# Note

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# Physiological effects of copper on the freshwater alga *Closterium ehrenbergii* Meneghini (Conjugatophyceae) and its potential use in toxicity assessments

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Although green algae of the genus *Closterium* are considered ideal models for testing toxicity in aquatic ecosystems, little data about the effects of toxicity on these algal species is currently available. Here, *Closterium ehrenbergii* was used to assess the acute toxicity of copper (Cu). The median effective concentration ( $EC_{50}$ ) of copper sulfate based on a dose response curve was 0.202 mg L<sup>-1</sup>, and reductions in photosynthetic efficiency ( $F_v/F_m$  ratio) of cells were observed in cultures exposed to Cu for 6 h, with efficiency significantly reduced after 48 h (p < 0.01). In addition, production of reactive oxygen species significantly increased over time (p < 0.01), leading to damage to intracellular organelles. Our results indicate that Cu induces oxidative stress in cellular metabolic processes and causes severe physiological damage within *C. ehrenbergii* cells, and even cell death; moreover, they clearly suggest that *C. ehrenbergii* represents a potentially powerful test model for use in aquatic toxicity assessments.

Key Words: chlorophyll fluorescence; Closterium ehrenbergii; copper; EC<sub>50</sub>; reactive oxygen species

#### INTRODUCTION

Microalgae play key roles in the primary productivity and biochemical cycles of aquatic systems. They are sensitive indicators of environmental change, and thus they have often been used to evaluate the impacts of metals, herbicides, and other pollutants in freshwater ecosystems (Qian et al. 2008); their physiology may be affected even under conditions of "no observable effect concentration" of a pollutant. In addition, the use of algae in toxicity assays confers numerous advantages: they are, for instance, easy to culture, requiring simple inorganic culture media, and they exhibit a rapid rate of growth and generational turnover (Lewis 1995).

Several physiological parameters are used as endpoints in algae-based ecotoxicological assessments,

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such as cell growth rate, biovolume, antioxidant system response, pigment production, and photosynthetic rate, which are compared and contrasted between treated and non-treated cells. Cell number is typically used as an indicator of growth in toxicological studies (Franklin et al. 2002), and many environmental contaminants affect the size and morphology of the tested cells via induction of oxidative stress (Sabatini et al. 2009). Chlorophyll autofluorescence (CAF) is an effective method for analyzing *in situ* photosynthetic efficiency, as well as to measure photosynthetic response to various stresses (Schreiber et al. 1995). Previous studies have reported the inhibition of microalgal photosynthesis by a wide variety of contaminants, leading to reductions in chlorophyll *a* (Chl *a*) con-

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centrations (Ferrat et al. 2003), but little is known about the characteristics of CAF. Moreover, studies have shown that biochemical parameters play important roles in antioxidant defense systems in algal cells, which are estimated via antioxidant enzymatic assays (i.e., catalase, peroxidase, and superoxide dismutase) (Manimaran et al. 2012), but direct information about the formation of reactive oxygen species (ROS) in microalgae is scarce.

The unicellular green alga Closterium ehrenbergii is found in freshwater ecosystems worldwide (Ichimura and Kasai 1984). This species is crescent-shaped (long, curved, and tapered at both ends) and is typically much larger (~300 µm in length) than most other unicellular microalgae (Lee et al. 2015), making observation of morphological and cellular changes relatively easy. It has long been used in research on algal sexual reproduction and cell cycling (e.g., Fukumoto et al. 1997), due to these features. In addition, previous studies have shown that C. ehrenbergii is very sensitive to a wide range of surfactants (Kim et al. 1998), and thus, recently it has been employed in additional toxicity evaluations and bioassays to detect the deleterious effects of hazardous substances on aquatic systems (Juneau et al. 2003, Sathasivam et al. 2016). However, to the best of our knowledge, there is little toxicological data available regarding the effects of various pollutants on Closterium species, and sensitivity of the species has not yet been compared to that of other microalgae (e.g., Aulacoseira granulate, Chlorella vulgaris, and Chlamydomonas sp.) by employing typical environmental contaminants, like heavy metals and pesticides.

