

Blood haematology, muscle pH and serum cortisol changes in pigs with different levels of drip loss

Nunyarat Koomkrong¹, Chaiwat Boonkaewwan¹, Watchara Laenoi², and Autchara Kayan^{1,*}

* Corresponding Author: Autchara Kayan Tel: +66-81-992-4645, Fax: +66-25-79-1120, E-mail: fagrark@ku.ac.th

¹ Department of Animal Science, Faculty of Agriculture, Kasetsart University, Bangkok 10900, Thailand

² Department of Animal Science, School of Agriculture and Natural Resources, University of Phayao, Phayao 56000, Thailand

Submitted Jan 21, 2017; Revised Apr 26, 2017; Accepted May 24, 2017

Objective: An experiment was conducted to study the blood haematology, muscle pH, and serum cortisol changes in pigs with different levels of drip loss.

Methods: Two groups (low and high) of 20 animals were selected from 100 pigs based on drip loss. All [Duroc× (Large White×Landrace)] pigs were slaughtered according to standard slaughtering procedures. At exsanguinations, blood samples were taken for the haematological parameters and serum cortisol analysis. The muscle samples were taken from *longissimus dorsi* muscle to evaluate the muscle pH and drip loss.

Results: Haematological parameters of low drip loss group showed higher content of white blood cells and monocytes than high drip loss group (p<0.05). The low drip loss group had higher muscle pH at 45 min (p<0.05) and 24 h (p<0.001) post-mortem than the high drip loss group. However, there was no significant difference in serum cortisol levels (p>0.05).

Conclusion: Drip loss is mainly affected by the muscle pH decline after slaughter and also might be affected by white blood cells and monocytes.

Keywords: Drip Loss; Haematology; Muscle pH; Serum Cortisol; Pig

INTRODUCTION

Meat quality is an indicator of acceptability or preference for the consumer. The important quality traits for fresh meat consist of color, water-holding capacity, texture and fat distribution [1]. Especially, water-holding capacity determined by drip loss is a main quality attribute of fresh meat. Drip loss has financial implications due to the loss of weight, reduced acceptance and rejection by consumers [2]. The large variations are affected by many factors in the whole meat production chain including physiological factors, rearing conditions and processing factors [2]. Particularly, the rate of pH decline is the key factor that correlates to drip loss [3,4]. pH is mainly influenced by pre-slaughter stress [5]. Stress before slaughter can cause a faster muscle glycogen breakdown and thus lower pH values in the muscle when the temperature of the carcass is still high. The combination of low pH and high temperature causes the denaturation of muscle proteins leading to reduction in the meat water holding capacity [6].

Haematology blood parameters are good indicators of the physiological and health status in animals [7,8]. It has been reported that haematological parameters could be employed to high-light the stress condition during transport [9,10]. Decrease in stress has been documented as one of the factors influencing on heterophils, lymphocytes and total white blood count levels [11]. Furthermore, cortisol is the main hormone of the hypothalamic-pituitary-adrenocortical (HPA) axis responding to stress [12]. Increase of serum cortisol was observed in pigs under stress conditions including high temperature and transportation [13]. Increased cortisol levels lead to increase post-mortem metabolism that could be reflected on meat quality [14]. Higher serum cortisol was related poor pork quality with a higher muscle temperature [15], faster muscle pH decline,

Copyright © 2017 by Asian-Australasian Journal of Animal Sciences

AJAS

higher drip loss and lightness in pork [16]. Therefore, the objective of this study was to investigate the blood haematology, pH muscle and serum cortisol changes in pigs with different levels of drip loss to obtain evidence that could be used for prediction water-holding capacity in meat.

MATERIALS AND METHODS

Animals and muscle sampling

A total of 100 three crossbred pigs [Duroc×(Large White× Landrace)] were obtained from a commercial slaughterhouse in Thailand. All pigs were slaughtered according to standard slaughtering procedures. The average of slaughter weight was $112.13\pm$ 4.81 kg. After electrical stunning, carcasses were scalded, cleaned, eviscerated and split. Two groups (low and high) of 20 animals were selected from 100 pigs based on drip loss. The muscle samples were immediately taken from the *longissimus dorsi* muscles at the 2nd to 6th of lumbar vertebrae for further analysis.

