



Correlations among Stress Parameters, Meat and Carcass Quality Parameters in Pigs

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ABSTRACT: Relationships among different stress parameters (lairage time and blood level of lactate and cortisol), meat quality parameters (initial and ultimate pH value, temperature, drip loss, sensory and instrumental colour, marbling) and carcass quality parameters (degree of rigor mortis and skin damages, hot carcass weight, carcass fat thickness, meatiness) were determined in pigs (n = 100) using Pearson correlations. After longer lairage, blood lactate (p<0.05) and degree of injuries (p<0.001) increased, meat became darker (p<0.001), while drip loss decreased (p<0.05). Higher lactate was associated with lower initial pH value (p<0.01), higher temperature (p<0.001) and skin blemishes score (p<0.05) and more developed rigor mortis (p<0.05), suggesting that lactate could be a predictor of both meat quality and the level of prelaughter stress. Cortisol affected carcass quality, so higher levels of cortisol were associated with increased hot carcass weight, carcass fat thickness on the back and at the sacrum and marbling, but also with decreased meatiness. The most important meat quality parameters (pH and temperature after 60 minutes) deteriorated when blood lactate concentration was above 12 mmol/L. (**Key Words:** Pig, Blood Lactate, Prelaughter Stress, Meat Quality, Carcass Quality)

INTRODUCTION

The production of high quality pork has been a constant objective of the pig industry for many decades. The main goal is to obtain pigs with high lean percentage and good meat quality traits at the same time. However, this goal has proved difficult to achieve (Kusec et al., 2003). Meat and carcass quality are complex traits and are influenced by many antemortem and postmortem factors (Sellier and Monin, 1994; Huff-Lonergan et al., 2002). The prediction and evaluation of pork quality is of particular importance if these traits are to be improved. Many physical and biochemical factors have been evaluated in an attempt to assess meat quality (Huff-Lonergan et al., 2002).

There are three important parameters to define pork quality: drip loss, ultimate pH and colour (Lee et al., 2000).

Using those parameters, meat can be classified into the five quality categories; PSE (pale, soft, exudative), normal or RFN (red, firm, nonexudative), DFD (dark, firm, dry), RSE (reddish-pink, soft, exudative) and PFN (pale, firm, non-exudative) meat (Kauffman et al., 1992). It is well known that changes in some meat quality traits can affect many other meat quality attributes and pork quality overall (Huff-Lonergan et al., 2002).

In addition, changes in physiological parameters affect meat quality. Increase in blood lactate concentration is associated with pre-slaughter stress, which has been shown to have a detrimental effect on pork quality (Hambrecht et al., 2005a, b; Edwards et al., 2010b). Hambrecht et al. (2004) determined that pigs exposed to aggressive handling just prior to stunning had a higher blood lactate concentration at slaughter and exhibited pork with higher drip loss and thus proposed that lactate was a potential indicator of both physical and psychological stress associated with the handling of pigs immediately before slaughter. Furthermore, the level of cortisol is an individual characteristic of each animal (Mormede, 2007) and it

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affects the amount of fat in the body, meatiness, and thus, carcass quality (Foury et al., 2005; 2007; Skrlep et al., 2009). Higher cortisol levels were associated with higher initial and ultimate pH values, and consequently with lower Minolta L* and b* values and drip loss after 24 and 48 hours suggesting that cortisol level at slaughter reflects on meat quality (Skrlep et al., 2009).

Considering relationships between biochemical traits and objective measures of quality, and complex biological mechanisms behind their development, the objective of this study was to determine to what extent different animal stress, meat quality and carcass quality traits were associated.

MATERIAL AND METHODS

Animals, housing and feeding

The experiment was conducted on 100 commercial market pigs (31 gilts, 51 barrows and 18 boars), six months old and with live weight between 115 and 130 kg. All animals were of the same origin (cross between Naima sows and hybrids P-76 PenArLan boars) and came from the same farm. Pigs were housed in the finishing facility on partially slatted floors, in pens with 20 animals per pen (stocking density = 1 m²/pig). Pigs were provided with *ad libitum* feed and water.

Pre-slaughter handling

Before transport feed and water were not withdrawn. Pigs from the same pen were transported together, without mixing with other pigs, in batches of 20 animals in a transportation trailer (stocking density was 0.45 m² per pig). The distance between pens and vehicle at loading was about 20 m and the slope of the loading ramp was 15°. Transport from farm to the abattoir lasted 15 minutes. The surface of the vehicle and the unloading area were on the same level. After unloading, pigs entered a 10 m long corridor that led to the lairage pens. During loading and unloading sticks and electric prods were used to move pigs.

