

## Invited Mini Review

The role of insulin/IGF-1 signaling in the longevity of model invertebrates, *C. elegans* and *D. melanogaster*Ozlem Altintas<sup>1,#</sup>, Sangsoon Park<sup>2,#</sup> & Seung-Jae V. Lee<sup>1,2,3,\*</sup><sup>1</sup>School of Interdisciplinary Bioscience and Bioengineering, <sup>2</sup>Department of Life Sciences, and <sup>3</sup>Information Technology Convergence Engineering, Pohang University of Science and Technology, Pohang 37673, Korea

**Insulin/insulin-like growth factor (IGF)-1 signaling (IIS) pathway regulates aging in many organisms, ranging from simple invertebrates to mammals, including humans. Many seminal discoveries regarding the roles of IIS in aging and longevity have been made by using the roundworm *Caenorhabditis elegans* and the fruit fly *Drosophila melanogaster*. In this review, we describe the mechanisms by which various IIS components regulate aging in *C. elegans* and *D. melanogaster*. We also cover systemic and tissue-specific effects of the IIS components on the regulation of lifespan. We further discuss IIS-mediated physiological processes other than aging and their effects on human disease models focusing on *C. elegans* studies. As both *C. elegans* and *D. melanogaster* have been essential for key findings regarding the effects of IIS on organismal aging in general, these invertebrate models will continue to serve as workhorses to help our understanding of mammalian aging. [BMB Reports 2016; 49(2): 81-92]**

## INTRODUCTION

The roundworm *Caenorhabditis elegans* and the fruit fly *Drosophila melanogaster* have been used as two most popular invertebrate models for studying aging and longevity (1, 2). In particular, their short lifespan together with their low cost and easy handling has established these invertebrates as excellent systems for research on molecular mechanisms regulating animal aging. Many important discoveries regarding evolutionarily conserved aging-regulatory pathways have been made using *C. elegans* and *D. melanogaster*. One of such pathways is the insulin/insulin-like growth factor (IGF)-1 signaling (IIS) pathway, which was first shown to regulate

longevity in *C. elegans*, and subsequently confirmed by using *D. melanogaster*. Importantly, the findings using these two invertebrate model organisms stimulated research on the role of IIS in mammalian aging, and led to discoveries showing that IIS also regulates aging in mammals, including mice and humans (3, 4). In this review, we will describe which components of IIS regulate lifespan, and how IIS modulates aging processes in these two model organisms. We will also review endocrine signaling and the importance of insulin-like peptides (ILPs) for systemic longevity regulation. Overall, our review will provide useful information regarding the conserved roles of IIS pathway in the aging of model organisms, which will eventually pave the way for understanding the mystery of human aging.

THE ROLE OF INSULIN/IGF-1 SIGNALING IN *C. elegans* AGINGInsulin/IGF-1 signaling pathway components that regulate the lifespan of *C. elegans*

The insulin/IGF-1 signaling (IIS) pathway contains many evolutionarily conserved components that regulate aging (Fig. 1). The gerontogenes *daf-2* and *age-1* encode the sole insulin/IGF-1 receptor and phosphatidylinositol-3-OH kinase (PI3K) (5, 6), respectively. DAF-2 and AGE-1 are two key upstream components of IIS that regulate various physiological aspects, including aging and adult lifespan. Two of the most important discoveries in the field of aging research were perhaps the findings demonstrating that inhibition of *daf-2* or *age-1* dramatically extended lifespan in *C. elegans* (7-9). These discoveries stimulated many subsequent studies on the role of IIS in lifespan regulation, not only in *C. elegans* but also in *D. melanogaster* and mammals.

IIS transduces signals through a combination of well-organized sequential events, depending on environmental conditions. Under favorable conditions, IIS is activated and this confers normal development and adult lifespan. Specifically, agonist insulin-like peptides (ILPs) bind to their receptor, DAF-2, which in turn recruits an insulin receptor substrate (IRS)/IST-1 (10). This leads to the activation of the AGE-1/PI3K, which increases the level of phosphatidylinositol (3,4,5)-trisphosphate (PIP<sub>3</sub>) (5, 11); this event is antagonistically balanced by DAF-18/PTEN phosphatase that promotes the conversion of PIP<sub>3</sub>

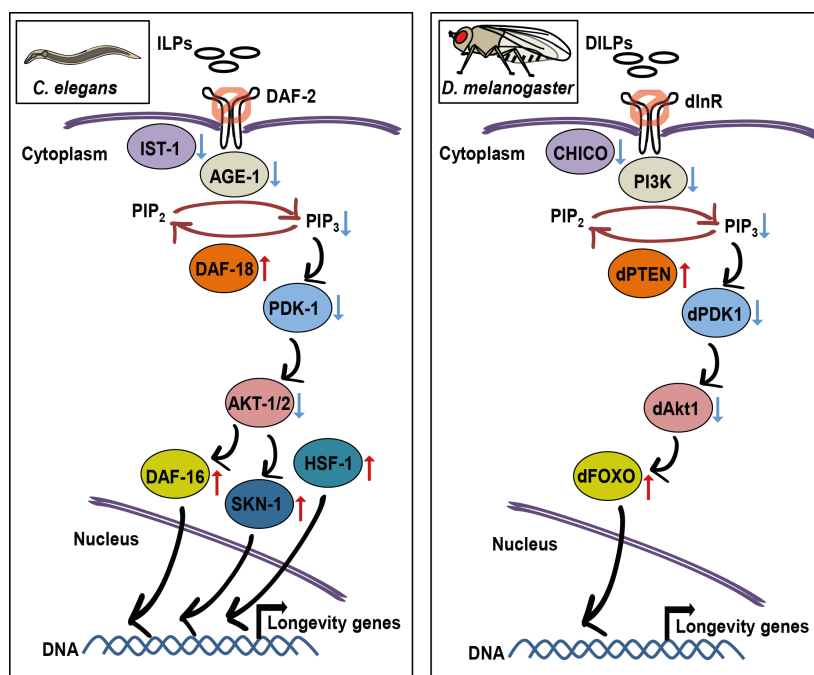
\*Corresponding author. Tel: +82-54-279-2351; Fax: +82-54-279-2199; E-mail: seungjaelee@postech.ac.kr

#These authors contributed equally to this work.

<http://dx.doi.org/10.5483/BMBRep.2016.49.2.261>

Received 18 December 2015

**Keywords:** Aging, *C. elegans*, *D. melanogaster*, Insulin/IGF-1 signaling, Longevity



**Fig. 1.** Conserved longevity-regulatory components of insulin/IGF-1 signaling pathway in *C. elegans* and *D. melanogaster*. Insulin-like peptides (ILPs in *Caenorhabditis elegans* and DILPs in *Drosophila melanogaster*) bind to insulin/IGF-1 receptor (DAF-2 in *C. elegans* and dInR in *D. melanogaster*) and lead to its phosphorylation. Inhibition of insulin/IGF-1 receptor results in decreased binding to the insulin receptor substrate (IST-1 in *C. elegans* and CHICO in *D. melanogaster*), which in turn decreases the activity of phosphoinositide-3 kinase (AGE-1 in *C. elegans* and PI3K in *D. melanogaster*) that converts PIP<sub>2</sub> to PIP<sub>3</sub>; conversely, the PTEN phosphatase (DAF-18 in *C. elegans* and dPTEN in *D. melanogaster*) functions to antagonize the activity of the phosphoinositide-3 kinase by converting PIP<sub>3</sub> to PIP<sub>2</sub>. Decreased PIP<sub>3</sub> levels lead to decreased activities of phosphoinositide-dependent kinase 1 (PDK-1 in *C. elegans* and dPDK1 in *D. melanogaster*) and the serine/threonine-specific protein kinase B (AKT-1/2 in *C. elegans* and dAkt1 in *D. melanogaster*), and the activation of downstream transcription factor FOXO (DAF-16 in *C. elegans* and dFOXO in *D. melanogaster*). Reduced insulin/IGF-1 signaling in *C. elegans* also increases the activities of heat shock transcription factor-1 (HSF-1) and SKN-1 (NRF2). These transcription factors regulate the expression of target genes, which contribute to longevity.

to phosphatidylinositol (4,5)-bisphosphate (PIP<sub>2</sub>) (12-19). The signals provided by PIP<sub>3</sub> activate the downstream kinase cascade, composed of 3-phosphoinositide-dependent protein kinase 1 (PDK-1) (20), protein kinase B (AKT-1/2) (21), and serum- and glucocorticoid-inducible kinase-1 (SGK-1) (22; but see also 23, 24). This in turn phosphorylates and inactivates DAF-16/FOXO transcription factor, by promoting its nucleus-to-cytosol translocation (22, 25-30). Conversely, in unfavorable conditions, IIS is down-regulated and leads to the activation of DAF-16/FOXO via enhancing its translocation from the cytoplasm to the nucleus, where it switches on the expression of genes that promote longevity. Thus, *C. elegans* IIS pathway acts as a system in which many components transduce signals to modulate the aging processes, depending on extracellular conditions.

