

Zerumbone, Sesquiterpene Photochemical from Ginger, Inhibits Angiogenesis

Ju-Hyung Park, Geun Mook Park, and Jin-Kyung Kim

Department of Biomedical Science, Catholic University of Daegu, Gyeongsan 712-702, Korea

Here, we investigated the role of zerumbone, a natural cyclic sesquiterpene of *Zingiber zerumbet* Smith, on angiogenesis using human umbilical vein endothelial cells (HUVECs). Zerumbone inhibited HUVECs proliferation, migration and tubule formation, as well as angiogenic activity by rat aorta explants. In particular, zerumbone inhibited phosphorylation of vascular endothelial growth factor receptor-2 and fibroblast growth factor receptor-1, which are key regulators of endothelial cell function and angiogenesis. *In vivo* matrigel plug assay in mice demonstrated significant decrease in vascularization and hemoglobin content in the plugs from zerumbone-treated mice, compared with control mice. Overall, these results suggest that zerumbone inhibits various attributes of angiogenesis, which might contribute to its reported antitumor effects.

Key Words: Angiogenesis, Zerumbone, Vascular endothelial growth factor receptor, Human umbilical vein endothelial cells

INTRODUCTION

Phytochemicals are non-nutritive compounds found in plants. They are attracting research interest because of their health-promoting effects [1]. Epidemiological studies have consistently indicated a relationship between copious consumption of fruits and vegetables and reduction in the risk of developing several chronic diseases [2]. These health benefits have spurred the widespread purchase and consumption of phytochemical-enriched foods. Most of the populace in developing countries mainly relies on herbs to treat disease, which are considered to be safer and more effective than synthetic drugs [3,4]. Still, how phytochemicals beneficially affect physiological conditions remains unclear.

Angiogenesis is the development of new blood vessels from pre-existing vessel. These complex and multistep process includes proliferation, migration and tube formation of endothelial cells [5]. The angiogenic process involves the activation, proliferation and migration of endothelial cells toward various angiogenic stimuli, such as vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF) [6-9]. Interestingly, many cancer cells can in-

duce secretion of several pro-angiogenic factors, which is commonly observed in most aggressive tumors [8]. Also, a tumor creates a favorable microenvironment that induces angiogenesis and facilitates cell invasion to promote continued tumor growth and metastasis [10,11]. Since angiogenesis is essential for tumor development and tumor vasculature is an optimal target for anti-cancer strategies, anti-angiogenic compounds may act as cancer treatment agents or adjuncts to standard chemotherapeutic regimens [12]. In this regard, non-toxic anti-angiogenic phytochemicals could have greater practical significance compared to non-selective cytotoxic therapies to control the tumor growth and metastasis by targeting angiogenesis.

Zerumbone ((2E, 6E, 10E)-2, 6, 9, 9-tetramethylcycloundeca-2, 6, 10-trien-1-one, Fig. 1) is a phytochemical that was first isolated from the essential volatile oil of rhizomes of *Zingiber zerumbet* [13]. Its potentially broad use in the

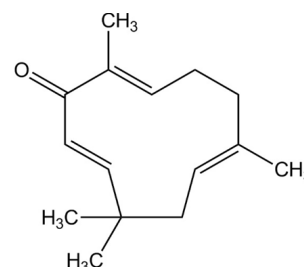


Fig. 1. Structure of Zerumbone.

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Corresponding to: Jin-Kyung Kim, Department of Biomedical Science, Catholic University of Daegu, 13-13 Hayang-ro, Gyeongsan 712-702, Korea. (Tel) 82-53-850-3774, (Fax) 82-53-850-3774, (E-mail) toto0818@cu.ac.kr

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ABBREVIATIONS: HUVECs, human umbilical vein endothelial cells; FGF, fibroblast growth factor; VEGF, vascular endothelial growth factor.

treatment of cancers, leukemia and virus infection has been indicated [14-16]. In addition, zerumbone has a variety of pharmacological effects, which include anti-oxidant, anti-inflammatory and anti-bacterial activity [17,18]. More recently, zerumbone was demonstrated to inhibit angiogenesis by blocking nuclear factor- κ B in pancreatic and gastric cancer [19,20]. But, the detail mechanisms of its anti-angiogenic effect on human endothelial cells remain unclear.

The present study reports that zerumbone can suppress the proliferation, migration and tube formation of endothelial cells *in vitro* and angiogenesis *in vivo* by blocking essential angiogenic signaling pathways.

