

Expressional Changes of Connexin Isoform Genes in the Rat Caput Epididymis Exposed to Flutamide or Estradiol Benzoate at the Early Postnatal Age

†Ki-Ho Lee

Department of Biochemistry and Molecular Biology, College of Medicine, Eulji University, Daejeon 301-746, Korea

ABSTRACT : Direct communication between neighboring cells through connexin (Cx)-based gap junction is a crucial biological manner to regulate functions of a tissue consisting of multi-cell types. The present research evaluated expressional changes of Cx isoforms in the caput epididymis of adult rat exposed to estradiol benzoate (EB) or flutamide (Flu) at the early postnatal age. A single subcutaneous administration of EB at a low-dose [0.015 µg/kg body weight (BW)] or a high-dose (1.5 µg/kg BW) or Flu at a low-dose (500 µg/kg BW) or a high-dose (5 mg/kg BW) was performed to an animal at 1 week of age. Quantitative real-time PCR analysis was employed to determine expressional changes of Cx isoforms. The transcript levels of Cxs30.3 and 37 were decreased by a low-dose EB treatment, while decreases of Cxs31, 31.1, 32, 40, and 45 transcript levels were observed with a low-dose EB treatment. The treatment of a high-dose EB resulted in expressional reduction of Cxs30.3, 31, 31.1, 37, 40, 43, and 45. The Flu treatment at a low dose caused increases of Cxs26, 37, and 40 transcript levels but decreases of Cxs31.1, 43, and 45 transcript levels. Increases of Cxs30.3, 31, 37, and 40 mRNA amounts were induced by a high-dose Flu treatment. However, exposure to a high-dose Flu produced expressional decreases of Cxs31.1, 32, and 43 in the adult caput epididymis. These observations suggest that exposure to EB or Flu at the neonatal period could lead to aberrant expression of Cx isoforms in the adult caput epididymis.

Key words : Gap junction, Connexin isoform, Estradiol benzoate, Flutamide, Real-time PCR, Caput epididymis

INTRODUCTION

The interaction between neighboring cells present in a multicellular tissue is chiefly achieved by three junctions, including tight junction, adherens junction, and gap junction (Lawrence et al., 1978). Of these junctions, gap junction involves in the direct cell-cell communication through the reciprocal exchange of materials, such as signaling molecules, ions, and small metabolites, between cells (Lawrence et al., 1978; Valiunas et al., 2005). A complex of six connexin (Cx)

subunits consists of a gap junctional channel, called a connexon, and there are at least 21 Cx isoforms identified in vertebrates (Goodenough & Paul, 2009). Even though some types of Cx isoforms are widely present in various tissues, the expression of other Cx isoforms is relatively restricted in certain cells (Meşe et al., 2007). In addition, the co-expression of multiple Cx isoforms within a single cell is frequently observed, including testis and epididymis of the male reproductive tract (Meşe et al., 2007).

The excurrent duct of male reproductive tract inclu-

Manuscript received September 21, 2017, Received in revised form September 24, 2016, Accepted September 26, 2017

† Corresponding Author : Ki-Ho Lee, Ph.D. Department of Biochemistry and Molecular Biology and Medical Sciences Research Institute, Eulji University, Daejeon 301-746, S. Korea. Tel: +82-42-259-1643, Fax: +82-42-259-1649, E-mail: kiholee@eulji.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

des the efferent ductules, epididymis, and vas deferens. The epididymis is the site in which spermatozoa released from the testis become mature and acquire fertilizing capacity (Robaire & Hermo, 1988). Histologically, the epididymis is a tubular structure having a lumen inside surrounded by a layer of epithelial cells (Robaire & Hermo, 1988). The epithelial cell layer is consisted of several types of cells, including principal, basal, narrow, and apical cells which have special morphological features and functions distinguishable from each other (Robaire & Hermo, 1988). In addition, the proportion of cell types present in the epididymal epithelial layer varies along the epididymal duct (Robaire & Hermo, 1988). Based on its structure and functions, the epididymis is divided into four different parts, initial segment, caput epididymis, corpus epididymis, and caudal epididymis (Robaire & Hermo, 1988). For example, the epithelium of caput epididymis is thicker than that of caudal epididymis (Robaire & Hermo, 1988). Also, the caput and corpus epididymis are histochemically specialized for luminal fluid absorption and secretion involved in sperm maturation, while the caudal epididymis is adapted for storage of sperm (Robaire & Hermo, 1988). Thus, it is generally considered that different compositions of epithelial cells would contribute to the regional difference of the epididymis, at least in part (Robaire et al., 2006).