Blood haematological and cortisol analysis

Blood samples were collected from each pig (n = 20) within 30 s during exsanguination after electrical stunning and sticking. The blood samples were collected using two types of tubes: the first tube was treated with ethylenediamine tetra-acetic acid (EDTA) to prevent blood coagulation, and the second tube was collected without EDTA. The first tube was used for haematological analysis including red blood cells (RBC), hemoglobin, hematocrit, mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), platelets, and white blood cells (WBC). These parameters were analyzed with hematological analyzer (Cell Dyn 3700, Wiesbaden, Hesse, Germany). The non-anticoagulated blood (second tube) was centrifuged at 3,000 rpm for 10 min to separate the serum. The serum samples were collected into microtubes and stored at -20°C until the determination of cortisol concentration by chemiluminescent microparticle immunoassay using IMMULITE 1000 automated immunoassay system (Siemens Healthcare Diagnostics Inc., Flanders, NJ, USA).

Muscle pH and drip loss analysis

The *longissimus dorsi* muscle pH was measured at 45 min and 24 h post-mortem using a pH meter with a spear-type electrode (pH Spear, Eutech Instruments, Singapore City, Singapore). Drip loss was scored base on a bag method with a size-standardized sample from *longissimus dorsi* muscle collected at 24 h post-mortem that was weighed, suspended in a plastic bag, held at 4°C for 48 h, and thereafter re-weighed. Drip loss was expressed as a percentage [17].

Statistical analysis

Haematological parameters and cortisol levels were described by descriptive statistics (mean, standard deviation, minimum, and maximum value). The effect of low and high drip loss groups on blood haematological parameters, muscle pH and cortisol levels was analyzed by *t*-tests of SAS (SAS Inst. Inc., Cary, NC, USA). Values of p<0.05 were considered to indicate statistically significant differences. The results are presented as least squares means with the standard errors.

RESULTS

Based on the data obtained, two drip loss groups were identified: low (<1.70%; n = 10) and high (>4.00%; n = 10) drip loss. Mean of the low and high drip loss groups was $1.19\%\pm0.20\%$ and 6.04% $\pm0.33\%$, respectively.

Blood haematological parameters

The mean values, standard deviations and overall ranges for blood haematological parameters of the pig population are shown in Table 1. The differences in values of haematological parameters between drip loss groups are presented in Table 2. The significant differences were observed in WBC and monocyte count (p<0.05). The low drip loss group had higher WBC (22.67 ± 2.58 vs $14.93\pm1.15\times10^3$ /mm³, respectively) and monocyte count ($3.30\%\pm0.30\%$ vs $2.40\%\pm0.27\%$, respectively). However, there were no significant differences between drip loss groups in RBC, hemoglobin, haematocrit, MCV, MCH, MCHC, platelets, neutrophils, eosinophils, lymphocytes and neutrophil:lymphocyte ratio.

Muscle pH

Muscle pH values in the low and high drip loss groups are presented in Figure 1. There were significant differences both in muscle pH at 45 min and 24 h post-mortem. The low drip loss group had a higher muscle pH at 45 min post-mortem than the high drip loss group (p<0.05). At 24 h post-mortem, the low drip loss group was at a higher muscle pH than the high drip loss

Table	1.	Characterization	of	blood	haemato	ogical	parameters in	ı piqs	(n = 20)

Parameter	Mean	SD	Min	Max
RBC ($\times 10^{6}$ /mm ³)	6.54	0.81	4.10	7.70
Hemoglobin (g/dL)	11.86	1.54	6.90	13.50
Haematocrit (%)	34.75	4.67	21.00	40.00
MCV (fL)	53.46	3.01	45.60	58.00
MCH (pg)	18.06	1.39	15.30	20.20
MCHC (g/dL)	33.76	1.30	31.00	36.00
Platelets ($\times 10^3$ /mm ³)	327.65	90.32	134.00	511.00
WBC ($\times 10^3$ /mm ³)	18.80	7.32	9.40	35.80
Neutrophils (%)	68.10	9.54	45.00	82.00
Eosinophils (%)	1.95	1.50	1.00	5.00
Lymphocytes (%)	27.10	9.31	15.00	51.00
Monocytes (%)	2.85	0.99	1.00	5.00
Neutrophil:lymphocyte	2.90	1.29	0.88	5.47

SD, standard deviation; RBC, red blood cells; MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; WBC, white blood cells.