METHODS

Materials

Zerumbone, heparin, dimethylsulfoxide (DMSO), HEPES solution and anti- β -actin antibody were obtained from Sigma-Aldrich (St. Louis, MO, USA). Medium 199 (M199), fetal bovine serum (FBS), penicillin and streptomycin were obtained from Hyclone (Logan, UT, USA). Endothelial cell growth supplement (ECGS) and matrigel were purchased from BD Bioscience (Becton, Dickinson, USA). Cell Titer 96[®] Aqueous One Solution Cell Proliferation assay kits and CytoTox 96[®] Non-Radioactive Cytotoxicity assay kits were purchased from Promega (Madison, MI, USA). Anti-phospho-VEGFR2 antibody, VEGFR2 antibody, FGFR antibody, recombinant mouse VEGF₁₆₄ and bFGF were obtained from Cell Signaling Technology (Danvers, MA, USA). Anti-phospho-FGFR antibody was obtained from R&D systems (Minneapolis, MN, USA). Recombinant human VEGF₁₆₅ and bFGF were purchased from Pepprotech (Rocky Hill, NJ, USA).

Cell culture

Human umbilical vein endothelia cells (HUVECs) were obtained from Merck Millipore (Darmstadt, Deutschland, Germany) and maintained in M199 medium containing 20% FBS, 50 μ g/ml ECGS, 100 μ g/ml heparin, 25 mM HEPES solution, 100 U/ml penicillin and 100 μ g/ml streptomycin (complement M199 medium) at 37°C in 5% CO₂. HUVECs were used at passages 3~5 for all the experiments.

Cell proliferation assay

The effect of zerumbone on the proliferation of HUVECs was measured by Cell Titer 96 Aqueous One solution cell proliferation assay (MTS assay). HUVECs were seeded at 5×10^3 per well in 96-well plate with 200 μ l of complete M199 medium for 24 h before treatment. Fresh complete M199 medium (200 μ l) containing various concentrations of zerumbone was added into each well to treat cells for 24 h. Cell Titer 96[®] Aqueous One Solution Cell Proliferation assay kits was used according to manufacturer's protocol to determine the number of viable cells. Absorbance was read at 490 nm using a microplate reader (Tecan, San Jose, CA, USA). Percentage of cell proliferation was determined using the following formula: (Absorbance test well/ Absorbance 0 μ M-treated well) $\times 100$.

Lactate dehydrogenase (LDH) toxicity assay

To determine of cytotoxicity of zerumbone on HUVECs, the LDH release assay was performed using a CytoTox 96[®] Non-Radioactive Cytotoxicity assay kits according to the manufacturer's instructions.

Wound healing migration assay

HUVECs were seeded into 48-well dishes and grew until 80~90% confluence. Sterilized one-milliliter pipette tip was used to generate wounding across the cell monolayer, and the debris was washed with PBS. Fresh complete M199 medium containing different concentrations of zerumbone was added to the scratched monolayers. Cells migrated into the wounded area were taken using an inverted microscope (Leica DM500, Leica Microsystems, Heerbrugg, Switzerland) at 40 \times magnification after 18 h of incubation. A total of nine areas were selected randomly in three wells of each group were quantified in each experiment. The assay was repeated three times independently. Data are expressed as the percent of control migration (0 μ M-treated cells). The assay was repeated three times independently.

Western blot analysis

HUVECs were treated with indicated concentration of zerumbone and recombinant human VEGF₁₆₅ (10 ng/ml) or bFGF (50 ng/ml) for 15 min. Whole cell lysates were extracted using PRO-PREP Protein Extraction Solution (iNtRON Biotechnology, Seongnam-Si, Korea) according to the manufacturer's instructions and separated by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The separated proteins were electrophoretically transferred onto nitrocellulose membranes. Immunoreactive bands were detected by incubating the samples with horseradish peroxidase-conjugated secondary antibodies and visualized using a Femto Supersignal Chemiluminescent Substrate (Thermo, Rockford, USA) and Davinch-Chemi CAS-400SM Western Imaging System (Davinch-K, Seoul, Korea). The density of bands was assessed by Total Lab software (Davinch-K).

