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Effect of dietary supplementation of fermented *Rhus verniciflua* on growth performance, apparent total tract digestibility, blood profile, and fecal microflora in weanling pigs

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Abstract

Fermented *Rhus verniciflua* (FRV) as feed additives act as excellent anti-diarrheal drugs as they increase the intestinal absorption rate therefore being indirectly associated with enhancing growth performance and increasing digestibility in livestock. A total of 80 weaned pigs with an average initial body weight (BW) of 6.82 ± 1.11 kg were used to evaluate a diet supplemented with FRV meal in a 6 week feeding trial with two dietary treatments [CON - basal diet; TRT - CON + 0.2% FRV] on growth performance, apparent total tract digestibility (ATTD), blood profiles, fecal microflora, and fecal score. Our results showed that the dietary supplementation of FRV improved ($p < 0.05$) average daily gain and gain : feed (G : F) ratio during days 15 - 42 and the overall experiment period and also increased the ATTD of dry matter ($p < 0.05$) at days 14 and 42. On the contrary, there was no effect ($p > 0.05$) on average daily feed intake, ATTD of nitrogen and energy, and blood profiles during the entire experiment. Moreover, dietary inclusion of FRV significantly increased fecal *Lactobacillus* ($p < 0.05$) counts and reduced the diarrhea during days 22 - 42. Thus, the results suggest that FRV can be used as a potential additive to improve growth performance and dry matter and to reduce diarrhea while having beneficial effects on fecal microflora in weanling pigs.

Keywords: diarrheal score, dry matter, fermented *Rhus verniciflua*, growth performance, weanling pigs

Introduction

It has been well documented that weaning is the most stressful event in a pig's lifetime due to the sudden shift from high protein, high fat, and high lactose milk to low protein, low fat, and high carbohydrates solid feed (Pluske et al., 1995; Turpin et al., 2016). This shift in the feed from liquid to solid appears to have adverse effects on gastro intestinal tract function. As a consequence, there



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is a substantial increase in pathogenic micro-organisms and a reduction in beneficial micro-organism which eventually contributes to intestinal and immune dysfunctions (Campbell et al., 2013). Due to an increase in the susceptibility to gram-negative bacterial infections (Nabuurs, 1995), weanling pigs are associated with growth retardation as well as an increase in both morbidity and mortality in pigs (Wilson et al., 1989). The ban on the sub-therapeutic use of antibiotics by the European Union in 2006 has driven the attention of researchers on finding alternatives from biological sources (Park et al., 2016a). Among different alternatives, the potential use of fermented phytogetic products for this purpose has been gaining momentum in recent years.

Toxicodendron vernicifluum (formerly *Rhus verniciflua*) a medicinal plant belonging to the anacardiaceous family, has been used as a preventive and therapeutic health measure in many East Asian countries especially, including Japan, China, and Korea (Lee et al., 2003). The sap of the lacquer tree (RV) is composed of urushiol (60 - 65%), glycoproteins (2.1 - 1.8%), flavonoids (1 - 2%), and gummy substances (6 - 7%) in addition to laccase (0.24%), stellacyanin, polysaccharides, peroxidase, and water (Yang et al., 2005). In addition to possessing anti-oxidant, anti-carcinogenic, anti-inflammatory, anti-bacterial, anti-apoptotic, immune-stimulating, and anti-viral properties, flavanoids from RV are known to overcome digestive disorders (Lee and Kim, 2000; Lee et al., 2002; Jang et al., 2005; Kim et al., 2010). The supplementation of RV in broilers improved fat digestibility and reduced serum levels of cholesterol and diglyceride with no effects on growth performance (Lohakare et al., 2006). It also improved the oxidative stability of meat in finishing pigs but without affecting carcass traits and performance (Song et al., 2008). In another study, it was indicated that the supplementation of RV meal in the diet of growing-finishing pigs improved carcass traits and decreased back fat thickness during refrigerated storage (Song et al., 2010).