During lairage, pigs were not mixed and stocking density was 0.70 m² per pig. Pigs were held in lairage less than 3 hours (slaughtered on the same day when they arrived, n = 28) or more than 14 hours (stayed overnight and slaughtered in the morning, n = 72). The exact length of lairage time was measured for each pig.

Between lairage pens and stunning area was 5-m single file corridor. During pig handling from lairage pens to the stunning area stick and electric prod were used randomly. After lairage pigs were head-only electrically stunned (50 Hz, 2 A and 220 V) in batches of 6 animals without restraining. Immediately after stunning pigs were bled on the floor and then hoisted on a rail. Following bleeding

carcasses were processed using conventional industry practice.

Blood sampling and determination of blood lactate and cortisol content

At slaughter blood samples were taken into plastic tubes and part of material was transferred to vacutainer tubes containing heparin (against blood coagulation). Thereupon blood lactate content was determined using a portable lactate analyzer (Lactate Scout, EKF Diagnostic, Magdeburg, Germany). The lactate analyzer was tested with a standard solution to ensure accuracy. After blood collection the vacutainer tubes were placed on ice and within 4 to 6 hours blood was centrifuged at 3,000 rpm for 3 minutes. Supernatants (plasma) were collected into microtubes and stored at -20°C until the determination of cortisol concentration by radioimmunoassay (RIA-CT Cortisol, INEP, Belgrade, Serbia).

Meat and carcass quality analyses

Meat quality measurements were carried out 60 minutes, 24 and 72 hours after slaughter on muscle *Longissimus dorsi* (LD), *pars lumbalis*. Values of pH 60 minutes and 24 hours postmortem (pH_{60min}, pH_{24h}), and temperature 60 minutes postmortem (t_{60min}) were measured using a pH-meter Testo 205 (Germany) calibrated with pH 4.00 and 7.00 phosphate buffer. Skin damages were assessed by three observers on three carcass regions (from head to back of shoulder, from back of shoulder to hind-quarters and region of hind-quarters) immediately after dressing using scores 1 (no damages), 2 (scratches or small wounds, less than 2 cm), 3 (bleeding wounds between 2 and 5 cm or a healed wounds of more than 5 cm) and 4 (deep and open wounds of more than 5 cm). The final skin damages score for each carcass was obtained by summing scores for the three regions, and ranged from 3 to 12. Degree of *rigor mortis* was estimated on right carcass side 3 hours *post mortem* by measuring degree of angle between body axis and foreleg (Davis et al., 1978). For that purpose photographic images were taken of carcasses, at distance of approximately 2 m and a height of 160 cm, parallel to the plane of the carcass and then the angle was calculated in AutoCAD. The size of the angle and intensity of rigor were inversely proportional, e.g. smaller angle meant a higher degree of *rigor mortis*. For determination of drip loss, colour and marbling, meat samples, 2.5 cm thick loin chops were taken 24 hours after slaughter from LD, between the 3rd and 4th lumbar vertebrae. Meat samples were weighed and stored for 48 hours at 4°C in a container (Honikel, 1998). After storage, meat samples were reweighed and drip loss (%) was calculated as the difference between sample weight before and after storage divided by the sample weight before

storage. Sensory and instrumental colour (Commission Internationale de l'Eclairage [CIE] $L^*a^*b^*$) (CIE, 1976) as well as marbling were determined at 24 hours postmortem, after approximately 60 minutes of blooming time (Honikel, 1998). L^* , a^* , and b^* (CIE, 1976) values were determined using a Minolta Chroma Meter CR-400 (Minolta Co., Ltd., Osaka, Japan) utilizing a 65 light source and a 2° observer. Colour, marbling and drip loss were analyzed in duplicate. An analytical panel of three trained members assessed sensory colour and marbling of meat samples by using the scaling method (NPPC, 2000).

Hot carcass weight and carcass fat thickness at two points (on the back between the 13th and 15th dorsal vertebrae and at the sacrum where is fat thickness thinnest) were measured. Meatiness (in percentages and kilograms) was determined according to regulation (Anon, 1985) on the basis of hot carcass weight and the sum of carcass fat thickness at two points (on the back and at the sacrum).