The expression and presence of *Cx* isoforms in the epididymis during postnatal development have been examined by several previous researches (Cyr et al., 1996; Dufresne et al., 2003; Han & Lee, 2013; Lee, 2013). It is generally agreed that the expression of *Cx* isoforms in the epididymis follows the age-specific and segmental-specific manners (Dufresne et al., 2003; Han & Lee, 2013; Lee, 2013). For example, the highest expression of *Cx26* in the caput epididymis is found at 5 months of age (Han & Lee, 2013), while the expression of *Cx26* in the corpus epididymis becomes increased until 25 days of age, followed by a gradual decrease of *Cx26* expression as aged (Lee, 2013). On the other hand, the expression of *Cx26* in the caudal epididymis

during postnatal development increases as aged and reaches the highest level at 2 years of age (Lee, 2013). However, even though expressional patterns of *Cx* isoforms in the epididymis are relatively well-documented, the cellular localization and expressional regulation of *Cx* isoforms in the epididymis have not been studied in detail. A significant decrease of *Cx43* expression in the epididymis, except the caudal epididymal part, has been observed by propylthiouracil-induced hypothyroidism at the neonatal age (St-Pierre et al., 2003). In addition, our previous researches have shown that expression of *Cx* isoforms in the corpus and caudal epididymis at the adult is modulated by exogenous treatments of estradiol benzoate (EB), a potent estrogen agonist, or flutamide (Flu), an antiandrogenic compound, at the neonatal age (Lee, 2015; Lee, 2016). These observations suggest that expression of *Cx* isoforms in the epididymis is regulated by intrinsic and/or exogenous compounds in a complicated manner. Yet, the effect of EB or Flu treatment on the expression of *Cx* isoforms in the caput epididymis has not been tested. Thus, in the present study, the expressional changes of *Cx* isoforms in the adult caput epididymis after the neonatal exposure to EB or Flu were examined at the transcript level.

MATERIALS AND METHODS

1. Experimental Animals and Design

Male pup rats were obtained from the delivery of pregnant Sprague Dawley rats acquired from Samtako (Osan, Korea). Once the arrival, each pregnant rat was individually separated and randomly designated to one of experimental groups, a control group (peanut oil), a low-dose estradiol benzoate treatment group [EB-L, 0.015 μg of EB/kg body weight (BW)], a high-dose EB treatment group (EB-H, 1.5 μg of EB/kg BW), a low-dose flutamide treatment group (Flu-L, 500 μg of Flu/kg BW), or a high-dose Flu treatment group (Flu-H, 5 mg of Flu/kg BW). Free access to drinking water and food for animals were permitted for whole

experimental period.

A subcutaneous injection of peanut oil for control group or pre-made EB or Flu solution for treatment groups was performed to an experimental animal at 1 week of age. The EB or Flu powder purchased from Tokyo Chemical Industry Co. (Tokyo, Japan) was dissolved in 100% EtOH, and the EB or Flu solution was then diluted in peanut oil to make working solution. An adequate amount of EB or Flu solution to be administered was calculated from body weight measured at 1 week of age. The maximum volume of the injected solution per animal was not over 0.05 mL. The number of animals utilized for the present study is as following; control group (n=6), EB-L group (n=6), EB-H group (n=6), Flu-L group (n=6), and Flu-H group (n=6). The present study was carried out in accordance with the guide for the care and use of laboratory animals of National Research Council

in S. Korea.

2. Procedures of Tissue Collection and Total RNA Extraction

Once the animal became 4 months of postnatal age, anesthetization by CO₂ stunning was performed. The male reproductive tract was taken out from an incision on lower abdomen and was placed on the cold PBS solution. The epididymis was first separated from the testis and other reproductive tract and was further dissected into different epididymal segments, including caput epididymis. The tissue was briefly washed in fresh cold PBS solution once and was quickly frozen in liquid nitrogen. The tissue was kept in -80°C until utilized for total RNA isolation.