However, there is still a limited amount of studies on this medicinal plant because it also contains a substance called urushiol that can cause allergic reactions or dermatitis (Kawai et al., 1992). We hypothesized that, if the RV extract was fermented, the negative effect due to urushiol could be reduced or eliminated, thereby enhancing the growth performance of animals. Thus, in this study, we investigated the efficacy of fermented *Rhus verniciflua* (FRV) meal as feed supplement on growth performance, apparent total tract digestibility (ATTD), blood profiles, fecal microflora, and fecal scores of weaning pigs.

Materials and Methods

The experiment was conducted at the Experimental Unit of the Dankook University (Anseodong, Cheonan, Choongnam, Korea). The protocol (Project No. PJ012067) for the current experiment was approved by the Animal Care and Use Committee of Dankook University.

Source of feed additive

The lacquer tree were initially placed in a machine where the conditions were controlled at 15 - 17% moisture, 70 - 90°C, and the water vapor pressure of 220 to 500 kPa, temperature at 115 - 125°C. Then, the materials were placed in an attrition mill to a size of 0.15 - 0.25 mm. The main composition of the herbs used in the current experiment was determined in accordance with the methods recommended by AOAC (2007). The lacquer tree was allowed to sun-dry for approximately 2 months. The sawdust obtained was then passed through a 2 - 3 mm mesh-screen, and this meal included 3.7 - 4.2% proteins, 5.1 - 6.3% fat, 63 - 79% neutral detergent fibre, and 1.3 - 2.1% ash, and the main

bioactive components were flavones, fisetin, arachidic acid, and butein (Cho et al., 2012) and used for supplement to the weanling pig diets.

Experimental design, animal, housing and diets

A total of 80 weaned pigs [(Landrace × Yorkshire) × Duroc, aged 24 - 25 days] with an average initial body weight (BW) of 6.82 ± 1.11 kg were used in a 42-day feeding trial. Pigs were randomly allotted to two experimental diets according to their BW and gender. There were eight replicate pens per treatment with five pigs per pen. Pigs were housed in an environmentally-controlled room. Each pen was equipped with one-sided, stainless steel self-feeder and a nipple drinker that allowed the pigs to have access to feed and water *ad libitum*. Dietary treatments consisted of

Table 1. Composition of basal diets (as-fed basis).

Items	1 - 7 days (Phase1)	8 - 21 days (Phase 2)	22 - 42 days (Phase 3)
Ingredients, g/kg			
Extruded corn	111.5	349.2	451.0
Extruded oat	100.0	-	-
Bakery by products	-	50.0	90.0
Soybean meal (44% CP)	80.0	200.0	296.5
Fermented soybean meal	78.0	82.0	-
Fish meal	50.0	40.0	25.0
Soy oil	41.5	48.0	30.0
Lactose	100.0	60.0	-
Whey	170.0	107.0	68.5
Milk product	130.0	20.0	20.0
Sugar	40.0	20.0	-
Plasma powder	65.0	-	-
L-Lysine HCl,	1.2	2.5	1.6
DL-Methionine, 99%	2.6	1.5	1.4
L-Threonine, 99%	7.7	0.8	-
Choline chloride, 25%	2.0	1.0	1.0
Vitamin premix ^y	1.0	1.0	1.0
Trace mineral premix ^z	2.0	2.0	2.0
Limestone	2.0	2.0	3.0
Salt	3.0	3.0	3.0
Calculated composition (%)			
ME (MJ/kg)	14.8	14.8	14.6
CP	220.0	210.0	205.0
Lysine	15.7	14.1	1.33
Methionine	6.0	4.9	0.47
Ca	8.0	7.8	0.75
Total P	7.6	7.6	0.64

^yProvided per kilogram of the complete diet: vitamin A, 11,025 IU; vitamin D3, 1,103 IU; vitamin E, 44 IU; vitamin K, 4.4 mg; riboflavin, 8.3 mg; niacin, 50 mg; thiamine, 4 mg; d-pantothenic acid, 29 mg; choline, 166 mg; vitamin B₁₂, 33 µg.

