

Research Report

Effects of Temperature and Ethylene Response Inhibitors on Growth and Flowering of Passion Fruit

Fang-Yin Liu^{1†}, Yung-Liang Peng^{2†}, and Yu-Sen Chang^{2*}¹Department of Horticulture, National Taitung Jr. College, No.889, Jhengci North Road, Taitung, 95045 Taiwan, ROC²Department of Horticulture and Landscape Architecture, National Taiwan University, No. 1, Section 4, Roosevelt Road, Taipei, 10617 Taiwan, ROC

Abstract: This study examined the effects of different day/night temperature regimes or silver ion on growth and flowering of passion fruit 'Tai-nung No.1'. Low temperature treatment (20/15°C) caused passion fruit cultivar 'Tai-nung No.1' to fail to flower. Flowering induction occurred within a temperature range of 20-30°C, with no significant difference in the days to first flower bud and the total number of flower buds between plants grown at 30/25°C and 25/20°C. However, plants grown at 30/25°C exhibited their first flower buds set on the higher nodes and had higher abortion rates of flower buds than those at 25/20°C. Plants grown at 30/25°C had the most rapid growth and the shortest plastochron. We also evaluated the effect of the ethylene response inhibitors silver nitrate (AgNO₃) and silver thiosulfate (STS) on growth and flowering of potted passion fruit 'Tai-nung No.1', when they were exposed to low temperature conditions (20/15°C) following chemical treatments (AgNO₃ or STS, at 0.5 or 1.0 mM). AgNO₃ and STS treatments induced flower formation and initial flower bud formation within approximately two weeks at 20/15°C whereas non-treated control plants exhibited no flower formation. ACC content and activity of ACC oxidase in the leaves of passion fruit 'Tai-nung No.1' exposed to low temperature conditions (20/15°C) were significantly inhibited by the ethylene inhibitor treatments. These results indicate that ethylene, which is produced under low temperature conditions, plays an important role in inhibiting flower formation in passion fruit.

Additional key words: ACC content, ACC oxidase activity, flower formation, silver nitrate, silver thiosulfate

Introduction

Passion fruit (*Passiflora* spp.) are popular in many subtropical and tropical countries. The results of previous studies have shown that the flowering of passion fruit is enhanced by appropriate regulation of environmental factors, such as photoperiod (Menzel and Simpson, 1988; Nave et al., 2010), temperature (Menzel et al., 1987), water content (Menzel et al., 1986; Staveley and Wolstenholme, 1990), and the application of plant growth regulators including aminoethoxyvinylglycine (AVG) and silver thiosulfate (STS) (Barbosa et al., 2001; Reis et al., 2003).

Seasonal temperature variations are usually less than 3-4°C in tropical areas (diurnal temperature variation is usually 8-10°C). Simon and Karnatz (1983) showed that

day/night temperatures of 25-30/20°C increased shoot growth and flowering, but reduced yields in purple passion fruit cuttings. Menzel et al. (1987) showed that the vegetative growth of purple × golden hybrids 'Purple Gold', 'E-23', and 'Lacey' was greater at 20/15°C, 25/20°C, and 30/25°C compared to those at 15/10°C under low solar radiation levels of 9.8 MJ m⁻² day⁻¹ (234 cal·cm⁻²·day⁻¹) (in winter in Queensland, Australia). For 'E-23' and 'Purple Gold', the increased vegetative growth at 25/20°C and 30/25°C compared to those at 20/15°C was associated with reduced flowering. The growth of shoot was greater at 33/28°C compared with 23/18°C and 28/23°C (day/night temperature) in the purple passion fruit (Utsunomiya, 1992).

Among the 400 species of *Passiflora*, there are 50-60 edible species of passion fruit but only half of them are

*Corresponding author: yschang@ntu.edu.tw

†These authors contributed equally to this work.

※ Received 22 May 2014; Revised 9 December 2014; Accepted 21 December 2014.

grown solely for their fruit, and among those that have been commercially cultivated are *P. edulis* Sim., *P. edulis* f. *flavicarpa* Deg., and their cross-breeding, i.e. 'Tai-nung No.1', *P. edulis* × *P. edulis* f. *flavicarpa*. Passion fruit 'Tai-nung No.1' tolerates a wide range of climatic conditions from sea-level to altitudes of up to 600 meters, and is the most popular passion fruit in Taiwan. Most *Passiflora* species, including passion fruit 'Tai-nung No.1' are not cold-tolerant for outdoor cultivation and require protected cultivation in cool temperature zones (Chang and Cheng, 1992). Many subtropical and tropical plants are chilling-sensitive; when stressed by low temperature, the tissues of such plants respond by producing ethylene (Max et al., 2011; Orihuel-Iranzo et al., 2010; Tacken et al., 2010). Several studies have demonstrated that the leaves of the *Passiflora* species stored at 0°C can release ethylene (Chen and Patterson, 1985), and the more sensitive species produced larger amounts of ethylene than those more tolerant to chilling (MacRae et al., 1986). Therefore, ethylene produced by plant tissue during cold winter conditions, is presumably an important inhibitor of growth and flowering in some *Passiflora* plants.

