

REVIEW ARTICLE

Salmonellosis in swine: Clinical perspectives

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Abstract

Salmonella is one of the most important food-borne zoonotic pathogens, causing acute or chronic digestive diseases such as enteritis. The acute form of enteritis is common in young pigs of 2 - 4 months of age. The main symptoms include high fever (41 - 42 °C), loss of appetite, and increased mortality within 2 - 4 days of onset of the disease. It is often the cause of increasing mortality, decreasing growth rate and reducing feed efficiency of piglets. In the case of chronic enteritis in pigs, the main symptom is weight loss due to the continuing severe diarrhea. *Salmonella enterica* serovar Typhimurium and *Salmonella enterica* serovar Choleraesuis are typical pig adapted serotypes, which cause one of four major syndromes: enteric fever, enterocolitis/diarrhea, bacteremia and chronic asymptomatic carriage. These syndromes cause a huge economic burden to swine industry by reducing production. Therefore, it is necessary that swine industries should strive to decrease Salmonellosis in pigs in order to reduce economic losses. There are several measures, such as vaccination to prevent salmonellosis, that are implemented differently from country to country. For the treatment of *Salmonella*, ongoing antibiotic treatment is needed. However constant doses of antibiotics can be a problem because of antibiotic resistance. Therefore, the focus should be made more on prevention than treatment. In this review, we addressed the basic information about *Salmonella*, route of infection, clinical symptoms, and prevention of Salmonellosis.

Keywords: prevention, productivity of the swine industry, *Salmonella*, Salmonellosis

Introduction

Salmonella is a non-lactose fermenting bacteria belonging to the family *Enterobacteriaceae*. *Salmonella* has fimbriae and peritrichous flagella, and it is a noncapsulated, gram-negative, short rod bacterium. Facultatively anaerobic *Salmonella* grows on defined media without special growth factors (Timoney et al., 1988). However, the quantification and isolation of *Salmonella* from feces is somewhat prone to failure since *Salmonella* is often less abundantly present among competitor bacterial species in feces. Multiple enrichments using specially formulated selective and differential media for the isolation of *Salmonella* have been devised in order to overcome this constraint. The two most useful of these are Xylose-Lysine-Deoxycholate (XLD) and xylose-lysine-tergitol (XLT) agars that have replaced MacConkey agar. In addition, the isolation of *Salmonella* in fecal contents



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using Rappaport-Vassiliadis R10 and tetrathionate broth has been effective. The optimum growth temperature for *Salmonella* is 37°C (Timoney et al., 1988; Busse, 1995; Nye et al., 2002; Korsak et al., 2004).

It has been known that medically important salmonella may be considered as a single species, known as *Salmonella enterica*, which has more than 2,500 different serotypes (Murray et al., 1999). Clinically familiar names, such as *Salmonella typhimurium* or *Salmonella* serotype typhimurium, which would be renamed *Salmonella enterica* serotype Typhimurium are being used in most medical laboratories (Timoney et al., 1988; Hohmann, 2001).

Salmonella serovars commonly found in swine are a major human health problem in the world. The most common serotypes associated with human *Salmonella* infections include following 4 serotypes, *S. enterica* serovar Typhimurium, *S. Heidelberg*, *S. Agona*, and *S. Infantis* (Foley and Nayak, 2008). As a common food borne diseases in humans, Salmonellosis is related to contaminated food, such as pork products that are an important source of *Salmonella* infection in humans. It has been shown that 9.6% of the pork in United States retail stores was contaminated with *Salmonella* (Duffy et al., 2000; Andino and Hanning, 2015).

The reduction of the opportunistic pathogens in the gastrointestinal tract of swine, including *Salmonella*, has been proposed as one of mechanisms for antibiotic-mediated growth promotion (AGPs). Nevertheless, it is not clear if the use of AGPs results in lower load or carriage of *Salmonella* (Dibner and Richards, 2005). If it is true, the reduction in opportunistic pathogens by AGPs is of importance in terms of accessing the mechanisms of growth promotion by AGPs as well as its relationship with public health.

Transmission, prevalence and clinical symptoms

Transmission

Transmission of *Salmonella* between pigs can occur through the fecal-oral and intranasal routes, involving colonization and dissemination of the gastrointestinal tract and organs such as lungs and tonsils. Also, airborne or direct nose-to-nose contact could increase the possibility of transmission of *Salmonella* in intensive pig production systems (Oliveira et al., 2007). In addition, several potential risk factors for *Salmonella* infection, such as poor management and hygiene have been identified (Table 1) (Wales et al., 2013). In general, all ages of pigs are susceptible to *Salmonella* infection, however, weaned and growing-finishing pigs are more prone to *Salmonella* infection (Meurens et al., 2009).