^zProvided per kilogram: Fe (as FeSO₄ × 7H₂O), 80 mg; Cu (as CuSO₄ × 5H₂O), 12 mg; Zn (as ZnSO₄), 85 mg; Mn (as MnO₂), 8 mg; I (as KI), 0.28 mg; and Se (as Na₂SeO₃ × 5H₂O), 0.15 mg.

CON, basal diet; TRT, and CON + 0.2% FRV. Diets were formulated to comply with National Research Council (NRC 2012) recommendations of nutrient requirements for swine (Table 1).

Sampling and measurements

Pigs were weighed at day 0, and week 2 and 6 of the experimental period, while feed intake was recorded on a per-pen basis to calculate average daily gain (ADG), average daily feed intake (ADFI), and gain : feed ratio (G : F). Chromium oxide (Cr_2O_3 , 2 g kg^{-1}) was added to diets as an indigestible marker to measure digestibility. Fresh fecal samples were collected directly via rectal massage from at least two pigs in each pen at days 14 and 42 of the experiment to determine the ATTD of dry matter (DM), energy (E), and nitrogen (N), according to AOAC (2007). All fecal and feed samples were stored at -20°C until analyzed. They were dried at 60°C for 72 h and ground to pass through a 1-mm screen. Chromium was analyzed by UV absorption spectrophotometry (Shimadzu UV-1201, Shimadzu, and Kyoto, Japan) using the method of Williams et al. (1962).

One gram of composite fecal sample from each pen was diluted with sterile saline (10^{-7} - 10^{-3}) and homogenized. Counts of viable bacteria in the fecal samples were determined by plating serial 10-fold dilutions (1% peptone solution) onto MacConkey agar plates and MRS agar plates (Difco, USA) to isolate *E. coli* and *Lactobacillus*, respectively. The number of colonies of *E. coli* and *Lactobacillus* was counted after incubation at 37°C for 38 h according to the method of Balasubramanian et al. (2016a). For blood characteristics, two pigs from each pen were randomly selected and blood samples were collected via anterior vena cava puncture at days 14 and 42, and collected into vacuum tubes containing K_2EDTA (Becton, Dickinson and Co., Franklin Lakes, NJ, USA). Red blood cells (RBC), white blood cells (WBC), IgG, and lymphocyte counts of whole blood samples were determined using an automatic blood analyzer (ADVIA 120, Bayer, Tarrytown, NY, USA) according to the method described by Balasubramanian et al. (2016b). Subjective diarrhea scores were recorded every 24 h from day 0 to day 42 by the same person and were based on the following: 1 = hard, dry pellets in a small hard mass; 2 = hard, formed stool that remains firm and soft; 3 = soft, formed, and moist stool that retains its shape; 4 = soft, unformed stool that assumes the shape of the container; 5 = watery, liquid stool that can be poured (Park et al., 2016b). Scores were recorded on a pen basis following observations of individual pig and of signs of stool consistency in the pen. The score is reported as average daily diarrhea of individual pig score.

Statistical analysis

All data were subjected to the GLM procedure of SAS/STAT[®] 9.2 (SAS Inst. Inc., Cary, NC) as a randomized complete block design. Each pen served as the experimental unit. Duncan's multiple range test was used to compare the means of the treatments (Duncan, 1955). Variability in the data was expressed as the pooled standard errors of means (SEM) and a probability value of $p < 0.05$ was considered to be statistically significant.

Results and Discussion

The use of phytogetic feed additives in the livestock industry shows an increasing trend due to their potential health benefits. Medicinal plants have positive biological activities and are recognized as natural substances (Bonneau and Laarveld, 1999; Chen et al., 2003). Guo et al. (2004) reported that some plants have beneficial effects such as

improving appetite and digestibility, stabilizing intestines by restraining viruses, increasing immunity by stimulating intestinal walls, and the production of digestive enzymes in animals.