Statistical analysis

Statistical analysis of the results was elaborated using software GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego CA, USA, www.graphpad.com. All parameters were described by descriptive statistics (mean, standard deviation, minimum and maximum value). Pearson's correlation was used to determine relationships among different stress, meat and carcass quality parameters. Student t-test was used for testing the effect of blood lactate concentration below and above 12 mmol/L on pH value and temperature measured 60 minutes postmortem.

RESULTS AND DISCUSSION

Characterization of the experimental population

Characterization of the experimental population is presented in Table 1. In this study, blood lactate ranged from 1.3 to 24.6 mmol/L which is in accordance with results of other authors, who found values ranging from 1.1 to 20.6 mmol/L (Edwards et al., 2010a) and from 4.0 to 19.7 mmol/L (Edwards et al., 2010b). Blood cortisol content varied within studies (Hambrecht et al., 2004; Foury et al., 2005; Hambrecht et al., 2005b), which was also observed in this experiment (3 to 248 nmol/L). Those variations in cortisol content could be explained by influence of species, breed, time of day, stress, meal, physical and sexual activity and change of environment. Values of pH after 60 minutes ranged from 5.64 to 6.81, while after 24 hours were from 5.26 to 5.93. The highest carcass temperature was 40.3°C and the highest drip loss of meat samples was 9.50%. There were carcasses without skin blemishes (score 3) in the experimental group, as well as those with a maximum score for skin blemishes (score

12). The CIE parameters ranged from 35.11 to 60.22 (CIE L^* value), from 4.15 to 11.83 (CIE a^* value) and from 2.23 to 7.41 (CIE b^* value), while sensory colour from 1.0 to 4.0.

The average hot carcass weight in this study was 93.46 kg, which was similar to most EU countries (less than 95 kg) (Hansson, 2003). Carcass fat thickness ranged from 12 to 36 mm on the back, and from 5 to 34 mm at the sacrum. The reason for those variations, especially at the sacrum, was due to the presence of barrows, gilts and boars in the study, since boars have the lowest backfat thickness (Jaturasitha et al., 2006). The average meatiness was 43.79%, which was lower compared to data from 16 EU countries, where average meatiness ranged from 55 to 55.99% (Hansson, 2003). Unlike in the EU, in this study muscle tissue from the belly was not included in determination of meat yield, so differences in meatiness recorded by these different measuring practices could be up to 10%.

The effect of lairage time

Relationships among lairage time, blood lactate and cortisol concentration, and meat quality parameters in pigs are shown in Table 2.

Table 1. Characterization of the experimental population (n = 100): Stress, meat and carcass quality parameters

Parameter	$\bar{X} \pm SD$	Min	Max
Blood lactate (mmol/L)	10.06±5.47	1.30	24.60
Plasma cortisol (nmol/L)	58.36±57.46	3.00	248.00
pH _{60 min} ¹	6.33±0.21	5.64	6.81
pH _{24h} ¹	5.55±0.13	5.26	5.93
t _{60 min} ² (°C)	38.58±0.74	37.10	40.30
Skin damages score	7.09±2.30	3.00	12.00
Rigor mortis (°)	124.80±4.62	115.10	136.60
Drip loss (%)	6.30±1.40	3.01	9.50
L^* value ³	50.20±3.02	35.11	60.22
a^* value ³	7.71±1.26	4.15	11.83
b^* value ³	4.21±0.96	2.23	7.41
Sensory colour ⁴	2.44±0.52	1.00	4.00
Marbling	1.30±0.38	1.00	2.50
HCW (kg)	93.46±11.25	54.30	115.60
FTB (mm)	21.94±5.90	12.00	36.00
FTS (mm)	14.51±5.23	5.00	34.00
Meatiness (kg)	40.99±4.37	31.80	52.20
Meatiness (%)	43.79±1.71	38.18	46.61

SD, standard deviation; HCW, hot carcass weight; FTB, fat carcass thickness on the back; FTS, fat carcass thickness at the sacrum.

¹ pH_{60min} and pH_{24h}, pH values measured 60 minutes and 24 hours postmortem.

² t_{60min}, meat temperature measured 60 minutes postmortem;

³ L^* , Lightness (a higher L^* value indicates a lighter colour); a^* , Redness (a higher a^* value indicates a redder colour); b^* , Yellowness (a higher b^* value indicates a more yellow colour);

⁴ Sensory colour scale: 1 = pale pinkish-gray to white; 6 = dark purplish-red.

