

Candelaria asiatica, an Ignored New Species from South Korea

Dong Liu and Jae-Seoun Hur

Korean Lichen Research Institute (KoLRI), Suncheon National University, Suncheon, Korea

ABSTRACT

The genus *Candelaria* is characterized by a micro-foliose to micro-fruticose thallus and contains eight species, two of which were reported in South Korea. During the excursion of a Korean lichen flora investigation, some suspected *Candelaria concolor* specimens were collected, and their morphological, chemical, molecular phylogenetic, and geographic analyses were conducted. The samples eventually proved to be a new species, *Candelaria asiatica*, which can be recognized by a small, yellow lobate thallus with a pulverulent surface, and a fragile lobe margin with blastidia or phyllidia-like lobules.

ARTICLE HISTORY

Received 28 March 2018
Revised 14 July 2018
Accepted 18 August 2018

KEYWORDS

Candelariaceae; taxonomy; phylogeny; Asia

1. Introduction

Candelaria A. Massal. is a small lichen genus characterized by a micro-foliose to micro-fruticose yellowish thallus and 8- or polyspored asci. This genus comprises eight species: *Candelaria antarctica* (Js. Murray) Poelt, *C. crawfordii* (Müll. Arg.) P.M. Jørg. & D.J. Galloway, *C. concolor* (Dicks.) Arnold, *C. fibrosa* (Fr.) Müll. Arg., *C. fibrosoides* M. Westb. & Frödén, *C. fruticans* Poelt & Oberw., *C. murrayi* Poelt, and *C. pacifica* M. Westb. & Arup, around the world, especially in the South American and African continents [1–3]. Molecular phylogenetic studies on the family Candelariaceae inferred that neither *Candelariella* nor *Candelaria* was monophyletic; the species of *Candelaria* were separated into three different clades [4]. The most well-known and widely distributed species of *Candelaria* are the sorediate lichen, *C. concolor* (Dicks.) Arnold, and the esorediate lichen, *C. fibrosa* (Fr.) Müll. Arg.; though they are grouped into one clade of *Candelaria* in the strict sense, both species are polyphyletic [4,5].

The type species, *C. concolor*, has been treated as the only common species of *Candelaria* in Europe, until Westberg [5] described a most similar species, *C. pacifica* M. Westb. & Arup from North America. The lichenologist was aware of a situation that the specimens recognized as *C. concolor* may not include only a single species. Thus, after further study, *C. concolor* became a rare lichen in Nordic countries [6], and some specimens collected in Austria, Sweden, and Germany [6–10] under the taxon *C. concolor* agreed with *C. pacifica*. In South

Korea, *C. concolor* and *C. crawfordii* have been reported [11–14]. Several specimens of suspected *C. concolor* were collected during a lichen flora survey in the freshwater areas of South Korea, and morphological, chemical, and phylogenetic analyses were conducted. As a result, all the specimens were found to belong to a distinct species different from *C. concolor*.

2. Materials and methods

2.1. Material and morphological studies

Three specimens in this study were collected from South Korea and deposited at Suncheon National University, Korean Lichen Research Institute (KoLRI). Morphological and anatomical characters were recorded under a dissecting microscope (Nikon SMZ 745 T, Tokyo, Japan) and an Olympus BX 50 microscope (Olympus, Tokyo, Japan), and images were captured using an HD-Measure LTHS-300 (Leetech Co., Seoul, South Korea) microscope and a Carl Zeiss MicroImaging with Axio Cam ERc 5 s imaging system (Carl Zeiss MicroImaging, GmbH 37081, Gottingen, Germany). The secondary metabolites were studied using the spot test (K = 10% aqueous KOH solution; C = saturated aqueous Ca(ClO)₂; KC = 10% aqueous KOH solution followed by saturated aqueous Ca(ClO)₂; PD = 5% alcoholic p-phenylenediamine solution) and thin-layer chromatography (TLC) in solvent C [15,16]. UV tests were performed

using a UV Chamber (CE07 21470) under long (366 nm) wavelength.