Prevalence

The prevalence of *Salmonella typhimurium* was higher (62.5%) than other *Salmonella* serovars, and the prevalence of *Salmonella* infection was 32.9% both in finishing pigs and weaned pigs in India (Kumar et al., 2014). Young pigs are more susceptible to Salmonellosis because of immature intestinal function, concurrent diseases, and changes of surrounding environment (Wales et al., 2013; Aiello et al., 2016). Of these reasons, it is advised that swine farmers should focus on both improved hygiene and husbandry of young piglets to prevent *Salmonella* infection.

Non-typhoidal salmonella (NTS) illness accounts for approximately one million cases in the United States alone (Scallan et al., 2011; Tamang et al., 2015). Moreover, *Salmonella* was the second most commonly reported zoonoses in the European Union and was the third most common reason of foodborne diseases in Korea. In Spain, García-Feliz et al. reported 43.1% of prevalence of *Salmonella* in finishing pig herds (García-Feliz et al., 2007; Tadee et al., 2014). However, the prevalence of *Salmonella* was 5.58% in fattening pigs in Germany (Visscher et al., 2011; Tadee et al.,

Table 1. Potential risk factors for *Salmonella* infection (Wales et al., 2013).

Type	Possible Sources
Housing	Type of pen (slatted, push = through, straw yard)
Management	Number of staff Number of pigs on farm Time as a pig farm Age groups housed separately Feed source Sick pens Reintegration of recovered pigs Bedding source Bedding type Feeder/drinker systems
Biosecurity/hygiene	Staff contact with other pigs Visitors who have contact with other pigs Water supply (mains or borehole) Drainage Presence of wildlife species Proximity of waterways, scrubland, pig wastes, cattle wastes, or sewage treatment/landfill sites Feed storage (capacity and sealed/open bins) Presence of biosecurity measures (boot dips, well wash, and visitor/staff clothing) Pigs loaded at perimeter Site contained within perimeter fence Rodent controls Disinfectant: type and frequency

2014).

S. typhimurium is most commonly attributed to swine *Salmonellosis* in Europe. It becomes a source of contamination of pork products, and it causes tremendous economic and animal welfare costs (Arce et al., 2014). In the pig farms of the Tarai region of Uttarakhand, India, the most prevalent *Salmonella* serotype was *Salmonella typhimurium* that accounts for about 62.5% of known serotypes (Kumar et al., 2014).

S. Choleraesuis is especially adapted to pigs and is rarely detected in other species, however, it can also cause systemic infection in humans (Chang et al., 2013). *Salmonella* serovar *Choleraesuis* var. *Kunzendorf* was the main cause of *S. Choleraesuis* outbreaks in pigs in Europe (Pedersen et al., 2015). Nevertheless, *S. Choleraesuis* infection in humans is not frequent in Canada or United Kingdom (Barrell, 1987; Briggs and Fratamico, 1999; Onyango et al., 2014). Interestingly, it was the second most common cause of human *Salmonellosis* in Taiwan (Chen et al., 1999; Onyango et al., 2014).

Clinical symptoms

The main route of *Salmonella* infection in humans is ingestion of contaminated food products or direct contact. It makes livestock species a potential source of *Salmonellosis* (Zhao et al., 2006a; Clothier et al., 2010). In swine, *Salmonella* infections such as *S. Choleraesuis* or *S. Typhimurium* produce a variety of disease syndromes including septicemia, enteritis, and respiratory disease (Boyen et al., 2008; Foley et al., 2008; Huang et al., 2009; Vigo et al.,

2009; Clothier et al., 2010). Piglets are especially vulnerable to *Salmonella* infection and this leads to high morbidity and mortality (Schierack et al., 2006; Wang et al., 2014).

Infections with *S. Typhisuis* and *S. Choleraesuis* usually result in swine typhoid, characterized by a severe systemic disease that may result in enteric and fatal systemic disease (Gray et al., 1996; Lichtensteiger and Vimr, 2003; Chiu et al., 2004; Chiu et al., 2005; Ku et al., 2005; Nishio et al., 2005; Zhao et al., 2006b; Boyen et al., 2008).

The main sign of *S. Typhimurium*, which is most commonly found in pigs, is usually diarrhea. However, *S. Typhimurium* infection in young pigs can cause significant disease such as increase of rectal temperature and occurrences of *S. Typhimurium* in the jejunum, cecum, tonsil, and mesenteric lymph node (Tanaka et al., 2010; Yin et al., 2014). Pigs can be asymptomatic with *S. Typhimurium* present in their tonsils, gut and gut-associated lymphoid tissues, and they can become *Salmonella* carriers (Berends et al., 1996; Verbrugge et al., 2011). When pigs are infected with *S. Typhimurium*, they suffer from enterocolitis which is one of the clinical signs of *S. Typhimurium* infection. Those pigs can also become chronic carriers (Nollet et al., 2005; Gradassi et al., 2013).