Three most important downstream lifespan-regulatory transcription factors of IIS that have been identified so far are DAF-16/FOXO, heat shock transcription factor 1 (HSF-1) and SKN-1/nuclear factor erythroid 2 (NRF2). DAF-16/FOXO

regulates aging processes downstream of the canonical IIS cascade as described above. In addition, Jun-N-terminal kinase (JNK/JNK-1) (31), AMP-activated protein kinase (AMPK/AAK-2) (32-34), and Ste20-like protein kinase (MST1/CST-1) (35) activate DAF-16/FOXO via phosphorylation. Other non-kinase proteins have been shown to regulate *C. elegans* DAF-16/FOXO. A serine/threonine-protein phosphatase 4-regulatory subunit SMK-1 (36), and an RNA helicase HEL-1 (37), extend longevity by acting together with DAF-16/FOXO. DAF-16/FOXO is acetylated by an acetyl-transferase CBP-1/CREB binding protein (CBP), whose inhibition leads to constitutive nuclear localization of DAF-16/FOXO (38). Host cell factor 1 (HCF-1) and enhancer of *akt-1* null 7 (EAK-7) are other regulatory factors that inhibit DAF-16/FOXO activity without altering its subcellular localization (39-41). DAF-16/FOXO also interacts with two highly homologous 14-3-3 protein family members, FTT-1/PAR-5 and FTT-2 (42, 43). The 14-3-3 proteins modulate the interaction between DAF-16/FOXO and other co-factors, such as SIR-2.1/sirtuin 1, an NAD-dependent

deacetylase (42, 44). These diverse interactions and post-translational modifications may help differentially regulate the activity of DAF-16/FOXO upon various environmental changes.

Downstream targets of DAF-16/FOXO were identified by using various approaches such as chromatin immunoprecipitation, bioinformatics, microarray and mRNA sequencing (31, 45-51). The DAF-16/FOXO target genes collectively contribute to longevity by enhancing cellular maintenance in animals with reduced IIS. Since many regulatory modes and targets of FOXO transcription factors are conserved among species, the longevity-regulatory modes of *C. elegans* DAF-16/FOXO are likely to be recapitulated in IIS-mediated longevity in mammals.

SKN-1, an oxidative stress-responsive NRF transcription factor, also contributes to the longevity conferred by reduced IIS (52, 53). Similar to DAF-16/FOXO, SKN-1 is sequestered in the cytoplasm by phosphorylation via the canonical IIS protein kinases, including AKT-1/-2 (52). SKN-1 mediates the expression of genes involved in detoxification and stress responses (52, 54-63). Overexpression of constitutively nuclear SKN-1 extends lifespan in a DAF-16/FOXO-independent manner (52). SKN-1 also promotes protein homeostasis through regulating proteasome production, which contributes to a longer lifespan (55, 57, 64). In addition, SKN-1 promotes longevity of animals with reduced IIS through remodeling of extracellular matrix (65).

HSF-1 is another important transcription factor acting downstream of IIS, and is essential for the longevity of animals with reduced IIS (66-70). The function of HSF-1 in promoting longevity and reducing proteotoxicity is closely associated with the conserved IIS pathway. Genetic inhibition of *hsf-1* accelerates tissue aging, thereby shortening the lifespan (71). Knockdown of *hsf-1* also suppresses the longevity phenotype of *daf-2* and *age-1* mutants; conversely, the overexpression of *hsf-1* is sufficient to extend lifespan (51, 66, 67, 69, 72, 73). HSF-1 binds to specific regions of DNA containing heat shock elements (HSEs) (74-76). The binding of HSF-1 to HSEs triggers the induction of genes encoding molecular chaperones, such as HSP-70 and HSP-16, whose overexpression extends lifespan (77, 78). Thus, HSF-1 appears to lead to longevity by up-regulating the chaperone network that enhances the proper folding of various proteins (66, 67). DDL-1 (the *C. elegans* homolog of human coiled-coil domain-containing protein 53: CCDC53), DDL-2 (the *C. elegans* homolog of human Wiskott-Aldrich syndrome protein and SCAR homolog: WASH2), and HSB-1 (heat-shock factor binding protein-1), form a complex with HSF-1 and regulate lifespan by inhibiting the activity of HSF-1 (69). Overall, HSF-1 and SKN-1 appear to promote longevity mainly through the induction of target genes that increase resistance to various stresses.

### Systemic regulation of insulin/IGF-1 signaling-mediated longevity in *C. elegans*

As the IIS pathway consists of many potential endocrine components, it is likely that IIS regulates lifespan in a systemic manner. The *C. elegans* genome encodes 40 ILPs, which

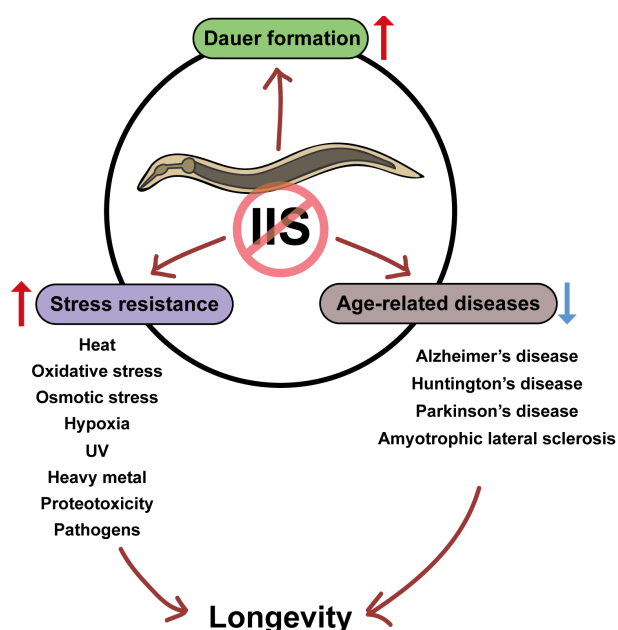
appear to act as extracellular endocrine signals in *C. elegans* (79, 80). Functional studies on several ILPs, including *ins-6* (81, 82), *ins-7* (47, 83, 84) and *daf-28* (80, 85, 86), have been conducted. However, the majority of the 40 ILPs, which potentially regulate longevity and development, are yet to be characterized in detail. This is perhaps because many possible combinations of the interactions between ILPs and DAF-2/insulin/IGF-1 receptor make it difficult to dissect the specific functions of each ILP. A recent study indicates that ILPs can function in a combinatorial manner to coordinate various physiological processes (87). This finding is different from the previous notion that ILPs generally confer a functional redundancy due to their structural similarities (79, 80, 88-91). Therefore, some individuals or a group of ILPs may have a profound effect on longevity.

Most of the ILPs are expressed in neurons, although some ILPs are expressed in non-neuronal tissues such as hypodermis and intestine (79-83, 88-90, 92-94). Overexpression of *ins-7* in the intestine decreases the activity of DAF-16/FOXO in non-intestinal tissues and shortens lifespan (83), suggesting an endocrine tissue-nonautonomous role of INS-7 in longevity. The expression of *daf-2* in neurons is largely responsible for the longevity of *daf-2* mutants (95, 96), pointing to the endocrine regulation of longevity by neuronal IIS. Together, it appears that IIS can systemically regulate lifespan from various tissues, via endocrine signaling.

### The role of insulin/IGF-1 signaling in *C. elegans* physiology and age-related disease models

*C. elegans* that is exposed to unfavorable environmental conditions such as reduced food availability, extreme temperatures and a high population during development, enters an alternative diapause stage called dauer (97, 98). IIS is one of extensively studied signaling pathways that govern this dauer developmental decision (Fig. 2). Genetic inhibition of *daf-2* or *age-1*, which extends adult lifespan, can cause constitutive dauer formation even under favorable conditions (97, 98). This dauer formation requires key downstream effectors in IIS, including DAF-18/PTEN and DAF-16/FOXO (97, 98). Reduced IIS activates a transcriptional program through DAF-16/FOXO, which leads to dauer formation. These findings raise the possibility that IIS may regulate dauer decision and lifespan using same effectors. However, the regulation of longevity and dauer formation by IIS can be uncoupled. Neuronal DAF-16/FOXO plays a more important role in the dauer decision than in lifespan regulation, whereas intestinal DAF-16/FOXO has a more profound effect on the lifespan extension than on the dauer decision (99). In addition, IIS pathway regulates the lifespan exclusively during adulthood, while it regulates the dauer formation during early larval development (100). Thus, spatiotemporal regulation of IIS differentially influences two separate aspects of animal physiology, development and adult lifespan.

IIS also regulates resistance to a variety of stresses. *C. elegans*



**Fig. 2.** The role of insulin/IGF-1 signaling in *C. elegans* physiology and age-related disease models. Insulin/IGF-1 signaling (IIS) regulates dauer formation, stress resistance, and the models of age-related diseases in *C. elegans*. Reduced IIS promotes dauer formation and enhances resistance to various external and internal stresses, and pathogens. Inhibition of IIS also ameliorates defects associated with various human disease models. These protective effects of reduced IIS contribute to organismal longevity.

with reduced IIS displays enhanced resistance to environmental stresses such as oxidative stress (Fig. 2) (52, 101-103), heat stress (104-106), hypoxic stress (107, 108), osmotic stress (109, 110), ultraviolet (UV) stress (36, 111), and heavy metal toxicity (112). Moreover, reduced IIS promotes better maintenance of internal homeostasis against cytosolic proteotoxicity (66, 113) and endoplasmic reticulum (ER) stress (114). The key downstream transcription factors of IIS that contribute to longevity, including DAF-16/FOXO (25, 26, 36, 66, 103, 106-116), HSF-1 (66, 67) and SKN-1 (52, 53), regulate these stress resistance phenotypes as well. Thus, proper regulation of IIS is crucial for the protection of *C. elegans* from both external and internal stresses.

