

Bactericidal Efficacy of a Disinfectant Composed of Povidone-iodine Against *Clostridium Perfringens* and *Mycobacterium Fortuitum*

Chun-Nam Cha¹, Eun-Kee Park², Youyoung Cho³, Chang-Yeul Yoo⁴, Engin Tutkun⁵, Suk Kim⁶, and Hu-Jang Lee^{6*}

¹Engineering Research Institute, Department of Industrial Systems Engineering,
Gyeongsang National University, Chinju 600-701, Korea

²Department of Medical Humanities and Social Medicine, College of Medicine, Kosin University, Busan 602-703, Korea

³Department of Nursing, Hanyeong College, Yeosu 550-704, Korea

⁴Department of Computer Information, Gyeongnam Provincial Namhae College, Namhae 668-801, Korea

⁵Ankara Occupational Diseases Hospital, Ministry of Health, Ankara 06200, Turkey

⁶Research Institute of Live Sciences, College of Veterinary Medicine, Gyeongsang National University, Chinju 600-701, Korea

(Received October 4, 2014/Revised November 19, 2014/Accepted December 9, 2014)

ABSTRACT - *Clostridium perfringens* (*C. perfringens*) and *Mycobacterium fortuitum* (*M. fortuitum*) are associated with considerable diseases in animals and human. In this study, the disinfection efficacy of a commercial disinfectant composed to povidone-iodine (PVI) was evaluated against *C. perfringens* and *M. fortuitum*. A bactericidal efficacy test by broth dilution method was used to determine the lowest effective dilution of the disinfectant following exposure to *C. perfringens* and *M. fortuitum* for 30 min at 4°C. The disinfectant and test bacteria were diluted with hard water (HW) or organic matter suspension (OM) according to treatment condition. On HW condition, the bactericidal activity of the disinfectant against *C. perfringens* and *M. fortuitum* was 50 and 80 fold dilutions, respectively. On OM condition, the bactericidal activity of the disinfectant against both *C. perfringens* and *M. fortuitum* was 15 fold dilutions. As the disinfectant composed to PVI possesses bactericidal efficacy against *C. perfringens* and *M. fortuitum*, the disinfectant solution can be used to control the spread of bacterial diseases.

Key words : Povidine-iodine, *Clostridium perfringens*, *Mycobacterium fortuitum*, Disinfectant efficacy

Clostridium perfringens (*C. perfringens*) is a Gram-positive, rod-shaped, anaerobic, spore-forming bacterium of the genus *Clostridium*¹⁾. The bacterium is ever present in nature and can be found as a normal component of decaying vegetation, marine sediment, and the intestinal tract of humans and other vertebrates²⁾.

C. perfringens is one of the most frequently isolated bacterial pathogens in foodborne disease outbreaks in humans^{3,4)}. *C. perfringens* is reported to be the third most common cause of foodborne illness in the world⁵⁾ and annually ranks as one of the most common causes of food poisoning in the industrialized world⁶⁻⁸⁾. Different meats, including poultry meat, have frequently been reported as the most common food vehicles^{5,9)}. The common occurrence of *C. perfringens* in raw meat and retail foods may be due to the contamination of carcasses and meat with the intestinal contents of the animals during the slaughtering process¹⁰⁾.

Mycobacterium fortuitum (*M. fortuitum*) is a rapidly growing mycobacterium and broadly presents world-wide in various habitats such as lakes, surface water, potable water and soil¹¹⁾. In addition, *M. fortuitum* has been detected in the environment of pig farms and unpasteurized milk¹²⁾. Furthermore, *M. fortuitum* causes nosocomial infections in human throughout contaminated water sources in hospitals¹³⁾. *M. fortuitum* isolations are not uncommon in normal or diseased domestic and pet animals. The organism has been isolated from skin and diseased organs and also from fluids and excretions of dogs, cats, horses, cattle, pigs and nearby birds and fish including salmonids^{12,14)}.