S. Choleraesuis usually causes necrotizing enterocolitis, apart from septicemia, characterized by hepatitis, pneumonia, and cerebral vasculitis and also causes systemic infections in humans. (Reed et al., 1986; Wilkins and Roberts, 1988; Chang et al., 2013).

Prevention

Management

The efforts made to reduce Salmonellosis incidence differ geographically. In the EU, efforts are focused on the process from farm to processing. On the other hand, in the USA, interventions are applied at the processing level (Rajic et al., 2007; Wilhelm et al., 2012).

Despite of the different focuses to reduce Salmonellosis, the main prevention involves measures to improve good management practices and post-harvest hygiene on farms (Mousing et al., 1997; Alban et al., 2002; Schwarz et al., 2011). These days, the addition of organic acid to feed or water and the administration of probiotics or prebiotics to pigs have emerged as methods to control *Salmonella* in swine farms. It has been shown that these can decrease the faecal excretion of *Salmonella* as well (Casey et al., 2007; Creus et al., 2007; De Busser et al., 2009; Martin-Pelaez et al., 2010; Schwarz et al., 2011; Mejicanos et al., 2016).

Another way to reduce *Salmonella* infection in swine farms is all-in-all-out system. Based on this system, all pigs should enter and leave from the facility together at the same time and this should be applied in mating, farrowing, and nursery units. This system can help to reduce the risk of *Salmonella* infection since the pigs raised under the same management system are considered to have a similar disease status (Scott et al., 2006; Padungtod et al., 2008).

In addition, it has been proven that fermented feed, *Lactobacillus casei*-fermented feed, are good for pigs and reduce *Salmonella* infection. It has also been shown that it was effective in reducing pig diarrhea and intestinal burden of *Salmonella* as well (Yin et al., 2014). Giving feed fermented liquid to pigs was shown to improve the performance of pigs of all ages (Missotten et al., 2015).

It was suggested that the use of a mixed-bacterial-species competitive-exclusion culture can reduce both load and carriage of *Salmonella* (Genovese et al., 2003). Reductions in the fecal shedding of *Salmonella* resulted in diminished horizontal transmission between pen- and littermates (Genovese et al., 2003).

The use of antibiotics is not recommended because of *Salmonella*'s intracellular persistence and antibiotic resistance. For this reason, vaccination may represent an attractive alternative for the reduction of *Salmonella* incidence in swine (Gradassi et al., 2013).

Vaccines

Even though, strict hygiene practices and rational husbandry resistance management are the best way to reduce the prevalence of *Salmonella*, this approach cannot be applied in countries where high prevalence of *Salmonella* infection is observed. In this case, vaccination is considered a major control measure to reduce *Salmonella* contamination at the early stages of meat production (Selke et al., 2007; Eddicks et al., 2009; Leyman et al., 2011; Xu et al., 2012; Pesciaroli et al., 2013).

Killed *Salmonella* vaccines have been used in pigs with variable results. This discrepancy between efficacy results of Killed *Salmonella* vaccines may be due to their poor ability to induce cell-mediated immunity in vaccinated pigs (Xu et al., 1993; Yamane et al., 2000; Davies and Breslin, 2003; Schwarz et al., 2011). On the other hand, attenuated live vaccine strains have been shown to offer better protection than Killed *Salmonella* vaccines (Haesebrouck et al., 2004). Live vaccine strains consist of strains that are attenuated, strains with a mutation in genes that are important for bacterial metabolism, or strains in which specific virulence genes are removed (Kramer et al., 1992; Lumsden and Wilkie, 1992; Mastroeni et al., 2001; Boyen et al., 2009; Schwarz et al., 2011).

Vaccination is one of the suggested methods to reduce the burden of *S. Typhimurium* on swine farms (Haesebrouck et al., 2004; Eddicks et al., 2009; Hotes et al., 2011; Pesciaroli et al., 2013) and it is one possible supplementary measure to reduce *Salmonella* infection. Nowadays, various vaccine studies have been conducted in pigs with various results (Roesler et al., 2004; Farzan and Friendship, 2010; Hotes et al., 2011; De Ridder et al., 2013a; De Ridder et al., 2013b). For example, SC54 vaccine, a *S. Choleraesuis* var. *kunzendorf* avirulent live strain, is effective to protect pigs against infection by virulent *S. Choleraesuis* and virulent *S. Typhimurium* (Kramer et al., 1992; Letellier et al., 2000).

Conclusion

Salmonella is a major concern to the swine industry because it leads to clinical illnesses in swine resulting in poor growth performance and profitability (Callaway et al., 2006). As such, *S. Typhimurium* infection in growing pigs causes growth retardation (Balaji et al., 2000; Turner et al., 2002a; Turner et al., 2002b). In the acute phase of the *Salmonella* infection, swines show a reduction of growth rate and develop a fever leading to lower profitability (Jenkins et al., 2004).