Bacteria serve as a major food source for *C. elegans*, and are likely to be abundant in the natural habitats of *C. elegans*, such as rotten fruits. Therefore, it seems likely that *C. elegans* constantly comes in contact with various bacterial species, which may include pathogenic bacteria. To combat infection by pathogens, *C. elegans* is equipped with an innate immune system, and IIS is one of the most prominent innate immune signaling pathways (117). *C. elegans* with reduced IIS displays enhanced pathogen resistance, which is mediated by DAF-16/FOXO, HSF-1, and SKN-1 (84, 118-122). Reduced IIS leads to the induction of several antimicrobial genes (47), and reduction in bacterial packing in the intestine (123).

Interestingly, *Pseudomonas aeruginosa*, a popular model bacterial pathogen in *C. elegans*, activates IIS to counteract the host immunity (124). Therefore, IIS may be located at the front of constant battles between the host *C. elegans* and its bacterial pathogens.

Because of its powerful genetics, *C. elegans* has also been widely used for modeling various human diseases, especially neurodegenerative diseases. The disease models of *C. elegans* were established by generating transgenic animals expressing various human disease-associated proteins; these include  $\beta$ -amyloid peptides ( $A\beta$ ) for Alzheimer's disease (125-127), polyglutamine (polyQ) proteins for Huntington's disease (128-132),  $\alpha$ -synuclein for Parkinson's disease (133-137), and a mutant superoxide dismutase 1 (SOD1) for amyotrophic lateral sclerosis (ALS) (138-141). The Alzheimer's disease model *C. elegans*, which expresses  $A\beta_{1-42}$  in body wall muscles, is paralyzed and displays the accumulation of protein aggregates (68, 125, 142). Reduced IIS relieves these phenotypes via activating DAF-16/FOXO and HSF-1 (68), and inducing autophagic degradation of the protein aggregates (142). Reduced IIS also suppresses the short lifespan of  $A\beta_{1-42}$ -expressing animals (68). The *C. elegans* model for Huntington's disease has been widely used for studying proteotoxicity caused by aggregation of polyQ proteins (113, 128-131, 143-150). The polyQ-expressing worms display progressive neurodegeneration, neuronal dysfunction, retarded development, and defective motility (113, 128-131, 143-150). The *daf-2* and *age-1* mutations ameliorate a gradual age-dependent increase in toxicity resulting from polyQ aggregation through HSF-1 and DAF-16/FOXO (66, 113, 132, 146, 149). Parkinson's disease patients suffer from degeneration of dopaminergic neurons, which display accumulated protein inclusions that contain  $\alpha$ -synuclein (151). Similarly, the *C. elegans* models for Parkinson's disease, which express wild-type or mutant human  $\alpha$ -synuclein proteins in neurons, display the loss of dopaminergic neurons (133, 134, 137, 152). Reduced IIS by *daf-2* mutations dramatically suppresses this neurodegeneration phenotype (152). ALS, which is characterized by progressive motor neuron degeneration (153), has also been studied using a *C. elegans* model (138-141). Familial ALS is associated with mutations in the gene encoding SOD1 (154, 155). Neuronal expression of a mutant human SOD1 causes locomotion defects (140) and paralysis (141) in *C. elegans*. *daf-2* mutations protect the ALS model worms from the paralysis (141). Collectively, the results using *C. elegans* models indicate that IIS plays a crucial role in the pathophysiology of a majority of neurodegenerative diseases (Fig. 2). These findings imply that IIS modulates protein homeostasis to regulate normal neuronal functions, which may be essential for a long and healthy life.

## INSULIN/IGF-1 SIGNALING PATHWAY AND *Drosophila melanogaster* AGING

### Insulin/IGF-1 signaling components implicated in the longevity of *D. melanogaster*

The IIS pathway of *Drosophila melanogaster* consists of many components (Fig. 1), including the insulin/IGF receptor (dInR), the insulin receptor substrate (CHICO), the phosphatidylinositol 3-kinase (PI3K) Dp110/p60, 3-phosphoinositide-dependent protein kinase 1 (dPDK1) and the protein kinase B (PKB), also known as dAkt1, and the transcription factor *Drosophila* FOXO (dFOXO) (156-171). The activation mechanism of the IIS pathway in *Drosophila* has substantial similarities to that in *C. elegans*. Basically, the activation of dInR leads to up-regulation of a cascade of intracellular phosphorylation events, subsequently leading to the phosphorylation of dFOXO protein (160-162). dInR conveys signals from *Drosophila* insulin-like peptides (DILPs), directly to PI3K or to CHICO, the insulin receptor substrate (156, 172). PI3K, which converts PIP<sub>2</sub> to PIP<sub>3</sub>, has a catalytic subunit, Dp110, and a regulatory subunit, Dp60 (158, 159). The action of PI3K is antagonized by the activity of dPTEN (173-175), which catalyzes PIP<sub>3</sub> to PIP<sub>2</sub>. PIP<sub>3</sub> acts as an intracellular second messenger that activates a cascade of protein kinases, including dPDK1 and PKB/dAkt, which subsequently lead to the phosphorylation and the nuclear exclusion of dFOXO (162, 170). Conversely, reduced IIS through *dInR* or *CHICO* mutations, or overexpression of *dPTEN*, causes the translocation of dFOXO from the cytoplasm to the nucleus, where it up-regulates genes involved in longevity and stress resistance (160, 162, 176, 177).

In *Drosophila*, the IIS pathway regulates various physiological processes, including lifespan, stress responses, growth and development. Genetic inhibition of negative regulators of dFOXO, including several DILPs (178, 179), dInR (180), the IRS/CHICO (181, 182), or 14-3-3 epsilon (183), extends the lifespan of *Drosophila*. Conversely, overexpression of antagonistic IIS regulators, such as dPTEN or dFOXO, also extends the lifespan and/or delay heart aging (177, 184-186; but see also 187). Overall, these findings using *Drosophila* have remarkable similarities with those of *C. elegans*, highlighting the evolutionarily conserved nature of lifespan regulation by IIS components.

### Endocrine regulation of lifespan by *Drosophila* insulin/IGF-1 signaling

The *Drosophila melanogaster* genome encodes eight DILPs (172, 188-190). The *dilp* genes display distinct temporal expression patterns. For example, *dilp2* is expressed from embryo to adult stages, whereas *dilp4* is expressed only during development prior to adulthood (172, 179, 191-195). In addition, the expression sites of the eight *dilp* genes are diverse (196). Notably, the major site of DILP production is the median neurosecretory cells (mNSCs) in the brain, also called insulin-producing cells (IPCs), where *dilp1*, *dilp2*, *dilp3* and

*dilp5* are expressed (172, 179, 191-194).

Cell non-autonomous regulation of lifespan by *Drosophila* IIS was proposed based on findings using tissue-specific overexpression of IIS components. Up-regulation of dFOXO in the adult head fat body is sufficient to promote longevity and oxidative stress resistance (177). Muscle-specific overexpression of either dPTEN, dFOXO, or 4E-BP (a dFOXO target), also significantly increases lifespan (184). The ablation of IPCs lengthens lifespan (179, 197), which corroborates the endocrine regulation of lifespan by IIS; this is also reminiscent of lifespan extension by the ablation of sensory neurons in *C. elegans* (reviewed in 198). Among the DILPs expressed in the IPCs, DILP2 has been extensively explored for its implication in lifespan regulation, since it has the highest homology with human insulin (172, 177-179, 199-201). The *dilp2* null-mutant flies live long (178), and the expression of *dilp2* in the IPCs is reduced by the activation of dFOXO (177, 199). Therefore, *Drosophila* IIS appears to regulate the expression of DILPs and longevity via a feedback mechanism. Furthermore, reduced *dilp2* expression and inhibited IIS in the fat body are associated with lifespan extension conferred by transgenic expression of a dominant-negative p53 (200). DILP6, which is predominantly produced in the fat body, is another endocrine lifespan regulator (202-204). Surprisingly, overexpression of *dilp6* in the abdominal fat body leads to the repression of *dilp2* in the brain, suggesting synergistic effects on lifespan regulation by a potential dInR antagonist DILP6 and an agonist DILP2 (204). Collectively, lifespan regulation by IIS is controlled systemically by the action of DILPs that transmit signals, at least between the brain and fat body.

## CONCLUSIONS

In this review, we have described findings regarding mechanisms by which IIS influences lifespan in two representative invertebrate models, *C. elegans* and *D. melanogaster*. The roles of many IIS components in aging are remarkably well conserved between *C. elegans* and *D. melanogaster*, and the intervention of the IIS leads to an extended lifespan in both animals. This suggests that the role of IIS in aging is likely to be conserved across phyla beyond these two species. Indeed, many findings using these invertebrate models have led to the discoveries demonstrating that changes in IIS can extend lifespan in mammals. For example, heterozygous IGF1 receptor-knockout mice (*Igf1r*<sup>+/-</sup>) live longer than wild-type (205), and lower circulating IGF1 level correlates with mouse longevity (206). In addition, genetic variants of IIS components, including IGF1 receptor and FOXO3A, are associated with human longevity (4, 207). Thus, the evidence for the evolutionarily conserved nature of IIS-mediated longevity is extremely strong, ranging from invertebrates to humans.