This study evaluated the effects of different day/night temperature regimes on growth and flower formation of *Passiflora* Tai-nung No.1 (*Passiflora edulis* × *P. edulis* f. *flavicarpa*) and also the effects of AgNO₃ and STS on growth and flowering of young, potted *Passiflora* 'Tai-nung No.1' plants grown at the low temperature regime (20/15°C).

Materials and Methods

Plant Material

Experiment 1. Passion fruit plants (Tai-nung No.1, *Passiflora edulis* × *P. edulis* f. *flavicarpa*) were obtained in June from a commercial passion fruit nursery and planted in 18-cm diameter pots containing 2.9 L of a 2:2:1 (v/v) mix of vermiculite, perlite, and peat moss, and then were grown in a phytotron (25/20°C day/night temperature) under natural light, at 60-80% RH. On 5 August, the plants had an average height of 50.2 cm and an average of 7.4 nodes when the various treatments started. The average PPFD (photosynthetic photon flux density) at noon was 400 μmol·m⁻²·s⁻¹. Plants were watered (to soil moisture content greater than 80%) when the surface of the medium became slightly dry. For slow-release fertilization, 3.0 g of Osmocote® (14:14:14 N:P:K; Scotts Company, Marysville, OH, USA) was applied to each pot at monthly intervals.

Experiment 2. *Passiflora* Tai-nung No.1 grafted seedlings were purchased from a nursery in Puli, Taiwan. These plants

were transplanted into 2.9 L plastic pots containing 1:1:1 (v/v) mix of vermiculite, perlite, and peat moss, then grown in an open area of the Experimental Farm at 20-28°C and an average noontime PPFD of 397.5 μmol·m⁻²·s⁻¹. For slow-release fertilization, 3.0 g of Osmocote® was applied to each pot at monthly intervals. After treatment, plants with average height of 52.4 cm and nodes of 7.9 were transferred to a phytotron at 20/15°C (day/night temperature), and average noon PPFD was 150 μmol·m⁻²·s⁻¹.

Effect of Temperature Variation on Growth and Flowering

Plants were grown in the phytotrons at 30/25°C, 25/20°C or 20/15°C (day/night temperature), and exposed to the day temperature for 12 h (6 a.m. to 6 p.m.). Each treatment was applied to four potted plants. Each vine was treated weekly with Peter's solution (20:20:20 N:P:K; Scotts Company, Marysville, OH, USA). The growth rate, plastochron (the interval of time between the appearances of successive leaf primordia at the shoot apex), days to first flower formation and the number of flower buds were recorded weekly (Experiment 1).

Treatment with Silver Ion under Low Temperature (20/15°C)

Plants were sprayed with AgNO₃ or STS both at 0.5 or 1.0 mM, or distilled water (control) and then subjected to low temperature conditions (20/15°C) for one month. STS and AgNO₃ stock solutions were prepared as follows: 0.1 M STS stock solution was prepared by dissolving 1.58 g of STS in 100 mL of water, whereas 0.1 M AgNO₃ stock solution was prepared by dissolving 1.7 g of AgNO₃ in 100 mL of water. The stock solutions were stored in the dark until needed to prepare the STS. Each treatment was applied to five potted plants. Growth rate, plastochron and internode length were determined, and the days to the first flower bud formation, the node number to initial flower bud were recorded (Experiment 2).

ACC Content

Detached passion fruit leaves after AgNO₃ or STS treatments were used. ACC (1-aminocyclopropene-1-carboxylate) content was assayed using the method described by Lizada and Yang (1979). For each sample 0.5 g fresh weight (FW) of leaves was washed in a test tube with 5 mL of 80% (v/v) ethanol for subsequent extraction of leaves in a hot water bath at 70°C for 20 min, followed by centrifugation. This procedure was repeated twice, and the mixture of the two extracts was further concentrated under vacuum until it became anhydrous. Distilled water was then added

to a final volume of 1.0 mL. All samples were then stored in test tubes at 0°C to be analyzed within 2 weeks.

To perform the analyses, two 0.5 mL aliquots of each sample were mixed with 0.1 mL 50 mM HgCl₂. Then 0.1 mL 100 mM ACC was added to one of these sub-samples. Nothing was added to the other. The final volumes were brought to 1.8 mL by adding distilled water and the mixtures were placed in test tubes, which were sealed with sleeve stoppers. While the tubes were submerged in crushed ice, 100 µL of a 2:1 (v/v) mixture of iced 5% (v/v) NaOCl and saturated NaOH was added to each test tube. The contents were vortex mixed for 5 sec., then each tube was placed in crushed ice for 2.5 min. They were then vibrated for 1 sec. and 1 mL of the gas was extracted for analysis using a GC-14A gas chromatograph (Shimadzu, Japan) fitted with a flame ionization detector. The carrier gas was nitrogen at a flow rate of 0.42 mL·s⁻¹ and the standard gas (Hsinan Inc., Taipei, Taiwan) was 1.0 µL·L⁻¹ ethylene.