2.2. DNA isolation, PCR, DNA sequencing, and sequence alignment

Total genomic DNA was extracted from the newly collected specimens using the NucleoSpin Plant II Kit (Clontech Laboratories, Mountain View, CA) following the manufacturer's instructions. The internal transcribed spacer (ITS) region and the large subunit of the ribosomal RNA (28S) were targeted via PCR using the primers pairs ITS4/ITS1F [17] and LR0R/LR5 [18,19], respectively. Protocols of PCR amplification were followed [20]. Sequencing was conducted by GenoTech (Daejeon, Korea) and Macrogen (Daejeon, Korea). Newly obtained sequences for the ITS region of *Candelaria* species (bold in Table 1) were assembled and edited using SeqMan and Mega 7.0 [21], and complemented with publicly available sequences of the family Candelariaceae (Table 1) into a matrix and aligned using Mafft v7.273 [22]. *Pycnora xanthococca* (Sommerf.) Hafellner was used as an outgroup

since *Pycnora* has been recognized as a possible sister clade to Candelariaceae [23].

2.3. Phylogenetic analysis

The matrix was analyzed under the criterion of maximum likelihood (ML) using RAxML v7.2.6 [24] with the GTR + I+G model. ML bootstrap values were estimated from consensus trees obtained from 1000 nonparametric bootstrapping pseudoreplicates; a clade of bootstrap value >70% indicates crudely supported. Bayesian inference (BI) was performed with MrBayes v3.1.2 [25], applying the best-fitted substitution models (GTR + T+G) based on the Akaike information criterion using jModelTest 3.7 [26]. Bayesian inferences were conducted using four chains and run for 2 million generations. Trees were sampled every 1000 generations with the first 25% of trees discarded. The remaining trees were used to generate a majority-rule consensus tree with posterior probabilities (PP); clades of PP value ≥ 0.95 were considered as significantly supported.

3. Results and discussion

The matrix contains 40 sequences in total, including 33 taxa of family Candelariaceae; after excluding the

Table 1. Voucher information of ITS sequences used in this study.

Taxon	ITS	Voucher information	Location
<i>Candelaria asiatica</i>	MG694269	D. Liu 171446 (KoLRI)	South Korea
<i>C. asiatica</i>	MG694270	D. Liu 171454 (KoLRI)	South Korea
<i>C. concolor</i>	EF535205	Westberg 454 (LD)	Mexico