Both *C. elegans* and *D. melanogaster* have been invaluable for the identification of IIS components and their roles in aging at the organism level. Still, much remains to be discovered

regarding the regulatory mechanisms of aging and longevity at the molecular level. As we can make new discoveries regarding organismal aging using these invertebrate models much faster than using vertebrates, *C. elegans* and *D. melanogaster* will continue to serve as indispensable tools for broadening our knowledge in aging. The progresses made by using these invertebrate models will eventually lead to the promotion of long and healthy human lives, and the prevention of age-associated diseases.

## ACKNOWLEDGEMENTS

We thank the Lee laboratory members for critical comments on the manuscript. This work was supported by Basic Research Laboratory Grants NRF-2012R1A4A1028200 and NRF-2013R1A1A2014754 funded by the Korean Government (MSIP) through the National Research Foundation of Korea (NRF) to S-J.V.L.

## REFERENCES

1. Lee Y, An S, Artan M et al (2015) Genes and Pathways That Influence Longevity in *Caenorhabditis elegans*; in Aging Mechanisms, Mori N and Mook-Jung I (eds), 123-169, Springer Japan
2. Giannakou ME and Partridge L (2007) Role of insulin-like signalling in *Drosophila* lifespan. Trends Biochem Sci 32, 180-188
3. Fontana L, Partridge L and Longo VD (2010) Extending healthy life span—from yeast to humans. Science 328, 321-326
4. Kenyon CJ (2010) The genetics of ageing. Nature 464, 504-512
5. Morris JZ, Tissenbaum HA and Ruvkun G (1996) A phosphatidylinositol-3-OH kinase family member regulating longevity and diapause in *Caenorhabditis elegans*. Nature 382, 536-539
6. Kimura KD, Tissenbaum HA, Liu Y and Ruvkun G (1997) *daf-2*, an insulin receptor-like gene that regulates longevity and diapause in *Caenorhabditis elegans*. Science 277, 942-946
7. Friedman DB and Johnson TE (1988) A mutation in the *age-1* gene in *Caenorhabditis elegans* lengthens life and reduces hermaphrodite fertility. Genetics 118, 75-86
8. Kenyon C, Chang J, Gensch E, Rudner A and Tabtiang R (1993) A *C. elegans* mutant that lives twice as long as wild type. Nature 366, 461-464
9. Klass MR (1983) A method for the isolation of longevity mutants in the nematode *Caenorhabditis elegans* and initial results. Mech Ageing Dev 22, 279-286
10. Wolkow CA, Munoz MJ, Riddle DL and Ruvkun G (2002) Insulin receptor substrate and p53 orthologous adaptor proteins function in the *Caenorhabditis elegans* *daf-2*/insulin-like signaling pathway. J Biol Chem 277, 49591-49597
11. Zhou K, Pandol S, Bokoch G and Traynor-Kaplan AE (1998) Disruption of *Dictyostelium* PI3K genes reduces [32P]phosphatidylinositol 3,4 bisphosphate and [32P]phosphatidylinositol trisphosphate levels, alters F-actin distribution and impairs pinocytosis. J Cell Sci 111 (Pt 2), 283-294
12. Ogg S and Ruvkun G (1998) The *C. elegans* PTEN homolog, DAF-18, acts in the insulin receptor-like metabolic signaling pathway. Mol Cell 2, 887-893
13. Gil EB, Malone Link E, Liu LX, Johnson CD and Lees JA (1999) Regulation of the insulin-like developmental pathway of *Caenorhabditis elegans* by a homolog of the PTEN tumor suppressor gene. Proc Natl Acad Sci U S A 96, 2925-2930
14. Mihaylova VT, Borland CZ, Manjarrez L, Stern MJ and Sun H (1999) The PTEN tumor suppressor homolog in *Caenorhabditis elegans* regulates longevity and dauer formation in an insulin receptor-like signaling pathway. Proc Natl Acad Sci U S A 96, 7427-7432
15. Rouault JP, Kuwabara PE, Sinilnikova OM, Duret L, Thierry-Mieg D and Billaud M (1999) Regulation of dauer larva development in *Caenorhabditis elegans* by *daf-18*, a homologue of the tumour suppressor PTEN. Curr Biol 9, 329-332
16. Dorman JB, Albinder B, Shroyer T and Kenyon C (1995) The *age-1* and *daf-2* genes function in a common pathway to control the lifespan of *Caenorhabditis elegans*. Genetics 141, 1399-1406
17. Larsen PL, Albert PS and Riddle DL (1995) Genes that regulate both development and longevity in *Caenorhabditis elegans*. Genetics 139, 1567-1583
18. Gottlieb S and Ruvkun G (1994) *daf-2*, *daf-16* and *daf-23*: genetically interacting genes controlling Dauer formation in *Caenorhabditis elegans*. Genetics 137, 107-120
19. Solari F, Bourbon-Piffaut A, Masse I, Payrastre B, Chan AM and Billaud M (2005) The human tumour suppressor PTEN regulates longevity and dauer formation in *Caenorhabditis elegans*. Oncogene 24, 20-27
20. Paradis S, Ailion M, Toker A, Thomas JH and Ruvkun G (1999) A PDK1 homolog is necessary and sufficient to transduce AGE-1 PI3 kinase signals that regulate diapause in *Caenorhabditis elegans*. Genes Dev 13, 1438-1452
21. Paradis S and Ruvkun G (1998) *Caenorhabditis elegans* Akt/PKB transduces insulin receptor-like signals from AGE-1 PI3 kinase to the DAF-16 transcription factor. Genes Dev 12, 2488-2498
22. Hertweck M, Gobel C and Baumeister R (2004) *C. elegans* SGK-1 is the critical component in the Akt/PKB kinase complex to control stress response and life span. Dev Cell 6, 577-588
23. Chen AT, Guo C, Dumas KJ, Ashrafi K and Hu PJ (2013) Effects of *Caenorhabditis elegans* *sgk-1* mutations on lifespan, stress resistance, and DAF-16/FoxO regulation. Aging Cell 12, 932-940
24. Xiao R, Zhang B, Dong Y et al (2013) A genetic program promotes *C. elegans* longevity at cold temperatures via a thermosensitive TRP channel. Cell 152, 806-817
25. Henderson ST and Johnson TE (2001) *daf-16* integrates developmental and environmental inputs to mediate aging in the nematode *Caenorhabditis elegans*. Curr Biol

