

Comparison of 12 Isoflavone Profiles of Soybean (*Glycine max* (L.) Merrill) Seed Sprouts from Three Different Countries

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ABSTRACT The levels of 12 isoflavones were measured in soybean (*Glycine max* (L.) Merrill) sprouts of 68 genetic varieties from three countries (China, Japan, and Korea). The isoflavone profile differences were analyzed using data mining methods. A principal component analysis (PCA) revealed that the CSRV021 variety was separated from the others by the first two principal components. This variety appears to be most suited for functional food production due to its high isoflavone levels. Partial least squares discriminant analysis (PLS-DA) and orthogonal projections to latent structures discriminant analysis (OPLS-DA) showed that there are meaningful isoflavone compositional differences in samples that have different countries of origin. Hierarchical clustering analysis (HCA) of these phytochemicals resulted in clusters derived from closely related biochemical pathways. These results indicate the usefulness of metabolite profiling combined with chemometrics as a tool for assessing the quality of foods and identifying metabolic links in biological systems.

Keywords : HPLC, isoflavones, partial least squares discriminant analysis, principal components analysis, soybean sprouts

In recent years, there has been increasing interest in soybeans (*Glycine max* (L.) Merrill) and related products, such as soybean sprouts (Kongnamul), soymilk, and tofu, due to their potential health benefits. Soybeans contain various nutrients, including protein, vitamins, minerals, and phenolic compounds. Especially, they are recognized as the richest sources of isoflavones, which are a group of phytoestrogens that have been shown to have beneficial effects on human health, including antioxidant activity, cancer prevention, and bone density improvement. Soybean isoflavones have been reported to reduce the risk of cardiovascular diseases, menopausal symptoms, osteoporosis, and certain types of cancers (Messina *et al.*, 1994; Anthony *et al.*, 1996; Sarkar & Li, 2003; Allred *et al.*, 2005). Several studies have shown that isoflavones have anticarcinogenic effects on hormone-related cancers (Allred *et al.*, 2005; Adlercreutz & Mazur, 1997). The various phy-

siological functions of isoflavones are mediated by mechanisms including estrogenicity, alteration of enzymatic activity in steroid biosynthesis, and activation of immune cell function (Omoni & Aluko, 2005; Barnes, 2010). These discoveries led to the use of soybeans as a source of isoflavones for the production of functional foods (Wang & Murphy, 1994; Setchell & Cole, 2003).

As diphenolic secondary metabolites, isoflavones can be classified into four groups according to their functional groups: glycosides (genistin, daidzin, and glycitin), malonyl-glycosides (malonylgenistin, malonyldaidzin, and malonyl-glycitin), acetylglycosides (acetylgenistin, acetyldaidzin, and acetylglycitin), and aglycones (genistein, daidzein, and glycitein) (Setchell & Cole, 2003). Isoflavones are generally synthesized from a branch of the phenylpropanoid pathway, in which phenylalanine is converted to genistein and daidzein

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by phenylalanine ammonia-lyase, chalcone synthase, and isoflavone synthase. The synthesized compounds can be conjugated sequentially with glucosyl and malonyl sidechains, and sequestered in vacuoles. However, the glycitein biosynthetic pathway is still largely unknown (Barnes, 2010; Yu *et al.*, 2003). Recent studies showed that the seed isoflavone content of soybean cultivars can be affected by their genetic variation, as well as environmental factors (precipitation, temperature, year, and location of cultivation) (Caldwell *et al.*, 2005; Hoeck *et al.*, 2000; Carrao-Panizzi *et al.*, 2009).

The soybean sprout is an important traditional vegetable food in China, Japan, and Korea, as it can be produced relatively easily at a low cost. In Korea, the mass production of soybean sprouts is an important agriculture business, with an annual consumption of more than 500,000 tons (Hwang, 2004). The impacts of the genetic varieties, cultivation condition, and crop management on isoflavone concentrations in soybean sprouts have been examined, in order to improve the quality and quantity of the sprouts. Previous studies reported that soybean sprouts have higher concentrations of isoflavones than the corresponding seeds (Kim *et al.*, 2006; Kim *et al.*, 2004a,b). The total isoflavone concentrations in 30 varieties of soybean sprouts produced in Korea are reported to vary from 768 to 3343 $\mu\text{g/g}$ (Kim *et al.*, 2003). The isoflavone levels in different parts of the sprout (cotyledons, hypocotyles, and roots) are also affected by the germination period (Kim *et al.*, 2004a,b; Phommalth *et al.*, 2008) and light condition during cultivation (Kim *et al.*, 2006; Lee *et al.*, 2007). Several studies have focused on the diversity of isoflavone compounds among soybean sprouts of different genotypes. However, to the best of our knowledge, a comparative study of the isoflavone composition in soybean sprouts with different countries of origin has not been reported.

In this study, we investigated the isoflavone contents in 68 varieties of soybean sprouts from three countries (China, Japan, and Korea), and analyzed their isoflavone profiles using data mining techniques, namely principal components analysis (PCA), partial least squares discriminant analysis (PLS-DA), orthogonal projections to latent structures discriminant analysis (OPLS-DA), Pearson's correlation analysis, and hierarchical clustering analysis (HCA) to distinguish possible compositional differences based on the country of origin.

MATERIALS AND METHODS

Cultivation of soybean sprouts

Sixty eight soybean germplasms collected from 3 different countries, Japan (12 varieties), China (15 varieties), and Korea (41 varieties), were distributed by the Rural Development Administration (RDA, Suwon, Gyonggi-do, Korea). Soybean seeds were first soaked in distilled water at room temperature for 4 h. Then the soaked soybean were cultivated in the dark in a greenhouse for 5 days, which is a popular standard method in Korea.

Isoflavone analysis

The harvested soybean sprouts were dried in a freeze-dryer (Freezeone 4.5; Labconco, Kansas, Missouri, USA) in vacuum, and then ground into powder. The extraction of isoflavones followed the method of Wang & Murphy (Wang & Murphy, 1994). 2 g of ground sample was mixed with 10 mL of acetonitrile and 2 mL of 0.1 N HCl, then stirred for 2 h at room temperature. The extracts were filtered through Whatman No. 42 filter paper (125 mm \times 100 circles, Maidstone, UK) and concentrated using a vacuum evaporator (EYELA; Tokyo Rikakikai, Co. Ltd, Japan) at 40°C. The residues were dispersed in 10 mL of methanol (100%, HPLC grade; J. T. Baker), and filtered through a 0.2- μm nylon membrane syringe filter. Isoflavone analysis was performed using a method modified from that of Lee *et al.* (Lee *et al.*, 2007). Standard isoflavone compounds were purchased from Wako Pure Chemical Industries Ltd. (Osaka, Japan). These standard compounds were dissolved separately in dimethylsulfoxide (DMSO; Sigma-Aldrich, USA) at several concentrations, and a high linearity ($r^2 > 0.997$) was obtained for each case. An HPLC system (Younglin YL 9100), with a UV/VIS detector (YL 9120) equipped with a YMC ODS AM-303 column (5 μm , 250 \times 4.6 mm I.D.) and an autosampler (YL 9150) was used. Solvent A was 0.1% glacial acetic acid in distilled water, and solvent B was 0.1% glacial acetic acid in acetonitrile (ACN). The mobile gradient used in this study was as follows: 0 min, 85% A:15% B; 0–50 min, 65% A:35% B; and 50–60 min, 65% A:35% B. The run time was 60 min, and the flow rate was 1 mL $\cdot\text{min}^{-1}$. The injection volume was 20 μL and the wavelength of the UV detector was 254 nm. Also, 4 groups isoflavones in soybean sprouts are shown in the

Supporting Information (Appendix 1, 2, 3, 4).

Statistical analysis

All experimental design of each soybean samples consisted of a completely randomized design with two replicates. All analyses were performed at least twice, and unit variance scaling was applied to all data prior to the analyses. PCA, PLS-DA, and OPLS-DA were performed using SIMCA-P version 13.0 (Umetrics, Umeå, Sweden) to evaluate differences among the multivariate data. The PCA and PLS-DA output consisted of score plots to visualize the contrast among the samples, and loading plots that explain the cluster separation. OPLS-DA was subsequently used to improve the separation. Pearson's correlation analysis was performed with the SAS 9.2 software package (SAS Institute, Cary, NC, USA). HCA and heat map visualisation of the correlation coefficients were performed using the software Multi-Experiment Viewer version 4.9.0 (<http://www.tm4.org/mev/>).

RESULTS

Isoflavone compositions of soybean sprouts

Soybean is a useful crop because it contains many compounds that have health benefits such as protein, isoflavones, fiber and other various phytochemicals (Kim *et al.*, 2012). We used HPLC analysis to measure the contents of 12 isoflavones in the soybean sprout samples. Quantitative data for the 68 varieties are shown in Table 1 and Supporting Information. Results observed in the present study regarding differences in total isoflavone contents between soybean genotypes. The total content of the 12 isoflavones was found to vary widely (325–1553 $\mu\text{g/g}$ dry weight (DW)) among different genotypes. However, no significant differences in the total isoflavone content and its chemical composition were observed among the three countries of origin.