Table 2. Influence of drip loss on blood haematological parameters in pigs

léanna	Drij			
items	Low (n = 10)	High (n = 10)	p-value	
RBC ($\times 10^{6}$ /mm ³)	6.39 ± 0.30	6.68 ± 0.20	0.440	
Hemoglobin (g/dL)	11.75 ± 0.55	11.98 ± 0.44	0.748	
Haematocrit (%)	34.40 ± 1.63	35.10 ± 1.39	0.747	
MCV (fL)	54.21 ± 0.69	52.71 ± 1.14	0.276	
MCH (pg)	18.26 ± 0.42	17.85 ± 0.47	0.526	
MCHC (g/dL)	33.69 ± 0.46	33.84 ± 0.38	0.805	
Platelets ($\times 10^3$ /mm ³)	407.69 ± 30.50	316.60 ± 27.66	0.598	
WBC ($\times 10^3$ /mm ³)	$22.67 \pm 2.58^{\circ}$	$14.93 \pm 1.15^{\text{b}}$	0.018	
Neutrophils (%)	69.70 ± 2.87	66.50 ± 3.23	0.468	
Eosinophils (%)	1.60 ± 0.43	2.30 ± 0.52	0.310	
Lymphocytes (%)	25.40 ± 2.60	28.80 ± 3.30	0.429	
Monocytes (%)	$3.30\pm0.30^{\text{a}}$	$2.40\pm0.27^{\text{b}}$	0.038	
Neutrophil:lymphocyte	3.12 ± 0.42	2.68 ± 0.40	0.470	

RBC, red blood cells; MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; WBC, white blood cells. ^{a,b} Values within a row with different superscripts differ significantly at p < 0.05.

group (p<0.001).

Serum cortisol levels

The result for serum cortisol levels for each group is shown in Figure 2. The low and high drip loss group had 7.66 ± 0.80 and $6.74\pm0.55 \ \mu g/dL$ of serum cortisol, respectively. There was no significant difference in serum cortisol levels between drip loss groups.

DISCUSSION

Blood haematological parameters

Blood haematological parameters are important in assessing the response of animals to various physiological situations [8]. Merck manual reported the normal range of values for pigs as follows: RBC, 5 to 8×10^6 /mm³; hemoglobin, 10 to 16 g/dL; hematocrit, 36% to 43%; MCV, 50 to 68 fL; MCH, 17 to 21 pg; MCHC, 30 to



Figure 2. Influence of drip loss on serum cortisol levels in piqs (n = 10 per group).

34 g/dL; platelets, 200 to 500×10^3 /mm³; WBC, 11 to 22×10^3 /mm³; neutrophils, 28% to 47%; eosinophils, 0.5% to 11%; lymphocytes, 39% to 62%; and monocytes, 2% to 10%. The average haematological parameters in this study were represented in the normal range except for neutrophils and lymphocytes. There were higher mean values of neutrophils and lower lymphocytes values in comparison to the normal range. This may have been influenced by stress prior to slaughter. An increase in neutrophil and a decrease in lymphocyte count occurs during stress [10,18]. Because stress stimulates the anterior pituitary gland to secrete adrenocorticotropic hormone which induces the adrenal cortex to produce glucocorticoids, involved in the mobilization of neutrophils will cause a decrease in lymphocytes. Neutrophils were significantly negatively correlated with lymphocytes (r = -0.95) [19].

Haematological traits are essential parameters for evaluating the health and physiological status of animals that can reflect the physiological responsiveness [7]. Haematological parameters have been commonly used as indicators of pre-slaughter stress [10]. Pre-slaughter stress is the main factor affecting meat quality [5] due to a faster muscle glycogen breakdown [6]. High pre-



Figure 1. Influence of drip loss on muscle pH at 45 min (A) and 24 h (B) post-mortem in *longissimus dorsi* muscle of pigs (* p<0.05; *** p<0.001) (n = 10 per group).

AJAS

slaughter stress was related to increased drip loss in pork [20]. This study indicated that pork with high water-holding capacity showed higher WBC and monocyte count. Previous study indicated that animals with high WBC have enhanced adaptability to environmental stresses [8]. Also, normal pigs had higher WBC, lymphocytes, and monocytes in comparison to pigs with a chronic stress syndrome due to failure to adapt to the new environmental [21]. In addition, WBC was positive correlated with monocytes (r = 0.58) [19]. However, it is unclear regarding haematological changes to the physiological response. Previous study has reported that the increasing in WBC may indicate that cell damage occurred during the pre-slaughter period. This resulted in an inflammation response which in turn led to an increase in the amount of WBC in the blood [9].