- 11, 1975-1980
26. Lee RY, Hensch J and Ruvkun G (2001) Regulation of *C. elegans* DAF-16 and its human ortholog FKHRL1 by the *daf-2* insulin-like signaling pathway. *Curr Biol* 11, 1950-1957
  27. Lin K, Hsin H, Libina N and Kenyon C (2001) Regulation of the *Caenorhabditis elegans* longevity protein DAF-16 by insulin/IGF-1 and germline signaling. *Nat Genet* 28, 139-145
  28. Cahill CM, Tzivion G, Nasrin N et al (2001) Phosphatidylinositol 3-kinase signaling inhibits DAF-16 DNA binding and function via 14-3-3-dependent and 14-3-3-independent pathways. *J Biol Chem* 276, 13402-13410
  29. Ogg S, Paradis S, Gottlieb S et al (1997) The Fork head transcription factor DAF-16 transduces insulin-like metabolic and longevity signals in *C. elegans*. *Nature* 389, 994-999
  30. Lin K, Dorman JB, Rodan A and Kenyon C (1997) *daf-16*: An HNF-3/forkhead family member that can function to double the life-span of *Caenorhabditis elegans*. *Science* 278, 1319-1322
  31. Oh SW, Mukhopadhyay A, Svrzikapa N, Jiang F, Davis RJ and Tissenbaum HA (2005) JNK regulates lifespan in *Caenorhabditis elegans* by modulating nuclear translocation of forkhead transcription factor/DAF-16. *Proc Natl Acad Sci U S A* 102, 4494-4499
  32. Curtis R, O'Connor G and DiStefano PS (2006) Aging networks in *Caenorhabditis elegans*: AMP-activated protein kinase (*aak-2*) links multiple aging and metabolism pathways. *Aging Cell* 5, 119-126
  33. Greer EL, Dowlathshahi D, Banko MR et al (2007) An AMPK-FOXO pathway mediates longevity induced by a novel method of dietary restriction in *C. elegans*. *Curr Biol* 17, 1646-1656
  34. Apfeld J, O'Connor G, McDonagh T, DiStefano PS and Curtis R (2004) The AMP-activated protein kinase AAK-2 links energy levels and insulin-like signals to lifespan in *C. elegans*. *Genes Dev* 18, 3004-3009
  35. Lehtinen MK, Yuan Z, Boag PR et al (2006) A conserved MST-FOXO signaling pathway mediates oxidative-stress responses and extends life span. *Cell* 125, 987-1001
  36. Wolff S, Ma H, Burch D, Maciel GA, Hunter T and Dillin A (2006) SMK-1, an essential regulator of DAF-16-mediated longevity. *Cell* 124, 1039-1053
  37. Seo M, Seo K, Hwang W et al (2015) RNA helicase HEL-1 promotes longevity by specifically activating DAF-16/FOXO transcription factor signaling in *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 112, E4246-E4255
  38. Chiang WC, Tishkoff DX, Yang B et al (2012) *C. elegans* SIRT6/7 homolog SIR-2.4 promotes DAF-16 relocalization and function during stress. *PLoS Genet* 8, e1002948
  39. Hu PJ, Xu J and Ruvkun G (2006) Two membrane-associated tyrosine phosphatase homologs potentiate *C. elegans* AKT-1/PKB signaling. *PLoS Genet* 2, e99
  40. Li J, Ebata A, Dong Y, Rizki G, Iwata T and Lee SS (2008) *Caenorhabditis elegans* HCF-1 functions in longevity maintenance as a DAF-16 regulator. *PLoS Biol* 6, e233
  41. Alam H, Williams TW, Dumas KJ et al (2010) EAK-7 controls development and life span by regulating nuclear DAF-16/FoxO activity. *Cell Metab* 12, 30-41
  42. Berdichevsky A, Viswanathan M, Horvitz HR and Guarente L (2006) *C. elegans* SIR-2.1 interacts with 14-3-3 proteins to activate DAF-16 and extend life span. *Cell* 125, 1165-1177
  43. Li J, Tewari M, Vidal M and Lee SS (2007) The 14-3-3 protein FTT-2 regulates DAF-16 in *Caenorhabditis elegans*. *Dev Biol* 301, 82-91
  44. Wang Y, Oh SW, Deplancke B, Luo J, Walhout AJ and Tissenbaum HA (2006) *C. elegans* 14-3-3 proteins regulate life span and interact with SIR-2.1 and DAF-16/FOXO. *Mech Ageing Dev* 127, 741-747
  45. Lee SS, Kennedy S, Tolonen AC and Ruvkun G (2003) DAF-16 target genes that control *C. elegans* life-span and metabolism. *Science* 300, 644-647
  46. Ookuma S, Fukuda M and Nishida E (2003) Identification of a DAF-16 transcriptional target gene, *scl-1*, that regulates longevity and stress resistance in *Caenorhabditis elegans*. *Curr Biol* 13, 427-431
  47. Murphy CT, McCarroll SA, Bargmann CI et al (2003) Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*. *Nature* 424, 277-283
  48. McElwee J, Bubbs K and Thomas JH (2003) Transcriptional outputs of the *Caenorhabditis elegans* forkhead protein DAF-16. *Aging Cell* 2, 111-121
  49. Golden TR and Melov S (2004) Microarray analysis of gene expression with age in individual nematodes. *Aging Cell* 3, 111-124
  50. Halaschek-Wiener J, Khattra JS, McKay S et al (2005) Analysis of long-lived *C. elegans* *daf-2* mutants using serial analysis of gene expression. *Genome Res* 15, 603-615
  51. Lee SJ, Murphy CT and Kenyon C (2009) Glucose shortens the life span of *C. elegans* by downregulating DAF-16/FOXO activity and aquaporin gene expression. *Cell Metab* 10, 379-391
  52. Tullet JM, Hertweck M, An JH et al (2008) Direct inhibition of the longevity-promoting factor SKN-1 by insulin-like signaling in *C. elegans*. *Cell* 132, 1025-1038
  53. An JH and Blackwell TK (2003) SKN-1 links *C. elegans* mesendodermal specification to a conserved oxidative stress response. *Genes Dev* 17, 1882-1893
  54. An JH, Vranas K, Lucke M et al (2005) Regulation of the *Caenorhabditis elegans* oxidative stress defense protein SKN-1 by glycogen synthase kinase-3. *Proc Natl Acad Sci U S A* 102, 16275-16280
  55. Kahn NW, Rea SL, Moyle S, Kell A and Johnson TE (2008) Proteasomal dysfunction activates the transcription factor SKN-1 and produces a selective oxidative-stress response in *Caenorhabditis elegans*. *Biochem J* 409, 205-213
  56. Oliveira RP, Porter Abate J, Dilks K et al (2009) Condition-adapted stress and longevity gene regulation by *Caenorhabditis elegans* SKN-1/Nrf. *Aging Cell* 8, 524-541
  57. Wang J, Robida-Stubbs S, Tullet JM, Rual JF, Vidal M and Blackwell TK (2010) RNAi screening implicates a SKN-1-dependent transcriptional response in stress resistance and longevity deriving from translation inhibition. *PLoS Genet* 6, e1001048

58. Staab TA, Griffen TC, Corcoran C, Evgrafov O, Knowles JA and Sieburth D (2013) The conserved SKN-1/Nrf2 stress response pathway regulates synaptic function in *Caenorhabditis elegans*. *PLoS Genet* 9, e1003354
59. Glover-Cutter KM, Lin S and Blackwell TK (2013) Integration of the unfolded protein and oxidative stress responses through SKN-1/Nrf. *PLoS Genet* 9, e1003701
60. Choe KP, Przybysz AJ and Strange K (2009) The WD40 repeat protein WDR-23 functions with the CUL4/DBP1 ubiquitin ligase to regulate nuclear abundance and activity of SKN-1 in *Caenorhabditis elegans*. *Mol Cell Biol* 29, 2704-2715
61. Park SK, Tedesco PM and Johnson TE (2009) Oxidative stress and longevity in *Caenorhabditis elegans* as mediated by SKN-1. *Aging Cell* 8, 258-269
62. Pang S, Lynn DA, Lo JY, Paek J and Curran SP (2014) SKN-1 and Nrf2 couples proline catabolism with lipid metabolism during nutrient deprivation. *Nat Commun* 5, 5048
63. Kell A, Ventura N, Kahn N and Johnson TE (2007) Activation of SKN-1 by novel kinases in *Caenorhabditis elegans*. *Free Radic Biol Med* 43, 1560-1566
64. Li X, Matilainen O, Jin C, Glover-Cutter KM, Holmberg CI and Blackwell TK (2011) Specific SKN-1/Nrf stress responses to perturbations in translation elongation and proteasome activity. *PLoS Genet* 7, e1002119
65. Ewald CY, Landis JN, Porter Abate J, Murphy CT and Blackwell TK (2015) Dauer-independent insulin/IGF-1 signalling implicates collagen remodelling in longevity. *Nature* 519, 97-101
66. Hsu AL, Murphy CT and Kenyon C (2003) Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science* 300, 1142-1145
67. Morley JF and Morimoto RI (2004) Regulation of longevity in *Caenorhabditis elegans* by heat shock factor and molecular chaperones. *Mol Biol Cell* 15, 657-664
68. Cohen E, Bieschke J, Perciavalle RM, Kelly JW and Dillin A (2006) Opposing activities protect against age-onset proteotoxicity. *Science* 313, 1604-1610
69. Chiang WC, Ching TT, Lee HC, Mousigian C and Hsu AL (2012) HSF-1 regulators DDL-1/2 link insulin-like signaling to heat-shock responses and modulation of longevity. *Cell* 148, 322-334
70. Seo K, Choi E, Lee D, Jeong DE, Jang SK and Lee SJ (2013) Heat shock factor 1 mediates the longevity conferred by inhibition of TOR and insulin/IGF-1 signaling pathways in *C. elegans*. *Aging Cell* 12, 1073-1081
71. Garigan D, Hsu AL, Fraser AG, Kamath RS, Ahringer J and Kenyon C (2002) Genetic analysis of tissue aging in *Caenorhabditis elegans*: a role for heat-shock factor and bacterial proliferation. *Genetics* 161, 1101-1112
72. Douglas PM, Baird NA, Simic MS et al (2015) Heterotypic signals from neural HSF-1 separate thermotolerance from longevity. *Cell Rep* 12, 1196-1204
73. Baird NA, Douglas PM, Simic MS et al (2014) HSF-1-mediated cytoskeletal integrity determines thermotolerance and life span. *Science* 346, 360-363
74. Amin J, Ananthan J and Voellmy R (1988) Key features of heat shock regulatory elements. *Mol Cell Biol* 8, 3761-3769
75. Kay RJ, Boissy RJ, Russnak RH and Candido EP (1986) Efficient transcription of a *Caenorhabditis elegans* heat shock gene pair in mouse fibroblasts is dependent on multiple promoter elements which can function bidirectionally. *Mol Cell Biol* 6, 3134-3143
76. Russnak RH and Candido EP (1985) Locus encoding a family of small heat shock genes in *Caenorhabditis elegans*: two genes duplicated to form a 3.8-kilobase inverted repeat. *Mol Cell Biol* 5, 1268-1278
77. Yokoyama K, Fukumoto K, Murakami T et al (2002) Extended longevity of *Caenorhabditis elegans* by knocking in extra copies of hsp70F, a homolog of mot-2 (mortalin)/mthsp70/Grp75. *FEBS Lett* 516, 53-57
78. Walker GA and Lithgow GJ (2003) Lifespan extension in *C. elegans* by a molecular chaperone dependent upon insulin-like signals. *Aging Cell* 2, 131-139
79. Pierce SB, Costa M, Wisotzkey R et al (2001) Regulation of DAF-2 receptor signaling by human insulin and *ins-1*, a member of the unusually large and diverse *C. elegans* insulin gene family. *Genes Dev* 15, 672-686
80. Li W, Kennedy SG and Ruvkun G (2003) *daf-28* encodes a *C. elegans* insulin superfamily member that is regulated by environmental cues and acts in the DAF-2 signaling pathway. *Genes Dev* 17, 844-858
81. Chen Z, Hendricks M, Cornils A, Maier W, Alcedo J and Zhang Y (2013) Two insulin-like peptides antagonistically regulate aversive olfactory learning in *C. elegans*. *Neuron* 77, 572-585
82. Cornils A, Gloeck M, Chen Z, Zhang Y and Alcedo J (2011) Specific insulin-like peptides encode sensory information to regulate distinct developmental processes. *Development* 138, 1183-1193
83. Murphy CT, Lee SJ and Kenyon C (2007) Tissue entrainment by feedback regulation of insulin gene expression in the endoderm of *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 104, 19046-19050
84. Kawli T and Tan MW (2008) Neuroendocrine signals modulate the innate immunity of *Caenorhabditis elegans* through insulin signaling. *Nat Immunol* 9, 1415-1424
85. Malone EA, Inoue T and Thomas JH (1996) Genetic analysis of the roles of *daf-28* and *age-1* in regulating *Caenorhabditis elegans* dauer formation. *Genetics* 143, 1193-1205
86. Malone EA and Thomas JH (1994) A screen for non-conditional dauer-constitutive mutations in *Caenorhabditis elegans*. *Genetics* 136, 879-886
87. Fernandes de Abreu DA, Caballero A, Fardel P et al (2014) An insulin-to-insulin regulatory network orchestrates phenotypic specificity in development and physiology. *PLoS Genet* 10, e1004225
88. Ritter AD, Shen Y, Fuxman Bass J et al (2013) Complex expression dynamics and robustness in *C. elegans* insulin networks. *Genome Res* 23, 954-965
89. Hung WL, Wang Y, Chitturi J and Zhen M (2014) A *Caenorhabditis elegans* developmental decision requires insulin signaling-mediated neuron-intestine communication. *Development* 141, 1767-1779
90. Chen Y and Baugh LR (2014) *Ins-4* and *daf-28* function redundantly to regulate *C. elegans* L1 arrest. *Dev Biol* 394, 314-326



