Effects of Draconis Resina on the Collagenase Activities and the Procollagen Synthesis in Hs68 Human Fibroblasts, and Tyrosinase Activity

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ABSTRACT

Objectives: Draconis Resina (DR), the resin of Daemonorops draco Bl., is used to circulate the blood and to stop bleeding. It also has been used to generate flesh including ulceration. The present study investigated the effects of DR extract on collagen metabolism in human fibroblasts and tyrosinase activity in mushroom tyrosinase.

Methods: The effect of DR extract on type I procollagen production (collagen type I synthesis) and collagenase (matrix metalloproteinase-1, henceforth referred as MMP-1) activity in human normal fibroblasts cell line, Hs68 cells after ultraviolet B (UVB, 312 nm) irradiation was measured using the enzyme-linked immunosorbent assay (ELISA). The tyrosinase activity was also measured to find out the whitening effects in mushroom tyrosinase by ELISA method.

Results: There was no cytotoxicity at DR extract at concentrations of 10 µg/ml, 30 µg/ml, and 100 µg/ml. DR extract significantly inhibited the increase of collagenase activity, whereas it did not show on the reduction of type I procollagen in UVB damaged Hs68 cells. DR extract did not reduce the L-DOPA oxidation. However, it significantly reduced the tyrosinase activity by DR extract at concentrations of 0.1 mg/ml, 1 mg/ml and 10 mg/ml.

Conclusions: In conclusion, DR showed the anti-wrinkle and whitening effects via the inhibition of collagenase production and the tyrosinase activity. These results suggest that DR may have potential as an anti-aging ingredient in cosmetic herb markets.

Key words: Draconis Resina, Elastase, Collagenase, Collagen, Hs68, Fibroblasts

Introduction

Draconis Resina (DR) is a resin obtained from Daemonorops draco Bl. Its effects are to activate blood circulation and relieve pain. It reduces bleeding and promotes generation of flesh in the cases of skin lesions and ulcerations1,2). The present drug was chosen from the preliminary screening tests (Data not shown).

The latest trend today is to look younger than one’s age. In these days, aging seems to be treated not as an inescapable destiny to accept but as a disease or a disorder to overcome. There are two major theories of aging: the programmatic theory that aging is an inherent genetic process, and the stochastic theory that aging represents random environmental damage. Processes that are associated with cellular damage and aging are the production of free radicals (a process much enhanced after ultraviolet irradiation) and an increasing number of errors during DNA replication. Cellular manifestations of intrinsic aging include decreased life span of cells, decreased responsiveness of cells to growth signals, which may reflect loss of cellular receptors for growth factors, and increased responsiveness to growth inhibitors. All these findings are more pronounced in cells derived from photodamaged skin3).

It has been shown that UV irradiation leads to the formation of reactive oxygen species (ROS) that activate the mitogen-activated protein (MAP) kinase pathway, which subsequently induces the expression and activation of matrix metalloproteinases (MMPs) in human skin in vivo4,5). MMPs including collagenase are considered key factors in the photoaging process.

DR is traditionally used in the recovery of skin

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lesions, It might prefigure the modern pharmacological anti-wrinkle effects of DR.

In the present study, we investigated the effect of DR on the changes of type I procollagen production and collagenase activity in human normal fibroblasts Hs68 after UVB irradiation. We also measured the tyrosinase activity in mushroom tyrosinase after treatment of DR.

Materials and Methods

1. Sample preparation

Draconis Resina was purchased from Omniherb (Korea). Draconis Resina extracts (DR) was prepared as follow, 100 g of Draconis Resina in 2,000 ml distilled water was heated in a heating extractor for 3 hours. The extract was filtered and concentrated by using the rotary evaporator. The extracts were lyophilized by using freeze dryer (10,3 g). The extract was dissolved in water and filtered three times through micro-filter paper and syringe filter (Whatman #2, 0.45 µm to 0.2 µm). Filtered material was placed in the disinfected vial and was sealed for further study.