To extract total RNA from the caput epididymis, the frozen tissue was completely homogenized in total RNA

Table 1. Oligonucleotide primers for real-time PCR analysis

Gene (GenBank ID)	Primer sequence (5' → 3')	T _m (°C)	PCR product size (bps)
Cx26 (NM_001004099)	(F) TCCTCTTCATCTTCCGCATC (R) CCGTTTCTTTTCGTGTCTCC	55	233
Cx30.3 (NM_053984)	(F) CCCAATGTCTGCTATGACGA (R) CACAGCAGCCTTGAAGATGA	57	243
Cx31 (NM_019240)	(F) TTGAGCGGTGTGAACCAGTA (R) TGTTGGAGATGGGGAAGAAG	57	193
Cx31.1 (NM_019241)	(F) CATCGTCTGCATCCTGCTTA (R) ATGAGGTCGCTTGAGAGGAA	55	165
Cx32 (NM_017251)	(F) AGAATCATGGTGCTGGTGGT (R) CCTCAAGCCGTAGCATTTTC	57	235
Cx37 (NM_021654)	(F) AGTGTCTGTACCTTGGATGCC (R) CAGCACACTTAGCCAAGAGC	51	223
Cx40 (NM_019280)	(F) ATACCATTACAGCCTGGTTGC (R) CGGCCTCTTTAGCTTTCTCA	57	189
Cx43 (NM_012567)	(F) AGCAAGCTAGCGAGCAAAC (R) GAGTTCATGTCCAGCAGCAA	55	151
Cx45 (NM_001085381)	(F) GATCATCCTGGTTGCTACTC (R) GATCCTCTTCATGGTCCTCT	51	173
<i>Ppia</i> (NM_07101.1)	(F) GGCAAATGCTGGACCAAACAC (R) TTAGAGTTGTCCACAGTCGGAGATG	59	196

Cx : connexin; *Ppia* : peptidylprolyl isomerase A (cyclophilin A).

extraction solution (iNtRON Biotech, Sungnam, Korea). Total RNA was precipitated by isopropanol, and total RNA pellet was washed with 70% DEPC-EtOH. Then, air-dried total RNA was dissolved in DEPC- dH₂O. The concentration of total RNA was assessed with NanoDrop Lite spectrophotometer (Thermo Scientific, Wilmington, DE). The quality of total RNA was evaluated by conventional agarose gel electrophoresis.

3. Construction of cDNA from Total RNA and Performance of Quantitative Real-time PCR

The first-stranded cDNA was generated with 1 µg of total RNA, oligo-dT primer, and ImProm-IITM reverse transcription system (Promega, Madison, WI) in a total volume of 20 µL. The reverse transcription reaction was carried out in a serial process of 25°C for 5 min, 42°C for 90 min, and 70°C for 15 min. The cDNA construction was used for quantitative real-time PCR analysis as below. Oligonucleotide primers utilized for quantitative real-time PCR analysis are listed in Table 1.

One microliter of cDNA, in addition to 10 pmol primer set, 10 µL PCR master mixture (Finnzymes, Espoo, Finland), and DNase-free dH₂O, was used to conduct the quantitative real-time PCR. The mixture of PCR in a total volume of 20 µL was placed in a thermocycler (Bio-Rad Laboratories, Hercules, CA) and was undergone a sequential procedure as followings; a pre-denaturation step at 95°C for 30 sec, 40 cycles of denaturation at 95°C for 30 sec, annealing at T_m for 30 sec, and extension at 72°C for 30 sec, and an additional extension step at 72°C for 5 min. For a quality control purpose of PCR, cyclophilin A (*Ppia*) was included in every reaction.

4. Statistical Analysis and Presentation of Data

For each experimental group, independently quadruplicated RT reactions and PCRs were performed to obtain a mean and a standard error. The experimental data were

presented in the relative expressional ratio between *Ppia* and *Cx* isoform. Statistical significances of *Cx* expression between control and EB or Flu treatment groups were evaluated by one-way ANOVA, followed by Duncan's test. If *P* is less than 0.05, it was regarded as the existence of statistical difference among experimental groups.

RESULTS

1. Expression of *Cx26* and *Cx30.3* in the Adult Caput Epididymis of Rat Exposed to EB or Flutamide at 1 Week of Postnatal Age

The treatment of EB at 1 week of age didn't influence on gene expression of *Cx26* in the adult caput epididymis (Fig. 1A). However, a significant increase of *Cx26* transcript level was observed with a low-dose Flu treatment, even though expression of *Cx26* was not significantly changed by a high-dose Flu treatment (Fig. 1A).

Expression of *Cx30.3* was significantly increased by a low-dose EB treatment, while a reduction of *Cx30.3* transcript level was detected by a high-dose EB treatment (Fig. 1B). The treatment of a low-dose Flu at 1 week of age didn't affect the expression of *Cx30.3* in the adult caput epididymis (Fig. 1B). However, a tremendous increase of *Cx30.3* transcript amount in the caput epididymis at the adult was observed with a high-dose Flu treatment at 1 week of age (Fig. 1B).