S. Typhimurium and *S. Choleraesuis* are typical pig-adapted serotypes. Infection by *Salmonella* results in economic losses in swine industries by causing serious clinical syndromes in weaned pigs. *Salmonella* infection in weaned pigs causes fever and continuous diarrhea. It is often the cause of increased mortality, decreased growth rate, and reduced feed efficiency. There are several measures, such as vaccination to prevent salmonellosis, that are implemented differently from country to country. For the treatment of *Salmonella*, ongoing antibiotic treatment is needed. However, constant doses of antibiotics can be a problem because of antibiotic resistance. Therefore, more focus should be made on prevention than treatment.

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References

- Alban L, Stege H, Dahl J. 2002. The new classification system for slaughter-pig herds in the Danish *Salmonella* surveillance-and-control program. *Preventive Veterinary Medicine* 53:133-146.
- Andino A, Hanning I. 2015. *Salmonella enterica*: survival, colonization, and virulence differences among serovars. *Scientific World Journal* 2015:520179.
- Aiello SE, Moses MA. 2016. The Merck Veterinary Manual 11th Edition. Merck and Co, Kenilworth, NJ.
- Arce C, Lucena C, Moreno A, Garrido JJ. 2014. Proteomic analysis of intestinal mucosa responses to *Salmonella enterica* serovar typhimurium in naturally infected pig. *Comparative Immunology, Microbiology and Infectious Diseases* 37:59-67.
- Balaji R, Wright KJ, Hill CM, Dritz SS, Knoppel EL, Minton JE. 2000. Acute phase responses of pigs challenged orally with *Salmonella typhimurium*. *Journal of Animal Science* 78:1885-1891.
- Barrell RA. 1987. Isolations of salmonellas from humans and foods in the Manchester area: 1981-1985. *Epidemiology and Infection* 98:277-284.
- Berends BR, Urlings HA, Snijders JM, Van Knapen F. 1996. Identification and quantification of risk factors in animal management and transport regarding *Salmonella* spp. in pigs. *International Journal of Food Microbiology* 30:37-53.
- Boyen F, Haesebrouck F, Maes D, Van Immerseel F, Ducatelle R, Pasmans F. 2008. Non-typhoidal *Salmonella* infections in pigs: a closer look at epidemiology, pathogenesis and control. *Veterinary Microbiology* 130:1-19.
- Boyen F, Pasmans F, Van Immerseel F, Donne E, Morgan E, Ducatelle R, Haesebrouck F. 2009. Porcine in vitro and in vivo models to assess the virulence of *Salmonella enterica* serovar Typhimurium for pigs. *Laboratory Animals* 43:46-52.
- Briggs CE, Fratamico PM. 1999. Molecular characterization of an antibiotic resistance gene cluster of *Salmonella typhimurium* DT104. *Antimicrobial Agents and Chemotherapy* 43:846-849.
- Busse M. 1995. Media for *Salmonella*. *International Journal of Food Microbiology* 26:117-131.
- Callaway TR, Morrow JL, Edrington TS, Genovese KJ, Dowd S, Carroll J, Dailey JW, Harvey RB, Poole TL, Anderson RC, Nisbet DJ. 2006. Social stress increases fecal shedding of *Salmonella typhimurium* by early weaned piglets. *Current Issues in Intestinal Microbiology* 7:65-71.
- Casey PG, Gardiner GE, Casey G, Bradshaw B, Lawlor PG, Lynch PB, Leonard FC, Stanton C, Ross RP, Fitzgerald GF, Hill C. 2007. A five-strain probiotic combination reduces pathogen shedding and alleviates disease signs in pigs challenged with *Salmonella enterica* Serovar Typhimurium. *Applied and Environmental Microbiology* 73:1858-1863.
- Chang CH, Chen YS, Chiou MT, Su CH, Chen DS, Tsai CE, Yu B, Hsu YM. 2013. Application of scutellariae radix, gardeniae fructus, and probiotics to prevent *Salmonella enterica* serovar choleraesuis infection in swine. *Evidence-Based Complementary and Alternative Medicine* 2013:568528.
- Chen YH, Chen TP, Tsai JJ, Hwang KP, Lu PL, Cheng HH, Peng CF. 1999. Epidemiological study of human salmonellosis during 1991-1996 in southern Taiwan. *The Kaohsiung Journal of Medical Sciences* 15:127-136.
- Chiu CH, Su LH, Chu C. 2004. *Salmonella enterica* serotype Choleraesuis: epidemiology, pathogenesis, clinical disease, and treatment. *Clinical Microbiology Reviews* 17:311-322.
- Chiu CH, Tang P, Chu C, Hu S, Bao Q, Yu J, Chou YY, Wang HS, Lee YS. 2005. The genome sequence of *Salmonella enterica*