2. Cell culture

Hs68 human fibroblasts (Health Protection Agency Culture Collections, UK) were cultured in Dulbecco's Modified Eagle's medium (Gibco, USA) containing 10% fetal bovine serum, 1% antibiotics at 37°C in a humidified atmosphere of 5% CO2. When cells reached above confluency (70–80%), subculture was conducted at a split ratio of 1:3.

3. UVB irradiation

A UVB lamp (Vilber Lourmat, France) was used as a UVB source. In brief, Hs68 cells were rinsed twice with phosphate–buffered saline (PBS, Sigma–Aldrich, USA), and all irradiations were performed under a thin layer of PBS (200 µl/well of 24 well plate). Immediately after irradiation (100 ml/cm²), fresh serum–free medium was added to the cells. After 24 hours incubation period, responses were measured. Mock–irradiated blanks followed the same schedule of medium changes without UVB irradiation.

4. Cell viability

General viability of cultured cells was determined by reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT, Sigma–Aldrich, USA) to formazan. The human fibroblast cells (Hs68) were seeded in 24–well plates at a density of 2×10⁵/ml per well and cultured at 37°C in 5% CO2. Cells were pretreated with the sample at a concentration of 10, 30, and 100 µg/ml for 24 hours prior to UVB irradiation. After UVB irradiation, cells were retreated with the sample and incubated for additional 24 hours, before being treated with 0.05 mg/ml (final concentration) of MTT. The blank and control group was cultivated without sample treatment. The cells were then incubated at 37°C for additional 4h. The medium containing MTT was discarded, and MTT formazan that had been produced was dissolved with 200 µl of dimethyl sulfoxide (DMSO, Sigma–Aldrich, USA). The absorbance was read at 595 nm with a reference wavelength of 690 nm. The cell viability being calculated as follows:

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\text{Cell viability (\%)} = \left(\frac{\text{OD595 of sample}}{\text{OD595 of control}}\right) \times 100
\]

5. Assays of collagen type I synthesis and collagenase inhibition

Hs68 human fibroblasts were inoculated into 24–well plate (2×10⁵ cells/well, Sigma–Aldrich, USA) and cultured at 37°C in 5% CO2. Cells were pretreated with the sample at a concentration of 10, 30, and 100 µg/ml for 24 hours prior to UVB irradiation. After UVB irradiation, cells were retreated with the sample and incubated for additional 24 hours. The blank and control group was cultivated without sample treatment. After culturing, the supernatant was collected from each well, and the amount of pro–collagen type I was measured with a procollagen type I C–peptide assay kit (Takara Bio, Japan). The activity of collagenase was measured with a MMP–1 human biotrak ELISA system (Amersham life science, USA).

6. Tyrosinase inhibition assay

Tyrosinase activity was determined essentially as previously described(6). The reaction mixtures were prepared by adding 40U of mushroom tyrosinase (Sigma–Aldrich, USA) to 20 µl of DR dissolved in distilled water (0.1, 1, and 10 mg/ml), and then adding 40 µl of 1.5 mM L-tyrosine (Sigma–Aldrich, USA) and 220 µl of 0.1 M sodium phosphate buffer (pH 6.5). The resulting mixture (300 µl) was incubated for 10 min at 37°C and then absorbance at 490 nm was measured. The same mixture, but without DR extract, was used as a control.
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7. Inhibition of L-DOPA oxidation

The inhibitory effect of DR on L-DOPA oxidation was determined according to the method of Joshi with a slight modification. 50 µl of DR dissolved in 0.1 M sodium phosphate buffer (0.1, 1, and 10 mg/ml) was added to 40 U of mushroom tyrosinase in 900 µl of 0.1 M sodium phosphate buffer (pH 6.5). After 6 min of incubation at 37°C, 3 mM of L-DOPA (Sigma–Aldrich, USA) was added. Then the mixture was incubated at 37°C for 15 min. Activities were quantified by measuring absorbance at 475 nm. The same mixture, but without DR extract, was used as a control.