<i>C. concolor</i>	FJ959355	Krumsvik s.n. (BG)	Norway
<i>C. concolor</i>	GU929921	Arup L07018 (LD)	Italy
<i>C. concolor</i>	GU929922	Arup L07001 (LD)	Italy
<i>C. crawfordii</i>	EF535204	Moberg & Santesson 8125b (UPS)	China. Yunnan,
<i>C. fibrosa</i>	EF535206	Worthington 21240 (COLO)	U.S.A. New Mexico
<i>C. fibrosa</i>	GU929923	Frödén 1670 (LD)	Argentina
<i>C. fibrosoides</i>	EF535211	Frödén 1513 (LD)	Peru.
<i>C. fibrosoides</i>	EF535212	Frödén 1512 (LD)	Peru.
<i>C. fruticans</i>	EF535207	Lassøe s.n. (C)	Ecuador
<i>C. pacifica</i>	EF535210	[John, Lich. Anatol. Exs. no. 16] (ASU)	Turkey
<i>C. pacifica</i>	GU929918	Westberg 967 (LD)	U.S.A. California
<i>Candelariella aggregata</i>	EF535156	Westberg 1281 (LD)	Canada
<i>Ca. antennaria</i>	EF535159	Westberg 1155 (LD)	U.S.A. Colorado
<i>Ca. aurella</i>	EF535163	Westberg 150 (LD)	U.S.A. Arizona
<i>Ca. biatorina</i>	EF535164	Westberg 1181 (LD)	U.S.A. Colorado
<i>Ca. blastidiata</i>	KX853128	Davydov 7716 (ALTB)	Russia
<i>Ca. borealis</i>	EF535168	Westberg 1079 (LD)	U.S.A. Colorado
<i>Ca. clarkii</i>	KR052104	Tripp & D'az 4876 (COLO)	U.S.A. Colorado
<i>Ca. complanata</i>	EF535173	Westberg 392 (LD)	Mexico
<i>Ca. granuliformis</i>	GU967376	Westberg 3128 (LD)	Sweden
<i>Ca. kansuensis</i>	EF535181	Wetmore 55470 (MIN)	U.S.A. Arizona
<i>Ca. lutella</i>	EF535182	Westberg 2808 (LD)	Norway
<i>Ca. medians</i>	EF535184	Arup L03165 (LD)	Sweden
<i>Ca. placodizans</i>	EF535188	Westberg 1083 (LD)	U.S.A. Colorado
<i>Ca. rosulans</i>	EF535191	Westberg 1146 (LD)	U.S.A. Colorado
<i>Ca. spraguei</i>	EF535194	Westberg 1037 (LD)	U.S.A. Colorado
<i>Ca. subdeflexa</i>	EF535197	Nash 38631 (ASU)	U.S.A. Arizona
<i>Ca. vitellina</i>	EF535199	Westberg 875 (LD)	U.S.A. Oregon
<i>Ca. xanthostigma</i>	EF535202	Westberg 1122 (LD)	U.S.A. Colorado
<i>Candelina mexicana</i>	EF535214	Westberg 388 (LD)	Mexico
<i>Can. submexicana</i>	EF535215	Westberg 408 (LD)	Mexico
<i>Placomaronea candelarioides</i>	FJ959350	Frödén 1720 (LD)	Argentina
<i>P. fuegiana</i>	FJ959351	Frödén 1786 (LD)	Argentina
<i>P. kaernefeltii</i>	FJ959354	Frödén 1503 (LD)	Chile
<i>P. mendozae</i>	EF535219	Westberg 833 (LD)	U.S.A. Arizona
<i>P. minima</i>	FJ959353	Frödén 1656 (LD)	Chile
<i>Pycnora xanthococca</i>	KF360412	E. Timdal 11646 (O L-163707)	Norway