Among 68 soybean sprouts resources, the total isoflavone compounds showed the highest concentrations in CSRV021 as $1553.21 \pm 4.73 \mu\text{g/g}$. Isoflavones were divided into four groups that were glycosides, malonylglycosides, acetylglycosides and aglycones. Among the four isoflavone group, the malonyl-conjugated group had the highest content in most varieties, followed by glycosides, aglycones, and acetylglycosides. The malonyl glycoside and glycoside contents

showed the highest in CSRV020 ($905.25 \pm 3.45 \mu\text{g/g}$, $383.25 \pm 1.35 \mu\text{g/g}$). The acetyl glycosides showed higher in CS02685 ($317.22 \pm 0.56 \mu\text{g/g}$), the aglycone group revealed higher in CSRV021 ($613.29 \pm 1.44 \mu\text{g/g}$). While, the glycoside group had the lowest concentrations in CSRV018 ($81.58 \pm 0.57 \mu\text{g/g}$), the malonyl glycoside contents showed the lowest in CS02685 ($133.79 \pm 0.26 \mu\text{g/g}$). And, the acetyl glycosides showed lower in CSRV032 ($8.76 \pm 0.02 \mu\text{g/g}$), the aglycone showed the lowest in CSRV033 ($21.79 \pm 0.59 \mu\text{g/g}$).

Classification of isoflavone profiles by multivariate analysis

To examine the differences in isoflavone profiles among the genetic varieties, PCA and PLS-DA were carried out. These two methods are commonly used as clustering techniques to identify how one group of samples differs from another, which variables contribute the most strongly to the difference, and whether the variables are correlated (Kim *et al.*, 2013; Park *et al.*, 2013). Fig. 1A shows the PCA results by plotting the principal component scores. The two highest-ranking principal components accounted for 51.6% (28.6% for component 1 (PC1) and 23.0% for component 2 (PC2)) of the total variance within the data set. Upon combining these two components, the CSRV021 sample (from Korea) appeared as an outlier. To identify the compounds that contributed most to the separation, the factor loadings in PC1 and PC2 were compared. The corresponding loading in PC1 was positive for all measured compounds except acetylgenistin (Fig. 1B). The variations in PC1 and PC2 were mainly attributed to genistin and daidzein, respectively.

However, PCA could not distinguish among the three countries of origin. Therefore, PLS-DA was conducted to enhance the weak separation obtained with the PCA model (Fig. 2). The corresponding loading indicated that the proportion of malonylglycitin was higher in most Chinese varieties than in the Korean ones, and that of malonylgenistin was higher in most Korean varieties than in Chinese varieties.

Subsequently, OPLS-DA method was applied to identify the major isoflavones responsible for the separation between Korean and Chinese or Japanese samples (Fig. 3). Because its integrated orthogonal signal correction (OSC) filter removes systematic spectral variation, OPLS-DA is often used in lieu of PLS-DA to identify a predictor for classification. The

Table 1. Content ($\mu\text{g/g}$ dry weight) of isoflavones in soybean sprouts^a.

IT No.	Origin	Glycoside ^b	Malonyl glycoside ^c	Acetyl glycoside ^d	Aglycone ^e	Sum
CS00732	China	267.35±5.95	753.10±1.26	60.80±3.39	117.21±0.18	1198.46±8.24
CS00733	China	231.88±2.04	683.65±5.09	39.10±0.12	43.57±0.59	998.20±2.59
CS00734	China	228.50±2.55	519.65±7.29	52.42±2.56	68.22±1.52	868.80±13.92
CS01248	China	317.07±3.65	659.01±1.86	15.81±0.54	45.81±0.26	1037.69±1.50
CS01250	China	108.14±0.15	191.10±0.00	135.04±0.76	483.79±0.79	918.07±0.12
CS01252	China	347.44±1.06	437.49±1.03	13.91±0.07	104.46±0.83	903.30±2.98
CS01277	China	212.27±7.29	300.11±0.44	18.93±0.87	89.38±0.40	620.68±9.01
CS01278	China	129.86±1.77	212.25±1.88	13.31±0.16	70.16±0.24	425.59±3.58
CS01381	China	133.21±0.83	290.75±1.51	18.40±0.55	60.16±1.57	502.52±0.22
CS01383	China	225.72±0.90	486.37±3.20	41.21±5.46	164.52±1.72	917.82±9.48
CS01385	China	192.79±1.27	237.99±0.68	56.67±0.08	236.29±0.36	723.74±0.15
CS01386	China	157.56±0.42	224.66±0.07	89.90±0.14	214.41±0.07	683.53±0.14
CS01567	China	267.62±2.89	681.02±0.36	30.73±0.85	61.67±0.07	1041.04±1.61
CS01804	China	103.70±0.72	189.33±2.10	53.29±2.22	102.69±0.81	449.02±4.23
CS02685	China	175.07±1.50	133.79±0.26	317.22±0.56	347.66±2.10	973.74±0.91
CS00736	Japan	257.42±1.66	953.89±4.26	35.02±1.93	72.33±0.79	1318.67±4.78
CS00738	Japan	126.78±0.58	380.66±3.44	10.63±0.41	62.54±0.34	580.60±2.11
CS00740	Japan	240.75±7.65	817.62±11.47	19.19±1.36	60.57±1.74	1138.13±22.22
CS02052	Japan	86.04±1.02	226.06±1.22	15.39±0.43	61.46±0.38	388.96±1.43
CS02059	Japan	146.87±1.80	182.15±1.29	9.99±0.21	65.59±0.02	404.59±3.33
CS02065	Japan	230.63±2.63	598.58±2.46	57.29±1.10	141.26±0.19	1027.76±6.39
CS02205	Japan	343.41±1.10	554.51±0.24	66.16±1.88	382.57±1.47	1346.65±4.68
CS02564	Japan	235.82±0.59	404.52±1.44	121.45±1.48	161.81±0.66	923.59±1.21
CS02587	Japan	139.90±0.21	166.60±0.52	8.85±1.33	35.26±0.05	350.61±1.07
CS02590	Japan	108.22±0.28	270.81±1.23	27.17±0.42	41.30±1.48	447.50±2.01
CS02599	Japan	166.73±0.13	596.37±0.74	34.02±0.56	66.18±1.56	863.31±0.38
CS02665	Japan	123.49±1.57	351.95±1.51	13.31±1.73	216.75±1.48	705.50±3.28
CS01407	Korea	377.17±9.62	802.34±2.44	16.92±0.21	83.22±0.27	1279.65±11.57
CS01408	Korea	294.90±9.56	383.39±6.45	41.31±0.58	214.60±1.66	934.19±2.03
CS01416	Korea	290.55±0.38	432.45±4.08	24.54±0.72	51.56±1.71	799.11±1.27
CS01426	Korea	312.37±0.79	421.15±0.84	37.64±0.51	97.49±2.15	868.65±4.30
CS01429	Korea	160.31±6.64	154.09±2.02	20.83±0.66	111.33±0.68	446.56±8.63
CS01924	Korea	189.82±2.63	393.18±1.71	238.62±2.89	260.35±1.01	1081.97±4.83
CS01949	Korea	328.75±0.94	741.49±2.73	24.05±0.08	53.01±0.68	1147.30±1.03
CS01972	Korea	184.42±1.20	405.65±3.46	16.56±0.07	67.19±0.54	673.81±2.87
CS02554	Korea	317.32±1.59	687.17±0.91	19.88±0.87	53.74±0.76	1078.12±4.13
CS02576	Korea	130.36±2.45	296.76±0.11	14.74±0.72	126.38±1.62	568.24±3.45
CS02594	Korea	236.27±0.11	390.86±2.59	27.61±0.12	39.35±0.90	694.08±3.73
CS02602	Korea	135.08±0.40	381.27±2.06	20.87±0.08	48.84±1.44	523.06±3.81
CS02603	Korea	169.97±0.37	597.35±1.07	75.34±4.43	122.96±0.30	965.62±4.03

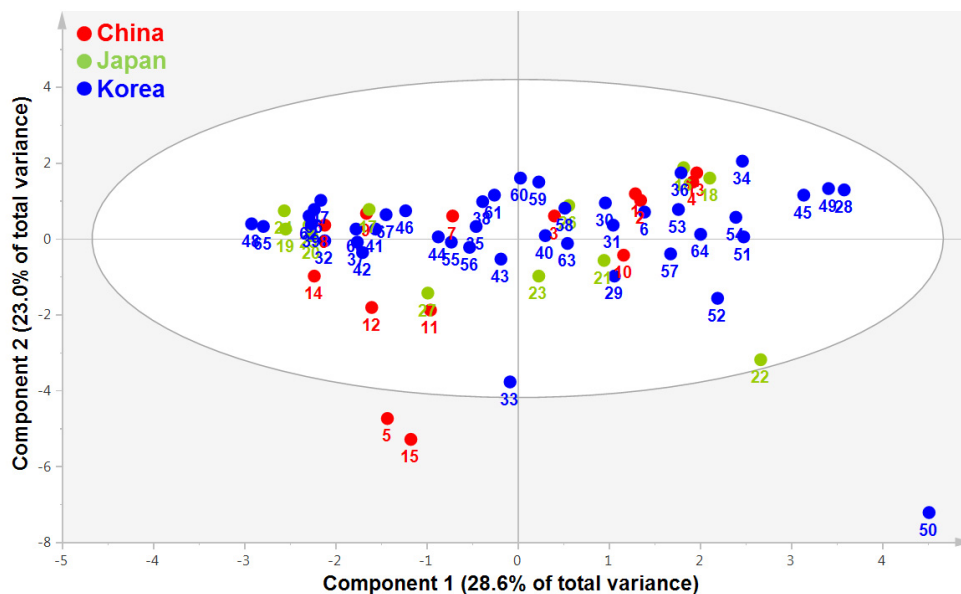
Table 1. Content ($\mu\text{g/g}$ dry weight) of isoflavones in soybean sprouts^a (Continued).