In the present study, we quantified the sub-lethal effects of exposure to a well-known pollutant (copper, Cu) on a variety of morphological and physiological parameters of a strain of the freshwater green alga *C. ehrenbergii*, focusing on chlorophyll fluorescence characteristics, photosynthetic efficiency, and ROS levels. We then investigated the potential of Korean *C. ehrenbergii* isolates as testing models for aquatic toxicity assessments via comparisons of the median effective concentrations (EC<sub>50</sub>) of the test chemical copper sulfate (CuSO<sub>4</sub>), which is widely used as a biocide for cleaning swimming pools, in aquaculture farms, and even for removing harmful algal blooms (Kim et al. 2007), despite evidence of contamination of aquatic ecosystems and possibly even drinking water.

#### **MATERIALS AND METHODS**

# Alga culture and median effective concentration $(EC_{50})$

Cells of Korean *C. ehrenbergii* (Ce-01; Environmental Bio Inc., Seoul, Korea) were cultured in C medium (Watanabe et al. 2000) under conditions of  $20 \pm 1^{\circ}$ C ambient temperature, a 12 : 12 h light-dark cycle, and a photon flux density of ~65 µmol photons m<sup>-2</sup> s<sup>-1</sup>.

The effects of Cu toxicity on *C. ehrenbergii* was assessed using  $CuSO_4$  at concentrations of 0.1, 0.5, 1.0, 2.5, and 5.0 mg L<sup>-1</sup>.  $CuSO_4$  was procured from a commercial source (cat. No. C1297; Sigma, St. Louis, MO, USA) and prepared for standard stock solutions; all dilutions were prepared from standard stock solutions and all exposures were repeated in triplicate. The percentile inhibition and 72 h median effective concentration (EC<sub>50</sub>-72h) were calculated following the Organisation for Economic Cooperation and Development testing guidelines (OECD 2006), with EC<sub>50</sub>-72h estimates derived from a sigmoidal dose–response curve and plotted using Origin v. 8.5 software (MicroCal Software Inc., Northampton, MA, USA).

#### Measurements of chlorophyll fluorescence

Photosynthetic efficiency  $(F_v / F_m)$  was measured using a Handy Plant Efficiency Analyser fluorometer (Hansatech Instruments Ltd., Norfolk, UK). To determine the photosynthetic parameters of *C. ehrenbergii*, 2 mL of each sample was collected using a specimen cup. The fluorescence parameters, minimal fluorescence in the darkadapted state  $(F_o)$ , maximal fluorescence in the darkadapted state  $(F_m)$ , and variable fluorescence  $(F_v; F_m - F_o)$ , were measured at 0, 6, 24, and 48 h following exposure to Cu concentrations of 0.1, 0.2, 0.4, and 1.0 mg L<sup>-1</sup>. Values of  $F_v$  and maximal quantum efficiency of photosystem II (PSII) photochemistry  $(F_v / F_m)$  were then derived from  $F_o$ and  $F_m$ .

#### **ROS** measurement

Dihydroxyrhodamine 123 (DHR123-D1054; Sigma) staining was used to measure ROS production, as DHR123 emits a green fluorescence when oxidized by ROS (Qin et al. 2008). Cells were independently treated with 0.2 mg  $L^{-1}$  Cu, with incubation periods of 6, 24, and 48 h. *C. ehrenbergii* cells were stained via exposure to DHR123 at a final concentration of 20 µM for 1 h, then harvested by centrifugation and twice rinsed with fresh C medium.

The cultures were then re-suspended in fresh C medium, and mounted onto a slide and sealed; culture slides were observed with a fluorescence microscope (Carl Zeiss Axioskop, Oberkochen, Germany) to determine the level of ROS production in the cells. ImageJ software (NIH, Bethesda, MD, USA) was used to quantify relative CAF and ROS levels from the fluorescent microscopic images.