Capillary tube formation assay

Matrigel was thawed at 4°C overnight, after which each well of 48-well plates were coated with 150 μ l matrigel and incubated at 37°C for 1 h. HUVECs (5×10^4 cells) were added in 500 μ l complete M199 medium with zerumbone at 37°C in a humidified 5% CO₂ atmosphere. Images of the formation of capillary-like structures were obtained after 18 h with a computer-assisted Leica DM500 microscope microscope at $\times 40$ magnification. Tubular structures were quantified by manually counting the numbers of connected cells in randomly selected fields; the total tube number in the control group (0 μ M-treated cells) was designated as 100%. Tube formation was measured by taking photographs with the Leica DM500 microscope using a $\times 40$ objective. The assay was repeated three times independently.

Rat aortic ring assay

48-well tissue culture plates were covered with 150 μ l matrigel and allowed to gel for 30 to 45 min at 37°C, 5%

CO₂. Thoracic aortas were excised from 8-week-old male Sprague-Dawley rats (Hyochang Science, Deagu, Korea), and the fibroadipose tissue was removed. The aortas were sectioned into 1 mm to 1.5 mm-long cross sections, rinsed several times with PBS, placed on matrigel in wells and covered with an additional 100 μ l of matrigel. The rings were cultured in 1.5 ml of M199 medium without serum for 24 h, and then the medium was replaced with 1.5 ml of complete M199 medium with indicated concentration of zerumbone. The medium was changed every 2 days. After 7 days of incubation, the rings were fixed with 4% PFA. The microvessel growth was measured in photographs taken with a Leica DM500 microscope using a $\times 40$ objective.

Matrigel plug assay

Four-week-old C57BL/6 male mice were obtained from Hyochang Science and were subcutaneously injected with 500 μ l of matrigel containing 100 ng/ml of recombinant mouse VEGF₁₆₄, 100 ng/ml of recombinant mouse bFGF and 25 U heparin with indicated concentration of zerumbone. The mice were sacrificed after 7 days and the matrigel plugs were immediately photographed and weighed. To quantitate the vascularisation of the plug, the amount of hemoglobin (Hb) accumulated in the plug was measured using Drabkin's reagent (Sigma) following the manufacturer's protocol.

In addition, the removed matrigel plugs were fixed in 4% PFA followed by embedding in paraffin. Sections were then stained with hematoxylin-eosin (H&E) staining. Red blood cells were measured in photographs taken with a Leica DM500 microscope using a $\times 40$ objective. The protocol was reviewed by the Institutional Animal Care and Use Committee (IACUC) of Catholic University of Daegu (Accredited Unit No. CUD-2013-024).

Statistical analysis

The data are means \pm SEM of at least three independent experiments. The values were evaluated by one-way analysis of variance (ANOVA) with Bonferroni multiple comparison post tests using GraphPad Prism 4.0 software (GraphPad Software Inc., San Diego, CA). p -values < 0.05 were considered statistically significant.

RESULTS

Zerumbone inhibits HUVEC migration and tube formation

We first assessed whether zerumbone treatment affected HUVECs proliferation using the MTS assay. Treatment with zerumbone induced a dose-dependent anti-proliferative effect in HUVECs (Fig. 2A). Zerumbone treatment at > 100 μ M inhibited the proliferation of HUVECs. To confirm whether the anti-proliferative effect of zerumbone was due to cytotoxicity, we next measured activity of LDH, a stable enzyme in the cell cytoplasm that is rapidly released into the culture medium upon damage of the plasma membrane. As shown in Fig. 2B, zerumbone had no effect on cytotoxicity in HUVECs suggesting that the anti-proliferative activity of zerumbone on HUVECs is not due to cytotoxicity.

Endothelial migration is the key process in angiogenesis.

To determine whether zerumbone could inhibit endothelial cell migration, wound-healing assay was employed to explore the changes in migration. As shown in Fig. 2C, the endothelial cell migration was significantly blocked by 100 and 200 μ M zerumbone in a concentration-dependent manner.