IT No.	Origin	Glycoside ^b	Malonyl glycoside ^c	Acetyl glycoside ^d	Aglycone ^e	Sum
CS02606	Korea	127.90±0.40	386.68±2.64	28.11±1.35	75.44±2.42	618.13±4.11
CS02615	Korea	114.97±1.67	331.68±1.36	23.95±1.36	99.13±0.1	569.74±1.26
CSRV001	Korea	186.18±0.43	414.07±1.72	26.08±0.26	116.44±1.72	742.77±0.17
CSRV003	Korea	184.90±1.67	382.25±9.17	50.40±1.24	84.78±1.14	702.33±9.89
CSRV015	Korea	333.17±0.38	770.91±4.43	22.20±1.22	93.46±0.43	1219.74±2.39
CSRV016	Korea	170.19±1.12	415.93±1.81	15.97±3.23	43.35±0.13	645.44±4.03
CSRV017	Korea	136.12±0.55	318.73±3.63	12.70±0.53	33.59±0.95	501.14±2.65
CSRV018	Korea	81.58±0.57	190.18±1.08	17.81±0.49	35.76±0.16	325.33±0.15
CSRV020	Korea	383.25±1.35	905.25±3.45	16.90±0.07	89.78±0.59	1395.18±1.44
CSRV021	Korea	317.47±4.20	601.69±0.41	20.76±1.56	613.29±1.44	1553.21±4.73
CSRV022	Korea	359.12±4.36	723.39±3.04	29.25±0.62	172.49±2.89	1284.24±10.91
CSRV023	Korea	288.55±1.86	693.60±5.20	14.98±1.81	233.97±5.22	1231.10±6.74
CSRV024	Korea	299.23±4.26	701.03±5.15	19.44±1.39	126.63±34.59	1146.33±34.09
CSRV027	Korea	386.39±3.87	542.08±5.09	29.94±1.04	115.38±2.01	1073.80±12.00
CSRV029	Korea	162.27±1.87	553.01±11.27	41.49±1.10	75.52±1.24	832.29±10.79
CSRV030	Korea	161.47±1.92	563.55±3.72	20.11±0.22	90.06±0.03	835.19±2.05
CSRV031	Korea	354.08±0.50	517.19±7.58	22.47±1.30	141.35±0.76	1035.09±6.02
CSRV032	Korea	218.67±3.84	585.95±11.18	8.76±0.02	67.00±0.46	880.38±15.47
CSRV033	Korea	250.19±0.84	525.29±1.11	17.79±0.55	21.79±0.59	815.06±0.87
CSRV034	Korea	225.96±4.00	531.79±7.32	9.94±0.12	23.30±2.41	791.00±13.85
CSRV035	Korea	238.40±2.54	450.28±4.64	16.33±0.26	49.51±0.53	754.51±2.37
CSRV036	Korea	142.61±1.00	222.02±0.39	16.34±1.58	42.52±0.40	423.49±2.57
CSRV037	Korea	274.96±2.87	411.92±0.19	15.35±0.77	124.75±0.83	826.98±3.13
CSRV038	Korea	336.82±1.45	677.63±1.03	47.47±0.83	80.38±0.41	1142.31±0.01
CSRV039	Korea	96.57±0.32	257.24±0.50	16.96±0.25	45.83±0.19	416.60±1.26
CSRV040	Korea	141.77±1.21	225.01±3.12	17.15±0.36	50.90±1.26	434.83±5.95
CSRV041	Korea	179.69±1.46	279.68±1.79	25.66±1.25	51.48±1.71	536.51±2.63
CSRV042	Korea	152.26±0.16	356.50±2.16	69.31±9.21	25.10±0.25	603.17±7.46

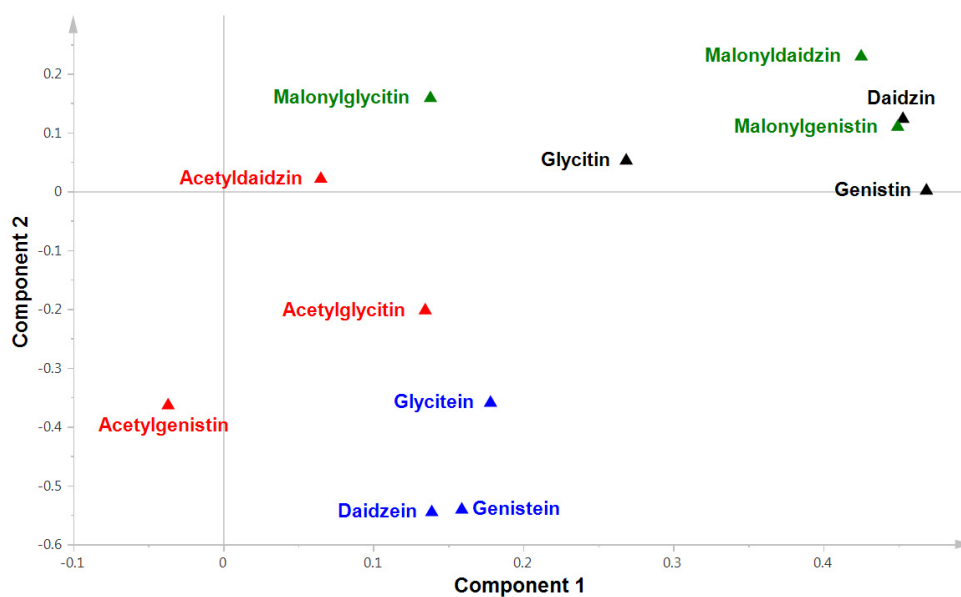
^aEach value represents the mean \pm standard deviation ($n = 2$). ^bGlycoside content is the sum of daidzin, glycitin, and genistin contents. ^cMalonylglycoside content is the sum of malonyldaidzin, malonylglycitin, and malonylgenistin contents. ^dAcetylglycoside content is the sum of acetyldaidzin, acetylglycitin, and acetylgenistin contents. ^eAglycone content is the sum of daidzein, glycitein, and genistein contents.

contribution of each variable to the separation was confirmed using the significance of the variable in the projection (VIP) value. VIP is a weighted sum of squares of the OPLS weight, and a value >1 is generally used as the criterion to identify the most important variables in a model. The variables that played the greatest role discriminating between Korean and Chinese samples were malonylgenistin, glycitin, malonylglycitin, acetylgenistin, and genistin (Fig. 3B). Separation between

Korean and Japanese samples was mainly attributable to malonylglycitin, genistin, acetylgenistin, and malonylgenistin (Fig. 3D). These results confirmed that among 12 isoflavones, malonylglycitin, genistin, acetylgenistin, and malonylgenistin differed significantly between Korean and Chinese or Japanese samples.



(A)



(B)

Fig. 1. Score (A) and loading (B) [black color representing glycosides (genistin, daidzin, and glycitin); green color representing malonylglycosides (malonylgenistin, malonyldaidzin, and malonylglycitin); red color representing acetylglycosides (acetylgenistin, acetylaidzin, and acetylglycitin), and blue color representing aglycones (genistein, daidzein, and glycitein)] plots of principal components 1 and 2 of the PCA results, for 12 isoflavones measured in 68 soybean sprout varieties. 1, CS00732; 2, CS00733; 3, CS00734; 4, CS01248; 5, CS01250; 6, CS01252; 7, CS01277; 8, CS01278; 9, CS01381; 10, CS01383; 11, CS01385; 12, CS01386; 13, CS01567; 14, CS01804; 15, CS02685; 16, CS00736; 17, CS00738; 18, CS00740; 19, CS02052; 20, CS02059; 21, CS02065; 22, CS02205; 23, CS02564; 24, CS02587; 25, CS02590; 26, CS02599; 27, CS02665; 28, CS01407; 29, CS01408; 30, CS01416; 31, CS01426; 32, CS01429; 33, CS01924; 34, CS01949; 35, CS01972; 36, CS02554; 37, CS02576; 38, CS02594; 39, CS02602; 40, CS02603; 41, CS02606; 42, CS02615; 43, CSRV001; 44, CSRV003; 45, CSRV015; 46, CSRV016; 47, CSRV017; 48, CSRV018; 49, CSRV020; 50, CSRV021; 51, CSRV022; 52, CSRV023; 53, CSRV024; 54, CSRV027; 55, CSRV029; 56, CSRV030; 57, CSRV031; 58, CSRV032; 59, CSRV033; 60, CSRV034; 61, CSRV035; 62, CSRV036; 63, CSRV037; 64, CSRV038; 65, CSRV039; 66, CSRV040; 67, CSRV041; 68, CSRV042.

