

Determination of best enrichment media for growth of *Salmonella* injured from cold temperature during process and storage

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저온저장으로 인해 손상된 살모넬라를 배양하기 위한 최적의 배지 선정에 관한 연구

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Abstract

This purpose of this study was to determine the best enrichment medium for rejuvenating and recovering *Salmonella* placed in cold temperature prior to the employment of the gold biosensor combined with a light microscopic imaging system. A mixture of nalidixic-resistant *Salmonella* Typhimurium and Enteritidis were inoculated onto chicken (1,000 CFU/chicken). After cold injury at 4°C for 24 hr, *Salmonella* on chicken was enriched for 6 hr with six non-selective media including buffered peptone water broth, lactose broth, brain heart infusion broth (BHI), universal pre-enrichment broth, nutrient broth, and tryptic soy broth, and five selective media including brilliant green broth (BG), rappaport-vassiliadis R10 broth, selenite cystine broth, selenite broth, and tetrathionate brilliant green broth (TBG) for the comparison of *Salmonella* growth. Various concentrations of *Salmonella* (10, 50, 100, 500, and 1,000 CFU/chicken) were then enriched for 6 hr in both BHI and BG media to select the best media. BHI was selected as the most effective non-selective enrichment medium, while BG was selected as the most effective selective enrichment medium. Finally, BHI medium was selected as the most efficient enrichment medium for *Salmonella* growth injured from cold temperature during processing or storage.

Key words : enrichment, *Salmonella*, injury, cold temperature, gold biosensor combined with light microscopic imaging system

Introduction

Salmonella has been one of the most prevalent foodborne pathogens, ranking higher in incidence than *Campylobacter*, *E. coli* O157:H7, *Listeria monocytogenes*, *Shigella*, *Vibrio*, *Yersinia*, *Cryptosporidium*, and *Cyclospora* (1-3). Approximately 1.4 million cases of *Salmonella* infections have been reported in the United States annually, and the annual cost for medical

treatment and lost productivity have been estimated to be 0.5 to 2.3 billion dollars (1). The major serotypes causing *Salmonella* infections are *S. enterica* serovar Typhimurium (20%), Enteritidis (15%), Newport (10%), Javiana (7%), and Heidelberg (5%), which combined account for 56% of all human cases (3,4).

Salmonella outbreaks has been frequently associated with meats, poultry, eggs, milk, and dairy products. However, outbreaks associated with poultry products have drawn the most attention because poultry consumption has consistently increased whereas beef consumption has consistently decreased (56.8 lbs of poultry per capita vs 64.5 lbs of beef per capita annually; poultry per capita consumption 2002). In addition, although several prevention strategies have been

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employed in the poultry industry, it is hard to prevent contamination during processes (5). Rose et al. (6) found that 14.4% of poultry products randomly collected from a federally inspected processing plant were contaminated with *Salmonella*. In addition, another study (7) also found that 35% of ground poultry products were contaminated with *Salmonella*. Therefore, on-site applicable rapid detection methods need to be performed in poultry plants for the inspection of poultry products (8,9).

A considerable number of studies have been focused on the development of on-site applicable rapid detection methods in order to overcome the limitations of existing rapid detection methods (10-12). Recently, our research group has developed a gold biosensor combined with a light microscope imaging system (LMIS) for foodborne pathogen detection. Previous studies demonstrated that the gold biosensor combined with LMIS was successfully able to detect *Listeria monocytogenes* and *Escherichia coli* O157:H7 in food (5,12,13). Using the same principle and methodologies, this system will be expanded to detect *Salmonella* in poultry as on-site applicable rapid method.