We next analyzed the effect of zerumbone on tube for-

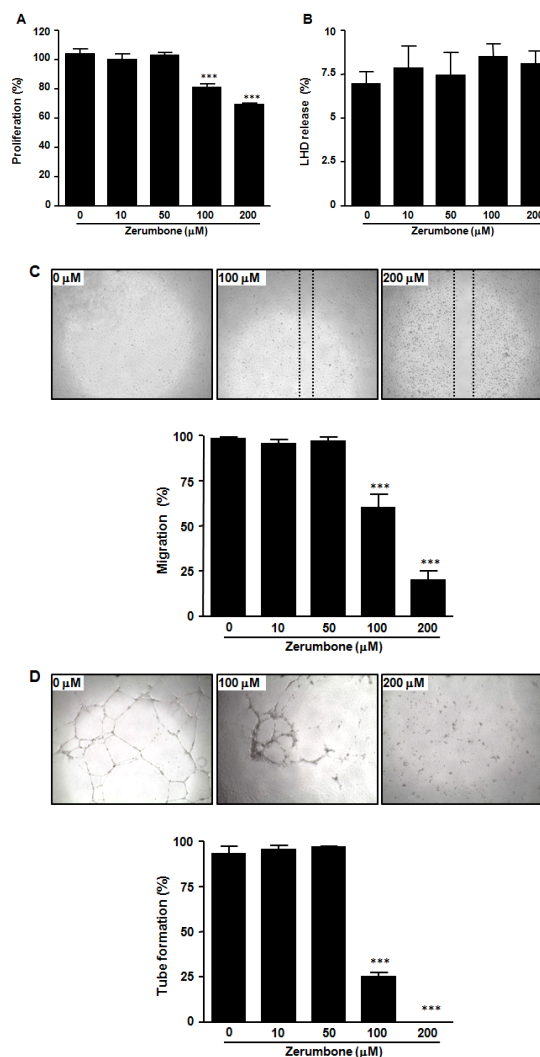


Fig. 2. Inhibition of endothelial cell proliferation, migration, and capillary-like tubule formation by zerumbone. HUVECs were plated in 96-well plates, allowed to attach overnight, and then cultured for 24 h with indicated concentration of zerumbone. (A) The proliferation and (B) LDH releases were determined as described in the Methods section. (C) For cell migration, a monolayer of inactivated HUVECs was wounded by scratching with a 0.1 ml pipette tip, and fresh medium containing indicated concentration of zerumbone was added. After 18 h, migration of HUVECs was measured as described in the Methods section. (D) For capillary-like tubule formation, HUVECs were seeded onto matrigel-coated 48-well plates and incubated with indicated concentration of zerumbone for 18 h. Endothelial tubules were photographed and quantitated. The results are means \pm SEM for at least three independent experiments. *** $p < 0.001$ versus 0 μ M zerumbone-treated cells.

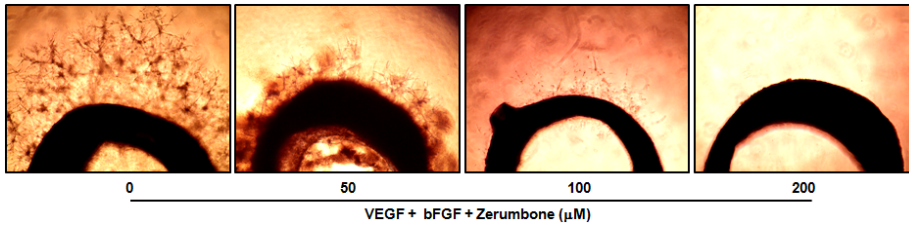


Fig. 3. Effect of zerumbone on microvessel outgrowth arising from rat aortic rings. Aortic rings isolated from SD rats were embedded in matrigel in 48-well plates, and then fed with medium containing zerumbone for 7 days. Photographs are representative of three independent experiments.

mation of HUVECs. Tube formation of endothelial cell recapitulates some angiogenesis steps, such as cell migration, alignment, formation of tubes and tube branching, and anastomosis with adjacent tubes [21]. Presently, when HUVECs were plated on matrigel in a short-term culture, they displayed alignment into networks of tubules (Fig. 2D). Zerumbone at 100 and 200 μM strongly disrupted HUVEC capillary morphogenesis (Fig. 2D). These results suggested that zerumbone inhibits endothelial cell differentiation and indicated its potential angiogenesis inhibition activity.

Zerumbone inhibits microvessel outgrowth from rat aortic ring

An *ex vivo* rat thoracic aorta ring assay was employed to evaluate the anti-angiogenic activity of zerumbone. The rat thoracic aorta ring model mimics the *in vivo* condition compared with the HUVECs tuber formation model in view of the multiple steps (sprouting, proliferation, migration and differentiation) involved in angiogenesis [22]. As predicted, zerumbone appeared to reduce microvessel outgrowth in a concentration-dependent manner, with angiogenesis being nearly abrogated at 100 and 200 μM (Fig. 3).