Epidemiological evidence suggests that *C. perfringens* enterotoxin (CPE) plays an important role in the pathogenesis of both food-borne and non-food-borne human gastrointestinal (GI) illnesses caused by *cpe*-positive *C. perfringens* type A isolates¹⁵⁾. Specifically, feces from patients suffering from GI illnesses caused by *cpe*-positive isolates contain CPE at concentrations that cause GI effects in animal models¹⁶⁾. A previous study demonstrated that *M. fortuitum* induced strong apoptosis in human acute monocytic leukemia cells (THP-1) and murine bone marrow-derived macrophages and dendritic cells¹⁷⁾. In murine infection

*Correspondence to: Hu-Jang Lee, College of Veterinary Medicine, Gyeongsang National University, 900 Gajwa-dong, Chinju 660-701, Korea
Tel: 82-55-772-2352, Fax: 82-55-772-2308
E-mail: hujang@gnu.ac.kr

model, *M. fortuitum* did not invade recombination activating genes (RAG) murine kidney cell line, while the bacilli infected and proliferated freely inside murine macrophages¹⁸.

In the food industry, thermal treatments typically applied during meat processing are sufficient to kill *C. perfringens* vegetative cells. However, its spores may survive and become highly committed to germinate, outgrow, and multiply to hazardous levels^{19,20}. These heat-activated spores are a major concern to food processors, especially during improper chilling after cooking or stored under abusive temperature, conditions that allow a rapid proliferation of *C. perfringens*²¹. In bovine milk, the presence of *Mycobacterium* spp. including *M. fortuitum* has emerged as a public-health concern, especially among individuals who consume raw milk and related dairy products²².

The wide use of antimicrobial agents in raises an important question regarding the emergence and spread of antimicrobial resistance primarily in the resident normal enteric flora²³. Antibiotic-resistance of *C. perfringens* and *M. fortuitum* isolates has been reported in several countries^{24,25}.

As antibiotic-resistant strains of *C. perfringens* and *M. fortuitum* have been increasing due to abuse and overuse of antibiotics, the effective cleaning and disinfection regimes are essential for the prevention of infections and outbreaks^{26,27}. As not all disinfectant are effective against the major pathogens including *C. perfringens* and *M. fortuitum*, the choice of disinfectants is critical in establishing a successful sanitation program. Biocides are often composed of a mixture of ingredients that act upon a wide range of cellular mechanisms and targets, which makes it difficult for bacteria to become resistant to biocides²⁸.

Several disinfectants including chlorine dioxide, betaine hydrochloride and propylene glycol have been used for decontamination of farmed animal and food borne diseases^{29,30}. However, there is not the efficacy test for the disinfectant composed of povidone-iodine (PVI) against *C. perfringens* and *M. fortuitum*. Therefore, this study was carried out to examine bactericidal efficacy of a disinfectant solution composed to PVI against *C. perfringens* and *M. fortuitum*.

Materials and methods

Bacteria and culture

C. perfringens (KVCC-BA1100001) and *M. fortuitum* (ATCC 6841) were obtained from the Korean Veterinary Culture Collection (KVCC, Anyang, Korea) and Rural Development Administration (RDA)-Genebank Information Center (Jeonju, Korea), respectively. The strains were maintained as frozen glycerol stock. *C. perfringens* and *M. fortuitum* were cultured in cooked meat medium (BD Korea

Co., Ltd., Incheon, Korea) and Middlebrook 7H9 broth (BD-Korea, Seoul, Korea), respectively, and incubated at 37°C under 5% CO₂ condition for 24 h and 7 days, correspondingly. After incubation, the number of bacteria in culture was measured using the McFarland method³¹.

Disinfectant

The active ingredient for the tested disinfectant solution, Betadine® Solution-Concentrate, is PVI 10 g per 100 mL. The disinfectant was provided by Korea Pharma Co., Ltd. (Seoul, Korea). The disinfectant solution was stored in the dark in room temperature and prepared for dilution on the day of evaluation. Determination of the bactericidal efficacy of the disinfectant was based on Animal and Plant Quarantine Agency (Anyang, Korea), Regulation No. 2013-34.

Diluents and treatment condition

Testing was based on bactericidal effects of disinfectant diluents in two treatment conditions (standard hard water (HW) condition and organic matter (OM) condition) and pathogen control (disinfectant negative control) in Table 1. HW, an ingredient of HW treatment condition, was made by adding anhydrous CaCl₂ 0.305 g and MgCl₂·6H₂O 0.139 g into one liter distilled water. Organic suspension, an ingredient of OM treatment condition, is a solution of 5% (w/v) yeast extract in HW. The test organism was prepared by titration of cultural broth into at least 10⁸ colony forming unit (CFU)/mL viable organisms with the same kind of diluents of treatment condition.

