



Administration of Alpha_{s1}-Casein Hydrolysate Increases Sleep and Modulates GABA_A Receptor Subunit Expression

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Abstract

Sleep is the most basic and essential physiological requirement for mental health, and sleep disorders pose potential risks of metabolic and neurodegenerative diseases. Tryptic hydrolysate of α_{s1} -casein (α_{s1} -CH) has been shown to possess stress relieving and sleep promoting effects. However, the differential effects of α_{s1} -CH on electroencephalographic wave patterns and its effects on the protein levels of γ -aminobutyric acid A (GABA_A) receptor subtypes in hypothalamic neurons are not well understood. We found α_{s1} -CH (120, 240 mg/kg) increased sleep duration in mice and reduced sleep-wake cycle numbers in rats. While α_{s1} -CH (300 mg/kg) increased total sleeping time in rats, it significantly decreased wakefulness. In addition, electroencephalographic theta (θ) power densities were increased whereas alpha (α) power densities were decreased by α_{s1} -CH (300 mg/kg) during sleep-wake cycles. Furthermore, protein expressions of GABA_A receptor β_1 subtypes were elevated in rat hypothalamus by α_{s1} -CH. These results suggest α_{s1} -CH, through GABA_A receptor modulation, might be useful for treating sleep disorders.

Key Words: Sleep, α_{s1} -CH, Electroencephalogram, GABA_A receptor

INTRODUCTION

Sleep is the most basic and essential physiological requirement for maintaining health, mental stability, and memory retrieval (Lo *et al.*, 2016; Schouten *et al.*, 2017). Based on electroencephalogram frequency-band rhythms, that is, delta (δ), theta (θ), alpha (α), beta (β), and gamma (γ) rhythms, sleep is classified into five stages, namely, non-rapid eye movement (NREM) sleep (stages I to IV) and rapid eye movement (REM) sleep (stage V), which occur in alternating cycles (Doroshenkov *et al.*, 2007), although recently an automatic, 6-stage, electroencephalographic sleep classification method was proposed (Diykh *et al.*, 2016). While δ rhythm dominates NREM sleep, θ rhythm is commonly observed during REM sleep (Doroshenkov *et al.*, 2007; Luppi *et al.*, 2017). Accordingly, electroencephalography (EEG) can be employed to identify sleep disorders and to aid the predictions of treatment outcomes in various psychiatric diseases (Olbrich *et al.*, 2015).

Sleep disorders not only reduce quality of life but also serve

as risk factors of dementia (Mishima, 2016) and metabolic diseases, like atherosclerosis (Tobaldini *et al.*, 2017), and hence, early intervention is clinically relevant as it potentially mitigates harmful consequences. Recently developed drugs that have been used to treat insomnia, but can have undesirable side effects (Kay-Stacey & Attarian, 2016). Furthermore, reports indicate lotus leaf extract augments hypnosis by binding to γ -aminobutyric acid A (GABA_A) receptor (Tian and Liu, 2015; Yan *et al.*, 2015), and that consuming dairy products supports sleep in a better way (Kitano *et al.*, 2014). Alpha (α)_{s1}-casein hydrolysate (α_{s1} -CH) is a milk protein with reported chronic stress relieving properties (Guesdon *et al.*, 2006; Kim *et al.*, 2007). However, although the tryptic hydrolysate of α_{s1} -casein appears to improve sleep quality (Dela Peña *et al.*, 2016), little data is available on the way it affects pentobarbital-induced sleep in mice or influences EEG band rhythms during stages of sleep. Furthermore, it has not been determined whether α_{s1} -CH mediates its hypnotic action in mice via GABA_A receptor in hypothalamus. Therefore, we investigated the effects of

Open Access <https://doi.org/10.4062/biomolther.2017.083>

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Received Apr 6, 2017 Revised Aug 29, 2017 Accepted Sep 19, 2017

Published Online Jan 9, 2018

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α_{s1} -CH on sleep duration, sleep quality as determined by electroencephalography, and on the protein expression of GABA_A receptor subunits ($\alpha 1$, $\beta 1$, $\gamma 3$) in the rat hypothalamus.