Zerumbone down-regulates signaling molecules

To investigate the molecular mechanism of the anti-angiogenic effect of zerumbone, the essential signal pathways of angiogenesis were explored. VEGFR2 is the primary receptor for VEGF that mediates angiogenic activity, endothelial cell proliferation, migration, differentiation and tube formation [23]. Appropriately we tested whether zerumbone interacted with the VEGF/VEGFR2 signaling pathway. VEGFR2 was phosphorylated by exogenous VEGF in HUVECs and zerumbone blocked this phosphorylation in a concentration-dependent manner (Fig. 4A).

In addition to VEGF, bFGF are an important class of angiogenic factors [24,25]. We examined whether zerumbone also inhibited phosphorylation of FGFR1 induced by bFGF stimulation. The observation of the markedly reduced levels of phosphorylated-FGFR1 in the presence of zerumbone (Fig. 4B) indicated that zerumbone-mediated interference with the VEGF and FGF signaling pathway might, at least in part, be the basis of the anti-angiogenic effect.

Zerumbone inhibits angiogenesis *in vivo*

To further examine the anti-angiogenic effects of zerumbone *in vivo*, we tested its effects using the mouse matrigel plug neovascularization assay. C57BL/6 mice received subcutaneous matrigel containing angiogenic factors (VEGF, bFGF and heparin) in the absence or presence of zerumbone. After 7 days, the plugs were removed and ma-

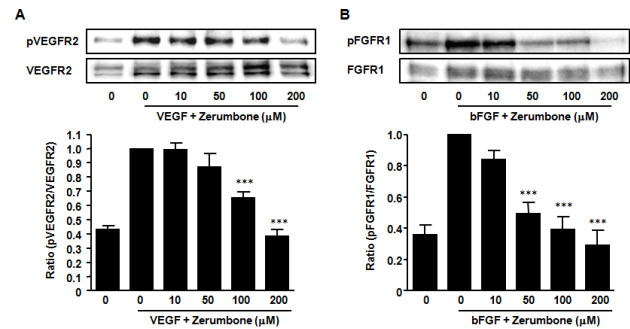


Fig. 4. Effects of zerumbone on VEGFR2- and FGFR1-mediated signaling pathways. (A) HUVECs were treated with various concentrations of zerumbone and stimulated with human VEGF (10 ng/ml) for 15 min. Phosphorylation of VEGFR2 by VEGF was determined using Western blot. (B) HUVECs were treated with zerumbone and stimulated with human bFGF (50 ng/ml) for 15 min. Phosphorylation of FGFR by bFGF were determined using Western blot. The bands were quantified using image analysis software and their relative intensity was expressed as fold against the image of the VEGF- or bFGF- stimulated cells. *** $p < 0.001$ versus VEGF or bFGF+0 μM zerumbone-treated cells.

trigel hemoglobin content was measured. The positive control group (VEGF, bFGF and heparin) displayed extensive neovascularization of the matrigel, as shown by the red color distributed in the whole plug (Figs. 5A, 5B). In contrast, matrigel implants treated with both angiogenic factor and zerumbone (200 μM) displayed strong inhibition of vascular development similar to that of the negative control (matrigel alone). In addition, H&E staining showed that the presence of red blood cells was markedly reduced in the zerumbone-containing plug (Fig. 5C). This observed *in vivo* anti-angiogenic activity was consistent with the *in vitro* anti-angiogenic effect of zerumbone.

DISCUSSION

Presently, zerumbone inhibited the proliferation, migration and capillary tube formation of HUVECs in a non-cytotoxic fashion (Fig. 2). Coincidentally, zerumbone potentially inhibited the outgrowth of new blood vessels in the rat aortic rings in a dose-dependent manner (Fig. 3). Zerumbone treatment also blocked angiogenesis in mice as demonstrated in the matrigel plug assay (Fig. 5). Our data might point toward an anti-angiogenic activity of zerumbone. However, it has to be mentioned that the *in vitro* data were gained at a dose of 100 and 200 μM . As demonstrated, lower than 100 μM of zerumbone hardly showed anti-angiogenic activity. Thus, 100 μM seems to be (near) the minimum dose to exert anti-angiogenic effect of zerumbone.