Following regulations in the US, poultry carcasses should be chilled to 4.4°C or lower for a certain period of time to ensure a high quality and safe product (8). Therefore, all the processes in poultry plants are performed in cold temperatures (approximately 4~6°C) in order to inhibit or minimize the microbial growth during processes (14). Cold temperatures delay or injure *Salmonella* growth because of the extension of its lag phase (15). The sluggish growing and/or viable-but-non-culturable state under unfavorable temperatures might provide false-negative results, so *Salmonella* needs to be enriched or rejuvenated prior to employing a gold biosensor combined with LMIS. Therefore, the objectives of this study was to determine the best enrichment medium for rejuvenating and increasing the number of *Salmonella* injured from cold temperatures for the employment of a gold biosensor combined with LMIS.

Method and Materials

Salmonella strains and culture conditions

Two strains of nalidixic-resistant *Salmonella enterica* serovar Typhimurium (ATCC13311), and Enteritidis were obtained from Auburn University (Auburn, AL, USA). Each strain of *Salmonella* was cultivated in 20 mL of tryptic soy broth (TSB, Difco Laboratories, Sparks, MD, USA) for 16

hr at 37°C with agitation. After incubation, the cultures were washed three times with phosphate buffered saline (PBS, pH 7.2, Sigma-Aldrich Co., St. Louis, MO, USA) by centrifugation at 5,000×g for 5 min. The collected bacterial cells were re-suspended in PBS and the bacterial concentration was adjusted to 10⁹ CFU/mL using a pre-constructed standard curve determined by the optical density at 640 nm. *Salmonella* mixture was prepared by mixing equal amounts of the adjusted bacterial suspension prior to serial dilution.

Preparation of non-selective and selective enrichment media

Buffered peptone water broth (BPW, EMD Science, Darmstadt, Germany), lactose broth (LB, EMD Science), brain heart infusion broth (BHI, EMD Science), universal pre-enrichment broth (UPB, Difco Laboratories), nutrient broth (NB, EMD Science), and TSB were prepared as pre-enrichment media following the manufacturers' recommendation. Brilliant green broth (BG, Difco Laboratories), rappaport-vassiliadis R10 broth (RV, Difco Laboratories), selenite cystine broth (SC, Difco Laboratories), selenite broth (SB, Difco Laboratories), and tetrathionate brilliant green broth (TBG, Difco Laboratories) were prepared as selective enrichment media following the manufactures' recommendation.

Efficiency of *Salmonella* growth injured from cold temperature

Chicken skins were randomly collected from Koch Food Company (Montgomery, AL, USA) and cut into approximately 25 g portions. The chicken skin was washed with 200 ppm chlorine solution (Sigma-Aldrich Co.) and washed 10 times with sterilized deionized water to eliminate existing microorganisms in chicken. Two hundred µL of *Salmonella* mixture (1,000 CFU/200 µL) were spread onto chicken skins for inoculation. An equal amount of PBS buffer was applied to chicken as a negative control. After attachment of *Salmonella* for 30 min under a safety cabinet, each inoculated chicken skin was put into a sterile stomach bag containing 100 mL of non-selective or selective media with 100 ppm nalidixic acid then placed at 4°C for 24 hr and blended in a stomacher (Seward 400, Seward Inc., Bohemia, NY, USA) at 260 rpm for 2 min. The blended suspension was transferred to an Erlenmeyer flask for incubating at 37°C in an orbital shaker at 250 rpm. The bacterial number was determined by spread-plate method using tryptic soy agar (TSA, Difco Laboratories) plates containing 100 ppm nalidixic acid at 2, 4, and 6 hr. The medium that promoted the most growth

of *Salmonella* was chosen as the bacterial enrichment medium for further study.

Comparison of *Salmonella* growth injured from cold temperature using BHI, and BG media

For the selection of the best media, various concentrations of *Salmonella* mixture were inoculated onto chicken and enriched in BHI and BG media. Two hundred μL of *Salmonella* mixture containing 10, 50, 100, 500, and 1,000 CFU were inoculated on a chicken sample, respectively, followed by the same procedures described in the section above. The medium that promoted the most growth of *Salmonella* was chosen as the best enrichment medium for the employment of a gold biosensor combined with LMIS.