91. Duret L, Guex N, Peitsch MC and Bairoch A (1998) New insulin-like proteins with atypical disulfide bond pattern characterized in *Caenorhabditis elegans* by comparative sequence analysis and homology modeling. *Genome Res* 8, 348-353
92. Michaelson D, Korta DZ, Capua Y and Hubbard EJ (2010) Insulin signaling promotes germline proliferation in *C. elegans*. *Development* 137, 671-680
93. Leinwand SG and Chalasani SH (2013) Neuropeptide signaling remodels chemosensory circuit composition in *Caenorhabditis elegans*. *Nat Neurosci* 16, 1461-1467
94. Ohta A, Ujisawa T, Sonoda S and Kuhara A (2014) Light and pheromone-sensing neurons regulates cold habituation through insulin signalling in *Caenorhabditis elegans*. *Nat Commun* 5, 4412
95. Wolkow CA, Kimura KD, Lee MS and Ruvkun G (2000) Regulation of *C. elegans* life-span by insulinlike signaling in the nervous system. *Science* 290, 147-150
96. Iser WB, Gami MS and Wolkow CA (2007) Insulin signaling in *Caenorhabditis elegans* regulates both endocrine-like and cell-autonomous outputs. *Dev Biol* 303, 434-447
97. Hu PJ (2007) Dauer. WormBook: the online review of *C. elegans* biology, 1-19
98. Riddle DL and Albert PS (1997) Genetic and Environmental Regulation of Dauer Larva Development; in *C. elegans* II, Riddle DL, Blumenthal T, Meyer BJ et al (eds), Cold Spring Harbor Laboratory Press, Cold Spring Harbor (NY)
99. Libina N, Berman JR and Kenyon C (2003) Tissue-specific activities of *C. elegans* DAF-16 in the regulation of lifespan. *Cell* 115, 489-502
100. Dillin A, Crawford DK and Kenyon C (2002) Timing requirements for insulin/IGF-1 signaling in *C. elegans*. *Science* 298, 830-834
101. Larsen PL (1993) Aging and resistance to oxidative damage in *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 90, 8905-8909
102. Vanfleteren JR (1993) Oxidative stress and ageing in *Caenorhabditis elegans*. *Biochem J* 292 (Pt 2), 605-608
103. Honda Y and Honda S (1999) The *daf-2* gene network for longevity regulates oxidative stress resistance and Mn-superoxide dismutase gene expression in *Caenorhabditis elegans*. *FASEB J* 13, 1385-1393
104. Gems D, Sutton AJ, Sundermeyer ML et al (1998) Two pleiotropic classes of *daf-2* mutation affect larval arrest, adult behavior, reproduction and longevity in *Caenorhabditis elegans*. *Genetics* 150, 129-155
105. Lithgow GJ, White TM, Melov S and Johnson TE (1995) Thermotolerance and extended life-span conferred by single-gene mutations and induced by thermal stress. *Proc Natl Acad Sci U S A* 92, 7540-7544
106. McColl G, Rogers AN, Alavez S et al (2010) Insulin-like signaling determines survival during stress via post-transcriptional mechanisms in *C. elegans*. *Cell Metab* 12, 260-272
107. Scott BA, Avidan MS and Crowder CM (2002) Regulation of hypoxic death in *C. elegans* by the insulin/IGF receptor homolog DAF-2. *Science* 296, 2388-2391
108. Mabon ME, Scott BA and Crowder CM (2009) Divergent mechanisms controlling hypoxic sensitivity and lifespan by the DAF-2/insulin/IGF-receptor pathway. *PLoS One* 4, e7937
109. Lamitina ST and Strange K (2005) Transcriptional targets of DAF-16 insulin signaling pathway protect *C. elegans* from extreme hypertonic stress. *Am J Physiol Cell Physiol* 288, C467-C474
110. Burkewitz K, Choe K and Strange K (2011) Hypertonic stress induces rapid and widespread protein damage in *C. elegans*. *Am J Physiol Cell Physiol* 301, C566-C576
111. Murakami S and Johnson TE (1996) A genetic pathway conferring life extension and resistance to UV stress in *Caenorhabditis elegans*. *Genetics* 143, 1207-1218
112. Barsyte D, Lovejoy DA and Lithgow GJ (2001) Longevity and heavy metal resistance in *daf-2* and *age-1* long-lived mutants of *Caenorhabditis elegans*. *FASEB J* 15, 627-634
113. Morley JF, Brignull HR, Weyers JJ and Morimoto RI (2002) The threshold for polyglutamine-expansion protein aggregation and cellular toxicity is dynamic and influenced by aging in *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 99, 10417-10422
114. Henis-Korenblit S, Zhang PC, Hansen M et al (2010) Insulin/IGF-1 signaling mutants reprogram ER stress response regulators to promote longevity. *Proc Natl Acad Sci U S A* 107, 9730-9735
115. Essers MA, de Vries-Smits LM, Barker N, Polderman PE, Burgering BM and Korswagen HC (2005) Functional interaction between beta-catenin and FOXO in oxidative stress signaling. *Science* 308, 1181-1184
116. Mueller MM, Castells-Roca L, Babu V et al (2014) DAF-16/FOXO and EGL-27/GATA promote developmental growth in response to persistent somatic DNA damage. *Nat Cell Biol* 16, 1168-1179
117. Ermolaeva MA and Schumacher B (2014) Insights from the worm: the *C. elegans* model for innate immunity. *Semin Immunol* 26, 303-309
118. Garsin DA, Villanueva JM, Begun J et al (2003) Long-lived *C. elegans daf-2* mutants are resistant to bacterial pathogens. *Science* 300, 1921
119. Kerry S, TeKippe M, Gaddis NC and Aballay A (2006) GATA transcription factor required for immunity to bacterial and fungal pathogens. *PLoS One* 1, e77
120. Singh V and Aballay A (2006) Heat-shock transcription factor (HSF)-1 pathway required for *Caenorhabditis elegans* immunity. *Proc Natl Acad Sci U S A* 103, 13092-13097
121. Papp D, Csermely P and Soti C (2012) A role for SKN-1/Nrf in pathogen resistance and immunosenescence in *Caenorhabditis elegans*. *PLoS Pathog* 8, e1002673
122. Evans EA, Chen WC and Tan MW (2008) The DAF-2 insulin-like signaling pathway independently regulates aging and immunity in *C. elegans*. *Aging Cell* 7, 879-893
123. Portal-Celhay C, Bradley ER and Blaser MJ (2012) Control of intestinal bacterial proliferation in regulation of lifespan in *Caenorhabditis elegans*. *BMC Microbiol* 12, 49
124. Evans EA, Kawli T and Tan M-W (2008) *Pseudomonas aeruginosa* suppresses host immunity by activating the DAF-2 insulin-like signaling pathway in *Caenorhabditis*