ACC Oxidase Activity

Detached passion fruit leaves after AgNO₃ or STS treatments were used. The ACC oxidase detection method was from a previously described method (Mekhedov and Kende,

1996; Ververdis and John, 1991). A 1.0 mL gas sample was injected into a GC-14A gas chromatograph (Shimadzu, Japan) fitted with a flame ionization detector.

Ethylene Production

Detached passion fruit leaf after temperatures (30/25°C, 25/20°C and 20/15°C) or AgNO₃ or STS treatments were placed in a 40 mL sealed plastic jar at 20-28°C for 2 h under cool-white fluorescent lamps providing 15-16 µmol·m⁻²·s⁻¹. A 1.0 mL gas sample was injected into a Shimadzu GC-14A gas chromatograph with a flame ionization detector. The carrier gas was nitrogen and the standard gas was 1 mg·L⁻¹ ethylene.

Experimental Design and Statistical Analyses

The experiments were arranged in a completely randomized design. The data were analyzed using ANOVA for comparing means of treatment variables at $p < 0.05$. CoStat 6.2 (CoHort Software, Monterey, CA, USA) was used for the analysis.

Results

Effects of Temperature Variation on Growth Rate

Shoot growth at 30/25°C was faster than either 25/20°C or 20/15°C, where the plants produced more leaves with shorter plastochrons and enhanced internode elongation (Table 1). With respect to the plastochron, leaf formation was slowest in the 20/15°C treatment (4.5 days), indicating that vegetative growth of the shoot increases with increasing temperatures (Table 1).

Effects of Temperature Variation on Flower Bud Formation

The mean days to the first flower bud and the total number of flower buds per plant did not significantly differ between 25/20°C and 30/25°C treatments (Table 2). However,

Table 1. Effects of day and night temperatures on shoot growth of potted 'Tai-nung No.1' passion fruit plants grown in phytotrons.

Temperature (°C)	Growth rate (cm/day)	Plastochron (days/node)	Internode length (cm)
30/25	5.5 a ^z	1.9 c	10.2 a
25/20	3.8 b	2.3 b	8.7 b
20/15	1.4 c	4.5 a	5.8 c
LSD _{0.05}	0.2	0.3	0.3

^zDifferent letters within columns indicate significantly different values according to LSD test at $p < 0.05$ ($n = 4$).

Table 2. Effects of growth temperature on floral bud formation of the potted 'Tai-nung No.1' passion fruit plants grown in phytotrons.

Temperature (°C)	Mean nodes to the first flower bud	Mean days to the first flower bud	No. of flower buds	Days from visible flower bud to anthesis	Flower buds aborted (%)
30/25	18.8 a ^z	20.9 a	7.3 a	35.6 a	68.3 a
25/20	15.3 b	18.7 b	7.1 b	30.6 b	38.7 b
20/15	0.0 c ^y	0.0 c	0.0 c	0.0 c	0.0 c
LSD _{0.05}	0.4	0.3	0.2	2.0	7.6

^zDifferent letters within columns indicate significantly different values according to LSD test at $p < 0.05$ ($n = 4$).

^yThe plants at 20/15°C had no flowers.

plants grown at 30/25°C had a higher rate of aborted flower buds than those grown at 25/20°C. No flower formation was observed on the plants at 20/15°C (Table 2).

Effects of Temperature Variations on Ethylene Production

Ethylene production in the leaves of passion fruit was significantly higher when plants were grown at the relatively lower temperature (20/15°C), compared to 25/20° and 30/25° temperatures (Fig. 1).

Effects of Silver Ion on Growth and Flowering

Shoot growth of passion fruit 'Tai-nung No.1' was significantly inhibited by both AgNO₃ and STS treatments at 20/15°C. The plants treated with AgNO₃ or STS had lower growth rates and higher plastochron values than controls (Table 3). Those treated with 1.0 mM AgNO₃ had the highest plastochron values followed by those treated with 1.0 mM STS, indicating that ethylene action inhibitors delayed leaf formation. However, AgNO₃ and STS enhanced flower formation in passion fruit compared to the untreated control where no flowers were formed (Table 4). Internode length were

not influenced (Table 3). All plants treated with AgNO₃ or STS formed their first flower buds approximately 2 weeks after treatment.