The *Rhus verniciflua* is a plant indigenous to Korea which has traditionally been used in herbal medicines. It can be easily found on mountain terrain in many provinces of Korea (Kim, 1996). The active components were found to be garbanzol, sulfuretin, fisetin, fustin, and mollisacasin in the methanolic extract (Park et al., 2004). In Korea, much attention has been paid to *Rhus verniciflua* Stokes (RVS) because of it is a well known antioxidant (Hong et al., 1999). However, applications of RV as a food ingredient are limited due to the presence of urushiol, which causes allergic reactions (Eisen et al., 1962). Therefore, removal of urushiol is important for use of the plant in the food industry. Processing using organic solvents, heat, and enzymes has been attempted for removal of urushiol (Jeong et al., 2016). We supplemented FRV in weaning pig diets in the present study to evaluate their performance with an adequate diet. The FRV supplements have been reported to have antioxidant, anti-inflammatory, anticancer, and neuro-protective effects (Choi et al., 2010; Liu et al., 2013). Upadhaya and Kim (2015) also reported that *Bacillus* sp.-fermented soybean meal improved ileal digestibility of amino acids and nutrients in weaning pigs. In the current study, the inclusion of FRV meal had significant effects ($p < 0.05$) on growth performance of ADG and G : F ratio of weaning pigs produced. Improvements in these variables occurred during days 15 - 42 and the overall experimental period; however, no significant effect was observed ($p > 0.05$) on ADFI during the entire experiment (Table 2). In many plants, some active substances are highly odorous or may taste hot or pungent, which may restrict their use for animal feeding purposes (Zhang et al., 2012). They produce dose related depression of palatability in pigs fed these substances (Jugl-Chizzola et al., 2006; Schone et al., 2006). However, in the current study, RVS did not affect the ADFI throughout the experimental period. Contrary to the findings of the current study, lacquer meal supplementation had no significant difference on growth performance in broilers (Lohakare et al., 2006) and grower-finisher pigs (Song et al., 2008; 2010) were reported. However, in the present study, differences in growth performance for ADG and the G : F ratio were observed throughout the experimental period. These different results may be due to interspecies differences.

Table 2. Effect of fermented *Rhus verniciflua* supplementation on growth performance in weanling pigs^y.

Items	CON	TRT	SEM ^z
Days 0 - 14			
ADG, g	243	263	9
ADFI, g	333	362	30
G : F	0.730	0.727	0.024
Days 15 - 42			
ADG, g	426b	479a	9
ADFI, g	667	688	22
G : F	0.639b	0.696a	0.02
Days 0 - 42			
ADG, g	365b	407a	7
ADFI, g	556	579	25
G : F	0.656b	0.703a	0.012

^yCON: basal diet, TRT: CON + 0.2% Fermented *Rhus verniciflua*, ADG: average daily gain, ADFI: average daily feed intake, G : F: gain : feed ratio.

^zSEM: Standard error of the means.

a,b: Means in the same row with different superscripts are significantly different ($p < 0.05$).

Furthermore, dietary supplementation also improved the ATTD of DM ($p < 0.05$) at day 14 and the end of the experiment (Table 3). In contrast, there was no effect ($p > 0.05$) on ATTD of N and E during entire experiment. Huang et al. (2012) reported that Chinese medicinal herb supplementation provided a healthy and functional intestine, which in turn enhanced nutrient digestibility. Improved digestive capacity in the small intestine may be considered an indirect side effect of phytogenic feed additives, stabilizing the microbial balance in the gut of monogastric animals (Hernandez et al., 2004). Various herbal feed additives have been demonstrated to improve digestive tract function by increasing the activity of digestive enzymes of gastric mucosa and the nutrient utilization of livestock (Bhatt, 2015). The reason for improvement in growth performance and digestibility could be due to the enhancement of active materials derived from this medicinal plant and reduction or elimination of the anti-nutritive factor such as urushiol after fermentation. Similarly, Mathivanan et al. (2006) also demonstrated better growth rate in broilers fed *A. niger* fermented guar meal.