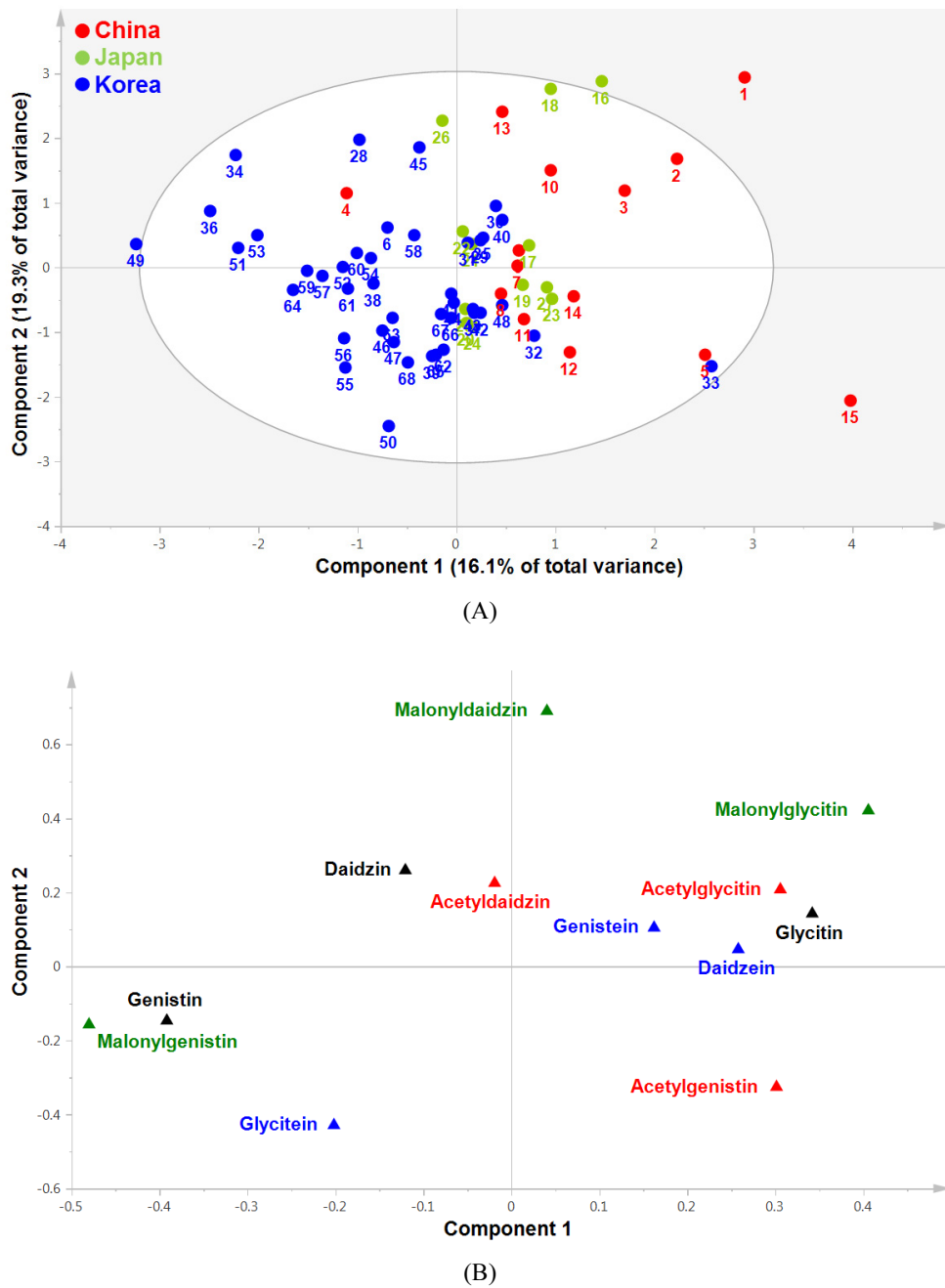


Fig. 2. Score (A) and loading (B) [black color representing glycosides (genistin, daidzin, and glycitin); green color representing malonylglycosides (malonylgenistin, malonyldaidzin, and malonylglycitin); red color representing acetylglycosides (acetylgenistin, acetyldaidzin, and acetylglycitin), and blue color representing aglycones (genistein, daidzein, and glycitein)] plots of principal components 1 and 2 of the PLS-DA results, for 12 isoflavones measured in 68 soybean sprout varieties. The numbers of the score plots represent the same compounds as those in Fig. 1.

Correlations among isoflavone profiles by HCA and Pearson’s correlation analysis

To examine detailed relationships among the levels of the 12 isoflavones identified in soybean sprouts, Pearson’s

correlation analysis and HCA on the accessions were performed. Correlation analysis is a useful technique for determining the strength of a relationship between two quantitative samples and can be used to detect associations

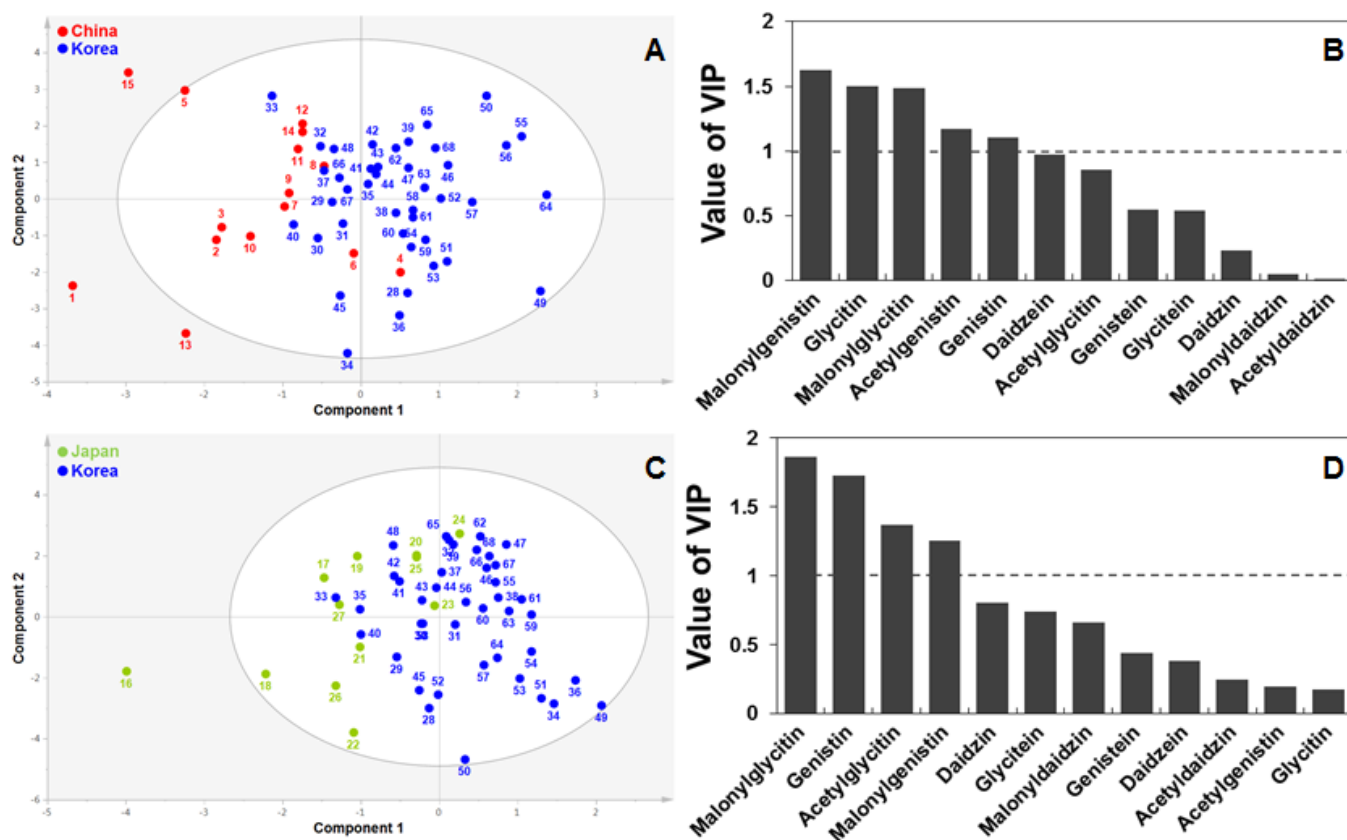


Fig. 3. OPLS-DA score plots and variable importance in the prediction (VIP) values obtained from isoflavones data sets for Korean and Chinese (A and B) and Korean and Japanese samples (C and D). The numbers of the score plots represent the same compounds as those in Fig. 1.

between metabolites in a biological system. Using the methods, strong correlation was found between metabolites that participate in closely related pathways (Fig. 4), which demonstrates the robustness of the present experimental system. For example, significant positive correlation was detected between genistein and daidzein ($r = 0.8902$), which are synthesized by 2-hydroxyisoflavone synthase. Genistin contents were positively correlated with malonylgenistin ($r = 0.8308$) and daidzin ($r = 0.7955$). HCA results obtained from Pearson's correlation coefficients revealed two clusters, which are marked with a dotted box in Fig. 4. One group contained glycosides and malonylglycosides, such as genistin, malonylgenistin, daidzin, and malonyldaidzin. The other group included the three aglycones (glycitein, daidzein, and genistein).