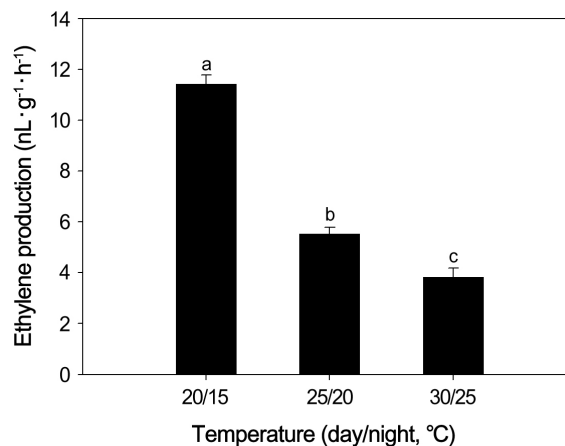


Fig. 1. Ethylene production from detached passion fruit leaves 7 days after being grown under the 30/25°C, 25/20°C, and 20/15°C (day/night) temperature conditions. Values represent means \pm SE (n=3). Different letters indicate significantly different values according to LSD test at $p < 0.05$.

Table 3. Effects of ethylene response inhibitors AgNO₃ and STS on shoot growth of potted 'Tai-nung No.1' passion fruit plants grown in a phytotron at 20/15°C day/night temperatures.

Treatments	Growth rate (cm·day ⁻¹) during:			Plastochron (days/node)	Internode length (cm)
	Early 14 days	Late 14 days	Whole period		
Control	0.7 a ^z	1.6 a	1.5 a	4.3 c	6.1 a
AgNO ₃ 0.5 mM	0.7 a	0.2 b	0.4 b	5.5 bc	5.6 a
AgNO ₃ 1.0 mM	0.5 a	0.1 b	0.3 b	11.1 a	5.2 a
STS 0.5 mM	0.2 a	0.3 b	0.5 b	7.0 b	5.9 a
STS 1.0 mM	0.3 a	0.1 b	0.6 b	10.2 a	5.9 a
LSD _{0.05}	0.7	0.9	0.6	2.7	1.8

^zDifferent letters within columns indicate significantly different values according to LSD test at $p < 0.05$ (n = 5).

Table 4. Effects of AgNO₃ and STS on floral bud formation of the potted 'Tai-nung No.1' passion fruit plants grown in a phytotron at 20/15°C day/night temperatures.

Treatments	Mean days to the first flower bud	Mean node no. to the first flower bud	No. of flower buds (per vine)
Control ^z	0.0 c ^y	0.0 b	0.0 b
AgNO ₃ 0.5 mM	16.2 ab	12.1 a	7.8 a
AgNO ₃ 1.0 mM	15.4 b	11.7 a	6.3 a
STS 0.5 mM	16.2 ab	12.5 a	7.8 a
STS 1.0 mM	17.0 a	11.9 a	6.8 a
LSD _{0.05}	1.1	2.9	3.1

^zThe control plants at 20/15°C had no flowers.

^yDifferent letters within columns indicate significantly different values according to LSD test at $p < 0.05$ (n = 5).

Effects of Silver Ion on ACC Content and ACC Oxidase under Low Temperature (20/15°C)

The ACC content and ACC oxidase activity in the leaves of passion fruit plants exposed to the 20/15°C temperature condition were significantly decreased by both AgNO₃ and STS treatments throughout 19 days of treatments (Fig. 2).

Effects on Ethylene Production under Low Temperature (20/15°C).

Low temperature treatment at 20/15°C obviously increased ethylene production during the first 7 days (Fig. 3). AgNO₃ and STS treatments significantly retarded ethylene production.

Discussion

This study revealed that temperature significantly affects vegetative growth and flowering of passion fruit. High

temperatures accelerate vegetative growth of passion fruit. These observations are consistent with those of Meinke and Karnatz (1990), Menzel et al. (1986, 1987), Simon and Karnatz (1983), and Utsunomiya (1992). Utsunomiya (1992) studied the growth and flowering response of the cool-loving purple passion fruit at 23/18°C, 28/23°C, and 33/28°C, and found that the lower temperature treatment (23/18°C) promoted flowering and fruit set, while the higher temperature treatments (28/23°C or 33/28°C) resulted in faster fruit development and better fruit quality. In this study, we investigated the growth and flowering response of the heat-tolerant 'Tai-ning No. 1' at 20/15°C, 25/20°C, and 30/25°C. We found that low temperature (20/15°C) inhibited flowering in 'Tai-ning No. 1' passion fruit, while higher temperatures (25/20°C and 30/25°C) promoted flowering. We also found that application of ethylene response inhibitor (AgNO₃ or STS) under low temperature induced flowering. Thus, the results of this study are significantly different from that found by Utsunomiya (1992). This significant difference in flowering is most probably due to different climatic preferences between these two different types of passion fruit, one cool-loving and the other heat-tolerant.