MATERIALS AND METHODS

Chemicals

Bovine α_{s1} -casein hydrolysate (α_{s1} -CH), commercialized as Lactium[®], was obtained from Ingredia (Arras, France). Pentobarbital sodium was obtained from Hanlim Pharm (Seoul). Diazepam and other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Animals and treatments

Male C57BL/6 mice (28-30 g) and Sprague-Dawley rats (male, 260-280 g) were purchased from the Orient Bio (Seoul, Korea) and allowed free access to water and food. Mice were grouped into six per cage, and maintained in an ambient atmosphere at 23°C under a 12 h diurnal light cycle. Rodents were divided into groups: 6 groups of mice for sleep testing, 3 groups of rats for EEG recording, and 3 groups of rats for western blotting. All behavioral experiments were carried out in a nearby room maintained where under the same environmental conditions. Experiments were conducted according to Animal Care and Use Guidelines of the School of Medicine, Ewha Womans University, Korea.

Mice were given a single dose (30-240 mg/kg, p.o.) of α_{s1} -CH or saline 30 min prior to an injection of pentobarbital sodium (42 mg/kg, i.p.) to determine the onset and duration of sleep, as previously described by Ma *et al.* (2009) with slight modification. Time elapsed between disappearance (sleep onset) and reappearance of righting reflex (up to a maximum of 2 h) was defined as sleep duration. Experiments were performed in mouse cages with aspen bedding. Animals that did not sleep within 15 min after pentobarbital injection were excluded. Rats were treated with α_{s1} -CH (150 or 300 mg/kg) orally once per day for 3 days before electroencephalography (EEG) or western blotting. EEG recordings were started at 2 hrs after last treatment, and hypothalami were collected at 6 hrs after last treatment for western blotting.

Electroencephalography

Rats were anesthetized with pentobarbital (50 mg/kg, i.p.) and a transmitter was implanted for EEG recording via telemetry as previously described (Sanford *et al.*, 2006). Briefly, in each case, the body of the transmitter was subcutaneously implanted just posterior to the scapula using three sutures for stabilization. The transmitter electrodes were led subcutaneously to the skull, and their bare ends were placed in contact with the dura through holes in the skull. Electrodes were anchored to the skull with screws and dental cement. All surgical procedures were performed stereotaxically under aseptic conditions.

For telemetric recording of cortical EEG signals, transmitter gain was set at -0.5/+0.5 volts per unit. Raw output signals, which ranged from 0.5 to 0.0 Hz, were processed using a Data Sciences analog converter and routed to an analog-to-digital (AD) converter (Eagle PC30, Data Sciences International, St. Paul, MN, USA), which digitized EEG and activity signals. Subsequently data were transferred to a computer and graphically displayed. An on-line fast Fourier transformation (FFT)

program was used to analyze EEG data and generate power density values from 0.0 to 20.0 Hz at a resolution of 0.5 Hz. FFT data were further averaged between 0 to 20 Hz at 10-s intervals. Sleep data and FFT results were saved to hard disk every 10 s for additional off-line analysis. Number of animal movements related to telemetry receiver generated transistor-transistor logic pulses were viewed as measures of activity. Data were gathered on the 1st and 3rd days after α_{s1} -CH treatment and percentage power densities were calculated. EEG signals were measured for 6 hrs between 11:00 am 5:00 pm. Each group contained 5-6 rats.

Determination of sleep behaviors using EEG signals

Times elapsed in wakefulness, NREM sleep, or REM sleep was determined using digitized data using animal sleep analysis software Sleep-Sign 2.1 (Kissei Comtec, Matsumoto, Japan). Briefly, this software identifies wakefulness as high-frequency, low-amplitude EEG, NREM sleep as spikes interspersed with slow waves, and REM sleep as δ -waves (0.75 to 4.0 Hz) with θ -wave activity (5.0 to 9.0 Hz) of peak frequency 7.5 Hz.