DISCUSSION

The isoflavone contents in soybeans are affected by

environmental and genotypes (Lee *et al.*, 2010). In previous report, the content and composition of isoflavone were reported to differ depending on soybean genotype and growth stage (Kim & Chung, 2007). And soybean isoflavones showed difference according to different environmental conditions such as, rainfall, planting dates, light intensity and temperature (Kim *et al.*, 2012). Kim *et al.* (Kim *et al.*, 2012) reported that isoflavone concentrations of soybeans affected by genotypes such as seed size and origins. In addition, isoflavone contents in soybean seeds were varied by commercial process in an industry and germination process (Kim *et al.*, 2006). Other previous studies have also shown that the isoflavone concentrations in soybean seeds altered by variety, seed weight, seed color, disease tolerance (Hoeck *et al.*, 2000; Lee *et al.*, 2010).

The measured isoflavone compositions are consistent with previous reports. For example, Chan *et al.*, reported that Chinese soybean sprouts contained malonylglycosides as the

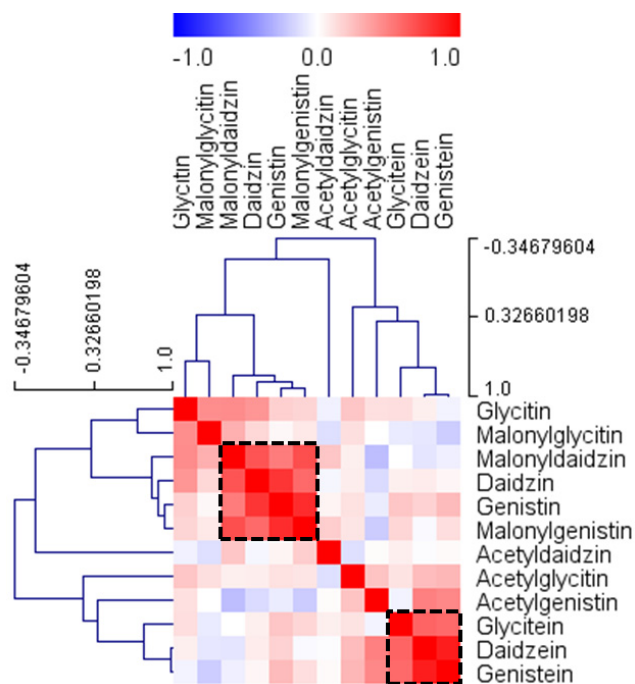


Fig. 4. Correlation matrix and cluster analysis results obtained from data on 12 isoflavones of soybean sprouts. Each square indicates the Pearson's correlation coefficient for a pair of compounds, and the value for the correlation coefficient is represented by the intensity of the blue or red color, as indicated on the color scale. Hierarchical clusters are represented by a cluster tree.

primary form of isoflavones (Chan *et al.*, 2009). Lee *et al.*, showed that glycosides and malonylglycosides were much more abundant than aglycones and acetylglycosides in Korean soybean sprouts (Lee *et al.*, 2007). Kim *et al.*, also reported that the malonyl-conjugated group showed highest concentrations in soybeans and soybean sprouts (Kim *et al.*, 2006). The malonylglycosides groups is major components in soybean germplasm (Kim *et al.*, 2012).

PLS-DA is preferred to PCA for sample discrimination, because the dimension reduction provided by PLS is guided explicitly by the among-groups variability (the origin of soybean sprouts, in this case) (Barker & Rayens, 2003; Brereton, 2009; Kim *et al.*, 2004a,b). Although the PLS-DA results still could not clearly distinguish the countries of origin, certain trends in the separation were identified by PC1. This variation was mainly attributable to malonylglycitin and malonylgenistin.

CONCLUSIONS

This study systematically measured 12 isoflavones (glycosides, malonylglycosides, acetylglycosides, and aglycones) in soybean sprouts, and demonstrated the diversity of isoflavone contents across genetic varieties from three countries. The total isoflavone level varied widely among the different genotypes, and the malonyl-conjugated group (the sum of malonyldaidzin, malonylglycitin, and malonylgenistin) had the highest content in most varieties. In previous reports, the malonylglycosides group of isoflavones revealed the highest in soybeans and soybean sprouts resources (Kim *et al.*, 2006). Other previous study indicated that isoflavone concentrations are influenced by the genotype (Kim & Chung, 2007). PCA, PLS-DA, and OPLS-DA were performed to identify differences among the samples, especially with regard to the country of origin. The PCA results revealed that the CSR021 was separated from the others by PC1 and PC2. This sample had the highest total isoflavone content among all varieties analyzed in this study. PLS-DA allowed the identification of some meaningful differences among the isoflavone profiles of samples originating from China, Japan, and Korea. The results of OPLS-DA demonstrated that among 12 isoflavones, malonylglycitin, genistin, acetylgenistin, and malonylgenistin were major isoflavones responsible for the discrimination between Korean and Chinese or Japanese samples. These conclusions suggest the usefulness of metabolite profiling combined with chemometrics as a tool for discriminating plant-based samples at the genotype level and assessing food quality. The results of this study also provide valuable information for future breeding programs, so as to increase the health benefits of isoflavones in soybean sprouts. The CSR021 variety, which contains a relatively high level of total isoflavones, should be of high dietary value. In conclusion, our results are similar to those of previous results that have observed that isoflavones in soybean sprouts are affected by genotypes. And the results of this study will be helpful to plant breeder and genetists for further investigating the basis of genetic control and selection of soybean isoflavones.

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Supplementary Appendix 1. Content ($\mu\text{g/g}$ dry weight) of Three Glycosides in Soybean Sprouts^a.

IT No.	Origin	Daidzin	Glycitin	Genistin	% ^b
CS00732	China	125.27 \pm 4.89	25.67 \pm 0.14	116.41 \pm 0.90	22.31
CS00733	China	72.43 \pm 0.63	47.63 \pm 0.17	111.82 \pm 1.24	23.23
CS00734	China	89.96 \pm 0.73	30.06 \pm 0.27	108.48 \pm 1.55	26.30
CS01248	China	134.64 \pm 0.61	20.12 \pm 2.15	162.31 \pm 0.88	30.56
CS01250	China	44.20 \pm 0.27	9.75 \pm 0.08	54.19 \pm 0.04	11.78
CS01252	China	156.27 \pm 1.04	20.11 \pm 0.07	171.06 \pm 0.06	38.46
CS01277	China	100.92 \pm 6.97	26.88 \pm 0.27	84.47 \pm 0.60	34.19
CS01278	China	66.52 \pm 1.42	12.18 \pm 0.21	51.16 \pm 0.14	30.51
CS01381	China	65.01 \pm 0.13	15.63 \pm 0.23	52.57 \pm 0.47	26.51
CS01383	China	110.04 \pm 0.56	23.65 \pm 0.18	92.03 \pm 0.15	24.60
CS01385	China	69.36 \pm 1.13	10.70 \pm 0.41	112.73 \pm 0.28	26.64
CS01386	China	50.41 \pm 0.75	13.71 \pm 0.01	93.45 \pm 0.32	23.05
CS01567	China	122.84 \pm 1.47	40.86 \pm 3.34	103.92 \pm 1.92	25.71
CS01804	China	25.13 \pm 0.32	8.24 \pm 0.89	70.33 \pm 0.49	23.10
CS02685	China	66.55 \pm 0.64	21.55 \pm 0.08	86.98 \pm 0.94	17.98
CS00736	Japan	110.15 \pm 0.71	20.84 \pm 0.12	126.44 \pm 0.83	19.52
CS00738	Japan	65.79 \pm 0.60	10.76 \pm 0.12	50.22 \pm 0.14	21.84
CS00740	Japan	98.37 \pm 0.64	34.47 \pm 5.73	107.90 \pm 1.28	21.15
CS02052	Japan	47.47 \pm 0.33	6.16 \pm 0.51	32.41 \pm 0.18	22.12
CS02059	Japan	54.44 \pm 0.75	4.28 \pm 0.10	91.15 \pm 0.95	36.30
CS02065	Japan	88.15 \pm 0.88	13.57 \pm 0.39	128.91 \pm 1.37	22.44
CS02205	Japan	139.39 \pm 0.41	7.76 \pm 0.00	196.26 \pm 0.69	25.50
CS02564	Japan	99.38 \pm 0.09	26.08 \pm 0.23	110.35 \pm 0.27	25.53
CS02587	Japan	74.87 \pm 0.05	11.44 \pm 0.08	53.59 \pm 0.24	39.90
CS02590	Japan	44.83 \pm 0.12	6.43 \pm 0.01	56.96 \pm 0.17	24.18
CS02599	Japan	73.92 \pm 0.44	12.60 \pm 0.59	80.21 \pm 0.27	19.31
CS02665	Japan	46.06 \pm 0.34	14.38 \pm 0.34	63.06 \pm 0.90	17.50
CS01407	Korea	176.80 \pm 8.82	25.34 \pm 0.07	175.04 \pm 0.72	29.47
CS01408	Korea	153.57 \pm 4.85	14.49 \pm 3.07	126.84 \pm 1.64	31.57
CS01416	Korea	141.88 \pm 0.02	29.70 \pm 0.21	118.97 \pm 0.57	36.36
CS01426	Korea	140.39 \pm 0.71	26.18 \pm 0.18	145.81 \pm 0.27	35.96
CS01429	Korea	69.06 \pm 1.81	19.49 \pm 4.13	71.76 \pm 0.70	35.89
CS01924	Korea	52.25 \pm 1.58	22.92 \pm 0.37	114.65 \pm 0.69	17.54
CS01949	Korea	146.66 \pm 0.91	18.40 \pm 1.06	163.70 \pm 1.08	28.65
CS01972	Korea	76.05 \pm 0.91	12.37 \pm 0.28	96.00 \pm 0.02	27.37
CS02554	Korea	125.06 \pm 0.73	12.31 \pm 0.01	179.95 \pm 0.84	29.43
CS02576	Korea	45.02 \pm 1.37	14.80 \pm 0.55	70.54 \pm 0.53	22.94
CS02594	Korea	92.04 \pm 0.15	12.51 \pm 0.03	131.71 \pm 0.24	34.04
CS02602	Korea	42.64 \pm 0.51	6.85 \pm 0.22	85.58 \pm 0.11	25.82
CS02603	Korea	72.11 \pm 0.04	18.53 \pm 0.01	79.33 \pm 0.32	17.60