In this study, plants grown at 25/20°C exhibited the earliest flower bud formation than those grown at higher (30/25°C) or lower (20/15°C) temperature regimes, and produced their first flower buds on lower nodes and had lower abortion rate of flower buds. The same effects were noted in tomatoes (Calvert, 1959; Hussey, 1963a; Ohyama et al., 2005). According to previous studies, flower formation

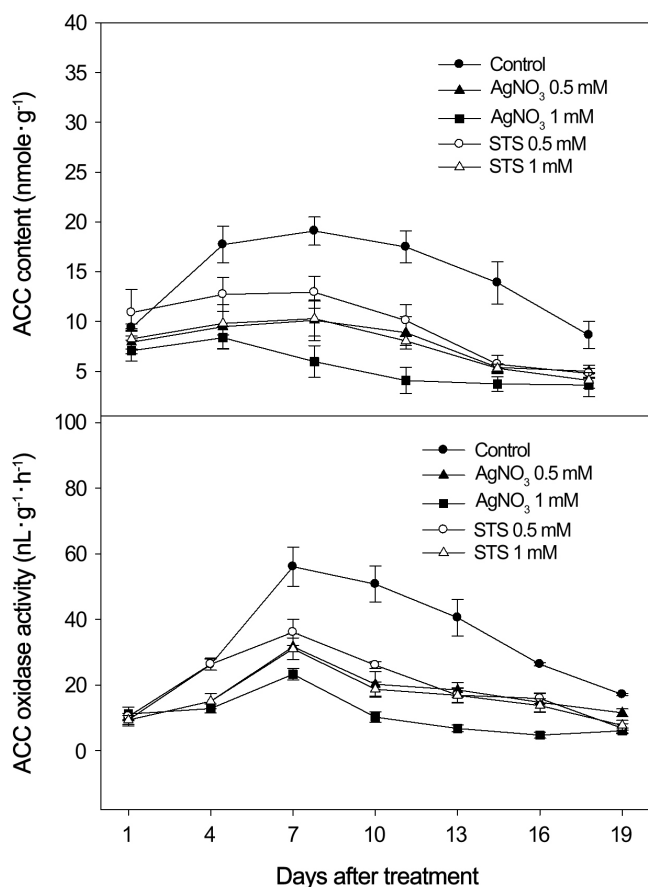


Fig. 2. Endogenous ACC content and the in vitro activity of ACC oxidase in the detached leaves of passion fruit after AgNO₃ or STS treatments. The plants were grown in a phytotron with 20/15°C (day/night) temperature conditions. Values represent means ± SE (n = 3).

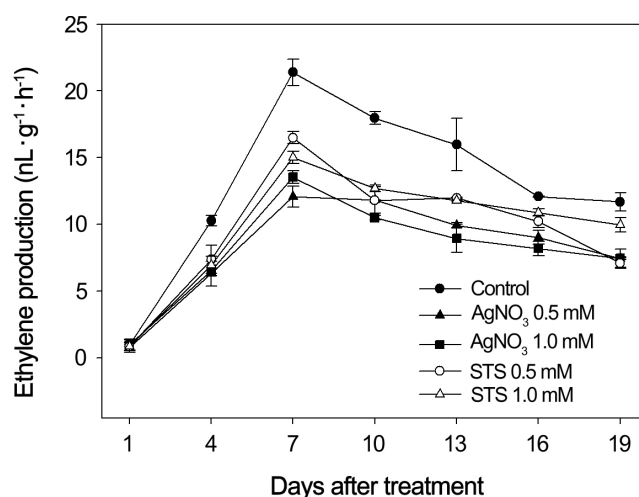


Fig. 3. Ethylene production from the detached leaves of passion fruit plants following AgNO₃ or STS treatments. The plants were grown in a phytotron with 20/15°C (day/night) temperature conditions. Values represent means ± SE (n = 3).

in passion fruit was affected by a critical range (20-30°C) of temperatures (Chang and Cheng, 1992). Beyond this range, no flower formation is induced (Chang and Cheng, 1992). This confirms the results of previous study demonstrating the inhibition of the growth and flowering of the passion fruit species during winter in Taiwan (Chang and Cheng, 1992).

An interesting question is why plants grown at higher temperatures form their initial flower buds on relatively higher nodes and have higher abortion rates of flower buds (Table 2). Hussey (1963a, 1963b) proposed that developing leaves and apical meristems of *Lycopersicon esculentum* compete vigorously for assimilates. In the case of tomato, high temperatures are beneficial for proliferation of leaf primordia, while low temperatures or the removal of young leaves may cause early transformation of apical meristems to inflorescence (Hussey, 1963a, 1963b; Sawhney, 1983). Thus we propose that passion fruit plants grown at high temperatures exhibit more vigorous vegetative growth by expending more nutrition on the vegetative growth of stems and leaves, which reduces nutrition available for flower formation. Therefore, more leaves are needed to produce more assimilates to support the formation and development of flower buds. Also plant hormone is related to floral induction. More vigorous growth produces more GA and/or auxin out of more young tissues, which delays floral initiation (Zhu and Davies, 1997). Additionally, treating tomato plants with gibberellic acid (GA) reduced flower bud formation and caused them to produce their first flower buds on higher nodes (Abdul and Harris, 1978; Sasaki et al., 2005). At high temperature, young leaves of tomato contained relatively higher GA content leading to enhanced plant growth, while flower formation and development were suppressed (Abdul and Harris, 1978; Beppu et al., 2001; Leonard and Kinet, 1982). The same effects were observed in grape (Ziv et al., 1981). Additionally, we found that treating passion fruit with paclobutrazol (gibberellin biosynthesis inhibitor) suppressed branch growth, caused the plants to produce their first flower buds on lower nodes, and promoted flower bud formation (data not shown). These observations may be associated with increases of GA in passion fruit grown at high temperatures. Based on the nutrient diversion hypothesis proposed by Sachs (1977), GA-induced vegetative growth caused the competition for nutrients between the formation of flower buds and the growth of branches and leaves; hence, enhanced growth of vegetative tissues will suppress the formation and development of flower buds.