2. Expression of *Cx31* and *Cx31.1* in the Adult Caput Epididymis of Rat Exposed to EB or Flutamide at 1 Week of Postnatal Age

Expression of *Cx31* in the adult caput epididymis was significantly decreased by a low-dose EB treatment (Fig. 2A). An additional reduction of *Cx31* transcript level was observed in a high-dose EB-treated group, as shown in Fig. 2A. There was no change of *Cx31* transcript amount by a low-dose Flu treatment, but a significant increase of *Cx31* expression was detected in a high-dose Flu-treated group (Fig. 2A).

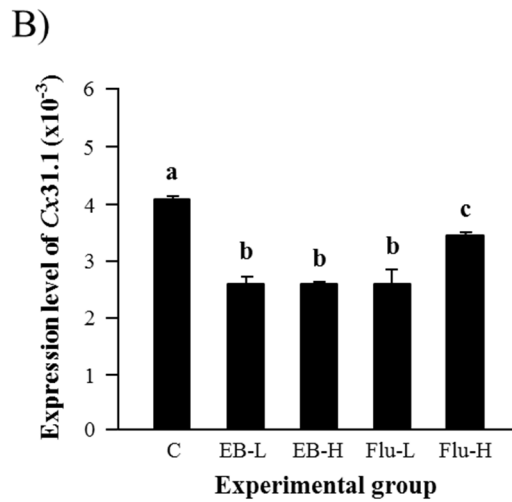
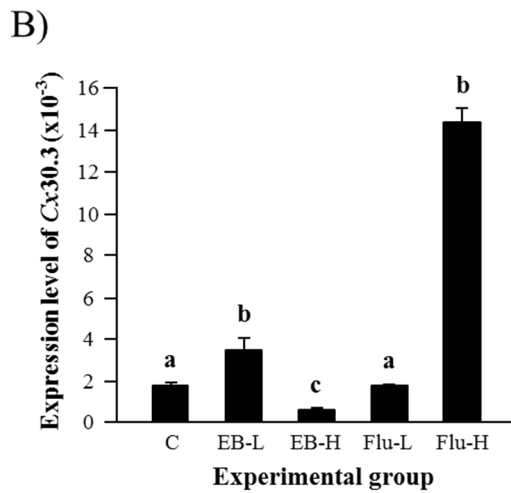
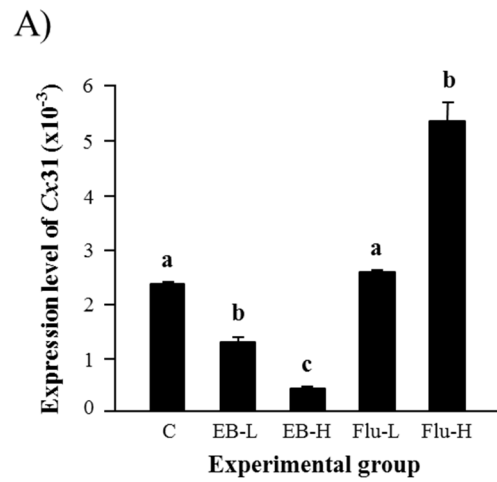
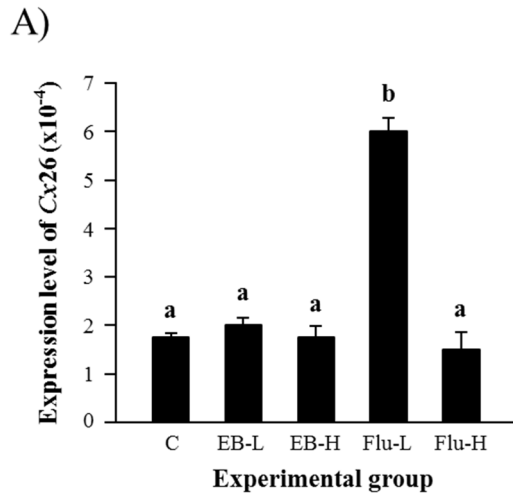


Fig. 1. Changes of Cx26 and Cx30.3 transcript levels in the caput epididymis of the adult rat exposed to estradiol benzoate (EB) or flutamide (Flu) at 1 week of postnatal age. Subcutaneous injection of EB at a low dose (0.015 mg/kg body weight, EB-L) or a high dose (1.5 mg/kg body weight, EB-H) or Flu at a low dose (500 mg/kg body weight, Flu-L) or at a high dose (50 mg/kg body weight, Flu-H) was carried out at 1 week of age. Peanut oil was administered into control group (Cont). Statistical differences among control and experimental groups in EB or Flu treatment group on transcript levels of Cx26 (A) or Cx30.3 (B) were present in different letters.