- serovar Choleraesuis, a highly invasive and resistant zoonotic pathogen. *Nucleic Acids Research* 33:1690-1698.
- Clothier KA, Kinyon JM, Frana TS. 2010. Comparison of *Salmonella serovar* isolation and antimicrobial resistance patterns from porcine samples between 2003 and 2008. *Journal of veterinary diagnostic investigation* 22:578-582.
- Creus E, Perez JF, Peralta B, Baucells F, Mateu E. 2007. Effect of acidified feed on the prevalence of *Salmonella* in market-age pigs. *Zoonoses and Public Health* 54:314-319.
- Davies R, Breslin M. 2003. Effects of vaccination and other preventive methods for *Salmonella enteritidis* on commercial laying chicken farms. *The Veterinary Record* 153:673-677.
- De Busser EV, Dewulf J, Nollet N, Houf K, Schwarzer K, De Sadeleer L, De Zutter L, Maes D. 2009. Effect of organic acids in drinking water during the last 2 weeks prior to slaughter on *Salmonella* shedding by slaughter pigs and contamination of carcasses. *Zoonoses and Public Health* 56:129-136.
- De Ridder L, Maes D, Dewulf J, Pasmans F, Boyen F, Haesebrouck F, Meroc E, Butaye P, Van der Stede Y. 2013a. Evaluation of three intervention strategies to reduce the transmission of *Salmonella* Typhimurium in pigs. *Veterinary Journal* 197:613-618.
- De Ridder L, Maes D, Dewulf J, Pasmans F, Boyen F, Haesebrouck F, Meroc E, Roels S, Leyman B, Butaye P, Van der Stede Y. 2013b. Effect of a DIVA vaccine with and without in-feed use of coated calcium-butyrate on transmission of *Salmonella* Typhimurium in pigs. *BioMed Central Veterinary Research* 9:243-6148-9-243.
- Dibner JJ, Richards JD. 2005. Antibiotic growth promoters in agriculture: history and mode of action. *Poultry Science* 84:634-643.
- Duffy EA, Belk KE, Sofos JN. 2000. United States Retail Pork Microbiological Baseline. In: Proceedings. Pork Quality and Safety Summit. pp. 305-309. National Pork Producers Council.
- Eddicks M, Palzer A, Hormansdorfer S, Ritzmann M, Heinritzi K. 2009. Examination of the compatibility of a *Salmonella* Typhimurium-live vaccine Salmoporc for three day old suckling piglets. *DTW. Deutsche Tierärztliche Wochenschrift* 116:249-254.
- Farzan A, Friendship RM. 2010. A clinical field trial to evaluate the efficacy of vaccination in controlling *Salmonella* infection and the association of *Salmonella*-shedding and weight gain in pigs. *Canadian Journal of Veterinary Research* 74:258-263.
- Foley SL, Lynne AM, Nayak R. 2008. *Salmonella* challenges: prevalence in swine and poultry and potential pathogenicity of such isolates. *Journal of Animal Science* 86:E149-62.
- Garcia-Feliz C, Collazos JA, Carvajal A, Vidal AB, Aladuena A, Ramiro R, de la Fuente M, Echeita MA, Rubio P. 2007. *Salmonella enterica* infections in Spanish swine fattening units. *Zoonoses and Public Health* 54:294-300.
- Genovese KJ, Anderson RC, Harvey RB, Callaway TR, Poole TL, Edrington TS, Fedorka-Cray PJ, Nisbet DJ. 2003. Competitive exclusion of *Salmonella* from the gut of neonatal and weaned pigs. *Journal of Food Protection* 66:1353-1359.
- Gradassi M, Pesciaroli M, Martinelli N, Ruggeri J, Petrucci P, Hassan WH, Raffatellu M, Scaglione FE, Ammendola S, Battistoni A, Alborali GL, Pasquali P. 2013. Attenuated *Salmonella enterica* serovar Typhimurium lacking the ZnuABC transporter: an efficacious orally-administered mucosal vaccine against salmonellosis in pigs. *Vaccine* 31:3695-3701.
- Gray JT, Stabel TJ, Fedorka-Cray PJ. 1996. Effect of dose on the immune response and persistence of *Salmonella choleraesuis* infection in swine. *American Journal of Veterinary Research* 57:313-319.
- Haesebrouck F, Pasmans F, Chiers K, Maes D, Ducatelle R, Decostere A. 2004. Efficacy of vaccines against bacterial diseases in swine: what can we expect? *Veterinary Microbiology* 100:255-268.
- Hohmann EL. 2001. Nontyphoidal salmonellosis. *Clinical Infectious Diseases* 32:263-269.
- Hotes S, Traulsen I, Krieter J. 2011. *Salmonella* control measures with special focus on vaccination and logistic slaughter procedures. *Transboundary and Emerging Diseases* 58:434-444.
- Huang TM, Lin TL, Wu CC. 2009. Serovar distribution and antimicrobial susceptibility of swine *Salmonella* isolates from clinically ill pigs in diagnostic submissions from Indiana in the United States. *Letters in Applied Microbiology* 48:331-336.