Light intensity is another important factor affecting flower

formation. In Experiment 1, the passion fruit plants were transferred to different temperature under high light intensity ($400 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) in the summer, and flower formation occurred more quickly at 30/25°C than at 25/20°C (data not shown). The number of flower buds did not significantly differ between 30/25°C and 25/20°C treatments. Additionally, no flower formation occurred among passion fruit plants transferred to three different temperature regimens (30/25°C, 25/20°C or 20/15°C) under low light intensity ($100\text{-}150 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) (data not shown). With low light intensity below a critical range, flowering of passion fruit was not induced at any temperature. Therefore, light intensity may moderate the impact of temperature on flowering of passion fruit. The same effects were observed in tomato (Calvert, 1959; Hussey, 1963a; Ohyama et al., 2005). In tomato, inflorescence occurred much earlier at higher temperatures (25°C) than at lower temperatures (15°C) under $318 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, but occurred more slowly at $159 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ (Hussey, 1963a). At low light intensity ($150 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$), floral development of tomato was enhanced as average air temperature decreased (Ohyama et al., 2005). High temperature apparently delayed flower formation and development under very low light intensity ($50 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) (Calvert, 1959).

Ethylene is a plant hormone that regulates many aspects of plant growth and development ranging from seed germination to organ senescence (Sisler and Yang, 1984). Ethylene promotes or inhibits vegetative growth (Jackson et al., 1981; Wheeler et al., 1996), promotes or inhibits the initiation of flowering (Elfving et al., 2004; Saltveit, 1999), and plays an important role in fruit maturity (Ritenour et al., 1999). Ethylene is also responsible for the transfer from the vegetative to the flowering stage in bougainvillea (Liu and Chang, 2011b). These indicate that ethylene is bi-directionally involved in regulating the flowering control of some plants. In Experiments 1 and 2, plants exhibited no flower formation at low temperature (20/15°C). The production of ethylene in the leaves of passion fruit 'Tai-nung No.1' at low temperature (20/15°C) was significantly higher than that at other temperatures (25/20°C and 30/25°C). All plants treated with AgNO_3 or STS formed their first flower buds starting approximately 2 weeks after treatment, even in the 20/15°C conditions. These data indicate that silver ion, an inhibitor of ethylene action, promotes flower formation in passion fruit at low temperatures. Additionally, ACC content, ACC oxidase activity and ethylene production in the leaves of passion fruit 'Tai-nung No.1' at low temperature (20/15°C) were significantly inhibited by those chemical treatments. These results suggest that suppressed vegetative growth led to greater availability of nutrients for reproductive

growth of plants under the low temperature condition. It seems that ethylene may play a positive role in the vegetative growth of passion fruit under the low temperature condition. The chemical treatment could enable transfer of plant nutrients to flower bud formation via reduction in ACC content, ACC oxidase activity and ethylene production.

Passion fruit leaves, stems or flowers produced similar amounts of ethylene by the end of the experiment (data not shown). Therefore, ethylene production in the passion fruit leaves is a good indicator for the entire shoot. The same effects were noted in bougainvillea (Liu and Chang, 2011a).

Although the number of flower buds in the STS-treated plants was comparable to that in AgNO₃-treated plants, the leaves of STS-treated plants exhibited less sign of chemical toxicity than those of AgNO₃-treated plants (data not shown). This indicated that the anionic complex STS, which is mobile and less phytotoxic in plant transport system (Veen, 1983, 1987), has less chemical toxicity than AgNO₃ on the leaves of *Passiflora* spp. Silver, applied in the form of thiosulfate, is a very effective inhibitor of ethylene responses, but this heavy metal cannot be used on food and feed, and has been objected by environmentalists. Recently, a non-toxic antagonist of ethylene action, 1-methylcyclopropene (1-MCP), was discovered and has since been widely used (Blankenship and Dole, 2003). Therefore, the use of AgNO₃ or STS can be substituted with 1-MCP for future applications to avoid silver ion toxicity.