Western blotting

Six hrs after the last administration of α_{s1} -CH, rats were decapitated and brains were quickly removed and chilled in ice-cold saline. Coronal sections were obtained using a rodent brain matrix (ASI Instruments, Warren, MI, USA). Hypothalami were dissected out, immediately frozen on dry ice, and stored at -80°C. Frozen tissue samples were homogenized in PRO-PREP protein-extraction solution (Intron Biotechnology Inc., Seongnam, Korea) and centrifuged at 13,000 rpm at 4°C for 20 min. Protein concentrations in supernatants were determined and 40 μ g aliquots were subjected to polyacrylamide gel electrophoresis. Proteins were then transferred to polyvinylidene fluoride membranes (Hybond-P; GE Healthcare, Amersham, UK) using a wet transfer system, and membranes were incubated with one of the following primary antibodies: rabbit anti-GABA_A $\alpha 1$ polyclonal antibody (diluted 1:2,000 in TBS containing 0.5% Tween 20; Abcam, Cambridge, UK); rabbit anti-GABA_A $\beta 1$ polyclonal antibody (diluted 1:2,500 in TBS containing 0.5% Tween 20); rabbit anti-GABA_A $\gamma 3$ polyclonal antibody (diluted 1:2,500 in TBS containing 0.5% Tween 20); rabbit anti-glutamic acid decarboxylase (GAD) polyclonal antibody (diluted 1:2,000 in TBS containing 0.5% Tween 20); and β -actin antibody. Membranes were then washed and incubated with the horseradish peroxidase-conjugated goat anti-rabbit secondary antibody (diluted 1:3,000 in TBS containing 0.5% Tween20). Immunoreactive bands were developed using a chemiluminescence detection kit (Roche Diagnostics, Mannheim, Germany) and quantitative analysis was performed by densitometric scanning.

Statistical analysis

Sleep onset and duration data were analyzed using analysis of variance (ANOVA). The Newman-Keuls test was used to perform intergroup comparisons. Values were expressed as means \pm SEM, and statistical significance was accepted for *p* values <0.05.

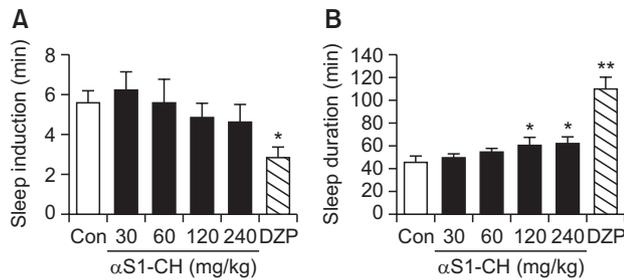


Fig. 1. Effects of α_{S1} -CH on the onset and duration of sleep in pentobarbital-treated mice. Mice were food-deprived for 12 h prior to being treated with α_{S1} -CH (30-240 mg/kg) or diazepam (DZP, 1mg/kg, i.p.). Sleep latency (A) and total sleeping time (B) were recorded for 120 min after injecting pentobarbital (42 mg/kg, i.p.). Columns contain mean values and SEMs (n=8-10) as determined by ANOVA. Comparisons were made using the Newman-Keuls test. * p <0.05, ** p -values of <0.01 were considered significant as compared with pentobarbital-treated controls.

RESULTS

Pretreatment with α_{S1} -CH prolonged sleep duration in mice

It has been demonstrated α_{S1} -casein hydrolysate protects rat from chronic mild stress-induced sleep disorders (Guesdon *et al.*, 2006). In our preliminary experiment, α_{S1} -CH showed an anxiolytic effect at relatively low doses (25, 50 mg/kg) in mice (data not shown). Therefore, we investigated whether α_{S1} -CH improves sleep duration in pentobarbital-treated mice. Mice treated with α_{S1} -CH in the dose range 30-240 mg/kg tended to have lower sleep-onset times (Fig. 1A). However, this effect of α_{S1} -CH was not significant as compared with pentobarbital controls. Diazepam (1 mg/kg, i.p.) exhibited significantly earlier sleep-onset times than that of control after pentobarbital treatment. One-way ANOVA showed significant differences in sleep induction between control and diazepam group [F (5,50)=5.0, p <0.01]. In contrast, the duration of sleep significantly elevated when mice were treated with α_{S1} -CH at higher doses (120 or 240 mg/kg, p.o.) [F (5,51)=15.02, p <0.01] (Fig. 1B).