Experimental procedures

For the efficacy test against *C. perfringens*, the disinfectant was diluted 40, 45, 50, 55, 60 and 65 times with HW, and diluted 12.0, 13.5, 15.0, 16.5, 18.0 and 19.5 times with OM, respectively. And the disinfectant for the test against *M. fortuitum* was diluted 64, 72, 80, 88, 96 and 104 times with HW, and diluted 12.0, 13.5, 15.0, 16.5, 18.0 and 19.5 times with OM, respectively.

To verify the lowest effective dilution of the disinfectant, five serial dilutions of the disinfectant were prepared and

Table 1. Experimental design for the determination of the bactericidal efficacy of the disinfectant composed to PVI

Treatment condition*	Contents according to treatment condition [†]			
	HW	OM	Disinfectant	Bacteria
HW condition	+	-	+	+
OM condition	+	+	+	+
Bacteria control	+	-	-	+

*HW: standard hard water, OM: organic matter.

[†]+: presence, -: absence

placed at 4°C prior to test reaction. 2.5 ml of each disinfectant dilution was mixed with the same amount of test organism followed by contact time of 30 min at 4°C. During this period, the mixture was shaken at 10 min interval. At the end of 30 min contact period, one mL of the mixture was neutralized with 9 mL of Nutrient broth containing 5% inactivated horse serum (BD Korea Co., Ltd., Incheon, Korea) at 37°C. 0.1 mL of the neutralized reaction mixture of *C. perfringens* and *M. fortuitum* was subcultured into 10 mL of recovery each cultural broth at 37°C, 5% CO₂ for 24 h and 7 days, respectively, in incubator. For the test of each bacteria control, 2.5 ml of hard water was mixed with the same amount of each test organism followed by contact time of 30 min at 4°C, and then all procedure were undertaken in parallel for the disinfection test.

The valid dilution of the disinfectant was determined that the greatest dilution showing no growth in four or more in the five replicates was confirmed. The final dilution time was statistically determined by a median value among three valid dilution of the triplicate test, but each value of which

should be within 20% experimental error. In each bacteria control, the number of bacterial growth in the five replicates was counted.

Results

Table 2 and 3 shows the final valid dilution of the disinfectant composed to PVI. When the bactericidal effect on HW condition was evaluated, the antibacterial activity of the disinfectant against *C. perfringens* and *M. fortuitum* showed on 50 and 80 fold dilutions, respectively. With the investigation of the bactericidal effect of the disinfectant on OM condition, both *C. perfringens* and *M. fortuitum* were inactivated on 15 fold dilutions. Because the organic material interferes with efficacy by either inactivating the disinfectant or blocking it from surface contact, the bactericidal activity of the disinfectant on the OM condition was lowered against pathogenic bacteria compared with HW conditions. In the bacterial control, the growth of *C. perfringens* and *M. fortuitum* was verified in all replicates.

Table 2. Final valid dilution of the disinfectant composed to PVI against *C. perfringens*

Bacterial strains	Treatment condition ¹⁾										
	HW			OM			BC				
	DT	1	2	3	DT	1	2	3	1	2	3
<i>C. perfringens</i>	40	×	×	×	12.0	×	×	×	+ [‡]	+	+
	45	×	×	×	13.5	×	×	×	+	+	+
	50	×	×	×	15.0	×	×	×	+	+	+
	55	o ²⁾	× ³⁾	o	16.5	o	o	×	+	+	+
	60	o	o	o	18.0	o	o	o	+	+	+
	65	o	o	o	19.5	o	o	o	+	+	+
	Valid	50			Valid	15.0			+		

¹⁾ HW: standard hard water; OM: organic matter; BC: bacterial control; DT: dilution time.

²⁾ o: growth, ×: growth inhibition.

³⁾ +: all growth in each replicate.

Table 3. Final valid dilution of the disinfectant composed to PVI against *M. fortuitum*

Bacterial strains	Treatment condition ¹⁾										
	HW			OM			BC				
	DT	1	2	3	DT	1	2	3	1	2	3
<i>M. fortuitum</i>	64	×	×	×	12.0	×	×	×	+ [‡]	+	+
	72	×	×	×	13.5	×	×	×	+	+	+
	80	×	×	×	15.0	×	o	×	+	+	+
	88	o ²⁾	× ³⁾	o	16.5	o	o	o	+	+	+
	96	o	o	o	18.0	o	o	o	+	+	+
	104	o	o	o	19.5	×	×	×	+	+	+
	Valid	80			Valid	15.0			+		

¹⁾ HW: standard hard water; OM: organic matter; BC: bacterial control; DT: dilution time.