Fig. 2. Changes of Cx31 and Cx31.1 transcript levels in the caput epididymis of the adult rat exposed to estradiol benzoate (EB) or flutamide (Flu) at 1 week of postnatal age. Subcutaneous injection of EB at a low dose (0.015 mg/kg body weight, EB-L) or a high dose (1.5 mg/kg body weight, EB-H) or Flu at a low dose (500 mg/kg body weight, Flu-L) or at a high dose (50 mg/kg body weight, Flu-H) was carried out at 1 week of age. Peanut oil was administered into control group (Cont). Statistical differences among control and experimental groups in EB or Flu treatment group on transcript levels of Cx31 (A) or Cx31.1 (B) were present in different letters.

The expression of Cx31.1 isoform in the adult caput epididymis was declined in all treatment groups (Fig. 2B). The

EB treatment resulted in a significant decrease of Cx31.1 transcript level, even though there was no significant

difference on *Cx31.1* expression between two doses (Fig. 2B). The Flu treatment also caused significant reduction of *Cx31.1* transcript level, but the effect of Flu treatment on *Cx31.1* expression was more severe with a low-dose treatment than a high-dose treatment (Fig. 2B).

3. Expression of *Cx32* and *Cx37* in the Adult Caput Epididymis of Rat Exposed to EB Or Flutamide at 1 Week of Postnatal Age

The expression of *Cx32* in the adult caput epididymis after the exposure to EB at 1 week of age was significantly decreased by a low-dose treatment, but not by a high-dose treatment (Fig. 3A). On the other hand, the treatment of a low-dose Flu didn't affect the expression of *Cx32*, but a significant decline of *Cx32* transcript level was detected with a high-dose Flu treatment (Fig. 3A).

The effect of EB or Flu treatment at 1 week of age on the expression of *Cx37* in the adult caput epididymis is shown in Fig. 3B. The treatment of a low-dose EB resulted in a significant increase of *Cx37* expression, but a high-dose EB treatment caused a decrease of *Cx37* transcript level (Fig. 3B). Exposure to a low-dose Flu at 1 week of age led into a major increase of *Cx37* expression in the adult caput epididymis, and a further rise of *Cx37* transcript level was induced by a high-dose Flu treatment (Fig. 3B).

4. Expression of *Cx40*, *Cx43*, and *Cx45* in The Adult Caput Epididymis of Rat Exposed to EB or Flutamide at 1 Week of Postnatal Age

Expression of *Cx40* in the adult caput epididymis was significantly down-regulated by the treatment of EB at 1 week of age, even though the level of *Cx40* transcript was not statistically different between a low-dose and a high-dose EB treatment (Fig. 4A). However, the Flu treatment at a low-dose resulted in a significant increase of *Cx40* transcript levels (Fig. 4A). The treatment of a high-dose Flu induced an additional surge of *Cx40* expressional level, as shown in Fig 4A.

