



The complete plastid genome of *Scopolia parviflora* (Dunn.) Nakai (Solanaceae)

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ABSTRACT: *Scopolia parviflora* of the family Solanaceae is an endemic species of Korea and a traditional Korean medicinal plant. The plastid genome was sequenced by next-generation sequencing (NGS) method. The characterized cp genome is 156,193 bp in size; the large single-copy (LSC) region is 86,364 bp, the inverted repeat (IR) is 25,905 bp, and the small single copy (SSC) region is 18,019 bp. The overall GC content of the plastid genome amounts to 37.61%. The cp genome contains 113 genes and 21 introns, including 80 protein-coding genes, four RNA genes, 30 tRNA genes, 20 group II introns, and one group I intron. A phylogenetic analysis showed that *Scopolia parviflora* was closely related to *Hyoscyamus niger*.

Keywords: Chloroplast, *Scopolia parviflora*, genome sequence, medicinal plant

Michikwangipul, *Scopolia parviflora* (Dunn.) Nakai (Dunn, 1912; Nakai, 1933) of the family Solanaceae, limitedly occurs in Korean peninsula, and is similar to *S. japonica* (Maximowicz, 1873) occurring in Japan. *S. parviflora* is distinguished from *S. japonica* by ITS and some phenetic characters (Hong and Paik, 2001; Kim et al., 2003). The members of the genus *Scopolia* are known as medicinal plants (Mino, 2002; Jung et al., 2003; Min et al., 2007). The genetic makeup of the plastid genome of *Scopolia* is poorly known. Here we sequenced the chloroplast genome of *Scopolia parviflora* as a representative of the genus *Scopolia*.

Materials and Methods

The plant material of Michikwangipul used in this study was collected from the wild population of Mt. Cheonma, Korea (N37° 40' 34.85" E127° 15' 33.00"). The voucher of the plant specimen (Parkjh 20150505-141) was deposited in NNIBR, Herbarium in Nakdonggang National Institute of Biological Resources. The total DNA was prepared as described by Lee et al. (2015). The Illumina paired-end (PE) genomic library of 200 bp was constructed and sequenced using an Illumina HiSeq

2000 platform. The plastid sequence was obtained using CLC Genomics Workbench version 7.05 as described by Jeong et al. (2014). Circular structures of each replicon were confirmed by polymerase chain reaction (PCR) amplification at their ends and by joining of Sanger sequence reads derived from the amplicons. The assemblies were further verified by examining paired-end distance and depth after re-mapping reads on the contig sequences. The BLAST searches of a large contig were verified to be plastid genomes. For gene annotation of organelle genomes, protein-coding and ribosomal RNA genes were annotated using DOGMA (<http://dogma.cccb.utexas.edu/>; Wyman et al., 2004). The boundaries of each annotated gene were manually determined by comparison with orthologous genes from other known cp genomes. Genes encoding tRNAs were first predicted using tRNAscan (http://lowelab.ucsc.edu/tRNA_scan-SE; Lowe and Eddy, 1997) and ARAGORN version 1.2 (<http://130.235.46.10/ARAGORN/>; Laslett and Canback, 2004), and were manually verified by predicting the tRNA secondary structure. Circular genome maps were drawn using GenomeVx (Conant and Wolfe, 2008) followed by manual modification. The sequencing data and gene annotations were submitted to GenBank with accession number

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Table 1. Gene list of *Scopolia parviflora* plastid.