Table 3. Effect of fermented *Rhus verniciflua* supplementation on apparent total tract digestibility of nutrients in weanling pigs^y.

Items, %	CON	TRT	SEM ^z
Day 14			
Dry Matter	82.25b	83.44a	0.21
Nitrogen	81.10	82.84	0.98
Energy	81.03	82.86	1.67
Day 42			
Dry Matter	81.28b	83.30a	0.20
Nitrogen	79.72	79.36	0.33
Energy	81.75	82.84	1.17

^yCON: basal diet, TRT: CON + 0.2% Fermented *Rhus verniciflua*.

^zSEM: Standard error of the means.

a,b: Means in the same row with different superscripts are significantly different ($p < 0.05$).

Our results revealed that immunity-related blood profiles (RBC, WBC concentration, lymphocyte percentage, and IgG) were not affected ($p > 0.05$) by FRV supplementation during the entire experimental period (Table 4) and stayed within the normal range (differences not significant). Therefore, these results specified that the supplementation in this study can stimulate the immune system to some extent (Dibner and Buttin, 2002). However, there are no studies so far to compare our blood profile results from pigs fed with FRV. Post-weaning diarrhea is one of the issues faced by the swine industry. In commercial practice, the use of different additives has been recommended as a way to help reduce diarrhea in piglets. With regards to fecal microbiota, the present study demonstrated a significant increase ($p < 0.05$) in *Lactobacillus* counts with FRV supplementation, but *E. coli* ($p > 0.05$) remained unaffected (Table 5). These results are in partial agreement with findings of Jeong and Kim (2015) who reported improved *Lactobacillus* counts in broiler fed fermented medicinal plants. In addition, Huang et al (2012) found that Chinese medicinal herbs used in their trial elicited a decrease in diarrhea scores in the first 10 days of the experimental period. Similarly, diarrhea scores ($p < 0.05$) showed significantly decreased effects during days 22 - 42 (Table 5) in the present study. However, Zhang et al. (2012) demonstrated that phytogenic supplementation had no effect on diarrhea score in weanling pigs. The reasons for this result may be that, within phytogenic feed additives, the content of active substances in products could vary

Table 4. Effect of fermented *Rhus verniciflua* supplementation on blood profiles in weanling pigs^y.

Items	CON	TRT	SEM ^z
Day 14			
WBC, 10 ³ /μL	16.99	18.21	1.46
RBC, 10 ⁶ /μL	6.91	6.71	0.43
Lymphocyte, %	50.0	51.2	0.9
IgG, mg/dL	203	201	12
Day 42			
WBC, 10 ³ /μL	23.44	24.57	1.66
RBC, 10 ⁶ /μL	6.51	6.07	0.34
Lymphocyte, %	65.4	65.7	2.5
IgG, mg/dL	291	286	31

^yCON: basal diet, TRT: CON + 0.2% Fermented *Rhus verniciflua*, WBC: white blood cells, RBC: red blood cells, IgG: Immunoglobulin G.

^zSEM: Standard error of the means.

Table 5. Effects of fermented *Rhus verniciflua* supplementation on fecal microbiota counts and fecal score in weanling pigs^x.

Items	CON	TRT	SEM ^y
Fecal microbial log10 cfu/g			
<i>Lactobacillus</i>	6.86b	7.48a	0.07
<i>E. coli</i>	6.40	6.41	0.12
Fecal score ^z			
0 - 21 days	2.47	2.86	0.13
22 - 42 days	3.25b	3.52a	0.16

^xCON : basal diet, TRT : CON + 0.2% Fermented *Rhus verniciflua*.

^ySEM : Standard error of the means.