Muscle pH

This study indicated that pork with lower drip loss showed higher muscle pH at 45 min and 24 h post-mortem than pork with higher drip loss. The previous study reported that drip loss was a negatively correlated to pH at 45 min and 24 h postmortem (r = -0.40and -0.50, respectively) [22]. Muscle pH is the best indicator of muscle to meat conversion including meat color, texture and moisture [5]. Once the muscle has shifted to use anaerobic glycolysis as a major energy-generating pathway, the pH is reflective of the accumulation of lactic acid within the muscle, resulting in the pH decline in post-mortem muscle. The muscle pH early postmortem (45 min) has been used to monitor the quality difference in fresh pork [3]. Drip loss varies due to post-mortem metabolism as a result of ATP degradation and the rate of acidification [4]. The faster pH decline caused denaturation of sarcoplasmic and myofibrillar proteins, resulting in reduced water holding capacity [23]. Furthermore, when the pH reaches the isoelectric point (pI) of the major proteins (pH = 5.4), the result is a reduced amount of water that can be attracted and held by the protein [24], and a reduced repulsion of structures within the myofibril. Water moves from the myofibril into the extramyofibrillar spaces, where it eventually is lost from the muscle cell [25].

Serum cortisol levels

Pre-slaughter stress stimulates the two main stress-responsive neuroendocrine systems including the HPA axis and the sympathetic nervous system. The activation of the HPA axis regulates secretion of cortisol in response to stress [26]. Pigs handled with high stress at pre-slaughter had higher levels of cortisol compared with low stress [27]. Serum cortisol levels increased during the journey (3.47 μ g/dL at loading and 8.52 μ g/dL at unloading) but decreased during the lairage (6.96 μ g/dL at exsanguination) [13]. Increased levels of serum cortisol related to increased blood glucose and lactate level, resulting in a faster muscle pH decline, high drip loss, and lightness in pork [16]. Previous studies have investigated the association between cortisol level and meat quality [12,16]. This study shown serum cortisol level did not differ between drip loss groups. The same result as the previous reports, as the level of cortisol was not associated with meat quality [15,28]. This is explained by the fact that the plasma clearance of cortisol is rapid [29]. In pigs, the cortisol levels rose immediately after the start of transport and decreased rapidly after unloading [30]. Of all stress indicators (lactate, cortisol, and catecholamines) measured at exsanguination, only blood lactate was strongly correlated with pork quality traits [20]. Therefore, this result indicates that measurement of serum cortisol levels may not be a good indicator for meat quality in pigs.

In conclusion, the present study indicated that the main factor affecting drip loss is the pH decline in post-mortem muscle. The low drip loss group had a higher muscle pH at 45 min and 24 h post-mortem than the high drip loss group. Moreover, blood haematological parameters including WBC and monocyte count affected drip loss. The low drip loss group had higher WBC and monocyte count than the high drip loss group. There was no effect of the serum cortisol levels on drip loss. Serum cortisol levels might be not the potential indicator parameters for drip loss in pork.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

ACKNOWLEDGMENTS

The authors are grateful to Department of Animal Science and Central Laboratory, Faculty of Agriculture, Kasetsart University for the utilization of laboratory facilities.

REFERENCES

- Joo ST, Kim GD, Hwang YH, Ryu YC. Control of fresh meat quality through manipulation of muscle fiber characteristics. Meat Sci 2013; 95:828-36.
- 2.den Hertog-Meischke MJA, van Laack RJLM, Smulders FJM. The water-holding capacity of fresh meat. Vet Quart 1997;19:175-81.
- 3.Huff-Lonergan E, Lonergan SM. New frontiers in understanding drip loss in pork: recent insights on the role of postmortem muscle biochemistry. J Anim Breed Genet 2007;124:19-26.
- 4.Żelechowska E, Przybylski W, Jaworska D, Santé-Lhoutellier V. Technological and sensory pork quality in relation to muscle and drip loss protein profiles. Eur Food Res Technol 2012;234:883-94.
- 5.Braden KW. Converting muscle to meat: the physiology of rigor. In: Kerth CR, editor. The science of meat quality. New York: John Wiley & Sons; 2013. p. 79-97.
- 6.Adzitey F, Nurul H. Pale soft exudative (PSE) and dark firm dry (DFD) meats: causes and measures to reduce these incidences - a mini review. Int Food Res J 2011;18:11-20.
- 7. Etim NN, Enyenihi GE, Williams ME, Udo MD, Offiong EEA. Haema-

tological parameters: indicators of the physiological status of farm animals. Br J Sci 2013;10:33-45.