Statistical analysis

The one-way analysis of variance (ANOVA) for more than two groups were performed to compare the means using the GraphPad and InStat v.3 programs (Graphpad, San Diego, CA, USA). Significant differences were determined at $p < 0.05$.

Results and Discussion

Effect of non-selective media on the growth of *Salmonella* injured from cold temperature

Poultry plants should be chilled to 4.4°C or lower for a certain period of time, because the minimum growth temperature of *Salmonella* in poultry was reported as 5°C (14). Therefore, a similar temperature condition was introduced in this study by placing chicken at 4°C for 24 hr after the inoculation of *Salmonella*. The media's effect on *Salmonella* growth after storing was then investigated using a series of non-selective enrichment media including TSB, BHI, LB, NB, UPB, and BPW (Fig. 1). As time increased, the concentration of *Salmonella* increased approximately 3~4 log CFU/chicken, 5 log CFU/chicken, and 6~7 log CFU/chicken at 2, 4, and 6 hr, respectively. However, there were no significant differences among non-selective media both 2 and 4 hr ($p < 0.05$). Significant differences in the growth number of *Salmonella* was obviously noticed at 6 hr. The number of *Salmonella* enriched in BHI at 6 hr was significantly greater than the number of *Salmonella* enriched in NB and UPB at 6 hr ($p < 0.05$). Although *Salmonella* enriched in TSB, BHI, BPW and LB did not demonstrate any significant differences among the media, BHI was selected as the best non-selective media because it caused

the highest increase in the number of *Salmonella*. The number of *Salmonella* increased up to (7.3 ± 0.2) log CFU/chicken, followed by TSB media [(7.1 ± 0.4) log CFU/chicken]. The result of this study was in good agreement with the result of Boer (17), which demonstrated that BHI and TSB exhibited significantly greater increases than other non-selective media. Since TSB and BHI contain similar nutrients such as casein, glucose, disodium phosphate and sodium chloride, except the meat substrate in BHI and soybean in TSB, the bacterial number enriched in both media did not show significant differences. Although UPB also possesses a similar nutrient composition to TSB and BHI, except magnesium sulfate, the bacterial numbers enriched in UPB were significantly lower than those enriched in TSB or BHI, presumably due to the relatively lower pH of UPB (pH 6.3) when compared to TSB or BHI (pH 7.3). Therefore, among the six non-selective media, BHI was selected as the most efficient non-selective media for further study.

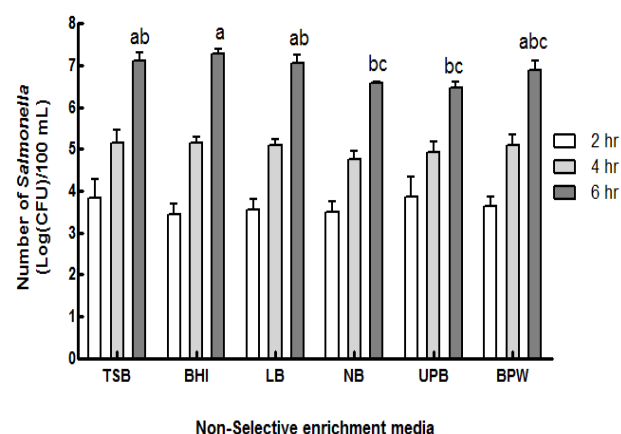


Fig. 1. Efficiency of non-selective enrichment media for *Salmonella* growth injured from cold temperature.

Salmonella-inoculated chicken (1,000 CFU/chicken) was exposed to 4°C for 24 hr prior to enrichment. Different letters (a,b,c) within the same group indicate significantly different means among the same enrichment periods at $p < 0.05$ ($N=10$). TSB, tryptic soy broth; BHI, brain heart infusion broth; LB, lactose broth; NB, nutrient broth; UPB, universal pre-enrichment broth; BPW, buffered peptone water.