8. Statistical analysis

The results were expressed as means ± standard error of the mean (SEM). Significances of changes were evaluated using the one–way ANOVA with Dunnett’s post–hoc test. Values of \( p < 0.05 \) were considered significant.

Results

1. Cytotoxicity of DR on Hs68 fibroblasts

To evaluate the cytotoxicity of DR extract in Hs68 cells, the extract was treated at 10, 30, and 100 µg/ml. The cell viability was determined by MTT assay, and calculated into 100% of control group. The cell viabilities were 92.9 ± 1.4% in DR extract at 10 µg/ml, 91.9 ± 0.9% in DR extract at 30 µg/ml, and 95.7 ± 2.8% in DR extract at 100 µg/ml. As a result, DR extract showed no cytotoxicity up to the 100 µg/ml (Fig. 1).

2. Effect of DR extract on collagen type I synthesis in Hs68 fibroblasts

To evaluate the amount of collagen type I synthesis that occurred upon exposure to the extract, collagen type I was quantitatively detected by using the procollagen type I C–peptide assay kit. As shown in Figure 2, the collagen concentration was 16.7 ± 0.3 ng/ml in DR extract at 10 µg/ml, 17.3 ± 0.6 ng/ml in DR extract at 30 µg/ml, and 14.2 ± 0.3 ng/ml in DR extract at 100 µg/ml. As a result, DR extract did not increase the collagen production.

3. Effect of DR extract on collagenase activity in Hs68 fibroblasts

To evaluate the collagenase activity, MMP–1 activity was quantitatively measured in cell lysates by MMP–1 assay kit. The activities of DR extract were calculated into 100% of control group (Fig. 3). DR extract significantly reduced the MMP–1 activity with 33.2 ± 3.1% at 10 µg/ml, with 15.3 ± 8.0% at 30 µg/ml, and with 7.8 ± 0.4% at 100 µg/ml.

4. Inhibitory effects of DR extract on tyrosinase activity in mushroom tyrosinase

Tyrosinase activities were measured in mushroom tyrosinase by ELISA method. The activities of DR extract
on tyrosinase activity was calculated into 100% of control group (Fig. 4). DR extract significantly reduced the tyrosinase activity at concentrations of 0.1, 1 and 10 mg/ml (21.8 ± 17.2%, 6.6 ± 7.8% and 1.0 ± 0.5%, p < 0.05, respectively).

5. Inhibitory effects of DR extract on L-DOPA oxidation in mushroom tyrosinase

L-DOPA oxidation activities were measured in mushroom tyrosinase by ELISA method. The activities of DR extract on L-DOPA oxidation were calculated into 100% of control group (Fig. 5). DR extract at 0.1, 1 and 10 mg/ml did not show any inhibitory activities in L-DOPA oxidation (72.6 ± 24.0%, 96.9 ± 18.7% and 88.4 ± 24.9% respectively).

Discussion

Dracoris Resina is a resin (Daemonorops draco Bl.)\(^9\). It is mainly used for external usage to stop bleeding and to promote the generation of flesh. Topical application treats both acute and chronic skin conditions. It stops bleeding in acute conditions, and promotes the healing of wounds in chronic conditions by facilitating the generation of flesh and prevents the spread of ulcers. It is also orally taken to activate blood circulation, to remove blood stasis, and to relieve pain. The internal use of DR treats both acute and chronic blood stasis, swelling, and pain\(^1,2\).