Bold were newly generated in this study, *C.* = *Candelaria*, *Ca.* = *Candelariella*, *Can.* = *Candelina*, *P.* = *Placomaronea*.

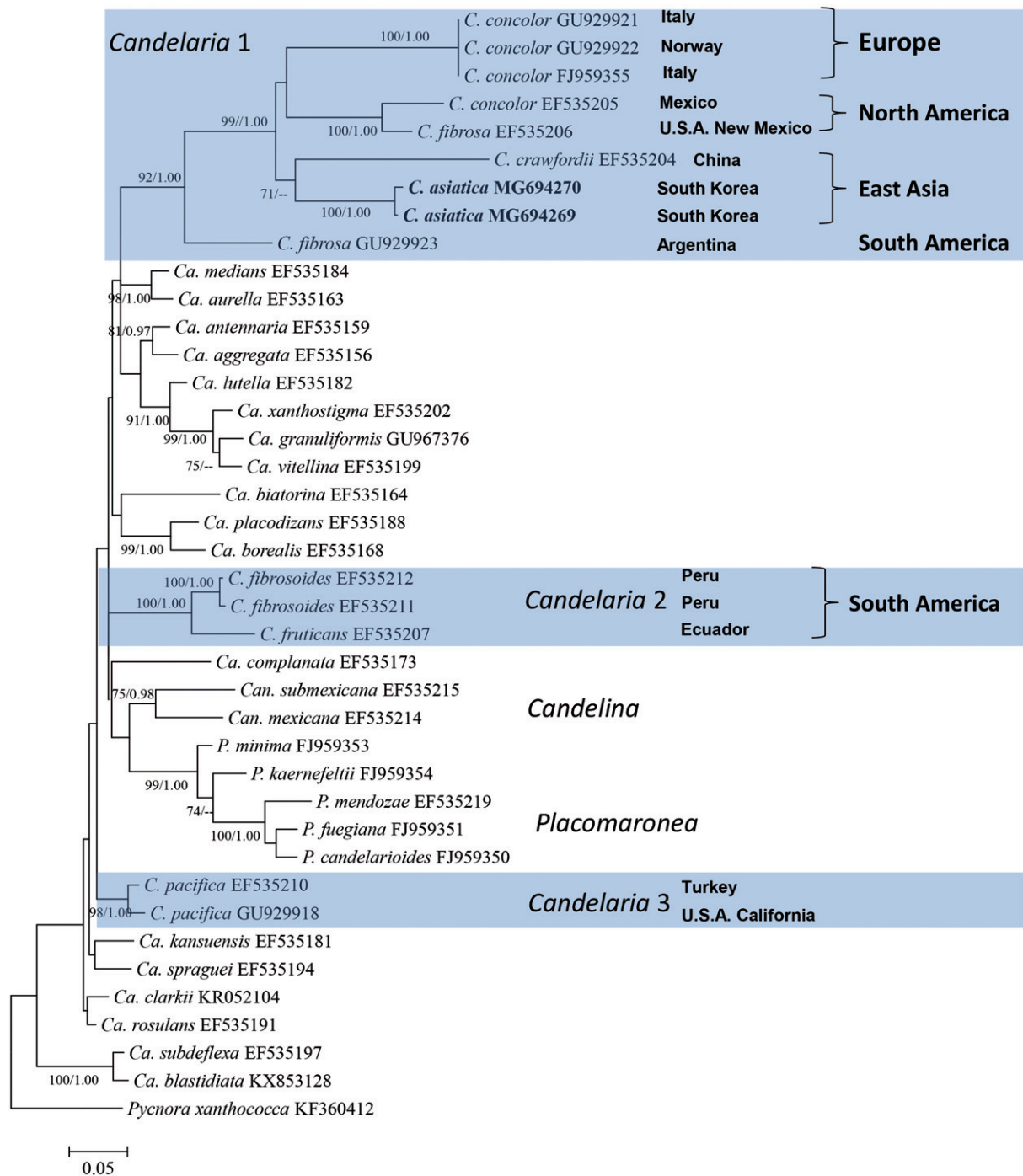


Figure 1. Most likely phylogenetic relationships among species of *Candelaria* inferred from ITS (nrDNA) sequences. Nodes supported by ML bootstrap values >70% and Bayesian posterior probabilities >0.95 are noted on the branches, and bootstrap values are erased on the lower supported clades. Clades of the genus *Candelaria* are highlighted in blue color.

ambiguous sites, 472 sets were reserved. Maximum likelihood and Bayesian analyses based on ITS show similar topology with lower support on the main branches and strong support on the most terminate branches (Figure 1). Both *Candelariella* and *Candelaria* are polyphyletic, whereas the genera *Candelina* and *Placomaronea* form a single clade, and are complementary sister groups with lower support.

Species of *Candelaria* separate into three distinct clades and fall into two morphological groups. They are group 1, *Candelaria* in the strict sense, containing the type species *C. concolor*, is characterized by

a well-developed lower cortex and polysporous asci, our newly described species *C. asiatica* robustly drops into this clade; group 2 is recognized by lacking a lower cortex and having 8-spored asci, and contains clades *Candelaria* 2 and *Candelaria* 3, including *C. fibrosoides*, *C. fruticans*, and *C. pacifica*.

In clade *Candelaria* 1, neither *C. concolor* nor *C. fibrosa* is monophyletic. Specimens of *C. concolor* collected from Europe form a single clade and are genetically distant from the specimen collected from North America, which is a sister group of *C. fibrosa*, which was collected from the same continent. In