^zSubjective fecal scores were recorded based on the following: 1 = hard, dry pellets in a small, hard mass; 2 = hard, formed stool that remains firm and soft; 3 = soft, formed, and moist stool that retains its shape; 4 = soft, unformed stool that assumes the shape of the container; 5 = watery, liquid stool that can be poured.

a,b: Means in the same row with different superscripts are significantly different ($p < 0.05$).

widely, depending on the plant part used (e.g. seeds, leaf, root or bark) harvesting season, and geographical origin (Windisch et al., 2008). The technique for processing (e.g. cold expression, steam distillation, and extraction with non-aqueous solvents) modifies the active substances and associated compounds within the final product; thus a detailed study of the active molecules is necessary and will be carried out in a future study. A possible explanation for the discrepancy in findings could be intrinsic and extrinsic factors including environment, diet, and nutritional status.

Conclusion

The result of the present study suggests that the addition of FRV meal supplement showed positive results in terms of growth performance, increased DM, beneficial effects on fecal microbiota counts, and decreased diarrheal score, indicating the potential of this medicinal plant as a substitute for antibiotics. However, in future experiments, a detailed analysis of the fermented product must be performed to better understand the role of fermentation in improving the feed value of this medicinal plant.

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References

- AOAC. 2007. Official methods of analysis, 18th ed., Association of Official Analytical Chemists, Gaithersburg, MD, USA.
- Balasubramanian B, Li T, Kim IH. 2016a. Effects of supplementing growing-finishing pig diets with *Bacillus spp.* probiotic on growth performance and meat-carcass grade quality traits. *Revista Brasileira de Zootecnia* 45:93-100.
- Balasubramanian B, Park JW, Kim IH. 2016b. Evaluation of the effectiveness of supplementing micro-encapsulated organic acids and essential oils in diets for sows and suckling piglets. *Italian Journal of Animal Science* 15:626-633.
- Bhatt N. 2015. Herbs and herbal supplements, a novel nutritional approach in animal nutrition. *Iranian Journal of Applied Animal Science* 5:497-516.
- Bonneau M, Laarveld B. 1999. Biotechnology in animal nutrition, physiology and health. *Livestock Production Science* 59:223-241.
- Campbell JM, Crenshaw JD, Polo J. 2013. The biological stress of early weaned piglets. *Journal of Animal Science and Biotechnology* 4:19.
- Chen HL, Li DF, Chang BY, Gong LM, Dai JG, Yi GF. 2003. Effect of Chinese herbal polysaccharides on the immunity and growth performance of young broilers. *Poultry Science* 82:364-370.
- Cho JH, Zhang S, Kim IH. 2012. Effects of anti-diarrhoeal herbs on growth performance, nutrient digestibility, and meat quality in pigs. *Asian-Australasian Journal of Animal Science* 25:1595-1604.
- Choi MJ, Lee SJ, Jang SH, Reza MA, Hong JH, Jung HK, Park SC. 2010. Biological activities and single oral dose toxicity in rat of fermented *Rhus verniciflua* extract. *Korean Journal of Veterinary Research* 50:187-195.
- Dibner JJ, Buttin P. 2002. Use of organic acid as a model to study the impact of gut microflora on nutrition and metabolism. *The Journal of Applied Poultry Research* 11:453-463.
- Duncan DB. 1955. Multiple range and multiple F test. *Biometrics* 11:1.
- Eisen EJ, Bohren BB, McKean HE. 1962. The Haugh unit as a measure of egg albumen quality. *Poultry Science* 41:1461-1468.
- Guo FC, Kwakkel RP, Soede J, Williams BA, Verstegen MW. 2004. Effects of a Chinese herb medicine formulation, as an alternative for antibiotics, on performance of broilers. *British Poultry Science* 45:793-797.
- Hernandez F, Madrid J, Garcia V, Orengo J, Megias MD. 2004. Influence of two plant extracts on broilers performance, digestibility, and digestive organ size. *Poultry Science* 83:169-174.
- Hong DH, Han SB, Lee CW, Park SH, Jeon YJ, Kim MJ, Kwak SS, Kim HM. 1999. Cytotoxicity of urushiols isolated from sap of Korean lacquer tree (*Rhus verniciflua* Stokes), *Archives of Pharmacal Research* 22:638-641.