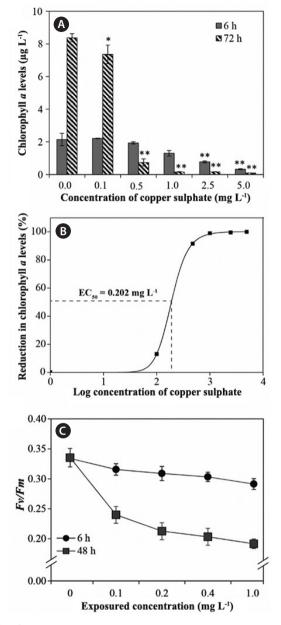
#### Statistical analysis

All data presented are mean values of triplicates. Oneway analysis of variance (ANOVA) with *post hoc* Student's Newmann-Keuls test using Graphpad InStat (Graphpad Software, Inc., San Diego, CA, USA) was used for comparisons between treated and untreated cultures, and p < 0.05 was used to determine significant differences between means.

#### **RESULTS AND DISCUSSION**

Chlorophyll pigments have often been used as indicators in the monitoring of environmental stressors of plants, including algae (Li et al. 2005). Exposure of Cu to C. ehrenbergii induced a wide range of responses, depending on the Cu concentration; whereas exposure of Cu for 6 h at the initial experimental concentrations (i.e., 0.1-1.0 mg L<sup>-1</sup>) had only slight effects on Chl *a* levels (Fig. 1A), and Chl a was significantly lower in cells exposed to relatively high concentrations of Cu (2.5 and 5.0 mg L<sup>-1</sup>) (p < 0.05). In addition, Chl a levels were markedly lower after exposed to Cu for 72 h, with reductions of 12.1, 91.3, 98.2, 98.1, and 98.9% at 0.1, 0.5, 1.0, 2.5, and 5.0 mg L<sup>-1</sup> of Cu concentrations, respectively. Similar results were reported by Chen et al. (2012), who found that Chl a concentrations significantly decreased in green algae exposed to Cu, a trend that was associated with increasing Cu concentrations (from 2 to 10 µM). Likewise, exposure of Cu resulted in marked reductions in Chl a concentration in the dinoflagellate Cochlodinium polykrikoides, an effect that was also more pronounced at higher Cu concentrations (Ebenezer et al. 2014). Overall, these results clearly demonstrate that the Korean strain of C. ehrenbergii used in this study is considerably more sensitive to Cu than other microalgae that have been tested previously (Cairns et al. 1978, Viana and Rocha 2005, Ebenezer and Ki 2013, Ebenezer et al. 2014).

Estimations of  $EC_{50}$  values are useful for identifying environmental contaminants that may inhibit algal growth (Antón et al. 1993). The  $EC_{50}$  of Korean *C. ehren*-



**Fig. 1.** Effect of different doses of copper to *Closterium ehrenbergii*. (A) Variation in chlorophyll *a* (Chl *a*) levels after 6 and 72 h exposure. (B) A dose response curve after 72 h exposure. (C) Variation in photosynthetic efficiency ( $F_v / F_m$  ratio) after 6 and 48 h exposure. Significant differences as determined by Student's Newmann-Keuls test are represented as \*p < 0.05, \*\*p < 0.01 level. Error bars represent ±standard deviation (n = 3).

*bergii* after exposure to Cu for 72 h was calculated 0.202 mg L<sup>-1</sup> (Fig. 1B). Previous studies have reported that  $EC_{50}$  for the green algae *Chlorella ellipsoidea*, *Chlorella vulgaris*, *Chlamydomonas* sp., and *Tetraselmis suecica* were 0.489, 0.200, 8.00, and 43.03 mg L<sup>-1</sup>, respectively (Young and Lisk 1972, Cairns et al. 1978, Ebenezer and Ki 2013,

Magdaleno et al. 2013). In addition,  $EC_{50}$  for diatoms *Planothidium lanceolatum*, *Ditylum brightwellii*, dinoflagellates *C. polykrikoides*, and *Prorocentrum minimum* after exposure to Cu were reported 0.620, 0.406, 0.633, and 1.07 mg L<sup>-1</sup>, respectively (Sbihi et al. 2012, Ebenezer et al. 2014, Guo et al. 2014, Lee et al. 2014). Comparisons of available  $EC_{50}$  values indicated that species used in this study was generally more sensitive to Cu than other algae (Table 1), suggesting that it is a reliable model organism for aquatic toxicity assessments (Kim et al. 1998, Juneau et al. 2003, Sathasivam et al. 2016).