Supplementary Appendix 1. Content ($\mu\text{g/g}$ dry weight) of Three Glycosides in Soybean Sprouts^a (Continued).

IT No.	Origin	Daidzin	Glycitin	Genistin	% ^b
CS02606	Korea	49.37±0.32	6.41±0.36	72.11±0.45	20.69
CS02615	Korea	40.12±0.71	9.13±0.03	65.73±0.94	20.18
CSRV001	Korea	61.57±0.23	21.05±0.19	103.55±0.39	25.07
CSRV003	Korea	78.17±0.38	11.84±0.27	94.88±1.02	26.33
CSRV015	Korea	135.38±0.27	34.24±0.33	163.55±0.22	27.31
CSRV016	Korea	52.07±0.49	10.49±0.18	107.63±0.45	26.37
CSRV017	Korea	45.44±0.10	10.09±0.36	80.59±0.10	27.16
CSRV018	Korea	31.00±0.37	6.59±0.24	44.00±0.44	25.08
CSRV020	Korea	154.42±0.44	12.24±0.07	216.59±0.98	27.47
CSRV021	Korea	103.75±0.60	25.87±2.41	187.85±1.18	20.44
CSRV022	Korea	156.49±2.18	8.25±0.09	194.38±2.27	27.96
CSRV023	Korea	104.43±1.71	10.95±0.14	173.16±0.29	23.44
CSRV024	Korea	116.66±2.87	9.70±0.67	172.88±0.72	26.12
CSRV027	Korea	181.36±1.12	28.89±0.26	176.14±2.49	35.98
CSRV029	Korea	33.59±2.37	6.57±0.99	122.11±3.24	19.50
CSRV030	Korea	36.33±0.23	4.89±1.12	120.25±0.57	19.33
CSRV031	Korea	158.54±0.04	6.59±0.10	188.95±0.56	34.21
CSRV032	Korea	85.31±4.24	16.29±1.35	117.06±0.95	24.84
CSRV033	Korea	96.60±0.26	13.49±0.11	140.10±0.69	30.70
CSRV034	Korea	93.98±2.50	15.13±0.46	116.86±1.03	28.57
CSRV035	Korea	99.23±0.40	13.79±1.37	125.38±0.77	31.60
CSRV036	Korea	61.51±0.70	11.56±0.20	69.55±0.09	33.67
CSRV037	Korea	107.75±1.48	20.94±0.47	146.28±0.93	33.25
CSRV038	Korea	135.56±0.57	8.59±0.04	192.67±0.83	29.49
CSRV039	Korea	35.02±0.90	2.85±0.14	58.69±0.44	23.18
CSRV040	Korea	66.94±1.50	13.01±0.08	61.82±0.37	32.61
CSRV041	Korea	69.41±0.75	18.13±0.15	92.15±0.56	33.49
CSRV042	Korea	37.79±2.24	7.01±1.58	107.46±0.49	25.25

^aEach value represents the mean \pm standard deviation (n = 2).

^bPercentage of three glycosides to total isoflavones.

Supplementary Appendix 2. Content ($\mu\text{g/g}$ dry weight) of Three Malonylglycosides in Soybean Sprouts^a.

IT No.	Origin	Malonyldaidzin	Malonylglycitin	Malonylgenistin	% ^b
CS00732	China	131.84±1.47	400.08±3.38	221.19±3.18	62.84
CS00733	China	107.84±0.73	188.83±2.05	386.99±2.31	68.49
CS00734	China	105.44±1.63	170.97±9.57	243.24±0.65	59.81
CS01248	China	181.39±0.59	46.45±0.01	431.17±1.28	63.51
CS01250	China	59.60±0.09	15.18±0.94	116.32±0.85	20.82
CS01252	China	126.31±0.58	39.64±0.51	271.53±0.06	48.43
CS01277	China	86.60±0.03	41.14±0.20	172.37±0.68	48.36
CS01278	China	82.87±0.42	24.75±0.83	104.63±0.63	49.87
CS01381	China	101.25±0.05	34.63±1.82	154.87±0.36	57.86
CS01383	China	187.62±0.78	52.23±0.67	246.51±1.76	52.99
CS01385	China	53.77±0.20	18.15±0.11	166.07±0.99	32.88
CS01386	China	38.73±0.01	16.69±0.07	169.24±0.01	32.87
CS01567	China	200.45±1.77	92.04±1.75	388.53±0.34	65.42
CS01804	China	33.88±0.02	13.61±0.57	141.84±1.54	42.17
CS02685	China	28.05±0.27	28.75±0.66	76.99±0.67	13.74
CS00736	Japan	179.23±0.36	369.41±5.74	405.25±1.12	72.34
CS00738	Japan	101.65±1.33	100.69±1.04	178.32±1.08	65.56
CS00740	Japan	221.82±6.42	160.04±1.70	435.77±3.35	71.84
CS02052	Japan	89.19±0.12	21.99±0.07	114.88±1.03	58.12
CS02059	Japan	46.88±0.11	14.32±0.92	120.95±0.26	45.02
CS02065	Japan	126.63±1.87	32.07±0.37	439.88±0.22	58.24
CS02205	Japan	133.53±0.37	24.48±0.98	396.49±0.37	41.18
CS02564	Japan	104.64±1.00	39.52±0.26	260.36±0.71	43.80
CS02587	Japan	45.32±0.25	21.15±0.62	100.13±0.89	47.52
CS02590	Japan	78.08±0.11	19.35±0.31	173.38±0.81	60.52
CS02599	Japan	174.43±0.31	38.13±0.09	383.81±0.34	69.08
CS02665	Japan	94.98±0.09	26.29±0.57	230.69±1.03	49.89
CS01407	Korea	210.36±1.19	46.61±0.30	545.38±1.55	62.70
CS01408	Korea	120.56±1.50	19.67±0.76	243.16±4.19	41.04
CS01416	Korea	125.85±0.61	40.38±0.25	266.22±3.22	54.12
CS01426	Korea	111.23±0.59	32.44±0.74	277.49±0.69	48.48
CS01429	Korea	37.57±1.23	21.70±0.55	94.83±0.23	34.51
CS01924	Korea	52.00±0.16	27.54±0.51	313.64±2.38	36.34
CS01949	Korea	214.18±0.66	20.76±0.55	506.55±2.61	64.63
CS01972	Korea	117.38±0.69	32.87±0.27	255.40±2.50	60.20
CS02554	Korea	168.17±1.81	27.28±0.03	491.72±0.87	63.74
CS02576	Korea	61.70±0.23	31.88±0.52	203.18±0.19	52.23
CS02594	Korea	105.54±0.57	28.65±0.59	256.67±1.43	56.31
CS02602	Korea	19.42±0.38	69.09±0.86	229.76±2.55	60.85
CS02603	Korea	168.75±0.20	65.04±1.64	363.57±0.37	61.86

Supplementary Appendix 2. Content ($\mu\text{g/g}$ dry weight) of Three Malonylglycosides in Soybean Sprouts^a (Continued).