- Huang CW, Lee TT, Shih YC, Yu B. 2012. Effects of dietary supplementation of Chinese medicinal herbs on polymorphonuclear neutrophil immune activity and small intestinal morphology in weanling pigs. *Journal of Animal Physiology and Animal Nutrition* 96:285-294.
- Jang HS, Kook SH, Son YO, Kim JG, Jeon YM, Jang YS, Choi KC, Kim J, Han SK, Lee KY, Park BK, Cho NP, Lee JC. 2005. Flavonoids purified from *Rhus verniciflua* Stokes actively inhibit cell growth and induce apoptosis in human osteosarcoma cells. *Biochimica et Biophysica Acta* 17:309-316.
- Jeong HJ, Park JA, Kim MJ. 2016. Optimization of the extraction process for fermented *Rhus verniciflua* stokes using response surface methodology. *Food Science and Biotechnology* 25:179-184.
- Jeong JS, Kim IH. 2015. Effect of fermented medicinal plants (*Gynura procumbens* *Rehmannia glutinosa*, *Scutellaria baicalensis*) as alternative performance enhancers in broilers. *The Journal of Poultry Science* 52:119-126.
- Jugl-Chizzola M, Ungerhofer E, Gabler C, Hagmuller W, Chiz-zola R, Zitterl-Eglseer K, Franz C. 2006. Testing of the palatability of *Thymus vulgaris* L. and *Origanum vulgare* L. as flavouring feed additive for weaner pigs on the basis of a choice experiment. *Berliner Und Munchener Tierarztliche Wochenschrift* 119:238-243.
- Kawai K, Nakagawa M, Kawai K, Miyakoshi T, Miyashita K, Asami T. 1992. Heat treatment of Japanese lacquerware renders it hypoallergenic. *Contact Dermatitis* 27:244-249.
- Kim JS, Kwon YS, Chun WJ, Kim TY, Sun JH, Yu CY, Kim MJ. 2010. *Rhus verniciflua* Stokes flavonoid extracts have anti-oxidant, anti-microbial and a glucosidase inhibitory effect. *Food Chemistry* 120:539-543.
- Kim TJ. 1996. Korea resource plants, Vol. II. pp 292-297. Seoul University Press, Seoul, Korea.
- Lee JC, Kim J, Jang YS. 2003. Ethanol eluted extract of *Rhus verniciflua* Stokes inhibits cell growth and induces apoptosis in human lymphoma cells. *International Journal of Biochemistry and Molecular Biology* 36:337-343.
- Lee JC, Lim KT, Jang YS. 2002. Identification of *Rhus verniciflua* Stokes compounds that exhibit free radical scavenging and anti-apoptotic properties. *Biochimica et Biophysica Acta* 1570:181-191.
- Lee JC, Lim KT. 2000. Screening of antioxidant and antimicrobial effects from *Rhus verniciflua* Stokes (RVS) ethanolic extract. *Food Science and Biotechnology* 9:139-145.
- Liu CS, Nam TG, Han MW, Ahn SM, Choi HS, Kim TY, Chun OK, Koo SI, Kim D. 2013. Protective effect of detoxified *Rhus verniciflua* Stokes on human keratinocytes and dermal fibroblasts against oxidative stress and identification of the bioactive phenolics. *Bioscience, Biotechnology, and Biochemistry* 77:1682-1688.
- Lohakare JD, Zheng J, Yun JH, Chae BJ. 2006. Effect of lacquer (*Rhus verniciflua*) supplementation on growth performance, nutrient digestibility, carcass traits and serum profile of broiler chickens. *Asian-Australasian Journal of Animal Science* 19:418-424.
- Mathivanan R, Selvaraj P, Nanjappan K. 2006. Feeding fermented soybean meal on broiler performance. *International Journal of Poultry Science* 5:868-872.
- Nabuurs MJA. 1995 Microbiological structural and functional changes of the small intestine of pigs at weaning. *Pig News and Information* 16:93-97.
- NRC. 2012. Nutrient requirements of swine, 11th ed. National Academy of Press, Washington, DC, USA.
- Park JW, Yun HM, Park JH, Lee IS, Kim IH. 2016a. Effect of supplementation *Oreganum aetheroleum* essential oil on growth performance in sows and growth performance, fecal score in weanling pigs. *Korean Journal of Agricultural Science* 43:794-801.
- Park KY, Jung GO, Lee KT, Choi JW, Choi MY, Kim GT, Jung HJ, Park HJ. 2004. Antimutagenic activity of