Effect of selective media on the growth of *Salmonella* injured from cold temperature

Since *Salmonella* co-exists with other competitive foodborne pathogens, it needs to be enriched using selective media only by multiplying the target pathogens while inhibiting other foodborne pathogens (17). *Salmonella* was enriched in BG, SC, SB, RV and TBG to determine the best efficient medium for *Salmonella* recovery injured in a cold environment (Fig. 2). As time increased, the number of *Salmonella* increased proportionally. However, there were no

significant differences among the media until 4 hr ($p < 0.05$). Unlike other media, the number of *Salmonella* enriched in SC did not increase obviously between 2 hr and 4 hr. Other media provided similar increase patterns in the number of *Salmonella*, which was proportional to the increase in time. At 6 h-enrichment period, it was noticed that there were significant differences among the selective media. The number of *Salmonella* in BG [(6.4±0.3) log CFU/chicken] was significantly greater than the number of *Salmonella* in SC [(5.5±0.2) log CFU/chicken], SB [(5.5±0.6) log CFU/chicken] or TBG [(4.4±0.1) log CFU/chicken] ($p < 0.05$). Although *Salmonella* in RV [(5.9±0.1) log CFU/chicken] was not significantly different from that in BG, BG media was demonstrated to be the most efficient selective media for *Salmonella* growth after cold injury. This result was in agreement with other studies (18-22).

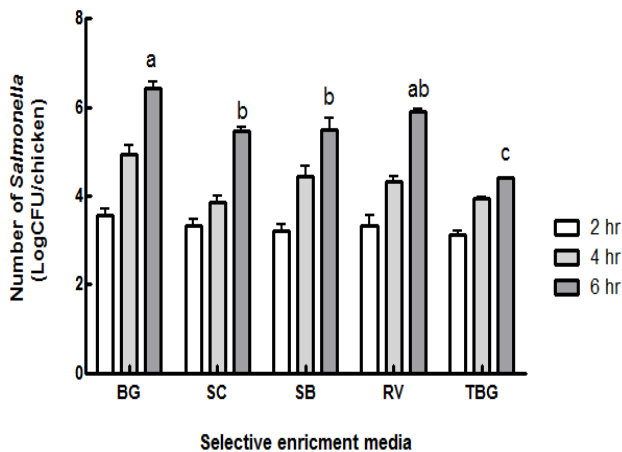


Fig. 2. Efficiency of selective enrichment media for *Salmonella* growth injured from cold temperature.

Salmonella-inoculated chicken (1,000 CFU/chicken) was exposed to 4°C for 24 hr prior to enrichment. Different letters (a,b,c) within same group indicate significantly different means among the same enrichment periods at $p < 0.05$ (N=10). BG, brilliant green broth; SC, selenite cystine broth; SB, selenite broth; RV, Rappaport-Vassilidis R10 broth; TBG, tetrathionate brilliant green broth.

Efficiency of *Salmonella* recovery inoculated with various concentrations using BHI, and BG media

Selected enrichment media (BHI and BG) were compared further for the determination of the best enrichment medium for *Salmonella* injured from cold temperatures. Various concentrations of *Salmonella* inoculated on chicken were enriched up to 6 hr and compared its detected number at each concentration (Fig. 3). During 2 hr incubation, the detected number of *Salmonella* increased as the inoculated concentration of *Salmonella* increased. Although, *Salmonella* was detected starting from the inoculated concentration at 50 CFU/chicken, however, no obvious increases in the number

of *Salmonella* were found. Presumably, *Salmonella* showed a relatively longer lag phase due to injury from the cold temperature for 24 hr. In addition, there were no significant differences in the number of *Salmonella* enriched in BHI and BG media.