In addition, most pollutants including heavy metals inhibit PSII activity and interfere with photosynthetic reactions (Giardi et al. 2001). The  $F_v / F_m$  ratio is a useful and easily measurable parameter for the physiological state of the photosynthetic system in intact algae cells (Chen et al. 2012). To access the effects of Cu on the photosynthetic systems of C. ehrenbergii, we measured a range of parameters associated with photosynthetic processes. The fluorescence kinetic curves (both  $F_a$  and  $F_m$ ) generated in the present study gradually decreased with increasing Cu concentrations and exposured time (data not shown).  $F_{\nu}$ /F<sub>m</sub> of C. ehrenbergii was considerably affected by Cu toxicity and also declined with increasing Cu concentration and exposured time (Fig. 1C). Values of  $F_u / F_m$  slightly fell after exposure for 6 h (Fig. 1C) but significantly declined after exposure for 48 h at all concentrations of Cu, indicating that chlorophyll content and / or photosynthetic electron transport were inhibited by exposure to Cu. Similar responses have been observed in other algae exposed to various environmental pollutants, although the extent of inhibition varied among species; for example, Mamboya et al. (1999) reported that the photosynthetic efficiency of

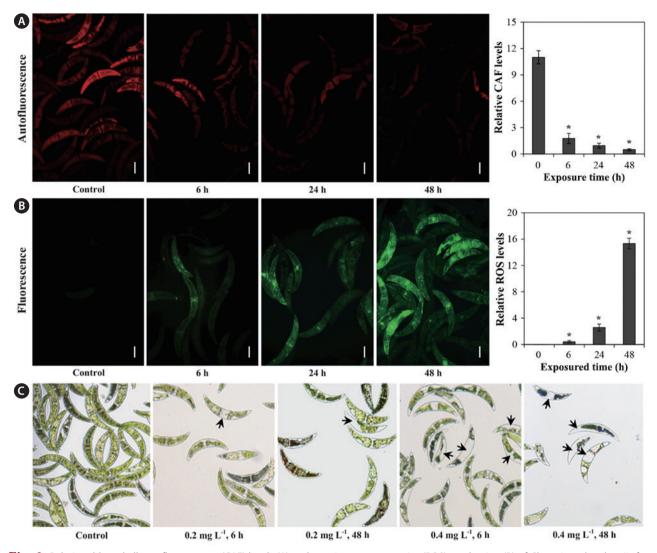
**Table 1.** The median effective concentration (EC<sub>50</sub>) values of *Closterium ehrenbergii* and other microalgae after expose to copper

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EC <sub>50</sub> (mg L <sup>-1</sup> )	Reference
0.202	This study
0.200	Young and Lisk (1972)
0.489	Magdaleno et al. (2013)
8.00	Cairns et al. (1978)
43.03	Ebenezer and Ki (2013)
2.58	Viana and Rocha (2005)
0.406	Lee et al. (2014)
0.620	Sbihi et al. (2012)
0.633	Ebenezer et al. (2014)
1.07	Guo et al. (2014)
	(mg L <sup>-1</sup> ) 0.202 0.200 0.489 8.00 43.03 2.58 0.406 0.620 0.633

the brown alga *Padina boergesenii* significantly declined in response to increasing Cu concentration and exposured time. Moreover, in chlorophytes *Chlorella vulgaris*, *Pseudokirchneriella subcapitata*, dinoflagellates *Cochlodinium polykrikoides*, and *P. minimum* exposed to 3  $\mu$ M, 250  $\mu$ M, 1.0 mg L<sup>-1</sup>, and 0.5 mg L<sup>-1</sup> of Cu, respectively, photosynthetic efficiency significantly declined in response to longer exposured times (Knauert and Knauer 2008, Guo et al. 2016*a*, 2016*b*). Such results not only demonstrate that exposure of Cu has an adverse effect on photosynthetic efficiency in many organisms, but also that the Korean *C. ehrenbergii* is a particularly sensitive to exposure of Cu (see Table 1).