- flavonoids from the heartwood of *Rhus verniciflua*. Journal of Ethno pharmacology 90:73-79.
- Park SW, Kim BH, Kim YH, Kim SN, Jang KB, Kim YH, Park JC, Song MH, Oh SN. 2016b. Nutrition and feed approach according to pig physiology. Korean Journal of Agricultural Science 43:750-760.
- Pluske JR, Williams IH, Aherne FX. 1995. Nutrition of the neonatal pig. In *The neonatal pig: development and survival*/edited by Varley MA. pp 187-235. CAB International, Wallingford, UK.
- Schone F, Vetter A, Hartung H, Bergmann H, Biertumpfel A, Richter G, Muller S, Breitschuh G. 2006. Effects of essential oils from fennel (*Foeniculi aetheroleum*) and caraway (*Carvi aetheroleum*) in pigs. Journal of Animal Physiology and Animal Nutrition 90:500-510. doi:10.1111/j.1439-0396.2006.00632.x.
- Song CH, Choi JY, Yoon SY, Yang YX, Shinde PL, Kwon IK, Kang SM, Lee SK, Chae BJ. 2008. Effects of lacquer (*Rhus verniciflua*) meal on carcass traits, fatty acid composition and meat quality of finishing pigs. Asian-Australasian Journal of Animal Science 21:1207-1213.
- Song CH, Kim JS, Shinde P, Kim YW, Kim KH, Kwon IK, Kang SM, Lee SK, Chae BJ. 2010. Effect of inclusion of lacquer (*Rhus verniciflua* stokes) meal on carcass traits and meat quality in growing-finishing pigs. Korean Journal for Food Science of Animal Resources 30:597-602.
- Turpin DL, Pieter L, Chen TY, Lines D, Pluske JR. 2016. Intermittent suckling causes a transient increase in cortisol that does not appear to compromise selected measures of piglet welfare and stress. Animals 6:24.
- Upadhaya SD, Kim IH. 2015. Ileal digestibility of nutrients and amino acids in unfermented, fermented soybean meal and canola meal for weaning pigs. Animal Science Journal 86:408-414.
- Williams CH, David DJ, Iismaa O. 1962. The determination of chromic oxide in faeces samples by atomic absorption spectrophotometry. Journal of Agricultural Science 59:381-385.
- Wilson AD, Stoke CR, Boure J. 1989. Effect of age on absorption and immune responses to weaning or introduction of novel dietary antigens in pigs. Research in Veterinary Science 46:180-186.
- Windisch W, Schedle K, Plitzner C, Kroismayr A. 2008. Use of phytogenic products as feed additives for swine and poultry. Journal of Animal Sciences 86:140-148.
- Yang J, Du Y, Huang R, Sun L, Liu H, Gao XJF. 2005. Kennedy Chemical modification and antitumour activity of Chinese lacquer polysaccharide from lac tree *Rhus vernicifera*, Carbohydrate Polymers 59:101-107.
- Zhang S, Jung JH, Kim HS, Kim BY, Kim IH. 2012. Influences of phytoncide supplementation on growth performance, nutrient digestibility, blood profiles, diarrhea scores and fecal microflora shedding in weaning pigs. Asian-Australasian Journal of Animal Sciences 25:1309-1315.