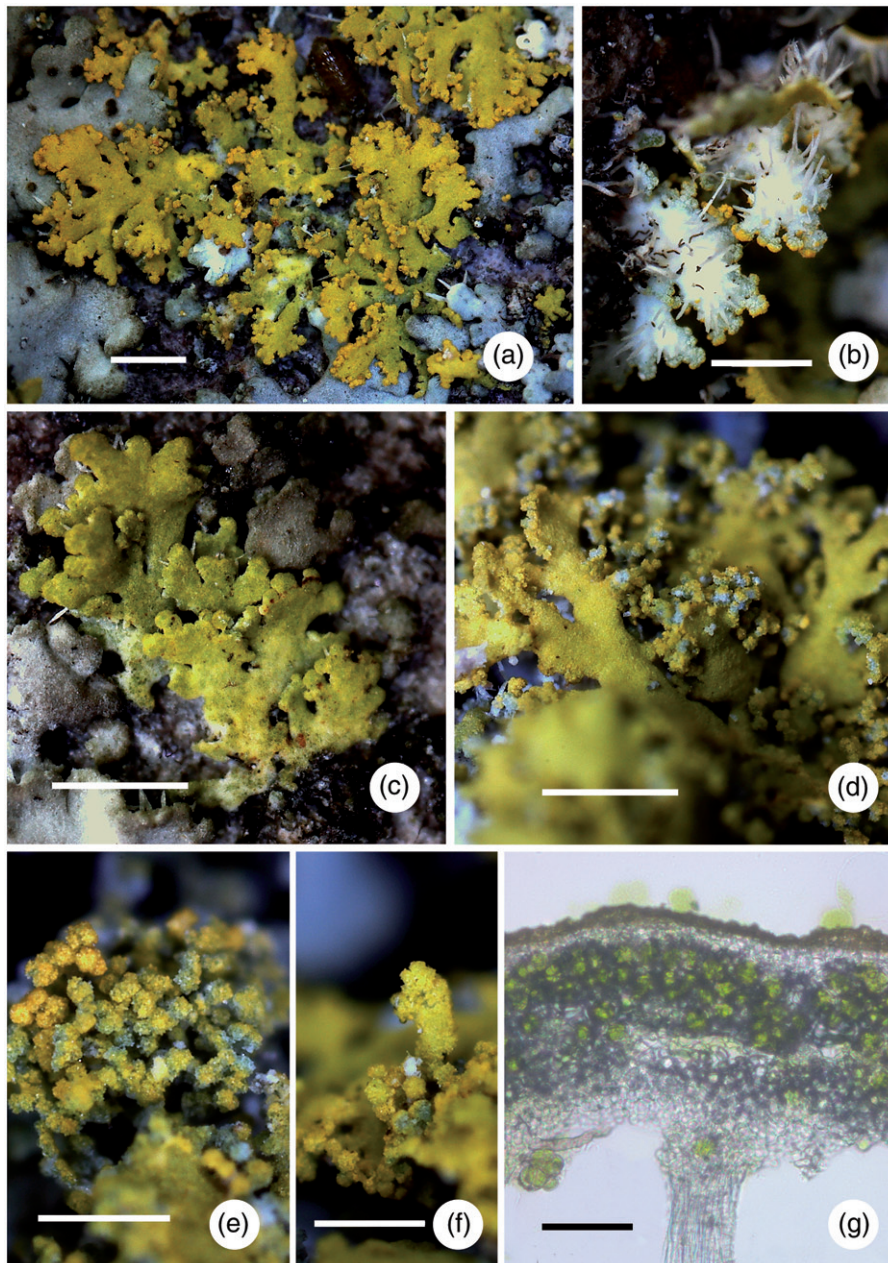


Figure 2. *Candelaria asiatica* (holotype). A, habit; B, lower surface; C, young thalli; D, lobes margin after blastidia or phyllidia-like lobules are dropped; E, F, blastidia or phyllidia-like lobules; G, transverse section of the thallus (Scale bars: A–F = 0.5 mm, G = 40 μ m).

addition, the specimen of *C. fibrosa* from South America is distant from the specimen collected in New Mexico. *Candelaria asiatica*, with two specimens, forms a robust clade and is the sister group of *C. crawfordii* with lower support, and both of them are collected from East Asia.

Unless additional morphological and anatomical characters are found in the future, it is difficult to clarify the relationship among species in the family Candelariaceae at a genetic level up to this study. Otherwise, geography probably plays a significant role in the genetic variation among species of *Candelaria*, which is more or less supported by this study. However, more sequence data and specimens from different continents should be

included in order to investigate the natural lineage of Candelariaceae.

New species

***Candelaria asiatica* D. Liu & J.-S. Hur, sp. Nov., (Figure 2)**

Mycobank No.: MB824810

Similar to *Candelaria concolor*, but differs in the lobe tips and margin, which is neither round nor smooth, and are often fragile or have blastidia, which makes them appear sorediate.

Type: South Korea, Jeollanam-do, Suncheon-si, Jungang-ro 225, beside the library of Sunchon National University, 34°58'01"N, 127°28'48"E, 47 m,

Table 2. Morphological difference among the species of *Candelaria* clade 1.