- Jenkins NL, Turner JL, Dritz SS, Durham SK, Minton JE. 2004. Changes in circulating insulin-like growth factor-I, insulin-like growth factor binding proteins, and leptin in weaned pigs infected with *Salmonella enterica* serovar Typhimurium. *Domestic Animal Endocrinology* 26:49-60.
- Korsak N, Degeye JN, Etienne G, China B, Daube G. 2004. Comparison of four different methods for *Salmonella* detection in fecal samples of porcine origin. *Journal of Food Protection* 67:2158-2164.
- Kramer TT, Roof MB, Matheson RR. 1992. Safety and efficacy of an attenuated strain of *Salmonella choleraesuis* for vaccination of swine. *American Journal of Veterinary Research* 53:444-448.
- Ku YW, McDonough SP, Palaniappan RU, Chang CF, Chang YF. 2005. Novel attenuated *Salmonella enterica* serovar Choleraesuis strains as live vaccine candidates generated by signature-tagged mutagenesis. *Infection and Immunity* 73:8194-8203.
- Kumar T, Rajora VR, Arora N. 2014. Prevalence of *Salmonella* in pigs and broilers in the Tarai region of Uttarakhand, India. *Indian Journal of Medical Microbiology* 32:99-101.
- Letellier A, Messier S, Lessard L, Quessy S. 2000. Assessment of various treatments to reduce carriage of *Salmonella* in swine. *Canadian Journal of Veterinary Research* 64:27-31.
- Leyman B, Boyen F, Van Parys A, Verbrugghe E, Haesebrouck F, Pasmans F. 2011. *Salmonella* Typhimurium LPS mutations for use in vaccines allowing differentiation of infected and vaccinated pigs. *Vaccine* 29:3679-3685.
- Lichtensteiger CA, Vimr ER. 2003. Systemic and enteric colonization of pigs by a *hilA* signature-tagged mutant of *Salmonella choleraesuis*. *Microbial Pathogenesis* 34:149-154.
- Lumsden JS, Wilkie BN. 1992. Immune response of pigs to parenteral vaccination with an aromatic-dependent mutant of *Salmonella typhimurium*. *Canadian Journal of Veterinary Research* 56:296-302.
- Martin-Pelaez S, Costabile A, Hoyles L, Rastall RA, Gibson GR, La Ragione RM, Woodward MJ, Mateu E, Martin-Orue SM. 2010. Evaluation of the inclusion of a mixture of organic acids or lactulose into the feed of pigs experimentally challenged with *Salmonella* Typhimurium. *Veterinary Microbiology* 142:337-345.
- Mastroeni P, Chabalgoity JA, Dunstan SJ, Maskell DJ, Dougan G. 2001. *Salmonella*: immune responses and vaccines. *Veterinary Journal* 161:132-164.
- Mejicanos G, Sanjayan N, Kim IH, Nyachoti CM. 2016. Recent advances in canola meal utilization in swine nutrition. *Journal of Animal Science and Technology* 58:7-016-0085-5. eCollection 2016.
- Meurens F, Berri M, Auray G, Melo S, Levast B, Virlogeux-Payant I, Chevaleyre C, Gerdts V, Salmon H. 2009. Early immune response following *Salmonella enterica* subspecies enterica serovar Typhimurium infection in porcine jejunal gut loops. *Veterinary Research* 40:5.
- Missotten JA, Michiels J, Degroote J, De Smet S. 2015. Fermented liquid feed for pigs: an ancient technique for the future. *Journal of Animal Science and Biotechnology* 6:4-1891-6-4. eCollection 2015.
- Mousing J, Jensen PT, Halgaard C, Bager F, Feld N, Nielsen B, Nielsen JP, Bech-Nielsen S. 1997. Nation-wide *Salmonella enterica* surveillance and control in Danish slaughter swine herds. *Preventive Veterinary Medicine* 29:247-261.
- Murray PR, Baron EJ, Tenover MA, Yolken RH. 1999. *Manual of clinical microbiology*. ASM Press, Washington, D.C.
- Nishio M, Okada N, Miki T, Haneda T, Danbara H. 2005. Identification of the outer-membrane protein PagC required for the serum resistance phenotype in *Salmonella enterica* serovar Choleraesuis. *Microbiology* 151:863-873.
- Nollet N, Maes D, Duchateau L, Hautekiet V, Houf K, Van Hoof J, De Zuttera L, De Kruif A, Geers R. 2005. Discrepancies between the isolation of *Salmonella* from mesenteric lymph nodes and the results of serological screening in slaughter pigs. *Veterinary Research* 36:545-555.
- Nye KJ, Fallon D, Frodsham D, Gee B, Graham C, Howe S, Messer S, Turner T, Warren RE. 2002. An evaluation of the performance