	Gene				
Genetic system genes					
Conserved off	<i>yef1</i>	<i>yef2</i> ×2	<i>yef3</i> **	<i>yef4</i>	<i>yef15</i>
Maturase K	<i>matK</i>				
RNA polymerase	<i>rpoA</i>	<i>rpoB</i>	<i>rpoC1</i> *	<i>rpoC2</i>	
Ribosomal protein					
Large subunits	<i>rpl2</i> *×2	<i>rpl14</i>	<i>rpl16</i> *	<i>rpl20</i>	<i>rpl22</i> <i>rpl23</i> ×2 <i>rpl32</i> <i>rpl33</i> <i>rpl36</i>
Small subunits	<i>rps2</i>	<i>rps3</i>	<i>rps4</i>	<i>rps7</i> ×2	<i>rps8</i> <i>rps11</i> <i>rps12</i> ** ×2 <i>rps14</i> <i>rps15</i>
	<i>rps16</i> *	<i>rps18</i>	<i>rps19</i>		
Photosynthesis genes					
Acetyl-CoA carboxylase	<i>accD</i>				
ATP-dependent Clp protease	<i>clpP</i> **				
ATP synthase	<i>atpA</i>	<i>atpB</i>	<i>atpE</i>	<i>atpF</i> *	<i>atpH</i> <i>atpI</i>
Cytochrome b	<i>petB</i> *				
Cytochrome b/f	<i>petD</i> *	<i>petG</i>	<i>petL</i>	<i>petN</i>	
Cytochrome f	<i>petA</i>				
Cytochrome C biogenesis	<i>ccsA</i>				
Membrane protein	<i>cemA</i>				
NADH dehydrogenase	<i>ndhA</i> *	<i>ndhB</i> * ×2	<i>ndhC</i>	<i>ndhD</i>	<i>ndhE</i> <i>ndhF</i> <i>ndhG</i> <i>ndhH</i> <i>ndhI</i>
	<i>ndhJ</i>	<i>ndhK</i>			
Photosystem I	<i>psaA</i>	<i>psaB</i>	<i>psaJ</i>	<i>psaC</i>	<i>psaI</i>
Photosystem II	<i>psbA</i>	<i>psbB</i>	<i>psbC</i>	<i>psbD</i>	<i>psbE</i> <i>psbF</i>
	<i>psbH</i>	<i>psbI</i>	<i>psbJ</i>	<i>psbK</i>	<i>psbL</i> <i>psbM</i> <i>psbN</i> <i>psbT</i> <i>psbZ</i>
Rubisco	<i>rbcL</i>				
Ribosomal RNA	<i>rrn16S</i> ×2	<i>rrn23S</i> ×2	<i>rrn4.5S</i> ×2	<i>rrn5S</i> ×2	
Transfer RNA	<i>tma_UGC</i> * ×2	<i>trnC_GCA</i>	<i>trnD_GUC</i>	<i>trnE_UUC</i>	<i>trnF_GAA</i> <i>trnG_GCC</i> <i>trnG_UCC</i> * <i>trnH_GUG</i>
	<i>trnI_CAU</i> ×2	<i>trnI_GAU</i> * ×2	<i>trnK_UUU</i> *	<i>trnL_CAA</i> ×2	<i>trnL_UAG</i> <i>trnM_CAU</i> <i>trnN_GUU</i> ×2 <i>trnP_UGG</i>
	<i>trnQ_UUG</i>	<i>trnR_ACG</i> ×2	<i>trmR_UCU</i>	<i>trnS_GCU</i>	<i>trnS_GGA</i> <i>trnS_UGA</i> <i>trnT_GGU</i> <i>trnT_UGU</i> <i>trnV_GAC</i> ×2
	<i>trnV_UAC</i> *	<i>trnW_CCA</i>	<i>trnY_GUA</i>		
Pseudo gene	<i>ψ-rps19</i>	<i>ψ-yef1</i>			

*intron ^ucp genome contains a copy of *rps12* exon 1 in LSC and two copies of *rps12* exon 2 and 3 in IR

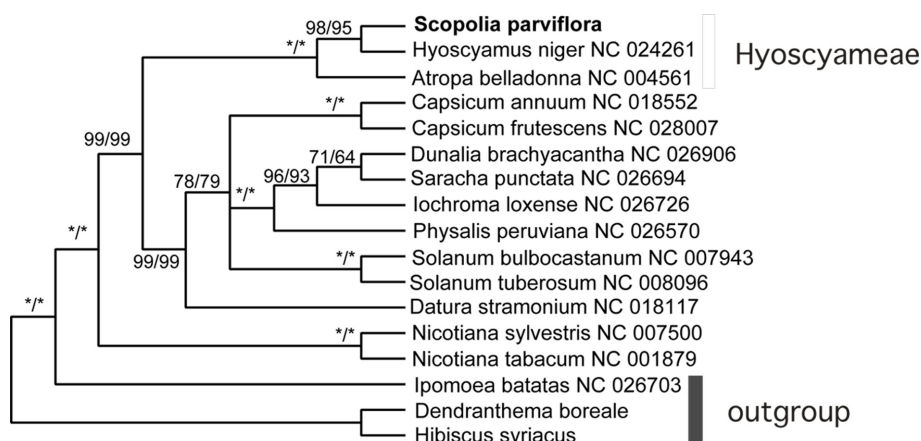


Fig. 2. Maximum parsimonious tree of 14 Solanaceae plastids, using seven protein coding gene (*psaA*, *psaB*, *psbA*, *psbB*, *psbC*, *psbD*, and *rbcL*) sequences. **/: Bootstrap value 100%/Jackknife value 100%.