- Etim NN, Williams ME, Akpabio U, Offiong EEA. Haematological parameters and factors affecting their values. Agric Sci 2014;2:37-47.
- 9.Wolmarans WJ. The effect of transport on live weight loss, meat quality and blood haematology in slaughter ostriches [master's thesis]. Stellenbosch, ZA: University of Stellenbosch; 2011.
- Giammarco M, Vignola G, Mazzone G, Fusaro I, Lambertini L. Haematological parameters as indicators of transport stress in rabbits. In: Proceedings 10th World Rabbit Congress; 2012 Sep 3-6; Sharm El Sheikh, EG: World Rabbit Science Association; 2012. p. 1033-7.
- Scope A, Filip T, Gabler C, Resch F. The influence of stress from transport and handling on hematologic and clinical chemistry blood parameters of racing pigeons (*Columba livia domestica*). Avian Dis 2002; 46:224-9.
- Škrlep M, Prevolnik M, Šegula B, Čandek-Potokar M. Association of plasma stress markers at slaughter with carcass or meat quality in pigs. Slov Vet Res 2009;46:133-42.
- 13. Averos X, Herranz A, Sanchez R, Comella JX, Gosalvez LF. Serum stress parameters in pigs transported to slaughter under commercial conditions in different seasons. Vet Med Czech 2007;52:333-42.
- Pighin DG, Cunzolo SA, Zimerman M, et al. Impact of adrenaline or cortisol injection on meat quality development of Merino hoggets. J Integr Agric 2013;12:1931-6.
- 15. Dokmanovic M, Baltic MZ, Duric J, et al. Correlations among stress parameters, meat and carcass quality parameters in pigs. Asian-Australas J Anim Sci 2015;28:435-41.
- 16. Choe JH, Kim BC. Association of blood glucose, blood lactate, serum cortisol levels, muscle metabolites, muscle fiber type composition, and pork quality traits. Meat Sci 2014;97:137-42.
- 17. Honikel KO, Kim CJ, Hamm R. Sarcomere shortening of prerigor muscles and its influence on drip loss. Meat Sci 1986;16:267-82.
- 18. Adenkola AY, Ayo JO, Sackey AKB, Adelaiye AB. Haematological and serum biochemical changes in pigs administered with ascorbic acid and transported by road for four hours during the harmattan season. J Cell Anim Biol 2009;3:21-8.
- 19. Oluwole OO, Omitogun GO. Haematological traits of Nigerian indi-

genous pig and its hybrid (50% Large White × 50 NIP) at post weaning ages. Am J Mol Biol 2016;6:45-52.

- 20. Hambrecht E, Eissen JJ, Nooijen RIJ, et al. Preslaughter stress and muscle energy largely determine pork quality at two commercial processing plants. J Anim Sci 2004;82:1401-9.
- Morrow-Tesch J, Andersson G. Immunological and hematological characterizations of the wasting pig syndrome. J Anim Sci 1994;72: 976-83.
- Ryu YC, Kim BC. The relationship between muscle fiber characteristics, postmortem metabolic rate, and meat quality of pig *longissimus dorsi* muscle. Meat Sci 2005;71:351-7.
- Scheffler TL, Gerrard DE. Mechanisms controlling pork quality development: the biochemistry controlling postmortem energy metabolism. Meat Sci 2007;77:7-16.
- 24. Huff-Lonergan E, Lonergan SM. Mechanisms of water-holding capacity of meat: the role of postmortem biochemical and structural changes. Meat Sci 2005;71:194-204.
- Apple JK, Yancey JWS. Water-holding capacity of meat. In: Kerth CR, editor. The science of meat quality. New York: John Wiley & Sons; 2013. p. 119-45.
- 26. Foury A, Lebret B, Chevillon P, et al. Alternative rearing systems in pigs: consequences on stress indicators at slaughter and meat quality. Animal 2011;5:1620-5.
- 27. Peres LM, Bridi AM, da Silva CA, et al. Effect of low or high stress in pre-slaughter handling on pig carcass and meat quality. R Bras Zootec 2014;43:363-8.
- Shaw FD, Trout GR, McPhee CP. Plasma and muscle cortisol measurements as indicators of meat quality and stress in pigs. Meat Sci 1995; 39:237-46.
- 29. McKay LI, Cidlowski JA. Pharmacokinetics of corticosteroids. In: Kufe DW, Pollock RE, Weichselbaum RR, et al., editors. Holland-frei cancer medicine. 6th ed. Hamilton, New Zealand: BC Decker; 2003.
- 30. Dalin AM, Magnusson U, Häggendal J, Nyberg L. The effect of transport stress on plasma levels of catecholamines, cortisol, corticosteroidbinding globulin, blood cell count, and lymphocyte proliferation in pigs. Acta Vet Scand 1993;34:59-68.