IT No.	Origin	Malonyldaidzin	Malonylglycitin	Malonylgenistin	% ^b
CS02606	Korea	93.31 \pm 0.40	51.88 \pm 2.20	241.49 \pm 0.04	62.56
CS02615	Korea	72.91 \pm 0.48	31.97 \pm 2.85	226.81 \pm 1.01	58.22
CSRV001	Korea	87.32 \pm 0.50	60.36 \pm 0.98	266.40 \pm 1.24	55.75
CSRV003	Korea	86.33 \pm 0.66	37.52 \pm 7.64	258.40 \pm 0.86	54.42
CSRV015	Korea	216.47 \pm 1.48	71.00 \pm 0.11	483.45 \pm 2.84	63.20
CSRV016	Korea	72.53 \pm 0.23	31.78 \pm 0.16	311.62 \pm 1.74	64.44
CSRV017	Korea	62.09 \pm 0.72	26.96 \pm 0.64	229.69 \pm 2.27	63.60
CSRV018	Korea	50.82 \pm 0.06	22.98 \pm 0.48	116.38 \pm 1.49	58.46
CSRV020	Korea	177.65 \pm 0.82	41.91 \pm 0.26	685.69 \pm 2.37	64.88
CSRV021	Korea	102.40 \pm 0.11	32.55 \pm 1.19	466.74 \pm 0.89	38.74
CSRV022	Korea	140.61 \pm 0.22	25.45 \pm 1.51	557.33 \pm 1.31	56.33
CSRV023	Korea	135.27 \pm 0.82	34.41 \pm 0.65	523.92 \pm 3.74	56.34
CSRV024	Korea	161.63 \pm 2.97	31.27 \pm 0.22	508.14 \pm 2.40	61.17
CSRV027	Korea	112.36 \pm 3.68	45.02 \pm 1.42	384.70 \pm 0.02	50.48
CSRV029	Korea	73.64 \pm 6.36	22.05 \pm 0.48	457.31 \pm 0.71	66.44
CSRV030	Korea	75.99 \pm 0.59	18.82 \pm 1.76	468.74 \pm 1.37	67.48
CSRV031	Korea	84.73 \pm 0.60	21.84 \pm 8.33	410.61 \pm 0.15	49.96
CSRV032	Korea	150.11 \pm 2.98	36.78 \pm 8.66	399.06 \pm 0.46	66.56
CSRV033	Korea	119.29 \pm 0.59	35.60 \pm 0.49	370.40 \pm 0.03	64.45
CSRV034	Korea	131.51 \pm 2.16	51.48 \pm 9.85	348.79 \pm 4.69	67.23
CSRV035	Korea	92.76 \pm 0.66	29.34 \pm 1.87	328.18 \pm 2.10	59.68
CSRV036	Korea	50.31 \pm 0.32	26.72 \pm 0.21	144.99 \pm 0.13	52.43
CSRV037	Korea	75.06 \pm 2.56	34.26 \pm 0.55	302.59 \pm 2.92	49.81
CSRV038	Korea	104.77 \pm 0.62	26.68 \pm 0.19	546.17 \pm 0.22	59.32
CSRV039	Korea	39.71 \pm 0.07	15.65 \pm 0.40	201.89 \pm 0.03	61.75
CSRV040	Korea	50.75 \pm 0.01	25.92 \pm 1.72	148.34 \pm 1.39	51.75
CSRV041	Korea	51.40 \pm 0.17	26.67 \pm 0.36	201.61 \pm 1.27	52.13
CSRV042	Korea	34.25 \pm 0.34	25.34 \pm 3.44	296.91 \pm 0.94	59.11

^aEach value represents the mean \pm standard deviation (n = 2).

^bPercentage of three malonylglycosides to total isoflavones.

Supplementary Appendix 3. Content ($\mu\text{g/g}$ dry weight) of Three Acetylglycosides in Soybean Sprouts^a.

IT No.	Origin	Acetylaidzin	Acetylglycitin	Acetylgenistin	% ^b
CS00732	China	3.77±0.01	6.65±0.18	50.38±3.22	5.07
CS00733	China	4.18±0.15	11.81±0.03	23.11±0.24	3.92
CS00734	China	4.72±0.22	8.09±0.57	39.61±2.91	6.03
CS01248	China	2.94±0.03	5.27±0.12	7.60±0.69	1.52
CS01250	China	5.79±0.06	4.53±0.10	124.71±0.80	14.71
CS01252	China	2.96±0.08	4.48±0.06	6.47±0.07	1.54
CS01277	China	2.26±0.23	3.03±0.27	13.64±0.38	3.05
CS01278	China	2.88±0.02	3.15±0.04	7.28±0.14	3.13
CS01381	China	5.85±0.21	4.64±0.01	7.91±0.33	3.66
CS01383	China	6.30±0.96	8.02±0.68	26.89±3.82	4.49
CS01385	China	5.73±0.07	5.19±0.67	45.75±0.66	7.83
CS01386	China	3.74±0.10	5.00±0.14	78.16±0.10	12.71
CS01567	China	10.58±0.69	2.99±0.16	17.16±0.00	2.95
CS01804	China	11.32±0.64	11.01±1.12	30.97±0.46	11.87
CS02685	China	6.54±0.47	7.03±0.06	303.65±0.15	32.58
CS00736	Japan	1.73±0.03	4.62±0.37	28.68±1.60	2.66
CS00738	Japan	2.70±0.16	4.62±0.17	3.31±0.74	1.83
CS00740	Japan	3.87±0.01	6.95±0.27	8.38±1.10	1.69
CS02052	Japan	2.94±0.13	5.77±0.13	6.68±0.18	3.96
CS02059	Japan	2.98±0.00	6.77±0.25	0.24±0.03	2.47
CS02065	Japan	3.29±0.25	11.55±0.34	42.45±0.51	5.57
CS02205	Japan	4.53±0.25	9.41±0.22	52.22±1.85	4.91
CS02564	Japan	4.74±0.13	3.81±0.18	112.90±1.78	13.15
CS02587	Japan	1.67±0.03	2.43±0.12	4.75±1.18	2.52
CS02590	Japan	4.93±0.45	4.32±0.01	17.92±0.04	6.07
CS02599	Japan	23.24±1.07	8.89±1.38	1.89±0.25	3.94
CS02665	Japan	2.27±0.43	6.94±0.11	4.09±1.19	1.89
CS01407	Korea	2.62±0.04	9.66±0.16	4.63±0.41	1.32
CS01408	Korea	3.03±0.52	8.01±0.84	30.27±0.26	4.42
CS01416	Korea	2.66±0.11	8.02±0.08	13.86±0.53	3.07
CS01426	Korea	1.31±0.00	6.72±0.15	29.61±0.66	4.33
CS01429	Korea	trace	3.07±0.83	17.75±0.17	4.66
CS01924	Korea	5.64±0.41	10.68±0.19	222.31±2.30	22.05
CS01949	Korea	15.43±0.31	trace	8.62±0.23	2.10
CS01972	Korea	4.03±0.14	9.60±0.20	2.93±0.01	2.46
CS02554	Korea	14.39±0.16	trace	5.49±0.70	1.84
CS02576	Korea	6.88±0.24	2.33±0.02	5.53±0.94	2.60
CS02594	Korea	5.60±0.09	3.13±0.03	18.88±0.25	3.98
CS02602	Korea	4.82±0.14	3.10±0.14	12.95±0.08	3.99
CS02603	Korea	7.75±0.23	4.44±0.12	63.14±4.08	7.80

Supplementary Appendix 3. Content ($\mu\text{g/g}$ dry weight) of Three Acetylglycosides in Soybean Sprouts^a (Continued).

IT No.	Origin	Acetylaidzin	Acetylglycitin	Acetylgenin	% ^b
CS02606	Korea	5.56±0.19	3.98±0.05	18.57±1.60	4.55
CS02615	Korea	5.37±0.16	5.71±0.29	12.86±1.49	4.20
CSRV001	Korea	5.73±0.06	5.94±0.05	14.40±0.26	3.51
CSRV003	Korea	4.68±0.03	4.65±0.02	41.07±1.20	7.18
CSRV015	Korea	3.58±0.15	6.85±0.27	11.77±0.80	1.82
CSRV016	Korea	2.84±0.22	3.71±0.39	9.43±2.61	2.47
CSRV017	Korea	3.71±0.24	0.49±0.00	8.50±0.29	2.53
CSRV018	Korea	7.70±0.05	5.37±1.71	4.74±1.27	5.47
CSRV020	Korea	7.58±0.05	2.15±0.07	7.18±0.06	1.21
CSRV021	Korea	7.88±0.28	7.03±0.13	5.85±1.15	1.34
CSRV022	Korea	6.16±0.21	2.39±0.30	20.70±0.11	2.28
CSRV023	Korea	2.67±0.11	6.14±0.37	6.18±1.33	1.22
CSRV024	Korea	8.41±0.79	2.41±0.23	8.61±0.83	1.70
CSRV027	Korea	3.14±0.18	5.85±0.39	20.95±0.47	2.79
CSRV029	Korea	2.45±0.12	4.90±0.05	34.14±1.17	4.99
CSRV030	Korea	4.32±0.14	6.56±1.02	9.22±1.37	2.41
CSRV031	Korea	3.50±0.14	8.88±1.04	10.09±2.20	2.17
CSRV032	Korea	2.20±0.03	6.57±0.01	trace	1.00
CSRV033	Korea	7.80±0.19	2.18±0.29	7.81±0.45	2.18
CSRV034	Korea	4.69±0.09	2.50±0.07	2.75±0.28	1.26
CSRV035	Korea	5.59±0.22	2.47±0.14	8.26±0.18	2.16
CSRV036	Korea	2.10±0.06	2.15±0.06	12.09±1.58	3.86
CSRV037	Korea	3.89±0.21	5.19±0.31	6.28±0.25	1.86
CSRV038	Korea	3.35±0.03	10.34±0.02	33.79±0.85	4.16
CSRV039	Korea	3.22±0.11	4.26±0.09	9.48±0.45	4.07
CSRV040	Korea	6.91±0.10	1.65±0.03	8.59±0.49	3.94
CSRV041	Korea	8.38±0.39	2.86±0.66	14.43±0.98	4.78
CSRV042	Korea	9.20±0.05	5.30±0.10	54.81±9.06	11.48

