 1985. Crop responses to elevated carbon dioxide concentrations. In: Strain, B.R., and J.D. Cure. (Eds.), Direct effects of increasing carbon dioxide on vegetation. US Department of Energy, Office of Energy Research, Office of Basic Energy Sciences, Carbon Dioxide Research Division. pp. 53-99.
- Campbell, W.J., L.H. Allen, and G. Bowes. 1990. Response of soybean canopy photosynthesis to CO₂ concentration, light, and temperature. J. Exp. Bot. 41: 427-433.
- Caporn, S.J.M. 1989. The effects of oxides of nitrogen and carbon dioxide enrichment on photosynthesis and growth of lettuce (*Lactuca sativa* L.). New Phytol. 111: 473-481.
- Farquhar, G.D., S. von Caemmerer, and J.A. Berry. 1980. A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species. Planta 149: 78-90.
- Gaudreau, L., J. Charbonneau, L.P. Vezina, and A. Gosselin. 1995. Effects of photoperiod and photosynthetic photon flux on nitrate content and nitrate reductase activity in greenhouse-grown lettuce. J. Plant Nutr. 18: 437-453.
- Goudriaan, J., H.H. van Laar, H. van Keulen, and W. Louwerse. 1985. Photosynthesis, CO₂ and plant production. In: Day, W., and R.K. Arkin. (Eds.), Wheat growth and modeling, vol. 86 (NATO ASI Series A). Plenum Press, New York. pp. 107-122.
- Green, C.F. 1987. Nitrogen nutrition and wheat growth in relation to absorbed solar radiation. Agric. Forest Meteorol. 41: 207-248.
- Hikosaka, K., T.O. Kumagai and A. Ito. 2016. Modeling canopy photosynthesis. In: Canopy Photosynthesis: From basics to applications, Springer. pp. 239-268.
- Inada, K. and Y. Yasumoto. 1989. Effects of light quality, day length and periodic temperature variation on the growth of lettuce and radish plants. Jpn. J. Crop Sci. 58: 689-694.
- Jung, D.H., D. Kim, H.I. Yoon, T.W. Moon, K.S. Park, and J.E. Son. 2016. Modeling the canopy photosynthetic rate of romaine lettuce (*Lactuca sativa* L.) grown in a plant factory

at varying CO₂ concentrations and growth stages. Hortic. Environ. Biotechnol. 57: 487-492.

- Jung, D.H., H.I. Yoon, and J.E. Son. 2017. Development of a three-variable canopy photosynthetic rate model of romaine lettuce (*Lactuca sativa* L.) grown in plant factory modules using light intensity, temperature, and growth stage. Protect. Hortic. Plant Fact. 26: 268-275.
- Kaitala, V., P. Hari, E. Vapaavuori, and R. Salminen. 1982. A dynamic model for photosynthesis. Ann. Bot. 50: 385-396.
- Kim, J.H., J.W. Lee, T.I. Ahn, J.H. Shin. K.S. Park, and J.E. Son. 2016. Sweet pepper (*Capsicum annuum* L.) canopy photosynthesis modeling using 3D plant architecture and light ray-tracing. Front. Plant Sci. 7: 1321.
- Kim, S.H., J.H. Jeong, and L.L. Nackley. 2013. Photosynthetic and transpiration responses to light, CO₂, temperature, and leaf senescence in garlic: Analysis and modeling. HortScience 138: 149-156.
- Kim, W.S. and J.H. Lieth. 2012. Simulation of year-round plant growth and nutrient uptake in *Rosa hybrida* over flowering cycles. Hortic. Environ. Biotechnol. 53: 193-203.
- Kozai, T. 2013. Resource use efficiency of closed plant production system with artificial light: Concept, estimation and application to plant factory. Proc. Jpn. Acad., Ser. B 89: 447-461.
- Leadley, P.W., J.F. Reynolds, R. Flagler, and A.S. Heagle. 1990. Radiation utilization efficiency and the growth of soybeans exposed to ozone: a comparative analysis. Agric. Forest Meteorol. 51: 293-308.
- Li, J., Z. Zou, and X. Wang. 2009. Effect of muskmelon leaf age on photosynthesis rate and other physiological parameters at different light density. Acta Hortic. 893: 785-790.
- Li, K., Z. Li, and Q. Yang. 2016. Improving light distribution by zoom lens for electricity savings in a plant factory with light-emitting diodes. Front. Plant Sci. 7: 92.
- Lin, W.C. 2001. Crop modelling and yield prediction for greenhouse-grown lettuce. In: IV International symposium on models for plant growth and control in greenhouses: modeling for the 21st century -agronomic and greenhouse crop models. pp. 159-164.
- Massa, G.D., H.H. Kim, R.M. Wheeler, and C.A. Mitchell. 2008. Plant productivity in response to LED lighting. Hort-Science 43: 1951-1956.
- Mills, E. 2012. The carbon footprint of indoor cannabis production. Energy Policy 46: 58-67.
- Mokhtarpour, H., C.B. Teh, G Saleh, A.B. Selamat, M.E. Asadi, and B. Kamkar. 2010. Non-destructive estimation of maize leaf area, fresh weight, and dry weight using leaf length and leaf width. Commun. Biometry Crop Sci. 5: 19-26.
- Monsi, M. 1960. Dry-matter reproduction in plants 1. Schemata of dry-matter reproduction. Bot. Mag. 861: 81-90.
- Niinemets, Ü. 2016. Leaf age dependent changes in withincanopy variation in leaf functional traits: a meta-analysis. J. Plant Res. 129: 313-338.