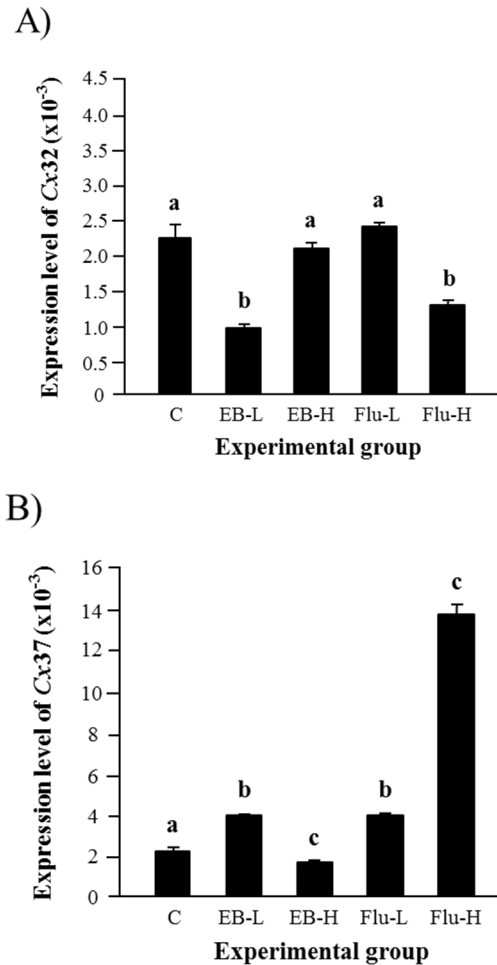


Fig. 3. Changes of *Cx32* and *Cx37* transcript levels in the caput epididymis of the adult rat exposed to estradiol benzoate (EB) or flutamide (Flu) at 1 week of postnatal age. Subcutaneous injection of EB at a low dose (0.015 mg/kg body weight, EB-L) or a high dose (1.5 mg/kg body weight, EB-H) or Flu at a low dose (500 mg/kg body weight, Flu-L) or at a high dose (50 mg/kg body weight, Flu-H) was carried out at 1 week of age. Peanut oil was administered into control group (Cont). Statistical differences among control and experimental groups in EB or Flu treatment group on transcript levels of *Cx32* (A) or *Cx37* (B) were present in different letters.

Expression of *Cx43* in the caput epididymis was not affected by a low-dose EB treatment (Fig. 4B). But, a significant decrease of *Cx43* transcript level was observed with a high-dose EB treatment (Fig. 4B). The treatment of Flu

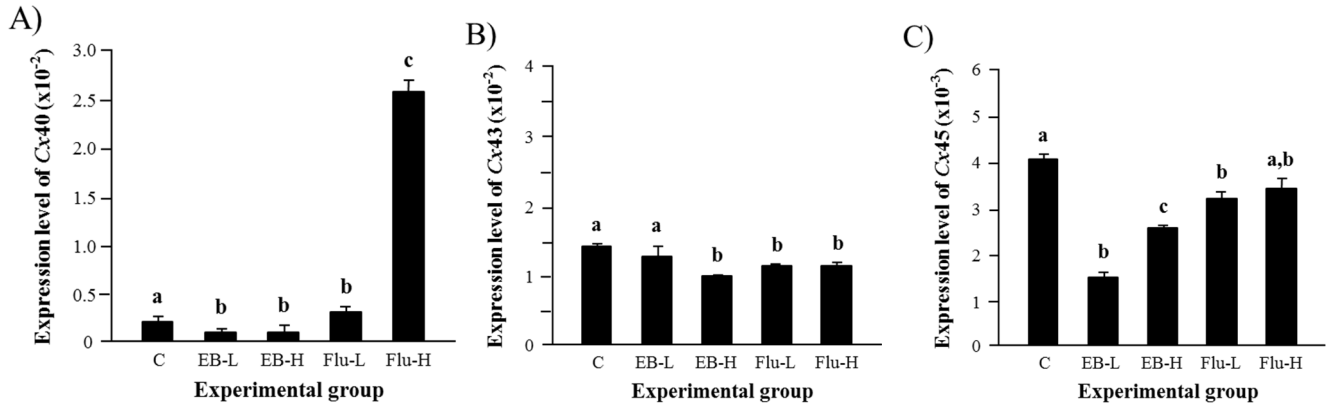


Fig. 4. Changes of Cx40, Cx43, and Cx45 transcript levels in the caput epididymis of the adult rat exposed to estradiol benzoate (EB) or flutamide (Flu) at 1 week of postnatal age. Subcutaneous injection of EB at a low dose (0.015 mg/kg body weight, EB-L) or a high dose (1.5 mg/kg body weight, EB-H) or Flu at a low dose (500 mg/kg body weight, Flu-L) or at a high dose (50 mg/kg body weight, Flu-H) was carried out at 1 week of age. Peanut oil was administrated into control group (Cont). Statistical differences among control and experimental groups in EB or Flu treatment group on transcript levels of Cx40 (A), Cx43 (B), or Cx45 (C) were present in different letters.

caused the reduction of Cx43 transcript level, although there was no significant difference on the expression of Cx43 between the treatments at a low-dose and a high-dose Flu (Fig. 4B).

The transcript level of Cx45 was significantly decreased by a low-dose EB treatment, as seen in Fig. 4C. The treatment of a high-dose EB also caused the reduction of Cx45 transcript level (Fig. 4C). However, the abundance of Cx45 transcript in a high-dose EB treatment group was higher than that in a low-dose EB treatment group (Fig. 4C). A low-dose Flu treatment led into a significant decrease of Cx45 expression in the adult caput epididymis, even though a high-dose Flu treatment didn't cause a change of Cx45 transcript level (Fig. 4C).