Rats pretreated with α_{S1} -CH had fewer sleep-wake cycles

Sleep-wake cycle disruption has been associated with stress, which suggests that reducing the number of sleep-wake cycles may provide relief from neurodegenerative diseases (Cedernaes *et al.*, 2017). Therefore, we investigated whether α_{S1} -CH could reduce the number of sleep-wake cycle disruptions in rats. In the preliminary test, administration of α_{S1} -CH at doses of 50 or 100 mg/kg did not significant affect sleep/wake cycles or EEG patterns in rats, and thus, the dose of α_{S1} -CH was increased to 150 or 300 mg/kg. We found α_{S1} -CH at 300 mg/kg significantly reduced the number of sleep-wake cycles by ~50% (Fig. 2). Furthermore, total time awake was reduced by α_{S1} -CH pretreatment and total asleep was increased (Fig. 3). Although REM sleep was decreased and NREM sleep was increased after treatment with α_{S1} -CH, no significant differences were found between treatment groups (Fig. 3).

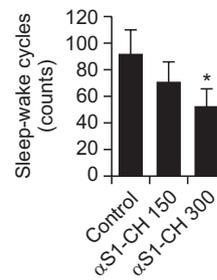


Fig. 2. Effects of α_{S1} -CH on sleep-wake counts. α_{S1} -CH (150, 300 mg/kg) was orally administered to rats once daily for 3 days. Sleep-wake cycles were measured by EEG for 6 hrs and analyzed using Sleep Sign 2.1 software. Values were compared using the Newman-Keuls test. Column contain mean values and SEMs (n=5-6). * p -values of <0.05 were considered significant as compared with naïve controls.

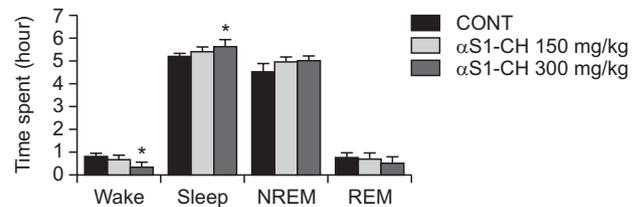


Fig. 3. Differential effects of α_{S1} -CH on sleeping stages in rats. Wakefulness, total sleep, REM sleep, and NREM sleep were determined after administering α_{S1} -CH (150, 300 mg/kg) orally to rats once daily for 3 days. Sleeping was determined using telemetric cortical EEG records and analyzed using Sleep Sign2.1 software. To compare each group versus naïve control, we used the Newman-Keuls test. Columns represent mean values and SEMs (n=5-6). * p <0.05 compared with naïve controls.

The effect of α_{S1} -CH on frequency bands of EEG during sleep-wake cycles

Protein α_{S1} -CH (150 or 300 mg/kg, p.o.) was administered to rats once per day for 3 days. Wakefulness, REM sleep and NREM sleep were monitored using the power densities of delta (δ), theta (θ), and alpha (α) frequency bands in rats treated without or with α_{S1} -CH (150 or 300 mg/kg). Whereas the percentage of θ power density was significantly increased by treatment with α_{S1} -CH (300mg/kg) in sleep-wake cycles, δ frequency bands showed negligible differences. On the other hand, treatment with α_{S1} -CH at 300 mg/kg decreased the percentage of α power density (Fig. 4).

α_{S1} -CH modulated the expression levels of β_1 and γ^3 subtypes of GABA_A receptor in the rat hypothalamus

GABA_A receptor subtypes in neuronal tissue have been reported to be targets for insomnia treatment (Luppi *et al.*, 2017). We investigated whether the protein expressions of the GABA_A receptor subunits α_1 , β_1 , and γ^3 were modulated in the hypothalami of rats treated with α_{S1} -CH. Treatment using α_{S1} -CH at 150 mg/kg or 300 mg/kg increased the protein expression of β_1 , but the protein expression level of α_1 and glutamic acid decarboxylase (GAD_{65/67}; catalyzes the formation of GABA in neuronal tissues) were unaltered (Fig. 5). Although the level of γ^3 tended to be elevated by α_{S1} -CH treatment, this was not statistically significant.