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- Olesen, J.E. and K. Grevsen. 1997. Effects of temperature and irradiance on vegetative growth of cauliflower (*Brassica oleracea* L. botrytis) and broccoli (*Brassica oleracea* L. italica). J. Exp. Bot. 48: 1591-1598.
- Pavlou, G.C., C.D. Ehaliotis, and V.A. Kavvadias. 2007. Effect of organic and inorganic fertilizers applied during successive crop seasons on growth and nitrate accumulation in lettuce. Sci. Hortic. 111: 319-325.
- Perez-Peña, J. and J. Tarara. 2015. A portable whole canopy gas exchange system for several mature field-grown grapevines. VITIS - J. Grapevine Res. 43: 1-7.

Shimizu, H., M. Kushida, and W. Fujinuma. 2008. A growth

model for leaf lettuce under greenhouse environments. Environ. Cont. Biol. 46: 211-219.

- Takahashi, N., P.P. Ling, and J.M. Frantz. 2008. Considerations for accurate whole plant photosynthesis measurement. Environ. Cont. Biol. 46: 91-101.
- Thornley, J.H.M. 1974. Light fluctuations and photosynthesis. Ann. Bot. 38: 363-373.
- Thornley, J.H.M. 1976. Mathematical models in plant physiology: A quantitative approach to problems in plant and crop physiology. Academic Press, London. p. 318.

광도, CO₂ 농도 및 정식 후 생육시기에 따른 식물공장 재배 상추의 군락 광합성 모델 확립

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적 요. 작물의 생산량은 광합성과 밀접한 관계가 있으며, 광합성 속도는 다양한 환경 요인에 의해 변화한다. 광합성 속도는 작물의 생육 상태나 생육 속도를 판단하는 지표로 사용되며, 작물 재배 시설을 구축하는 데 고 려해야 하는 중요한 요인이다. 이 연구의 목적은 광도, CO₂ 농도 및 생육 단계에 의해 변화하는 로메인 상추 의 군락 광합성 속도 모델을 개발하는 것이다. 군락 광합성 속도는 정식 후 5, 10, 15, 20 일차에서 5단계의 CO₂ 농도(600-2,200µmol·mol⁻¹)와 5단계의 광조건(60-340µmol·m⁻²·s⁻¹)이 처리된 3개의 밀폐 아크릴 챔버(1.0 × 0.8 × 0.5m) 내에서 측정하였다. 먼저 세 가지 환경 요인을 사용하는 식들을 곱하여 만든 단순곱 모델을 구성 하였다. 이와 동시에 생육 시기에 따라 변화하는 광화학 이용효율과 카르복실화 컨덕턴스, 호흡에 의한 이산화 탄소 발생 속도를 포함하는 수정 직각쌍곡선 모델을 구성하여 단순곱 모델과 비교하였다. 검증 결과, 단순곱 모델의 R²는 0.923이었으며, 수정 직각쌍곡선 모델의 R²는 0.941을 나타내었다. 따라서 수정 직각쌍곡선 모델 이 광도, CO₂ 농도, 생육 단계의 3 변수에 따른 군락 광합성 속도를 표현하는 데 더욱 적합한 것으로 판단하 였다. 본 연구에서 개발된 군락 광합성 모델은 식물공장에서 상추 재배를 위해 생육 단계별로 설정해야 할 최 적의 광도와 CO₂ 농도를 결정하는 데 도움이 될 것으로 생각된다.

추가 주제어: 생육 단계, 광합성 모델, 직각쌍곡선 모델, 회귀 모델