- of XLD, DCA, MLCB, and ABC agars as direct plating media for the isolation of *Salmonella enterica* from faeces. *Journal of Clinical Pathology* 55:286-288.
- Oliveira CJ, Garcia TB, Carvalho LF, Givisiez PE. 2007. Nose-to-nose transmission of *Salmonella* Typhimurium between weaned pigs. *Veterinary Microbiology* 125:355-361.
- Onyango DM, Ndeda VM, Wandili SA, Wawire SA, Ochieng P. 2014. Antimicrobial profile of *Salmonella enterica* serotype Choleraesuis from free-range swine in Kakamega fish market, western Kenya. *Journal of Infection in Developing Countries* 8:1381-1390.
- Padungtod P, Kadohira M, Hill G. 2008. Livestock production and foodborne diseases from food animals in Thailand. *The Journal of Veterinary Medical Science* 70:873-879.
- Pedersen K, Sorensen G, Lofstrom C, Leekitcharoenphon P, Nielsen B, Wingstrand A, Aarestrup FM, Hendriksen RS, Baggesen DL. 2015. Reappearance of *Salmonella serovar* Choleraesuis var. Kunzendorf in Danish pig herds. *Veterinary Microbiology* 176:282-291.
- Pesciaroli M, Gradassi M, Martinelli N, Ruggeri J, Pistoia C, Raffatellu M, Magistrali CF, Battistoni A, Pasquali P, Alborali GL. 2013. *Salmonella* Typhimurium lacking the Znuabc transporter is attenuated and immunogenic in pigs. *Vaccine* 31:2868-2873.
- Rajic A, Waddell LA, Sargeant JM, Read S, Farber J, Firth MJ, Chambers A. 2007. An overview of microbial food safety programs in beef, pork, and poultry from farm to processing in Canadian *Journal of Food Protection* 70:1286-1294.
- Reed WM, Olander HJ, Thacker HL. 1986. Studies on the pathogenesis of *Salmonella typhimurium* and *Salmonella choleraesuis* var kunzendorf infection in weanling pigs. *Veterinary Research American Journal of Veterinary Research* 47:75-83.
- Roesler U, Marg H, Schroder I, Mauer S, Arnold T, Lehmann J, Truyen U, Hensel A. 2004. Oral vaccination of pigs with an invasive gyrA-cpxA-rpoB *Salmonella* Typhimurium mutant. *Vaccine* 23:595-603.
- Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, Jones JL, Griffin PM, 2011. Foodborne illness acquired in the United States-major pathogens. *Emerging Infectious Diseases* 17:7-15.
- Schierack P, Nordhoff M, Pollmann M, Weyrauch KD, Amasheh S, Lodemann U, Jores J, Tachu B, Kleita S, Bliklager A, Tedin K, Wieler LH. 2006. Characterization of a porcine intestinal epithelial cell line for in vitro studies of microbial pathogenesis in swine. *Histochemistry and Cell Biology* 125:293-305.
- Schwarz P, Kich JD, Kolb J, Cardoso M. 2011. Use of an avirulent live *Salmonella* Choleraesuis vaccine to reduce the prevalence of *Salmonella* carrier pigs at slaughter. *The Veterinary Record* 169:553.
- Scott A, Zepeda C, Garber L, Smith J, Swayne D, Rhorer A, Kellar J, Shimshony A, Batho H, Caporale V, Giovannini A. 2006. The concept of compartmentalisation. *Revue Scientifique et Technique* 25:873-9, 881-7, 889-95.
- Selke M, Meens J, Springer S, Frank R, Gerlach GF. 2007. Immunization of pigs to prevent disease in humans: construction and protective efficacy of a *Salmonella enterica* serovar Typhimurium live negative-marker vaccine. *Infection and Immunity* 75:2476-2483.
- Tadee P, Kumpapong K, Sinthuya D, Yamsakul P, Chokesajjawatee N, Nuanualsuwan S, Pornsukarom S, Molla BZ, Gebreyes WA, Patchanee P. 2014. Distribution, quantitative load and characterization of *Salmonella* associated with swine farms in upper-northern Thailand. *Journal of Veterinary Science* 15:327-334.
- Tamang MD, Gurung M, Nam HM, Moon DC, Kim SR, Jang GC, Jung DY, Jung SC, Park YH, Lim SK. 2015. Prevalence and characterization of *Salmonella* in pigs from conventional and organic farms and first report of *S. serovar* 1,4,[5],12:i:- from Korea. *Veterinary Microbiology* 178:119-124.
- Tanaka T, Imai Y, Kumagae N, Sato S. 2010. The effect of feeding lactic acid to *Salmonella typhimurium* experimentally infected swine. *The Journal of Veterinary Medical Science* 72:827-831.