In conclusion, temperature has significant effects on the growth and flowering of *Passiflora*. Ethylene production is easily induced by low temperature in *Passiflora* plants, which are especially chilling-sensitive plants (Chen and Patterson, 1985). Our study suggests that ethylene is an important inhibitor of flowering in *Passiflora* spp. Using ethylene action inhibitors offers the possibility of inducing flower formation under low temperature conditions in *Passiflora* plants.

Literature Cited

- Abdul, K.S. and G.P. Harris. 1978. Control of flower number in the first inflorescence of tomato (*Lycopersicon esculentum* Mill.): the role of gibberellins. *Ann. Bot.* 42:1361-1367.
- Barbosa, W.M., W.C. Otoni, M. Camelossi, E. Silva, A.A. Azevedo, and G. Vieira. 2001. Rhizogenesis in *in vitro* shoot cultures of passionfruit (*Passiflora edulis* f. *flavicarpa* Deg.) is affected by ethylene precursors and by inhibitors. *Int. J. Hort. Sci.* 7:47-51.
- Beppu, K., T. Suehara, and I. Kataoka. 2001. Embryo sac development and fruit set of 'Satohnoshiki' sweet cherry as affected by temperature, GA₃, and paclobutrazol. *J. Jpn. Soc. Hort. Sci.* 70:157-162.
- Beyer, E.M. 1976. A potent inhibitor of ethylene action in plants. *Plant Physiol.* 58:268-271.
- Blankenship, S.M. and J.M. Dole. 2003. 1-Methylcyclopropene: a review. *Postharvest Biol. Technol.* 28:1-25.
- Calvert, A. 1959. Effect of the early environment on development of flowering in the tomato. *J. Hort. Sci.* 32:9-17.
- Chang, Y.S. and C.Y. Cheng. 1992. Effects of temperature and light on growth and flower formation of passionfruit. *J. Chinese Soc. Hort. Sci.* 38:30-36.
- Chen, Y.Z. and B.D. Patterson, 1985. Ethylene and 1-aminocyclopropane-1-carboxylic acid as indicators of chilling sensitivity in various plant species. *Aust. J. Plant Physiol.* 12:377-385.
- Elfving, D.C., D.B. Visser, M.D. Whiting, and G.A. Lang. 2004. Growth and flowering responses of sweet cherry cultivars to prohexadione-calcium and ethephon. *Acta Hort.* 636:75-82.
- Hussey, G. 1963a. Growth and development in the young tomato. I. The effect of temperature and light intensity on growth of the shoot apex and leaf primordia. *J. Exp. Bot.* 14:316-325.
- Hussey, G. 1963b. Growth and development in the young tomato. II. The effect of defoliation on the development of the shoot apex. *J. Exp. Bot.* 14:326-333.
- Jackson, M.B., M.C. Drew, and S.C. Giffard. 1981. Effects of applying ethylene to the root system of *Zea mays* on growth and nutrient concentration in relation to flooding tolerance. *Physiol. Plant* 52:23-28.
- Leonard, M. and J.M. Kinet. 1982. Endogenous cytokinin and gibberellin levels in relation to inflorescence development in tomato. *Ann. Bot.* 50:127-130.
- Liu, F.Y. and Y.S. Chang. 2011a. Effects of shoot bending on ACC content, ethylene production, growth and flowering of bougainvillea. *Plant Growth Regul.* 63:37-44.
- Liu, F.Y. and Y.S. Chang. 2011b. Ethephon treatment promotes flower formation in bougainvillea. *Bot. Stud.* 52:183-189.
- Lizada, M.C.C. and S.F. Yang. 1979. A simple and sensitive assay for 1-aminocyclopropane-1-carboxylic acid. *Anal. Biochem.* 100:140-145.
- MacRae, E.A., A.K. Hardacre, and I.B. Ferguson. 1986. Comparison of chlorophyll fluorescence with several other techniques used to assess chilling sensitivity in plants. *Physiol. Plant.* 67:659-665.
- Max, G., V. Acuña, W.V. Biasi, E.J. Mitchama, and D. Holcroft. 2011. Fruit temperature and ethylene modulate 1-MCP response in 'Bartlett' pears. *Postharvest Biol. Technol.* 60:17-23.
- Meinke, H. and A. Karnatz. 1990. Influence of air and soil temperatures on grafted and self-rooted *Passiflora* hybrids. *Sci. Hort.* 43:237-246.
- Mekhedov, S.L. and H. Kende. 1996. Submergence enhances expression of a gene encoding 1-aminocyclopropane-1-carboxylate oxidase in deepwater rice. *Plant Cell Physiol.* 37:531-534.
- Menzel, C.M. and D.R. Simpson. 1988. Effect of continuous shading