- elegans*. PLoS Pathog 4, e1000175
125. Link CD (1995) Expression of human  $\beta$ -amyloid peptide in transgenic *Caenorhabditis elegans*. Proc Natl Acad Sci U S A 92, 9368-9372
  126. Fay DS, Fluet A, Johnson CJ and Link CD (1998) In Vivo Aggregation of  $\beta$ -Amyloid Peptide Variants. J Neurochem 71, 1616-1625
  127. Link CD, Taft A, Kapulkin V et al (2003) Gene expression analysis in a transgenic *Caenorhabditis elegans* Alzheimer's disease model. Neurobiol Aging 24, 397-413
  128. Faber PW, Alter JR, MacDonald ME and Hart AC (1999) Polyglutamine-mediated dysfunction and apoptotic death of a *Caenorhabditis elegans* sensory neuron. Proc Natl Acad Sci U S A 96, 179-184
  129. Satyal SH, Schmidt E, Kitagawa K et al (2000) Polyglutamine aggregates alter protein folding homeostasis in *Caenorhabditis elegans*. Proc Natl Acad Sci U S A 97, 5750-5755
  130. Parker JA, Connolly JB, Wellington C, Hayden M, Dausset J and Neri C (2001) Expanded polyglutamines in *Caenorhabditis elegans* cause axonal abnormalities and severe dysfunction of PLM mechanosensory neurons without cell death. Proc Natl Acad Sci U S A 98, 13318-13323
  131. Brignull HR, Moore FE, Tang SJ and Morimoto RI (2006) Polyglutamine proteins at the pathogenic threshold display neuron-specific aggregation in a pan-neuronal *Caenorhabditis elegans* model. J Neurosci 26, 7597-7606
  132. Mohri-Shiomi A and Garsin DA (2008) Insulin signaling and the heat shock response modulate protein homeostasis in the *Caenorhabditis elegans* intestine during infection. J Biol Chem 283, 194-201
  133. Lakso M, Vartiainen S, Moilanen AM et al (2003) Dopaminergic neuronal loss and motor deficits in *Caenorhabditis elegans* overexpressing human alpha-synuclein. J Neurochem 86, 165-172
  134. Cao S, Gelwix CC, Caldwell KA and Caldwell GA (2005) Torsin-mediated protection from cellular stress in the dopaminergic neurons of *Caenorhabditis elegans*. J Neurosci 25, 3801-3812
  135. Kuwahara T, Koyama A, Gengyo-Ando K et al (2006) Familial Parkinson mutant  $\alpha$ -synuclein causes dopamine neuron dysfunction in transgenic *Caenorhabditis elegans*. J Biol Chem 281, 334-340
  136. Kuwahara T, Koyama A, Koyama S et al (2008) A systematic RNAi screen reveals involvement of endocytic pathway in neuronal dysfunction in  $\alpha$ -synuclein transgenic *C. elegans*. Hum Mol Genet 17, 2997-3009
  137. Hamamichi S, Rivas RN, Knight AL, Cao S, Caldwell KA and Caldwell GA (2008) Hypothesis-based RNAi screening identifies neuroprotective genes in a Parkinson's disease model. Proc Natl Acad Sci U S A 105, 728-733
  138. Oeda T, Shimohama S, Kitagawa N et al (2001) Oxidative stress causes abnormal accumulation of familial amyotrophic lateral sclerosis-related mutant SOD1 in transgenic *Caenorhabditis elegans*. Hum Mol Genet 10, 2013-2023
  139. Gidalevitz T, Krupinski T, Garcia S and Morimoto RI (2009) Destabilizing protein polymorphisms in the genetic background direct phenotypic expression of mutant SOD1 toxicity. PLoS Genet 5, e1000399
  140. Wang J, Farr GW, Hall DH et al (2009) An ALS-linked mutant SOD1 produces a locomotor defect associated with aggregation and synaptic dysfunction when expressed in neurons of *Caenorhabditis elegans*. PLoS Genet 5, e1000350
  141. Li J, Huang KX and Le WD (2013) Establishing a novel *C. elegans* model to investigate the role of autophagy in amyotrophic lateral sclerosis. Acta Pharmacol Sin 34, 644-650
  142. Florez-McClure ML, Hohsfield LA, Fonte G, Bealor MT and Link CD (2007) Decreased insulin-receptor signaling promotes the autophagic degradation of  $\beta$ -amyloid peptide in *C. elegans*. Autophagy 3, 569-580
  143. David DC, Ollikainen N, Trinidad JC, Cary MP, Burlingame AL and Kenyon C (2010) Widespread protein aggregation as an inherent part of aging in *C. elegans*. PLoS Biol 8, e1000450
  144. Wang H, Lim PJ, Yin C, Rieckher M, Vogel BE and Monteiro MJ (2006) Suppression of polyglutamine-induced toxicity in cell and animal models of Huntington's disease by ubiquitin. Hum Mol Genet 15, 1025-1041
  145. Wang H, Lim PJ, Karbowski M and Monteiro MJ (2009) Effects of overexpression of huntingtin proteins on mitochondrial integrity. Hum Mol Genet 18, 737-752
  146. Steinkraus KA, Smith ED, Davis C et al (2008) Dietary restriction suppresses proteotoxicity and enhances longevity by an *hsf-1*-dependent mechanism in *Caenorhabditis elegans*. Aging Cell 7, 394-404
  147. Faber PW, Voisine C, King DC, Bates EA and Hart AC (2002) Glutamine/proline-rich PQE-1 proteins protect *Caenorhabditis elegans* neurons from huntingtin polyglutamine neurotoxicity. Proc Natl Acad Sci U S A 99, 17131-17136
  148. Parker JA, Metzler M, Georgiou J et al (2007) Huntingtin-interacting protein 1 influences worm and mouse pre-synaptic function and protects *Caenorhabditis elegans* neurons against mutant polyglutamine toxicity. J Neurosci 27, 11056-11064
  149. Parker JA, Arango M, Abderrahmane S et al (2005) Resveratrol rescues mutant polyglutamine cytotoxicity in nematode and mammalian neurons. Nat Genet 37, 349-350
  150. Bates EA, Victor M, Jones AK, Shi Y and Hart AC (2006) Differential contributions of *Caenorhabditis elegans* histone deacetylases to huntingtin polyglutamine toxicity. J Neurosci 26, 2830-2838
  151. Schapira AH and Jenner P (2011) Etiology and pathogenesis of Parkinson's disease. Mov Disord 26, 1049-1055
  152. Knight AL, Yan X, Hamamichi S et al (2014) The glycolytic enzyme, GPI, is a functionally conserved modifier of dopaminergic neurodegeneration in Parkinson's models. Cell Metab 20, 145-157
  153. Pasinelli P and Brown RH (2006) Molecular biology of amyotrophic lateral sclerosis: insights from genetics. Nat Rev Neurosci 7, 710-723
  154. Rosen DR, Siddique T, Patterson D et al (1993) Mutations in Cu/Zn superoxide dismutase gene are associated