Measurement of CAF is an efficient means of assessing physiological status in microalgae, which are capable of autofluorescence due to the presence of photosynthetic pigments (Trampe et al. 2011), as autofluorescence enables distinction between damaged and undamaged cells (Sato et al. 2004). As can be seen in Fig. 2A, relative CAF levels decreased with exposured time. In addition, ROS production for C. ehrenbergii was determined after exposure to 0.2 mg L<sup>-1</sup> Cu for 6, 24, and 48 h (Fig. 2B); compared to controls, red fluorescence (autofluorescence) significantly decreased, whereas green fluorescence slowly increased with increasing exposured time, suggesting overproduction of ROS after exposure to Cu (Fig. 2A & B). The relative ROS level in C. ehrenbergii cells rose with increasing exposured time to Cu (Fig. 2B). Guo et al. (2014) reported that green fluorescence intensity significantly increased in the dinoflagellate P. minimum exposed to  $CuSO_4$  (p < 0.01), and gradually rose with increasing exposured time. In addition, Ishikawa et al. (1993) demonstrated that H<sub>2</sub>O<sub>2</sub> is generated in chloroplasts and mitochondria, and immediately diffuses from these organelles to the cytosol. Algal cell toxicity can be promoted by the reaction of Cu (II) with H<sub>2</sub>O<sub>2</sub>, which further induces oxidative damage in algae cells (Chen et al. 2012). Moreover, we also observed changes in cell size and morphology, fragmentation of intracellular chloroplast in some algal cells, and loss of some cell contents (i.e., pigments) (Fig. 2C); these observations are consistent with the harmful effects described above.

According to United States Environmental Protection Agency guidelines, the general standard for maximum discharge of Cu into the environment is 3.0 mg L<sup>-1</sup> (U.S. EPA 1986); however, we found that growth rates, Chl *a* levels, and photosynthetic efficiency were reduced, and ROS production enhanced for *C. ehrenbergii* exposed to levels of Cu as low as 1.0 mg L<sup>-1</sup>, with effects intensifying with increasing Cu concentrations. In a previous study,



**Fig. 2.** Relative chlorophyll autofluorescence (CAF) levels (A) and reactive oxygen species (ROS) production (B) of *Closterium ehrenbergii* after expose to 0.2 mg  $L^{-1}$  copper at different intervals (6, 24, and 48 h). (C) Morphological changes of *Closterium ehrenbergii* after 6 and 48 h exposure to 0.2 and 0.4 mg  $L^{-1}$  of copper as seen by light microscope. Arrows represent chloroplast damage or loss of pigment. Significant differences as determined by Student's Newmann-Keuls test are represented as \*p < 0.01 level. Error bars represent ±standard deviation (n = 10). Scale bars represent: A & B, 50 µm.

Sathasivam et al. (2016) demonstrated that growth rates, cell division, pigment levels, and CAF intensity decreased for Korean *C. ehrenbergii* exposed to NaOCl, whereas production of antioxidant enzymes increased, even at relatively lower contaminant concentrations. Moreover, expression of many stress-related genes (i.e., heat shock proteins, superoxide dismutase, glutathione S-transferase) increased for *C. ehrenbergii* exposed to Cu and other environmental pollutants (unpublished data). It is clear that *C. ehrenbergii* is extremely sensitive to a variety of environmental contaminants, and that the discharge limit for Cu concentrations established by the U. S. EPA is not

low enough to avoid damage to the green alga *C. ehrenbergii* and possibly to other microalgae in environments.

## CONCLUSION

The Korean freshwater green alga *C. ehrenbergii* exhibited a dose-dependent response when exposed to a typical pollutant copper. Comparisons of *C. ehrenbergii*'s  $EC_{50}$ values (0.202 mg L<sup>-1</sup>) to those of other organisms clearly demonstrated this species extreme sensitivity to Cu exposure. In addition, exposure to 1.0 mg L<sup>-1</sup> of Cu significantly inhibits production of Chl *a*, reduces photosynthetic efficiency, and induces the generation of intracellular ROS, which may disrupt cell membrane functioning to cell death. General discharge standards of Korea limit Cu concentrations to 3.0 mg L<sup>-1</sup>, and thus this level may be high enough to potentially cause severe harmful to *C. ehrenbergii* and other aquatic organisms. From these results, we can conclude that the Korean *C. ehrenbergii* strain represents a useful model organism for aquatic toxicity assessments, and as such can provide a wealth of information pertinent to general risk assessments.

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