- on growth, flowering and nutrient uptake of passionfruit. *Sci. Hortic.* 35:77-88.
- Menzel, C.M. and D.R. Simpson. 1994. Passionfruit, p. 225-241. In: B. Schaffer and P.C. Andersen (ed.). *Handbook of environmental physiology of fruit crops. Vol. II: Sub-tropical and tropical crops.* CRC Press, Florida.
- Menzel, C.M., D.R. Simpson, and A.J. Dowling. 1986. Water relations in passionfruit: effect of moisture stress on growth, flowering and nutrient uptake. *Sci. Hortic.* 29:239-249.
- Menzel, C.M., D.R. Simpson, and C.W. Winks. 1987. Effect of temperature on growth, flowering and nutrient uptake of passionfruit. *Sci. Hortic.* 31:259-268.
- Nave, N., E. Katz, N. Chayut, S. Gazit, and A. Samach. 2010. Flower development in the passionfruit *Passiflora edulis* requires a photoperiod-induced systemic graft-transmissible signal. *Plant Cell Environ.* 33:2065-2083.
- Ohyama, K., Y. Omura, and T. Kozai. 2005. Effect of air temperature regimes on physiological disorders and floral development of tomato seedlings grown under continuous light. *HortScience* 40:1304-1306.
- Orihuel-Iranzo, B., M. Miranda, L. Zacarías, and M.T. Lafuente. 2010. Temperature and ultra-low oxygen effects and involvement of ethylene in chilling injury of 'Rojo Brillante' persimmon fruit. *Food Sci. Technol.* 16:159-167.
- Reis, L.B., V.B.P. Neto, E.A.T. Picoli, M.G.C. Costa, M.M. Reso, C.R. Carvalho, F.L. Finger, and W.C. Otoni. 2003. Axillary bud development of passionfruit as affected by ethylene precursor and inhibitors. *In Vitro Cell Dev. Biol.* 39:618-622.
- Ritenour, M.A., C.H. Crisosto, D.T. Garner, G.W. Cheng, and J.P. Zoffoli. 1999. Temperature, length of cold storage and maturity influence the ripening rate of ethylene-preconditioned kiwifruit. *Postharvest Biol. Technol.* 15:107- 115.
- Sachs, R.M. 1977. Nutrient diversion: a hypothesis to explain the chemical control of flowering. *HortScience* 12:220-222.
- Saltveit, M.E. 1999. Effect of ethylene on quality of fresh fruits and vegetables. *Postharvest Biol. Technol.* 15:279-292.
- Sasaki, H., T. Yano, and A. Yamasaki. 2005. Reduction of high temperature inhibition in tomato fruit set by plant growth regulators. *JARQ* 39:135-138.
- Sawhney, V.K. 1983. The role of temperature and its relationship with gibberellic acid in the development of floral organs of tomato (*Lycopersicon esculentum*). *Can. J. Bot.* 61:1258-1265.
- Simon, P. and A. Karnatz. 1983. Effect of soil and air temperature on growth and flower formation of purple passionfruit (*Passiflora edulis* Sims var. *edulis*). *Acta Hortic.* 139:83-90.
- Sisler, E.C. and S.F. Yang. 1984. Ethylene, the gaseous plant hormone. *BioScience* 34:234-238.
- Staveley, G.W. and B.N. Wolstenholme. 1990. Effect of water stress on growth and flowering of *Passiflora edulis* Sims. grafted to *P. caerulea* L. *Acta Hort.* 275:551-557.
- Tacken, E., H. Ireland, K. Gunaseelan, S. Karunairetnam, D. Wang, K. Schultz, J. Bowen, R.G. Atkinson, J.W. Johnston, J. Putterill, R.P. Hellens, and R.J. Schaffer. 2010. The role of ethylene and cold temperature in the regulation of the apple POLYGALACTURONASE1 gene and fruit softening. *Plant Physiol.* 153:294-305.
- Utsunomiya, N. 1992. Effect of temperature on shoot growth, flowering and fruit growth of purple passionfruit (*Passiflora edulis* Sims var. *edulis*). *Sci. Hortic.* 52:63-68.
- Veen, H. 1983. Silver thiosulphate: an experimental tool in plant science. *Sci. Hortic.* 20:211-224.
- Veen, H. 1987. Use of inhibitors of ethylene action. *Acta Hortic.* 201:213-222.
- Ververdis, P. and P. John. 1991. Complete recovery in vitro of ethylene-forming enzyme activity. *Phytochemistry* 30:725-727.
- Wheeler, R.M., B.V. Peterson, J.C. Sager, and W.M. Knott. 1996. Ethylene production by plants in a closed environment. *Adv. Space Res.* 18:193-196.
- Zhu, Y.X. and P.J. Davies. 1997. The control of apical bud growth and senescence by auxin and gibberellin in genetic lines of peas. *Plant Physiol.* 113:631-637.
- Ziv, M.M., Z. Bernstein, and S. Lavee. 1981. Necrosis in grapevine buds (*Vitis vinifera* cv. Queen of Vineyard). II. Effect of gibberellic acid (GA₃) application. *Vitis* 20:105-114.