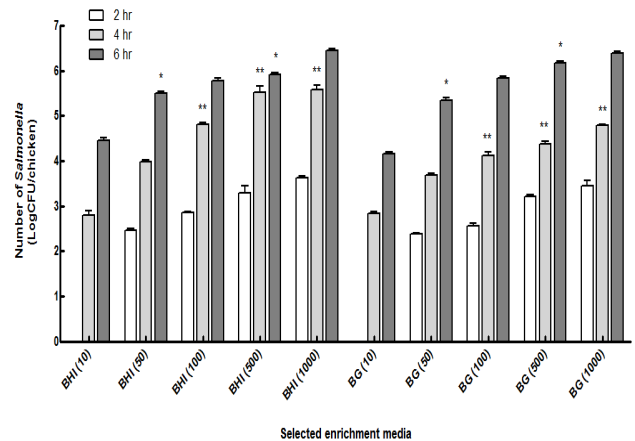


Fig. 3. Efficiency of selective media for the growth of injured *Salmonella* inoculated with various concentration on chicken.

Salmonella-inoculated chickens (10, 50, 100, 500, and 1,000 CFU/chicken) were exposed to 4°C for 24 hr prior to enrichment. Different symbols (*, **) within the same group indicate significantly different means among the same enrichment periods between BHI and BG at $p < 0.05$ (N=10). BHI, brain heart infusion broth; BG, brilliant green broth.

From the 4 hr incubation, the significant differences were observed starting from the inoculated concentration of 100 CFU/chicken between two media ($p < 0.05$). The number of *Salmonella* enriched in BHI was significantly greater than the number of *Salmonella* enriched in BG. The maximum growth of *Salmonella* was found in BHI media with (5.6±0.2) log CFU/chicken from an initial concentration of 3 log CFU/chicken. Furthermore, 4 hr incubation in both BHI and BG enriched the number of *Salmonella* up to the detectable levels (3 log CFU/chicken) of the gold biosensor combined with LMIS method except for 10 CFU/chicken (5). During 6 hr incubation period, the number of *Salmonella* enriched in BHI starting with 50 cells was significantly greater than the number of *Salmonella* enriched in BG ($p < 0.05$). In the meantime, the number of *Salmonella* enriched in BHI starting with 500 cells was significantly lower than the number of *Salmonella* enriched in BG ($p < 0.05$). Except for those instances, there were no significant differences between BHI and BG media among various concentrations of *Salmonella*. Even at the smallest inoculation (10 CFU), the *Salmonella* concentration was reached up to approximately 4 log CFU/chicken, which was a detectable level for the developed biosensor method. In conclusion, BHI medium was selected

as the most efficient enrichment medium for *Salmonella* growth injured from cold temperatures during processing or storage.

요 약

본 연구의 목적은 가금류에서 문제가 되고 있는 살모넬라 신속검출을 위해 광학현미경 기반 이미징 시스템 적용에 앞서서 냉장온도로부터 손상된 살모넬라를 증균시키기 위한 최적의 배지를 선정하는 것이다. 대표적인 식중독 유발 균주인 *S. Typhimurium*과 *S. Enteritidis*를 닭에 도말 하였고 4°C에서 24시간 동안 저장한 후 BPW, LB, BHI, UPB, NB, 그리고 TSB 배지와 BG, RV, SC, SB 그리고 TBS 선택배지에 각각 6시간 동안 배양하였다. 배양 후, 2시간마다 살모넬라 균수를 측정하였고 그 결과 BHI와 BG를 선택되었다. 최종적으로 최적의 배지 선택을 위해 다양한 농도의 살모넬라를 닭에 각각 도말 후 6시간 동안 배양하면서 균수를 변화를 측정하였다. 그 결과 BHI가 냉해로부터 손상된 살모넬라를 증균시키기 위한 가장 최적의 배지로 선정되었으며 본 연구의 결과는 광학현미경 기반 이미징 시스템을 활용한 신속검출법 적용을 위한 증균배지로 이용될 것이다.

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