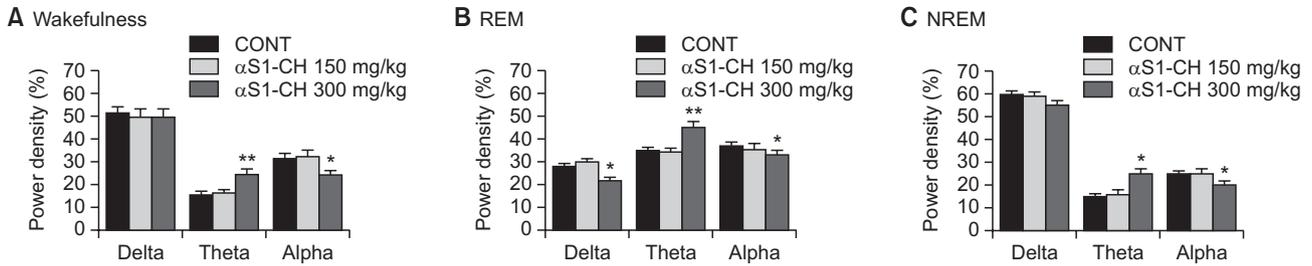


Fig. 4. The effects of α_{s1} -CH on EEG power densities during wakefulness (A), REM sleep (B) and NREM sleep (C) in rats. α_{s1} -CH (150, 300 mg/kg) was orally administrated to rats once daily for 3 days. Power densities were classified as δ -wave, θ -wave, and α -wave densities. The Newman-Keuls test was used to compare groups versus the naïve control group. Columns represent mean values and SEMs (n=5-6). *p*-values of <0.05 or <0.01 were considered significant (as indicated by * or **, respectively).

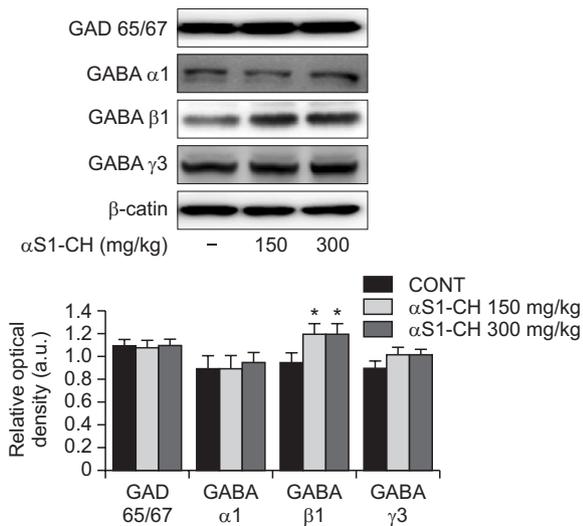


Fig. 5. Modulation of glutamic acid decarboxylate (GAD) and GABA_A receptor subunits in the rat hypothalamus by α_{s1} -CH (150, 300 mg/kg). After three days of consecutive treatment by α_{s1} -CH, hypothalamic tissues were isolated, homogenized, and immunoblotted. Immunoreactive band intensities were measured by densitometry. Protein levels were normalized versus GAPDH. Columns contain mean values and SEMs (n=3). The Newman-Keuls test was used to compare groups versus the naïve control group. **p*-values of < 0.05 were considered significant.

DISCUSSION

Considering the importance of nutrition-based hypnosis over that of recently developed drugs with undesired side effects, our report on α_{s1} -CH has merit with respect to prolonged duration of sleep, and fewer sleep-wake cycles, which suggests a new avenue for developing alternative therapeutic options against the insomnia experienced during stressful conditions. Sleep disorders are associated not only with mental problems (Yu *et al.*, 2013) but are also linked to various health conditions, such as, metabolic disease and reduced testosterone levels accompanied by altered sexual behavior (Alvarenga *et al.*, 2015). Recently, a disordered protein architecture of receptors was suggested to be related to sleep problems (Tou and Chen, 2014), which implies a complex mechanism underlies insomnia. Several drugs that ameliorate insomnia have been developed, but many are associated with unwanted side

effects (Kripke, 2016; Sirdifield *et al.*, 2017). Alternative options have been sought, such as, acupuncture (Lee and Lim, 2016) and the use of natural products (Shi *et al.*, 2014). Milk contains a wide variety of bioactive peptides, including those in tryptic hydrolysate of α_{s1} -casein, which has been reported to modulate the architecture of sleep (Dela Peña *et al.*, 2016).

Little is known of EEG band rhythms and membrane receptor expressions in hypothalamic neurons, and EEG parameters are considered essential during sleep examinations and are used to evaluate sleep patterns or problems. Therefore, we investigated the effects of α_{s1} -casein in mouse model of pentobarbital-induced sleep.