- with familial amyotrophic lateral sclerosis. *Nature* 362, 59-62
155. Cudkowicz M, McKenna-Yasek D, Sapp P et al (1997) Epidemiology of mutations in superoxide dismutase in amyotrophic lateral sclerosis. *Ann Neurol* 41, 210-221
  156. Bohni R, Riesgo-Escovar J, Oldham S et al (1999) Autonomous control of cell and organ size by CHICO, a *Drosophila* homolog of vertebrate IRS1-4. *Cell* 97, 865-875
  157. Verdu J, Buratovich MA, Wilder EL and Birnbaum MJ (1999) Cell-autonomous regulation of cell and organ growth in *Drosophila* by Akt/PKB. *Nat Cell Biol* 1, 500-506
  158. Weinkove D, Neufeld TP, Twardzik T, Waterfield MD and Leever SJ (1999) Regulation of imaginal disc cell size, cell number and organ size by *Drosophila* class I(A) phosphoinositide 3-kinase and its adaptor. *Curr Biol* 9, 1019-1029
  159. Leever SJ, Weinkove D, MacDougall LK, Hafen E and Waterfield MD (1996) The *Drosophila* phosphoinositide 3-kinase Dp110 promotes cell growth. *EMBO J* 15, 6584-6594
  160. Junger MA, Rintelen F, Stocker H et al (2003) The *Drosophila* forkhead transcription factor FOXO mediates the reduction in cell number associated with reduced insulin signaling. *J Biol* 2, 20
  161. Kramer JM, Davidge JT, Lockyer JM and Staveley BE (2003) Expression of *Drosophila* FOXO regulates growth and can phenocopy starvation. *BMC Dev Biol* 3, 5
  162. Puig O, Marr MT, Ruhf ML and Tjian R (2003) Control of cell number by *Drosophila* FOXO: downstream and feedback regulation of the insulin receptor pathway. *Genes Dev* 17, 2006-2020
  163. Staveley BE, Ruel L, Jin J et al (1998) Genetic analysis of protein kinase B (AKT) in *Drosophila*. *Curr Biol* 8, 599-602
  164. Poltilove RM, Jacobs AR, Haft CR, Xu P and Taylor SI (2000) Characterization of *Drosophila* insulin receptor substrate. *J Biol Chem* 275, 23346-23354
  165. Petruzzelli L, Herrera R, Arenas-Garcia R, Fernandez R, Birnbaum MJ and Rosen OM (1986) Isolation of a *Drosophila* genomic sequence homologous to the kinase domain of the human insulin receptor and detection of the phosphorylated *Drosophila* receptor with an anti-peptide antibody. *Proc Natl Acad Sci U S A* 83, 4710-4714
  166. Fernandez-Almonacid R and Rosen OM (1987) Structure and ligand specificity of the *Drosophila melanogaster* insulin receptor. *Mol Cell Biol* 7, 2718-2727
  167. Fernandez R, Tabarini D, Azpiazu N, Frasch M and Schlessinger J (1995) The *Drosophila* insulin receptor homolog: a gene essential for embryonic development encodes two receptor isoforms with different signaling potential. *EMBO J* 14, 3373-3384
  168. Marin-Hincapie M and Garofalo RS (1999) The carboxyl terminal extension of the *Drosophila* insulin receptor homologue binds IRS-1 and influences cell survival. *J Biol Chem* 274, 24987-24994
  169. Franke TF, Tartof KD and Tsichlis PN (1994) The SH2-like Akt homology (AH) domain of c-akt is present in multiple copies in the genome of vertebrate and invertebrate eucaryotes. Cloning and characterization of the *Drosophila melanogaster* c-akt homolog Dakt1. *Oncogene* 9, 141-148
  170. Cho KS, Lee JH, Kim S et al (2001) *Drosophila* phosphoinositide-dependent kinase-1 regulates apoptosis and growth via the phosphoinositide 3-kinase-dependent signaling pathway. *Proc Natl Acad Sci U S A* 98, 6144-6149
  171. Linossier C, MacDougall LK, Domin J and Waterfield MD (1997) Molecular cloning and biochemical characterization of a *Drosophila* phosphatidylinositol-specific phosphoinositide 3-kinase. *Biochem J* 321 (Pt 3), 849-856
  172. Brogiolo W, Stocker H, Ikeya T, Rintelen F, Fernandez R and Hafen E (2001) An evolutionarily conserved function of the *Drosophila* insulin receptor and insulin-like peptides in growth control. *Curr Biol* 11, 213-221
  173. Gao X, Neufeld TP and Pan D (2000) *Drosophila* PTEN regulates cell growth and proliferation through PI3K-dependent and -independent pathways. *Dev Biol* 221, 404-418
  174. Goberdhan DC, Paricio N, Goodman EC, Mlodzik M and Wilson C (1999) *Drosophila* tumor suppressor PTEN controls cell size and number by antagonizing the Chico/PI3-kinase signaling pathway. *Genes Dev* 13, 3244-3258
  175. Huang H, Potter CJ, Tao W et al (1999) PTEN affects cell size, cell proliferation and apoptosis during *Drosophila* eye development. *Development* 126, 5365-5372
  176. Bai H, Kang P, Hernandez AM and Tatar M (2013) Activin signaling targeted by insulin/dFOXO regulates aging and muscle proteostasis in *Drosophila*. *PLoS Genet* 9, e1003941
  177. Hwangbo DS, Gershman B, Tu MP, Palmer M and Tatar M (2004) *Drosophila* dFOXO controls lifespan and regulates insulin signalling in brain and fat body. *Nature* 429, 562-566
  178. Gronke S, Clarke DF, Broughton S, Andrews TD and Partridge L (2010) Molecular evolution and functional characterization of *Drosophila* insulin-like peptides. *PLoS Genet* 6, e1000857
  179. Broughton SJ, Piper MD, Ikeya T et al (2005) Longer lifespan, altered metabolism, and stress resistance in *Drosophila* from ablation of cells making insulin-like ligands. *Proc Natl Acad Sci U S A* 102, 3105-3110
  180. Tatar M, Kopelman A, Epstein D, Tu MP, Yin CM and Garofalo RS (2001) A mutant *Drosophila* insulin receptor homolog that extends life-span and impairs neuroendocrine function. *Science* 292, 107-110
  181. Clancy DJ, Gems D, Harshman LG et al (2001) Extension of life-span by loss of CHICO, a *Drosophila* insulin receptor substrate protein. *Science* 292, 104-106
  182. Tu MP, Epstein D and Tatar M (2002) The demography of slow aging in male and female *Drosophila* mutant for the insulin-receptor substrate homologue *chico*. *Aging Cell* 1, 75-80
  183. Nielsen MD, Luo X, Biteau B, Syverson K and Jasper H (2008) 14-3-3ε antagonizes FoxO to control growth, apoptosis and longevity in *Drosophila*. *Aging Cell* 7,

- 688-699
184. Demontis F and Perrimon N (2010) FOXO/4E-BP signaling in *Drosophila* muscles regulates organism-wide proteostasis during aging. *Cell* 143, 813-825
  185. Giannakou ME, Goss M, Junger MA, Hafen E, Leevers SJ and Partridge L (2004) Long-lived *Drosophila* with over-expressed dFOXO in adult fat body. *Science* 305, 361
  186. Wessells RJ, Fitzgerald E, Cypser JR, Tatar M and Bodmer R (2004) Insulin regulation of heart function in aging fruit flies. *Nat Genet* 36, 1275-1281
  187. Ford D, Hoe N, Landis GN et al (2007) Alteration of *Drosophila* life span using conditional, tissue-specific expression of transgenes triggered by doxycycline or RU486/Mifepristone. *Exp Gerontol* 42, 483-497
  188. Broeck JV (2001) Neuropeptides and their precursors in the fruitfly, *Drosophila melanogaster*. *Peptides* 22, 241-254
  189. Colombani J, Andersen DS and Léopold P (2012) Secreted peptide Dilp8 coordinates *Drosophila* tissue growth with developmental timing. *Science* 336, 582-585
  190. Garelli A, Gontijo AM, Miguela V, Caparros E and Dominguez M (2012) Imaginal discs secrete insulin-like peptide 8 to mediate plasticity of growth and maturation. *Science* 336, 579-582
  191. Cao C and Brown MR (2001) Localization of an insulin-like peptide in brains of two flies. *Cell Tissue Res* 304, 317-321
  192. Ikeya T, Galic M, Belawat P, Nairz K and Hafen E (2002) Nutrient-dependent expression of insulin-like peptides from neuroendocrine cells in the CNS contributes to growth regulation in *Drosophila*. *Curr Biol* 12, 1293-1300
  193. Rulifson EJ, Kim SK and Nusse R (2002) Ablation of insulin-producing neurons in flies: growth and diabetic phenotypes. *Science* 296, 1118-1120
  194. Lee KS, Kwon OY, Lee JH et al (2008) *Drosophila* short neuropeptide F signalling regulates growth by ERK-mediated insulin signalling. *Nat Cell Biol* 10, 468-475
  195. Grönke S and Partridge L (2010) The functions of insulin-like peptides in insects; in IGFs: Local Repair and Survival Factors Throughout Life Span, Clemmons DR, Robinson I and Christen Y (eds), 105-124, Springer
  196. Nassel DR, Liu Y and Luo J (2015) Insulin/IGF signaling and its regulation in *Drosophila*. *Gen Comp Endocrinol* 221, 255-266
  197. Haselton A, Sharmin E, Schrader J, Sah M, Poon P and Fridell Y-WC (2010) Partial ablation of adult *Drosophila* insulin-producing neurons modulates glucose homeostasis and extends life span without insulin resistance. *Cell Cycle* 9, 3135-3143
  198. Jeong DE, Artan M, Seo K and Lee SJ (2012) Regulation of lifespan by chemosensory and thermosensory systems: findings in invertebrates and their implications in mammalian aging. *Front Genet* 3, 218
  199. Wang MC, Bohmann D and Jasper H (2005) JNK extends life span and limits growth by antagonizing cellular and organism-wide responses to insulin signaling. *Cell* 121, 115-125
  200. Bauer JH, Chang C, Morris SNS et al (2007) Expression of dominant-negative Dmp53 in the adult fly brain inhibits insulin signaling. *Proc Natl Acad Sci U S A* 104, 13355-13360
  201. Broughton S, Alic N, Slack C et al (2008) Reduction of DILP2 in *Drosophila* triages a metabolic phenotype from lifespan revealing redundancy and compensation among DILPs. *PLoS One* 3, e3721
  202. Okamoto N, Yamanaka N, Yagi Y et al (2009) A fat body-derived IGF-like peptide regulates postfeeding growth in *Drosophila*. *Dev Cell* 17, 885-891
  203. Slaidina M, Delanoue R, Gronke S, Partridge L and Léopold P (2009) A *Drosophila* insulin-like peptide promotes growth during nonfeeding states. *Dev Cell* 17, 874-884
  204. Bai H, Kang P and Tatar M (2012) *Drosophila* insulin-like peptide-6 (*dilp6*) expression from fat body extends lifespan and represses secretion of *Drosophila* insulin-like peptide-2 from the brain. *Aging cell* 11, 978-985
  205. Holzenberger M, Dupont J, Ducos B et al (2003) IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. *Nature* 421, 182-187
  206. Yuan R, Tsaih SW, Petkova SB et al (2009) Aging in inbred strains of mice: study design and interim report on median lifespans and circulating IGF1 levels. *Aging Cell* 8, 277-287
  207. Tazearslan C, Cho M and Suh Y (2012) Discovery of functional gene variants associated with human longevity: opportunities and challenges. *J Gerontol A Biol Sci Med Sci* 67, 376-383