Taxon	Lobe	Upper surface	Asexual propagules	Distribution
<i>C. asiatica</i>	Shallowly to deeply branched, narrower (up to 0.47 mm)	Rough	Blastidia or phyllidia-like lobules present	South Korea
<i>C. concolor</i>	Deeply branched, narrower (up to 0.5 mm)	Smooth	Soredia granular	Asia, America, and Europe
<i>C. crawfordii</i>	Shallowly branched, wider (up to 0.7 mm)	Smooth	Soralia labriform, with soredia on the upturned lower surface.	Asia, Australia
<i>C. fibrosa</i>	Wider (up to 2 mm)	Smooth or wrinkled	Soredia absent	Pantropical region

on *Cerasus* sp. or lichenicolous, 30 Nov 2017, D. Liu 171454 (holotype, KoLRI!). Accession Number: ITS = MG694270, 28S = MH101755.

Etymology: The epithet “*asiatica*,” from Asia, refers to the geographic distribution of this species.

Morphology: Thallus corticolous or lichenicolous, scattered, mini squamulose to small foliose or sub-fruticose, 0.2–1.2 (–2.5) cm across, adnate, irregular, or rosettes, usually aggregates into extensive colonies covering the substrate; lobes dorsiventral, linear, irregular branched (2–4 times), 0.1–0.47 mm, adnate to erect, lobe tip slightly upturned, crenate; lobe surface color various, yellow to greenish yellow in the center, then becoming bright yellow toward the lobe tip, not smooth and pulverulent; margins usually have blastidia or phyllidia-like lobules (lower cortex poor development, Figure 2(C)), 40–120 µm wide (Figure 2(D,E,F)), loosely adnate lobe tips, which makes the thallus margin or tips pseudocyphellae-like, blastidia and lobulates absent when the colonization is very small or young; thallus 100–130 µm thick; upper cortex (pseudocortex) distinct, 8–16 µm thick, cortex cells 4–7 µm wide; algae chlorococcoid, very unevenly distributed below the upper cortex; medula white but not well-developed; the lower cortex is usually present at the center of the thallus, lacking at lobe tip; lower surface white, covered with white rhizines; apothecia and pycnidia not seen.

Chemistry: K–, KC–, C–, PD–; calycin and pulvinic acid as major substances.

Ecology: *Candelaria asiatica* was found growing on *Cerasus* sp., together with or directly on *Physciella melanchra* (Hue) Essl., and under open areas.

Remarks: *Candelaria asiatica* is characterized by yellow distinctly foliose to sub-fruticose thallus, usually irregular branched, with a pulverulent surface, slightly upturned and rough lobe tips, and a fragile lobe margin with blastidia or phyllidia-like lobules. *Candelaria asiatica* resembles *C. concolor*, but can be distinguished by the rough and pulverulent surface and phyllidia-like lobules, whereas *C. concolor* has a smooth, flattened surface and thicker thallus [5]. *Candelaria asiatica* is currently only known from Asia, whereas *C. concolor* occurs in Europe and America. Furthermore, the cortical hyphae of *C. concolor* are somewhat gelatinized in Stapper’s

figures [9], compared to those of *C. asiatica*, which are non-gelatinized. *Candelaria asiatica* differs from *C. crawfordii* and *C. fibrosa* by its narrower lobes, and blastidia or phyllidia-like lobules are frequently present on well-developed thallus, especially in old or rosulate individuals, while usually absent on squamulose or young individuals. The differences among the species of *Candelaria* clade 1 are listed in Table 2.

Specimens examined: South Korea. Jeollanam-do, Suncheon-si, Jungang-ro 225, Suncheon National University, 34°58′01″N, 127°28′48″E, 47 m, on *Cerasus* sp., 1 Dec 2017, D. Liu 171453; Gangwon-do, Jeongseon-gun, Gohan-eup, 37°11′22″N, 128°53′29″E, 882 m, on a pine tree, together with *Normandina pulchella*, 22 Sep 2016, D. Liu 171446, Accession number: ITS = MG694269, 28S = MH101754.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by a grant from the Korean National Research Resource Center Program (NRF-2017M3A9B8069471) through National Research Foundation of Korea and the Korean Forest Service Program through the Korea National Arboretum (KNA1-1-22, 17-2).