We found α_{s1} -CH (30-240 mg/kg) did not modulate sleep onset, but that at 120 or 240 mg/kg it prolonged sleep duration in mice. Similarly, α_{s1} -CH at 300 mg/kg reduced the number of sleep-wake counts nearly by a half in rats. Moreover, total sleeping time was increased but wakefulness was diminished by α_{s1} -CH at 300 mg/kg. Together, these findings strongly support previous findings that suggested the tryptic hydrolysate of α_{s1} -casein had sleep promoting properties. Pena *et al.* also showed that EEG δ waves increased in NREM sleep whereas α waves decreased (Dela Peña *et al.*, 2016). In the present study, we also found the power density of θ waves were significantly increased and α densities significantly decreased by α_{s1} -CH (300 mg/kg) in rats. It has been known δ waves are slow waves related to the governance of sleep, and that α waves are high frequency waves related to sedatives and hypnotics (Stahl, 2008). Interestingly, it was reported that δ rhythm is predominantly seen during NREM sleep in contrast to θ rhythm, which is usually observed during REM sleep (Luppi *et al.*, 2017). In general, in our EEG signals, θ waves were significantly enhanced during REM sleep, NREM sleep, and wakefulness when rats were treated with α_{s1} -CH (300 mg/kg), which indicates higher concentrations of α_{s1} -CH influence EEG signals. In contrast to θ rhythms, α rhythms are present during waking (Doroshenkov *et al.*, 2007), and in the present study were found to be decreased by pretreatment with α_{s1} -CH at higher concentration (300 mg/kg). This result may seem contradictory given the aforementioned EEG patterns of REM and NREM, and we cannot provide an explanation for this result. However, Rajaratnam *et al.* showed that melatonin administration does not significantly change δ or α activities in man (Rajaratnam *et al.*, 2004), and suggested melatonin facilitates rather than induces sleep. This might also be the case for α_{s1} -CH.

Despite controversies regarding the properties, functions,

and subunit arrangements of GABA_A receptors, they have been established to be pentameric ligand-gated channels that negatively mediate neurotransmission in the central nervous system, (Puthenkalam *et al.*, 2016; Wongsamitkul *et al.*, 2016). Dela Peña *et al.* (2016) suggested GABA_A receptor subunits play a role in mediating α_{S1} -CH induced sleeping based on results obtained using bicuculline, a competitive GABA_A receptor antagonist, and that the dose-dependent increase in chloride ion influx induced by α_{S1} -CH in cultured human neuroblastoma cells was blocked by bicuculline. In the present study, we found the protein expression of the β_1 receptor subunit of GABA_A was increased in the hypothalami of rats treated with α_{S1} -CH (150, 300 mg/kg), but that α_1 and GAD^{65/67} protein levels were unchanged. The activation of GAD plays an important role in the GABAergic system because GABA is generated from glutamate by the action of GAD. In the present study, protein levels of GAD^{65/67} were unaltered by α_{S1} -CH administration, suggesting α_{S1} -CH might not modulate GABA generation. Nevertheless, our report indicates the importance of β_1 subunits of GABA_A receptor in α_{S1} -CH enhanced sleep, though further studies are warranted on receptor subtypes and their arrangements in GABA_A receptor (Mohler *et al.*, 2005; Wongsamitkul *et al.*, 2016), especially since Liang and Marks (2014) observed the involvement of the GABA_A γ^2 receptor subunit in REM sleep.

In the present study, we found α_{S1} -CH significantly enhanced pentobarbital-induced sleep duration in mice, increased total sleep, and EEG θ wave during sleep in rats. Given increased protein expressions of GABA_A receptor β_1 subunits after α_{S1} -CH treatment observed in rats, further work is required to explore other GABA_A receptor subtypes and their arrangements to clearly delineate the sleep-enhancing effect of α_{S1} -CH. Nonetheless, our findings suggest α_{S1} -CH dietary supplementation could be deployed to treat sleep disorders.

ACKNOWLEDGMENTS

This research was supported by a grant from the National Research Foundation of Korea (NRF) funded by the Korean Ministry of Science, ICT & Future Planning (MRC, 2010-0029355) and by the Chungbuk Bio International R&D Project (2014-1-01).

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