The skin aging is one of the most obvious evidence of aging. The skin is increasingly exposed to ambient UV–irradiation thus increasing risks for photooxidative damage with long–term detrimental effects like photoaging, characterized by wrinkles, loss of skin tone and resilience. Photoaged skin displays alterations in the cellular component and extracellular matrix with accumulation of disorganized elastin and its microfibrillar component fibrillin in the deep dermis and a severe loss of interstitial collagens, the major structural proteins of the dermal connective tissue. It has been shown that UV irradiation leads to the formation of reactive oxygen species (ROS) that activate the mitogen–activated protein (MAP) kinase pathway, which subsequently induces the expression and activation of MMPs in human skin in vivo\(^4,5\). MMPs are known to be over–expressed in human fibroblasts within hours after exposure to UV irradiation. Therefore, MMPs are considered key regulators in the photoaging process. Inhibiting the major collagen–degrading enzymes like MMPs would be a useful agent for anti–aging.

In order to evaluate the cytotoxicity of DR, samples were prepared at various concentrations and used to treat human fibroblasts (Hs68). There was no cytotoxicity in all treated concentrations. The cells treated by DR 300 µg/ml showed a slight cytotoxicity.

Collagens are synthesized as precursor molecules, called procollagens. These molecules contain additional peptide sequences, usually referred to as ‘propeptides’, at both the amino–terminal end and the carboxy–terminal end. These propeptides are cleaved from the collagen triple–helix molecule during its secretion, after which the triple–helix collagens are polymerized into extracellular collagen fibrils. Thus, the amount of free propeptide stoichiometrically reflects the amount of collagen molecules synthesized\(^9\). Collagen is a group of naturally occurring proteins. In nature, it is found exclusively in animals, especially in the flesh and connective tissues of mammals\(^9\). It is the main component of connective tissue, and is the most abundant protein in mammals, making up about 25% to 35% of the whole–body protein content\(^9\). Collagen, in the form of elongated fibrils, is mostly found in fibrous tissues such as tendon, ligament and skin, and is also abundant in cornea, cartilage, bone, blood vessels, the gut, and intervertebral disc. In muscle tissue it serves as a major component of endomysium. Collagen constitutes 1% to 2% of muscle tissue, and accounts for 6% of the weight of strong, tendinous muscles\(^11\). Collagen occurs in many places throughout the body. So far,
only 29 types of collagen have been identified and described, Over 90% of the collagen in the body, however, is of type I, II, III, and IV. Among them, collagen type I is placed at skin, tendon, vascular, ligature, organs, and bone (main component of bone). Collagen-related diseases most commonly arise from genetic defects or nutritional deficiencies that affect the biosynthesis, assembly, posttranslational modification, secretion, or other processes involved in normal collagen production. In this study, the amount of collagen type I was slightly increased at all concentrations of DR. However, there was no statistical significance.

To evaluate the collagenase activity, MMP-1 activity was quantitatively measured. DR significantly reduced the MMP-1 activity at all concentrations in a dose-dependent manner. The MMP-1 reducing activities at 30 and 100 µg/ml are below the activity of blank group. It might be thought that DR might inhibited the naturally induced collagenase activity as well.

The activities of DR on tyrosinase activity were significantly effective at 0.1, 1 and 10 mg/ml.

**Conclusion**

In this study, I determined the amounts of type I collagen synthesis and collagenase activity. And the tyrosinase activity and L-DOPA oxidation were measured in DR-treated groups.

1. DR showed no cytotoxicity up to the effective concentration for the measuring the amounts of type I collagen synthesis and collagenase activity in Hs68 cells.

2. The collagen amounts of DR 10 µg/ml, 30 µg/ml and 100 µg/ml treated group did not increase.

3. DR significantly reduced the MMP-1 activity at concentrations of 10 µg/ml, 30 µg/ml, and 100 µg/ml in a dose dependent manner.

4. DR significantly reduced the tyrosinase activity at concentrations of 0.1 mg/ml, 1 mg/ml and 10 mg/ml in a dose dependent manner.

5. DR 0.1 mg/ml, 1 mg/ml and 10 mg/ml treated groups did not show any inhibitory activity in L-DOPA oxidation.

These results suggest that DR may have potential as an anti-aging ingredient in cosmetic herbal drugs, I think further studies will be needed to unravel exactly under the molecular mechanisms.

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**References**