^aEach value represents the mean \pm standard deviation (n = 2).

^bPercentage of three acetylglycosides to total isoflavones.

Supplementary Appendix 4. Content ($\mu\text{g/g}$ dry weight) of Three Aglycones in Soybean Sprouts^a.

IT No.	Origin	Daidzein	Genistein	Glycitein	% ^b
CS00732	China	109.45±0.44	2.68±0.05	5.07±0.31	9.78
CS00733	China	36.93±0.28	2.54±0.19	4.10±0.13	4.37
CS00734	China	52.27±1.15	3.39±0.02	12.56±0.34	7.85
CS01248	China	28.88±0.03	1.74±0.05	15.19±0.33	4.41
CS01250	China	404.84±0.89	8.94±0.10	70.00±0.00	52.70
CS01252	China	80.33±0.32	2.76±0.02	21.36±0.49	11.56
CS01277	China	71.23±0.44	2.31±0.04	15.84±0.01	14.40
CS01278	China	49.12±0.02	4.01±0.04	17.03±0.22	16.49
CS01381	China	45.93±0.28	2.12±0.04	12.11±1.89	11.97
CS01383	China	123.87±0.74	9.72±0.54	30.92±0.43	17.93
CS01385	China	179.73±0.11	4.09±0.01	52.47±0.25	32.65
CS01386	China	169.43±0.22	2.11±0.00	42.88±0.15	31.37
CS01567	China	43.47±0.76	1.95±0.02	16.25±0.71	5.92
CS01804	China	74.36±0.02	2.38±0.09	25.95±0.88	22.87
CS02685	China	260.48±0.87	3.29±0.20	83.88±1.04	35.70
CS00736	Japan	57.14±0.09	1.94±0.13	13.26±0.58	5.49
CS00738	Japan	48.77±0.18	3.37±0.02	10.40±0.18	10.77
CS00740	Japan	37.57±1.52	1.92±0.13	21.08±0.10	5.32
CS02052	Japan	44.90±0.14	3.13±0.00	13.43±0.52	15.80
CS02059	Japan	49.93±0.31	1.74±0.06	13.92±0.23	16.21
CS02065	Japan	109.50±0.47	2.53±0.01	29.22±0.29	13.74
CS02205	Japan	285.85±0.96	4.73±0.08	91.99±0.43	28.41
CS02564	Japan	123.45±0.11	5.29±0.02	33.07±0.53	17.52
CS02587	Japan	25.39±0.34	1.96±0.01	7.91±0.28	10.06
CS02590	Japan	21.25±0.81	3.23±0.06	16.82±2.22	9.23
CS02599	Japan	48.96±1.74	3.80±0.06	13.42±0.12	7.67
CS02665	Japan	164.00±1.70	8.04±0.22	44.71±0.44	30.72
CS01407	Korea	66.05±0.08	3.19±0.20	13.98±0.02	6.50
CS01408	Korea	176.45±0.18	6.78±0.04	31.36±1.53	22.97
CS01416	Korea	39.96±0.35	2.82±0.19	8.79±1.17	6.45
CS01426	Korea	75.06±0.63	2.15±0.10	20.28±1.42	11.22
CS01429	Korea	92.97±1.26	1.92±0.05	16.44±0.63	24.94
CS01924	Korea	193.98±0.87	3.24±0.12	63.13±0.03	24.06
CS01949	Korea	36.13±0.22	2.15±0.04	14.72±0.50	4.62
CS01972	Korea	48.94±0.16	3.48±0.04	14.77±0.33	9.97
CS02554	Korea	34.68±0.04	2.24±0.05	16.83±0.76	4.98
CS02576	Korea	100.13±0.36	4.63±0.36	21.62±0.90	22.24
CS02594	Korea	26.53±0.58	3.30±0.15	9.52±0.16	5.67
CS02602	Korea	33.80±1.62	3.52±0.04	11.52±0.23	9.34
CS02603	Korea	94.70±0.24	5.16±0.58	23.10±0.04	12.73

Supplementary Appendix 4. Content ($\mu\text{g/g}$ dry weight) of Three Aglycones in Soybean Sprouts^a (Continued).

IT No.	Origin	Daidzein	Genistein	Glycitein	% ^b
CS02606	Korea	51.55 \pm 3.09	4.85 \pm 0.09	19.04 \pm 0.59	12.20
CS02615	Korea	66.06 \pm 0.02	6.59 \pm 0.12	26.48 \pm 0.51	17.40
CSRV001	Korea	77.37 \pm 0.31	14.33 \pm 0.18	24.75 \pm 1.86	15.68
CSRV003	Korea	61.57 \pm 0.56	4.62 \pm 0.13	18.58 \pm 0.46	12.07
CSRV015	Korea	70.70 \pm 0.20	5.65 \pm 0.77	17.11 \pm 0.13	7.66
CSRV016	Korea	30.15 \pm 0.48	3.03 \pm 0.06	10.17 \pm 0.41	6.72
CSRV017	Korea	26.34 \pm 1.07	1.96 \pm 0.01	5.28 \pm 0.13	6.70
CSRV018	Korea	22.60 \pm 0.28	2.15 \pm 0.13	11.00 \pm 0.02	10.99
CSRV020	Korea	64.78 \pm 0.08	7.25 \pm 0.05	17.75 \pm 0.72	6.43
CSRV021	Korea	432.70 \pm 0.73	55.24 \pm 0.17	125.35 \pm 0.88	39.49
CSRV022	Korea	120.94 \pm 1.92	1.75 \pm 0.02	49.79 \pm 1.00	13.43
CSRV023	Korea	143.64 \pm 0.26	8.22 \pm 0.14	82.11 \pm 5.10	19.00
CSRV024	Korea	98.26 \pm 0.56	3.50 \pm 0.06	24.87 \pm 34.09	11.01
CSRV027	Korea	102.16 \pm 1.70	9.18 \pm 0.14	4.04 \pm 0.17	10.74
CSRV029	Korea	44.25 \pm 0.47	4.77 \pm 0.20	26.49 \pm 0.57	9.07
CSRV030	Korea	51.83 \pm 0.11	4.44 \pm 0.06	33.80 \pm 0.09	10.78
CSRV031	Korea	100.56 \pm 0.46	4.16 \pm 0.13	36.64 \pm 0.18	13.66
CSRV032	Korea	49.03 \pm 0.68	3.18 \pm 0.11	14.80 \pm 1.04	7.61
CSRV033	Korea	14.01 \pm 0.00	3.67 \pm 0.10	4.12 \pm 0.49	2.67
CSRV034	Korea	15.87 \pm 0.74	3.15 \pm 1.18	4.29 \pm 0.49	2.94
CSRV035	Korea	41.47 \pm 0.52	2.28 \pm 0.10	5.76 \pm 0.08	6.56
CSRV036	Korea	33.13 \pm 0.58	5.11 \pm 0.14	4.27 \pm 0.32	10.04
CSRV037	Korea	98.47 \pm 0.26	10.91 \pm 0.32	15.36 \pm 0.25	15.08
CSRV038	Korea	55.89 \pm 0.19	5.18 \pm 0.15	19.31 \pm 0.45	7.04
CSRV039	Korea	33.43 \pm 0.09	2.32 \pm 0.19	10.09 \pm 0.10	11.00
CSRV040	Korea	44.63 \pm 1.27	3.18 \pm 0.06	3.09 \pm 0.05	11.70
CSRV041	Korea	41.27 \pm 1.54	4.08 \pm 0.18	6.13 \pm 0.36	9.59
CSRV042	Korea	17.04 \pm 0.08	2.72 \pm 0.31	5.35 \pm 0.01	4.16

^aEach value represents the mean \pm standard deviation (n = 2).

^bPercentage of three aglycones to total isoflavones.