References

- [1] Westberg M, Frödén P. *Candelaria fibrosoides*—a new species from Peru. *Bibl Lichenol.* 2007;95: 549–554.
- [2] Kärnefelt I, Westberg M. *Candelaria fruticans* found in southern Africa. *Mycotaxon.* 2001;80: 465–467.
- [3] Jorgensen P, Galloway D. Notes on *Candelaria crawfordii*. *Lichenologist.* 1992;24:407–410.
- [4] Westberg M, Arup U, Kärnefelt I. Phylogenetic studies in the Candelariaceae (lichenized Ascomycota) based on nuclear ITS DNA sequence data. *Mycol Res.* 2007;111:1277–1284.
- [5] Westberg M, Arup U. *Candelaria pacifica* sp. nova (Ascomycota, Candelariales) and the identity of *Candelaria vulgaris*. *Bibl Lichenol.* 2011;106: 353–364.

- [6] Westberg M, Arup U. *Candelaria concolor*—a rare lichen in the Nordic countries. *Graphis Scripta*. 2010;22:38–42.
- [7] Neuwirth G. Revision of the lichen genus *Candelaria* (Ascomycota, Candelariales) in Upper Austria. *Stapfia*. 2014;101:39–46.
- [8] Dolnik C. *Candelaria pacifica* und andere bemerkenswerte Flechten aus Schleswig-Holstein. *Kieler Notizen Zur Pflanzenkunde*. 2013;39:11–18.
- [9] Stapper NJ. Illustrierte Bestimmungshilfe zur Unterscheidung von *Candelaria concolor* und *Candelaria pacifica*. *Arch Lichenol*. 2012;7:1–12.
- [10] Bomble FW. *Candelaria pacifica* und *Xanthomendoza borealis* im Aachener Raum – neu für Deutschland. *Veröff Bochumer Bot Ver*. 2012;4:1–8.
- [11] Moon KH. Lichen-forming and lichenicolous fungi of Korea. Incheon, Korea: National institute of biological resources; 2013. (In Korean)
- [12] Kashiwadani H, Moon K-H, Inoue M, et al. Lichens of the Cheju Island, Republic of Korea I. The macrolichens. *National Science Museum Monographs*. 2002;22:115–135.
- [13] Park YS. The macrolichens flora of Korea. *Bryologist*. 1990;93:105–160.
- [14] Aptroot A, Moon KH. 114 New reports of microlichens from Korea, including the description of five new species, show that the microlichen flora is predominantly Eurasian. *Herzogia*. 2014;27:347–365.
- [15] Elix JA. A catalogue of standardized chromatographic data and biosynthetic relationships for lichen substances. Canberra: The Author; 2014.
- [16] Orange A, James P, White F. *Microchemical methods for the identification of lichens*. 2nd ed. London: British Lichen Society; 2010.
- [17] White TJ, Bruns T, Lee S, et al. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR Protocols: a Guide to Methods and Applications*. 1990;18:315–322.
- [18] Rehner SA, Samuels GJ. Taxonomy and phylogeny of *Gliocladium* analysed from nuclear large subunit ribosomal DNA sequences. *Mycol Res*. 1994;98:625–634.
- [19] Vilgalys R, Hester M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J Bacteriol*. 1990;172:4238–4246.
- [20] Liu D, Wang XY, Li JW, et al. Contributions to the lichen flora of the Hengduan Mountains, China (6): revisional study of the genus *Canoparmelia* (lichenized Ascomycota, Parmeliaceae). *Plant Diversity Resour*. 2014;36:781–787.
- [21] Kumar S, Stecher G, Tamura K. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol*. 2016;33:1870–1874.
- [22] Katoh K, Standley DM. A simple method to control over-alignment in the MAFFT multiple sequence alignment program. *Bioinformatics*. 2016;32:1933–1942.
- [23] Bendiksby M, Timdal E. Molecular phylogenetics and taxonomy of *Hypocenomyce sensu lato* (Ascomycota: Lecanoromycetes): Extreme polyphyly and morphological/ecological convergence. *Taxon*. 2013;62:940–956.
- [24] Stamatakis A. RAxML Version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics*. 2014;30:1312–1313.
- [25] Huelsenbeck JP, Ronquist F. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics*. 2001;17:754–755.
- [26] Posada D. jModelTest: phylogenetic model averaging. *Mol Biol Evol*. 2008;25:1253–1256.