²⁾ o: growth, ×: growth inhibition.

³⁾ +: all growth in each replicate.

Discussion

The disinfectant composed to PVI is a potential anti-bacterial disinfectant. PVI is a stable chemical complex of polyvinylpyrrolidone (povidone, PVP) and elemental iodine³². In addition, PVI is a highly efficient broad-spectrum germicidal agent and effective against bacteria, viruses, fungi, and protozoa³³. It is widely-used for topical cleansing and wound treatment. PVI releases free iodine, which has an important role in the bactericidal effect of PVI solution through the oxidizing effects of released iodine on proteins and fatty acids³⁴. Similarly, through the cytotoxic effects of free iodine, PVI is also an effective tumoricidal agent that may be used as an irrigation fluid to eradicate free cancer cells during head, neck, and colorectal cancer surgery³⁵⁻³⁷. In addition, in contrast to various antibiotic substances which act on the cell walls, PVI not only destroys bacteria, but also effectively inhibits the release of pathogenic factors such as exotoxins, endotoxins and tissue-destroying enzymes³⁸.

Although the slow release of iodine from the PVI complex in solution minimizes iodine toxicity towards mammalian cells, the iodine is delivered to the bacterial cell surface where it penetrates the cell membrane and inactivates key cytosolic proteins, fatty acids, and nucleotides³⁹. In the previous study⁴⁰, the oral toxicity LD₅₀ of PVI for both rats and mice was 10 g/kg body weight, and PVI containing available iodine 6 g/L was non-irritating to rabbit skin and eye.

To determine a practical minimal disinfecting concentration for 10% PVI over different contact times and temperatures, Heiner et al.⁴¹ exposed *Escherichia coli* to various dilutions of 10% PVI for 5, 15, and 30 min at 10, 20, and 30°C, neutralized with 0.5% sodium thiosulfate, and determined mean viable colony forming units (CFUs). In results, no CFUs were observed after exposure to the 1:100 dilutions and after 15 min of exposure to the 1:1,000 dilutions across experimental temperatures. In addition, Udombijitkul et al.⁴² carried out the sporicidal efficacy of disinfectants used in food-processing facilities and domestic kitchens against different *C. perfringens* strains on stainless steel surfaces. Therefore, treatment with 25 ppm iodophore sanitizer (1.6% iodine) for 10 min was achieved 2.7 log *C. perfringens* spore reduction, which was the highest reduction than other disinfectants composited to quaternary ammonium compounds or hydrogen peroxide. Phillips and von Reyn⁴³ reported that PVI at the concentration of 50-150 ppm as free iodine is known for their superior disinfectant activity against *Mycobacterium* spp. including *M. fortuitum*.

With the consideration of previous studies, the disinfectant composed to PVI is a more effective and safe disinfectant than quaternary ammonium compounds and hydrogen peroxide against *C. perfringens* and *M. fortuitum*.

In the present study, the disinfectant efficacy of the disinfectant composed to PVI has limitation that the results are based on *in vitro* test. Organic material in suspension (OM condition) could not represent all possible parameters of *C. perfringens* and *M. fortuitum* contaminated environment.

As the efficacy of the disinfectant composed to PVI against *C. perfringens* and *M. fortuitum* was investigated *in vitro*, a controlled field trial is required to determine whether use of the disinfectant will be able to reduce new pathogenic bacteria infection in animal farm and food industry area.