DISCUSSION

Expressional alternation of Cx isoforms in the adult corpus and caudal epididymis by EB or Flu treatment at neonatal age has been previously examined (Lee, 2015; Lee, 2016). The current research has investigated the effect of EB or Flu treatment at 1 week of postnatal age on the expression

of Cx isoforms in the caput epididymis at the adult. The findings are summarized as follows: (1) the exposure to EB at a low dose results in significant decreases of the expression of most Cx isoforms in the caput epididymis, except Cx30.3 and Cx37 which the expression has become increased and Cx26 and Cx43 with no significant expressional changes: (2) the treatment of EB at a high dose at 1 week of postnatal age generally causes significant decreases of most Cx isoforms in the caput epididymis at the adult, but not Cx26 and Cx32 showing no significant change of the expressional level: (3) the effect of a low-dose Flu treatment leads to increases of Cx37 and Cx40 expression, decreases of Cx31.1, Cx43, and Cx45 expression, and no expressional change of Cx26, Cx30.3, Cx31, and Cx32: and (4) the administration of a high-dose Flu at 1 week of age results in significant expressional increases of Cx30.3, Cx31, Cx37, and Cx40 and significant expressional decreases of Cx31.1, Cx32, and Cx43 in the adult caput epididymis.

It is well known that the epididymis is a representative androgen-responsive tissue in the male reproductive tract (Robaire & Hinton, 2006). Testosterone and dihydrotestosterone play important roles on the proper differentiation

and maturation of the epididymis (Robaire & Hinton, 2006). However, Atanassova et al (2001) have demonstrated that expression of estrogen receptor α (ER α) in the epididymis is segmental-specific and postnatal age-specific. Also, others have shown the aberrant gene expression in the epididymis of the adult after the exposure to EB during neonatal period (Putz et al., 2001). Moreover, Growska et al (2014) have found aberrant gene expression in the epididymis exogenously treated with steroidal and/or anti-steroidal compounds. Thus, these findings suggest that the epididymis could be a target tissue of not only androgen but also estrogen. Indeed, our previous researches have also shown abnormal expression of Cx isoforms in the adult corpus and caudal epididymis exposed with EB or Flu at the neonatal age (Lee, 2015; Lee, 2016). However, it seems that expressional regulation of Cx isoforms in different epididymal parts does not fit into a uniformed manner. For instance, with a low-dose EB treatment at 1 week of age, there is no expressional change of Cx43 in the caput epididymis of the adult. But, in the same circumstance, expression of Cx43 is greatly increased in the corpus epididymis, but not in caudal epididymis (Lee, 2015; Lee, 2016). In addition, even though the treatment of a low-dose Flu at 1 week of age causes a significant increase of Cx32 expression in the adult caput epididymis, the same treatment does not give an influence on the expression of Cx32 in adult corpus and caudal epididymis (Lee, 2015; Lee, 2016). Due to the complexity of data obtained, it is hard to conclude the role of estrogen and/or androgen on the expression of each Cx isoform in the epididymis at this point. But, it seems to be clear that exogenous exposure to EB or Flu at the early postnatal age affects the expression of Cx isoforms in the entire epididymal segments. Also, it is likely that the expressional regulation of Cx isoform by androgenic and/or estrogenic substances is segmental-specific in the epididymis. Like promoter assay and gene expression analysis, detailed molecular approaches would provide valuable information to explain differential ex-

pressional regulation of each Cx isoform at the different epididymal parts.

The expression of androgen receptor (AR) and ER in the epididymis during postnatal development is fluctuated and shows segmental-specific patterns (Atanassova et al., 2001; You & Sar, 1998). Additionally, until 14 days of postnatal age, the differentiation of epithelial cells in the epididymis does not begin (Robaire et al., 2006). At the 3th week of postnatal age, columnar cells are derived from undifferentiated epithelial cells, and the columnar cell becomes either the basal or principal cell around 4th week of postnatal age (Robaire et al., 2006). The appearance of other cell types, including narrow and clear cells, start after 35 days of postnatal age (Robaire et al., 2006). Because the postnatal maturation of the epididymis is dependent on androgenic and/or other various hormonal factors, any elements affecting hormonal homeostasis during the early postnatal period could give an influence on the differentiation of the epithelial cell in the epididymis and furthermore numbers of epithelial cell types in the adult epididymis. Cyr et al (1996) have demonstrated the limited localization of Cx43 between basal and principal cells, but not between adjacent principal cells. Thus, if EB or Flu treatment at 1 week of postnatal age influences on the differentiation of the epithelial cells in the epididymis, it is supposed that the proportion of epithelial cell types in the caput epididymis at the adult would be changed and thus the expression of Cx isoforms, including Cx43, would be altered.