- Timoney JF, Gillespie JH, Scott FW, Barlough JE. 1988. Hagan and Bruner's microbiology and infectious diseases of domestic animals. Cornell University Press, Ithaca, New York.
- Turner JL, Dritz SS, Higgins JJ, Minton JE. 2002a. Effects of *Ascophyllum nodosum* extract on growth performance and immune function of young pigs challenged with *Salmonella typhimurium*. *Journal of Animal Science* 80:1947-1953.
- Turner JL, Dritz SS, Higgins JJ, Herkelman KL, Minton JE. 2002b. Effects of a Quillaja saponaria extract on growth performance and immune function of weanling pigs challenged with *Salmonella typhimurium*. *Journal of Animal Science* 80:1939-1946.
- Verbrugge E, Boyen F, Van Parys A, Van Deun K, Croubels S, Thompson A, Shearer N, Leyman B, Haesebrouck F, Pasmans F. 2011. Stress induced *Salmonella* Typhimurium recrudescence in pigs coincides with cortisol induced increased intracellular proliferation in macrophages. *Veterinary Research* 42:118-9716-42-118.
- Vigo GB, Cappuccio JA, Pineyro PE, Salve A, Machuca MA, Quiroga MA, Moredo F, Giacoboni G, Cancer JL, Caffer IG, Binsztein N, Pichel M, Perfumo CJ. 2009. *Salmonella enterica* subclinical infection: bacteriological, serological, pulsed-field gel electrophoresis, and antimicrobial resistance profiles--longitudinal study in a three-site farrow-to-finish farm. *Foodborne Pathogens and Disease* 6:965-972.
- Visscher CF, Klein G, Verspohl J, Beyerbach M, Stratmann-Selke J, Kamphues J. 2011. Serodiversity and serological as well as cultural distribution of *Salmonella* on farms and in abattoirs in Lower Saxony, Germany. *International Journal of Food Microbiology* 146:44-51.
- Wales A, Weaver J, McLaren IM, Smith RP, Mueller-Doblies D, Davies RH. 2013. Investigation of the distribution of *Salmonella* within an integrated pig breeding and production organisation in the United Kingdom. *ISRN Veterinary Science* 2013:943126.
- Wang J, Hu G, Lin Z, He L, Xu L, Zhang Y. 2014. Characteristic and functional analysis of a newly established porcine small intestinal epithelial cell line. *PLoS One* 9:e110916.
- Wilhelm B, Rajic A, Parker S, Waddell L, Sanchez J, Fazil A, Wilkins W, McEwen SA. 2012. Assessment of the efficacy and quality of evidence for five on-farm interventions for *Salmonella* reduction in grow-finish swine: a systematic review and meta-analysis. *Preventive Veterinary Medicine* 107:1-20.
- Wilkins EG, Roberts C. 1988. Extraintestinal salmonellosis. *Epidemiology and Infection* 100:361-368.
- Xu HR, Hsu HS, Moncure CW, King RA. 1993. Correlation of antibody titres induced by vaccination with protection in mouse typhoid. *Vaccine* 11:725-729.
- Xu XG, Zhao HN, Zhang Q, Ding L, Li ZC, Li W, Wu HY, Chuang KP, Tong DW, Liu HJ. 2012. Oral vaccination with attenuated *Salmonella enterica* serovar Typhimurium expressing Cap protein of PCV2 and its immunogenicity in mouse and swine models. *Veterinary Microbiology* 157:294-303.
- Yamane Y, Leonard JD, Kobatake R, Awamura N, Toyota Y, Ohta H, Otsuki K, Inoue T. 2000. A case study on *Salmonella enteritidis* (SE) origin at three egg-laying farms and its control with an *S. enteritidis* bacterin. *Avian Diseases* 44:519-526.
- Yin F, Farzan A, Wang QC, Yu H, Yin Y, Hou Y, Friendship R, Gong J. 2014. Reduction of *Salmonella enterica* serovar typhimurium DT104 infection in experimentally challenged weaned pigs fed a lactobacillus-fermented feed. *Foodborne Pathogens and Disease* 11:628-634.
- Zhao S, McDermott PF, Friedman S, Abbott J, Ayers S, Glenn A, Hall-Robinson E, Hubert SK, Harbottle H, Walker RD, Chiller TM, White DG. 2006a. Antimicrobial resistance and genetic relatedness among *Salmonella* from retail foods of animal origin: NARMS retail meat surveillance. *Foodborne Pathogens and Disease* 3:106-117.
- Zhao SH, Kuhar D, Lunney JK, Dawson H, Guidry C, Uthe JJ, Bearson SM, Recknor J, Nettleton D, Tuggle CK. 2006b. Gene expression profiling in *Salmonella* Choleraesuis-infected porcine lung using a long oligonucleotide microarray. *Mammalian Genome : Official Journal of the International Mammalian Genome Society* 17:777-789.