요 약

Clostridium perfringens (*C. perfringens*)와 *Mycobacterium fortuitum* (*M. fortuitum*)은 동물과 사람에서 심각한 질병과 관련이 있는 세균들로 알려져 있다. 본 연구에서는, povidone-iodine을 주성분으로 하는 소독제의 살균효과를 *C. perfringens*와 *M. fortuitum*을 대상으로 평가하였다. 소독제의 살균효과는 배지희석법을 이용하여, 대상 세균들을 4°C에서 소독제에 30분 동안 노출시킨 다음, 가장 낮은 소독제의 살균 희석배수를 결정하였다. 소독제는 경수와 유기물로 희석하였으며, 경수 조건에서, *C. perfringens*와 *M. fortuitum*에 대해 효과적인 소독제 희석배수는 각각 50과 80배이었다. 유기물 조건에서는, *C. perfringens*와 *M. fortuitum*에 대해 효과적인 소독제 희석배수는 모두 15배로 나타났다. 이상의 결과로부터, povidone-iodine을 주성분으로 하는 소독제는 *C. perfringens*와 *M. fortuitum*에 대해 살균효과를 갖는 것으로 확인되었으며, *C. perfringens*와 *M. fortuitum*에 의한 질병의 확산을 방지하기 위해 사용될 수 있을 것으로 사료된다.

Acknowledgements

This work was financially supported by Mundipharma Korea Co., Ltd. (Seoul, Korea).

References

1. Ryan, K.J.: *Clostridium, Peptostreptococcus, Bacteroides, and Other Anaerobes*. In Sherris Medical Microbiology - An Introduction to Infectious Disease, 4th Ed. (Ryan, K.J. and Ray, C.G. eds.) McGraw-Hill, New York, pp. 309-326 (2004).
2. Borah, D., Solanki, V. and Mishra, V.K.: Protein and molecular characterization of *Clostridium* spp. isolated from contaminated food and soil samples. *Int. J. Appl. Biol. Pharm. Technol.* **2**, 189-193 (2011).
3. Fisher, D.J., Miyamoto, K., Harrison, B., Akimoto, S., Sarker, M.R. and McClane, B.A.: Association of beta2 toxin production with *Clostridium perfringens* type A human gastrointestinal disease isolates carrying a plasmid enterotoxin gene.