In conclusion, based on previous and present findings, the expression of Cx isoforms in the epididymis is differentially regulated in segment-specific manner. Even though a precise regulatory mechanism of expression of Cx isoforms in the epididymis has not been provided, it is evident that the exposure to estrogenic and/or anti-androgenic substance during the early postnatal development could impact on the expression of normal expression of Cx isoforms in the adult epididymis and presumably epididymal functions at the adult.

ACKNOWLEDGEMENT

This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (20090067187 and 2017030209).

REFERENCES

- Atanassova N, McKinnell C, Williams K, Turner KJ, Fisher JS, Saunders PT, Millar MR, Sharpe RM (2001) Age-, cell- and region-specific immunoexpression of estrogen receptor alpha (but not estrogen receptor beta) during postnatal development of the epididymis and vas deferens of the rat and disruption of this pattern by neonatal treatment with diethylstilbestrol. *Endocrinology* 142: 874-886.
- Cyr DG, Hermo L, Laird DW (1996) Immunocytochemical localization and regulation of connexin43 in the adult rat epididymis. *Endocrinology* 137:1474-1484
- Dufresne J, Finnson KW, Gregory M, Cyr DG (2003) Expression of multiple connexins in the rat epididymis indicates a complex regulation of gap junctional communication. *Am J Physiol Cell Physiol* 284:C33-43.
- Goodenough DA. and Paul DL (2009) Gap junctions. *Cold Spring Harb. Perspect. Biol* 1:a003061.
- Gorowska E, Zarzycka M, Chojnacka K, Bilinska B, Hejmej A (2014) Postnatal exposure to flutamide affects CDH1 and CTNNB1 gene expression in adult pig epididymis and prostate and alters metabolism of testosterone. *Andrology* 2:186-197.
- Han S-Y, Lee K-H (2013) The expression patterns of connexin isoforms in the rat caput epididymis during postnatal development. *J Ani Sci Tech* 55: 249-255.
- Lawrence TS, Beers WH, Gilula NB (1978) Transmission of hormonal stimulation by cell-to-cell communication. *Nature* 272: 501-506.
- Lee K-H (2013) Differential expression of multiple connexins in rat corpus and cauda epididymis at various postnatal stages. *J Ani Sci Tech* 55:521-530.
- Lee K-H (2015) Modification of gene expression of connexins in the rat corpus epididymis by estradiol benzoate or flutamide exposure at the early neonatal age. *Dev Reprod* 19:69-77.
- Lee K-H (2016) Changes in expression of connexin isoforms in the caudal epididymis of adult Sprague-Dawley rats exposed to estradiol benzoate or flutamide at the neonatal Age. *Dev Reprod* 20:237-245.
- Meşe G, Richard G, White TW (2007) Gap junctions: Basic structure and function. *J Invest Dermatol* 127:2516-2524.
- Putz O, Schwartz CB, LeBlanc GA, Cooper RL, Prins GS (2001) Neonatal low- and high-dose exposure to estradiol benzoate in the male rat: II. Effects on male puberty and the reproductive tract. *Biol Reprod* 65:1506-1517.
- Robaire B, Hermo L (1988) Efferent ducts, epididymis, and vas deferens: structure, functions, and their regulation. In: Knobil E et al (eds.). *The Physiology of Reproduction*. Raven Press, New York, NY, pp 999-1080.
- Robaire B, Hinton BT, Orgebin-Crist MC (2006) The epididymis. In: Knobil E and Neil J (eds.) *The Physiology of Reproduction*, Elsevier, New York, NY pp 1071-1148.
- St-Pierre N, Dufresne J, Rooney AA, Cyr DG (2003) Neonatal hypothyroidism alters the localization of gap junctional protein connexin 43 in the testis and messenger RNA levels in the epididymis of the rat. *Biol Reprod* 68:1232-1240.
- Valiunas V, Polosina YY, Miller H, Potapova IA, Valiuniene L, Doronin S, Mathias RT, Robinson RB, Rosen MR, Cohen IS, Brink PR (2005) Connexin-specific cell-to-cell transfer of short interfering RNA by gap junctions. *J Physiol* 568:459-468.
- You L, Sar M (1998) Androgen receptor expression in the testes and epididymides of prenatal and postnatal Sprague-Dawley rats. *Endocrine* 9:253-261.