single group II intron: *rpoC1.i*, *rpl2.i*, *rpl16.i*, *rps16.i*, *atpF.i*, *petB.i*, *petD.i*, *ndhA.i*, *ndhB.i*, *trnA_ugc.i*, *trnG_ucc.i*, *trnI_gau.i*, *trnK_uuu.i*, and *trnV_uac.i*. Among the 20 group II introns, the intron in *rps12*, between exons 1 and 2, is trans-splicing, while the other 19 group II introns are cis-splicing.

Seventeen genes, five introns, and parts of three genes and an intron are found within the IR, which has two copies. These 17 genes include six protein-coding genes (*ndhB*, *rpl2*, *rpl23*, *rps7*, *yef2*, *yef15*), all four rRNA genes (*16S*, *23S*, *4.5S*, *5S*), and seven tRNA genes (*trnA_ugc*, *trnI_cau*, *trnI_gau*, *trnL_caa*, *trnN_guu*, *trnR_acg*, *trnV_gac*). The five introns are *ndhB.i*, *rpl2.i*, *trnA_ugc.i*, *trnI_gau.i*, and *rps12.i2*. The IR contains the 5' end of *yef1* at the border with the SSC, resulting in one intact *yef1* and a 1,473-bp ψ -*yef1* in the cp-genome. The IR also contains the 5' end of *rps19* at the border with the LSC, resulting in one intact *rps19* and a 84-bp ψ -*rps19* in the cp-genome. In addition, the IR contains parts of the *rps12* gene. This *rps12* gene consists of three exons, *rps12.e1*, *rps12.e2*, and *rps12.e3*, *rps12.e1* is in the LSC, but *rps12.e2* and *rps12.e3* are in the IR. Thus, the genome contains a single copy of *rps12.e1* but has two copies of *rps12.e2* and *rps12.e3*. A cis-splicing group II intron, *rps12.i2*, intervenes between *rps12.e2* and *rps12.e3*, but a trans-splicing intron, *rps12.ilt*, occurs between *rps12.e1* and *rps12.e2*. The *rps12.ilt* is split into two pieces, *rps12.ilt1* and *rps12.ilt2*, because the *rps12* gene is transcribed in two separate operons, the *clpP* operon (*clpP*-*rps12.e1*-*rps12.ilt1*-*rpl20*) and the 3' *rps12* operon (*rps12.ilt2*-*rps12.e2*-*rps12.i2*-*rps12.e3*-*rps7*-*ndhB*).

Currently, more than 20 plastid genomes have been deposited in Genbank from 10 genera of Solanaceae. Phylogenetic analysis showed that *Scopolia parviflora* formed a strong clade with *Hyoscyamus niger* (Sanchez-Puerta & Abbona, 2014) and *Atropa belladonna* (Schmitz-Linneweber et al., 2002), and

that *Hyoscyamus niger* was the closest to *Scopolia parviflora*. The results support monophyly of the tribe Hyoscyameae of Solanaceae. The three plants are known to be highly toxic and are also used as medicine. In contrast, another toxic plant, *Datura* (Yang et al., 2014), clad with potato (Fig. 2).

In Korea, two species of the genus *Scopolia* have been documented. One is purple flowered *Scopolia parviflora* (Dunn.) Nakai (1933) and the other is yellow flowered *S. lutescens* Y. Lee (1993). In contrast, purple flowered *Scopolia japonica* Maxim. (1873) and yellow flowered *Scopolia japonica* Maxim. f. *lutescens* Sugim. (1977) also occur in Japan. Currently, ITS sequence analysis suggested that *Scopolia parviflora* and *S. japonica* were clearly distinguished, but that *Scopolia parviflora* and *S. lutescens* Y. Lee were indistinguishable (Kim et al., 2003). Plastid DNA sequence of *Scopolia japonica* from Japanese collection (voucher Tsugaru & Sawada, 17731) was only available in *ndhF* (Genbank EU580945). Without 162 ambiguous sequence of EU580945, *Scopolia japonica* had 5 SNPs in *ndhF* distinguished from *Scopolia parviflora*. Further study of comparative plastid genomics would help our understanding on the relationship among the *Scopolia* species.

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