- Vet. Microbiol.* **56**, 747-762 (2005).
4. Young, M.K., Smith, P., Holloway, J. and Davison, R.P.: An outbreak of *Clostridium perfringens* and the enforcement of food safety standards. *Communic. Dis. Intell.* **32**, 462-465 (2008).
 5. Osman, K.M., Soliman, Y.A., Amin, Z.M.S. and Aly, M.A. K.: Prevalence of *Clostridium perfringens* type A isolates in commercial broiler chickens and parent broiler breeder hens in Egypt. *Rev. Sci. Tech. Off. Int. Epiz.* **31**, 931-941 (2012).
 6. Eriksen, J., Zenner, D., Anderson, S.R., Grant, K. and Kumar, D.: *Clostridium perfringens* in London, July 2009: two weddings and an outbreak. *Euro surveill.* **15**, pii=19598 (2010).
 7. Miki, Y., Miyamoto, K., Kaneko-Hirano, I., Fujiuchi, K. and Akimoto, S.: Prevalence and characterization of enterotoxin gene-carrying *Clostridium perfringens* isolates from retail meat products in Japan. *Appl. Environ. Microbiol.* **74**, 5366-5372 (2008).
 8. Morris, W.E. and Fernández-Miyakawa, M.E.: Toxins of *Clostridium perfringens*. *Rev. Argent Microbiol.* **41**, 251-260 (2009).
 9. Cooper, K.K., Theoret, J.R., Stewart, B.A., Trinh, H.T., Glock, R.D. and Songer, J.G.: Virulence for chickens of *Clostridium perfringens* isolated from poultry and other sources. *Anaerobe* **16**, 289-292 (2010).
 10. Wen, Q. and McClane, B.A.: Detection of enterotoxigenic *Clostridium perfringens* type A isolates in American retail foods. *Appl. Environ. Microbiol.* **70**, 2685-2691 (2004).
 11. Smith, M.B., Schnadig, V.J., Boyars, M.C. and Woods, G.L.: Clinical and pathologic features of *Mycobacterium fortuitum* infections. An emerging pathogen in patients with AIDS. *Am. J. Clin. Pathol.* **116**, 225-232 (2001).
 12. Bercovier, H. and Vincent, V.: Mycobacterial infections in domestic and wild animals due to *Mycobacterium marinum*, *M. fortuitum*, *M. chelonae*, *M. porcinum*, *M. farcinogenes*, *M. smegmatis*, *M. scrofulaceum*, *M. xenopi*, *M. kansasii*, *M. simiae* and *M. genavense*. *Rev. Sci. Tech.* **20**, 265-290 (2001).
 13. Talaat, A.M., Trucksis, M., Kane, A.S. and Reimschuessel, R.: Pathogenicity of *Mycobacterium fortuitum* and *Mycobacterium smegmatis* to goldfish, *Carassius auratus*. *Vet. Microbiol.* **66**, 151-64 (1999).
 14. Cadmus, S.I., Adesokan, H.K., Okker, M. and Jahans, K.: *Mycobacterium fortuitum* from lesions of slaughtered pigs in Ibadan, Nigeria. *Rev. Sci. Tech.* **29**, 705-711 (2010).
 15. Fernández Miyakawa, M.E., Pistone Creydt, V., Uzal, F.A., McClane, B.A. and Ibarra, C.: *Clostridium perfringens* enterotoxin damages the human intestine *in vitro*. *Infect. Immun.* **73**, 8407-8410 (2005).
 16. Birkhead, G., Vogt, R.L., Heun, E.M., Snyder, J.T. and McClane, B.A.: Characterization of an outbreak of *Clostridium perfringens* food poisoning by quantitative fecal culture and fecal enterotoxin measurement. *J. Clin. Microbiol.* **26**, 471-474 (1988).
 17. Bohsali, A., Abdalla, H., Velmurugan, K. and Briken, V.: The non-pathogenic mycobacteria *M. smegmatis* and *M. fortuitum* induce rapid host cell apoptosis via a caspase-3 and TNF dependent pathway. *BMC Microbiol.* **10**, 237 (2010).
 18. Parti, R.P., Srivastava, S., Gachhui, R., Srivastava, K.K. and Srivastava, R.: Murine infection model for *Mycobacterium fortuitum*. *Microbes Infect.* **7**, 349-355 (2005).
 19. Udombijitkul, P., Paredes-Sabja, D. and Sarker, M.R.: Inhibitory effects of nisin against *Clostridium perfringens* food poisoning and nonfood-borne isolates. *J. Food Sci.* **77**, M51-M56 (2012).
 20. Sarker, M.R., Shivers, R.P., Sparks, S.G., Juneja, V.K. and McClane, B.A.: Comparative experiments to examine the effects of heating on vegetative cells and spores of *Clostridium perfringens* isolates carrying plasmid genes versus chromosomal enterotoxin genes. *Appl. Environ. Microbiol.* **66**, 3234-3240 (2000).
 21. Reddy Velugoti, P., Rajagopal, L., Juneja, V. and Thippareddi, H.: Use of calcium, potassium, and sodium lactates to control germination and outgrowth of *Clostridium perfringens* spores during chilling of injected pork. *Food Microbiol.* **24**, 687-694 (2007).
 22. Franco, M.M., Paes, A.C., Ribeiro, M.G., de Figueiredo Pantoja, J.C., Santos, A.C., Miyata, M., Leite, C.Q., Motta, R.G. and Listoni, F.J.: Occurrence of mycobacteria in bovine milk samples from both individual and collective bulk tanks at farms and informal markets in the southeast region of Sao Paulo, Brazil. *BMC Vet. Res.* **9**, 85 (2013).
 23. Slavić, D., Boerlin, P., Fabri, M., Klotins, K.C., Zoethout, J.K., Weir, P.E. and Bateman, D.: Antimicrobial susceptibility of *Clostridium perfringens* isolates of bovine, chicken, porcine, and turkey origin from Ontario. *Can. J. Vet. Res.* **75**, 89-97 (2011).
 24. Gholamiandehkordi, A., Eeckhaut, V., Lanckriet, A., Timmermont, L., Bjerrum, L., Ducatelle, R., Haesebrouck, F. and Van Immerseel, F.: Antimicrobial resistance in *Clostridium perfringens* isolates from broilers in Belgium. *Vet. Res. Commun.* **33**, 1031-1037 (2009).
 25. Gillespie, S.H., Basu, S., Dickens, A.L., O'Sullivan, D.M. and McHugh, T.D.: Effect of subinhibitory concentrations of ciprofloxacin on *Mycobacterium fortuitum* mutation rates. *J. Antimicrob. Chemother.* **56**, 344-348 (2005).
 26. Kassaiy, Z.G., El Hakim, R.G., Rayya, E.G., Shaib, H.A. and Barbour, E.K.: Preliminary study on the efficacy and safety of eight individual and blended disinfectants against poultry and dairy indicator organisms. *Vet. Ital.* **43**, 821-830 (2007).
 27. Sabagh, B.P., Souto Ada, S., Reis, L.M., Silva, S.A., Pereira, D.C., Neves Mde, C., Pinheiro, R.R., Duarte, R.S., Miyazaki, N.H. and Bôas, M.H.: Comparative study with two different enrichments in the culture media used in the disinfectant efficacy assay. *J. Microbiol. Methods* **88**, 255-262 (2012).
 28. Meckes, M.C. and Rhodes, E.R.: Evaluation of bacteriological indicators of disinfection for alkaline treated biosolids. *J. Environ. Eng. Sci.* **3**, 231-236 (2004).
 29. Shams, A.M., O'Connell, H., Arduino, M.J. and Rose, L.J.: Chlorine dioxide inactivation of bacterial threat agents. *Let. Appl. Microbiol.* **53**, 225-230 (2011).
 30. Lindstedt, M., Allenmark, S., Thompson, R.A. and Edebo, L.:

- Antimicrobial activity of betaine esters, quaternary ammonium amphiphiles which spontaneously hydrolyze into non-toxic components. *Antimicrob. Agents Chemother.* **34**, 1949-1954 (1990).
31. McFarland, J.: The nephelometer: an instrument for estimating the number of bacteria in suspensions used for calculating the opsonic index and for vaccines. *JAMA* **49**, 1176-1178 (1907).
 32. Zaid, A.: Formulation and evaluation of the chemical stability of povidone-iodine in some trademarks cleaning formulations. *Int. J. Pharm. Pharm. Sci.* **5**, 46-48 (2013).
 33. Cunliffe, P.J. and Fawcett, T.N.: Wound cleansing: the evidence for the techniques and solutions used. *Prof. Nurse* **18**, 95-99 (2002).
 34. Art, G.: Combination povidone-iodine and alcohol formulations more effective, more convenient versus formulations containing either iodine or alcohol alone. *J. Infus. Nurs.* **28**, 314-320 (2005).
 35. Hah, J.H., Roh, D.H., Jung, Y.H., Kim, K.H. and Sung, M.W.: Selection of irrigation fluid to eradicate free cancer cells during head and neck cancer surgery. *Head Neck* **34**, 546-550 (2012).
 36. Pattana-arun. J. and Wolff. B.G.: Benefits of povidone-iodine solution in colorectal operations: science or legend. *Dis. Colon. Rectum.* **51**, 966-971 (2008).
 37. Basha, G., Ghirardi, M., Geboes, K., Yap, S.H. and Penninckx, F.: Limitations of peritoneal lavage with antiseptics in prevention of recurrent colorectal cancer caused by tumor-cell seeding: experimental study in rats. *Dis. Colon. Rectum.* **43**, 1713-1718 (2000).
 38. Reimer, K., Wichelhaus, T.A., Schäfer, V., Rudolph, P., Kramer, A., Wutzler, P., Ganzer, D. and Fleischer, W.: Antimicrobial effectiveness of povidone-iodine and consequences for new application areas. *Dermatology* **204**, 114-120 (2002).
 39. Durani, P. and Leaper, D.: Povidone-iodine: use in hand disinfection, skin preparation and antiseptic irrigation. *Int. Wound J.* **5**, 376-387 (2008).
 40. Xia, Y., Wei, L., Gao, X., Wu, H., Zheng, H. and Yu, Z.: Experimental study on toxicity of povidone iodine disinfectant. *Chin. J. Disinfect.* **25**, 263-267 (2005).
 41. Heiner, J.D., Hile, D.C., Demons, S.T. and Wedmore, I.S.: 10% povidone-iodine may be a practical field water disinfectant. *Wilderness Environ. Med.* **21**, 332-336 (2010).
 42. Udampijitkul, P., Alnoman, M. and Sarker, M.R.: Inactivation strategy for *Clostridium perfringens* spores adhered to food contact surfaces. *Food Microbiol.* **34**, 328-336 (2013).
 43. Phillips, M.S. and von Reyn, C.F.: Nosocomial infections due to nontuberculous mycobacteria. *Clin. Infect. Dis.* **33**, 1363-1374 (2001).