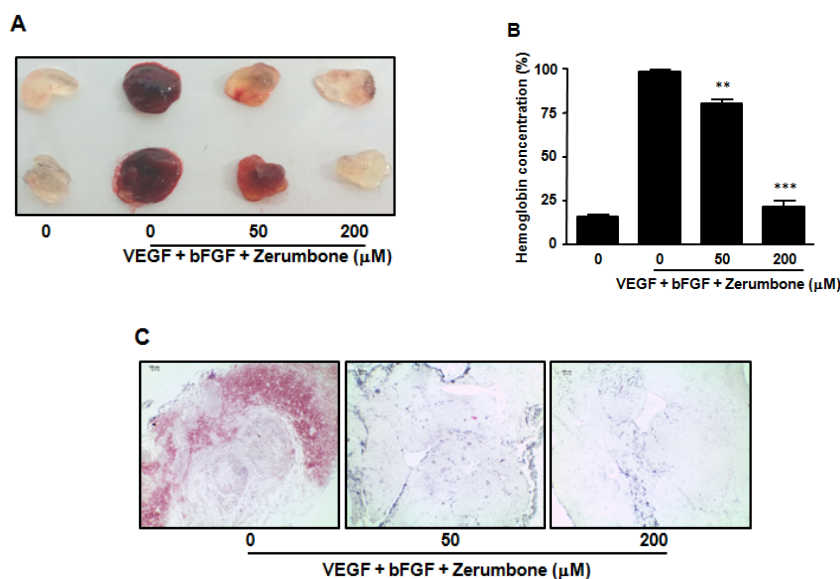


Fig. 5. Effects of zerumbone *in vivo* angiogenesis by matrigel plug assay. (A) Matrigel containing mouse VEGF (100 ng/ml), mouse bFGF (100 ng/ml), and zerumbone was subcutaneously injected into C57BL/6J mice. After 7 days, matrigels were removed and photographed. (B) Paraffin sections of matrigel plugs were stained with hematoxylin and eosin. Original magnification, $\times 100$; Scale bar 100 μm . (C) The quantification of neovascularization on the matrigel was performed by measuring the hemoglobin content. The results are reported as the mean \pm SEM of three independent experiments. Statistical significance is based on the difference when compared with non-treated animals, ** $p < 0.01$, *** $p < 0.001$.

Angiogenesis is vital for the growth and development of an organism, as well as in wound repair. The human body regulates angiogenesis by balancing stimulatory and inhibitory factors. Because this process is essential for healing, growth, development and maintenance, angiogenesis is a therapeutic target in various diseases. Among these diseases, cancer is associated with angiogenesis [26]. Angiogenesis is important for multiple steps in the progression of cancer pathophysiology, extending from the initial development of the tumor to the growth in the local environment invasion and extravasation into the vasculature. Anti-angiogenic agents are attractive for the treatment of several cancers [12]. The anti-angiogenic activity of zerumbone may be at least a partial basis of the compound's anti-cancer activity.

A variety of different growth factors can promote angiogenesis. One of the most important is VEGF. VEGF is a soluble protein secreted by most types of cells, but not endothelial cells [5]. Endothelial cells express receptors for this growth factor [27]. VEGF binding normally triggers the phosphorylation of its receptor, which in turn initiates an intracellular cascade of signaling events leading to physiological pro-angiogenic responses. Among their receptors, VEGFR2 phosphorylation may represent an important mechanism for the control of angiogenesis [7,27]. Inhibition of VEGFR2 phosphorylation by zerumbone might be a potential strategy to regulate angiogenesis. Therefore, we examined the effects of zerumbone on VEGFR2 expression and the VEGFR2 phosphorylation in HUVECs. Zerumbone at concentrations of 100 and 200 μM inhibited VEGF-induced VEGFR2 phosphorylation in HUVECs without any change the total VEGFR2 expression (Fig. 4A). These results implicate interference with VEGFR phosphorylation in the anti-angiogenic effect of zerumbone.

While the exact role of bFGF in tumor angiogenesis remains elusive, this potent mitogen of vascular endothelial cells and fibroblasts is believed to cooperate with VEGF during blood vessel development [28]. bFGF mediates its activity by primarily binding with FGFR1 and recruitment of vascular sprouting [24,25]. Since VEGFR2 and FGFR1 directly influence tumor angiogenesis, simultaneous in-

hibition of these two receptor tyrosine kinases is postulated to enhance antitumor activity. To this end, small molecule inhibitors of these receptors that inhibit VEGFR2 and FGFR1 have been described [29-31]. Furthermore, since anti-angiogenic therapies are directed toward highly regulated endothelial cells of the host, rather than genetically unstable tumor cells, it is anticipated that selective VEGFR2 and FGFR1 inhibition will lead to cancer therapeutics that have a lower risk of developing classical drug resistance compared to traditional chemotherapy. In this regard, zerumbone might be an ideal agent to regulate angiogenesis because it blocks the VEGFR2 and FGFR1 signaling pathways that are essential for angiogenesis.

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