Table 2. Correlations among lairage duration, stress and meat quality parameters

Parameters	LT	Lac	Cor	pH _{60min} ¹	pH _{24h} ¹	t _{60min} ²	SB	RM	DL	L* ³	a* ³	b* ³	SC	Mar
LT	1.0	0.21*	-0.12	0.02	0.0004	-0.14	0.35***	-0.27**	-0.23*	-0.39***	-0.25*	-0.40***	0.20*	0.26**
Lac		1.0	-0.01	-0.34**	0.03	0.37***	0.20*	-0.20*	-0.13	0.08	0.05	0.01	-0.07	0.22*
Cor			1.0	-0.15	0.06	0.23*	0.07	0.01	-0.10	-0.10	0.20	0.13	0.07	0.22†
pH _{60min}				1.0	-0.09	-0.16†	0.05	0.05	-0.12	-0.19†	-0.23*	-0.15	0.14	-0.09
pH _{24h}					1.0	-0.06	0.02	-0.10	-0.10	-0.002	0.09	0.005	-0.12	0.27**
t _{60min}						1.0	-0.14	-0.07	-0.17	0.15	0.16	0.13	-0.01	0.12
SB							1.0	0.002	0.05	-0.31**	-0.12	-0.30**	0.12	0.07
RM								1.0	0.19†	0.12	-0.006	0.08	-0.11	-0.08
DL									1.0	0.28**	0.02	0.12	-0.20*	-0.21*
L*										1.0	0.16	0.60***	-0.60***	-0.29**
a*											1.0	0.72***	0.20*	0.16
b*												1.0	-0.25*	-0.05
SC													1.0	0.30**
Mar														1.0

LT, lairage time; Lac, lactate blood concentration; Cor, cortisol blood concentration; SB, skin blemishes score; RM, rigor mortis; DL, drip loss; SC, sensory colour; Mar, marbling.

¹ pH_{60min} and pH_{24h}, pH values measured 60 minutes and 24 hours postmortem.

² t_{60min} - Meat temperature measured 60 minutes postmortem.

³ L*, lightness (a higher L* value indicates a lighter colour); a*, redness (a higher a* value indicates a redder colour); b*, yellowness (a higher b* value indicates a more yellow colour).

† p<0.1; * p<0.05; ** p<0.01; *** p<0.001.

With longer lairage time, blood lactate level ($r = 0.21$, $p < 0.05$) and skin blemishes ($r = 0.35$, $p < 0.001$) increased, carcasses had a greater degree of rigor mortis ($r = -0.27$, $p < 0.01$) and meat released less water ($r = -0.23$, $p < 0.05$). In pigs with longer lairage times, meat became darker, as shown by L* values ($r = -0.39$, $p < 0.001$) and sensory color ($r = 0.20$, $p < 0.05$), as well as less red ($r = -0.25$, $p < 0.05$) and yellow ($r = -0.40$, $p < 0.001$).

Although lairage time did not have a significant effect on cortisol concentration, other authors found the opposite (Perez et al., 2002; Warriss, 2003a), as cortisol decreased during lairage, particularly at night when pigs rested and became calm. On the other hand, in this study, lairage time was positively correlated with blood lactate, which, as a measure of stress, indicated that a longer lairage caused a higher level of stress. This could be explained by after longer lairage pigs were more susceptible to stress due to longer period of experiencing stressors in lairage (rough handling, fights, change of environment, food deprivation, etc.) (Stam et al., 2000; Bruijnzeel et al., 2001; Stam et al., 2002). Lairage time did not have a significant effect on initial or ultimate pH values, although higher pH values were found after longer lairage times in other studies (Perez et al., 2002; Hoffman and Fisher, 2010). This is explained by the fact that reserves of glycogen become depleted during lairage and after a longer lairage there is a lower muscle glycogen content and consequently higher pH value of meat. As in our study, Hoffman and Fisher (2010) determined lower drip loss after longer lairage. In addition, longer lairage increased the level of skin blemishes on the carcass. The percentage of skin blemishes is higher after a longer lairage time because it increases the aggressiveness of the pigs (Nanni Costa et al., 2002; Perez et al., 2002;

Warriss, 2003a; Guardia et al., 2009). Since the rate of rigor mortis is positively correlated with the content of cortisol and lactate as indicators of stress (Warriss et al., 2003b), a greater degree of rigor mortis after longer lairage indicated a more stressful procedure. Lairage time affected meat colour, so after longer lairage meat became darker, less red and yellow which was also observed by Nanni Costa et al. (2002) and Hoffman and Fisher (2010).

The relationship among stress and meat quality parameters

Increased levels of blood lactate were correlated with lower pH values after 60 minutes ($r = -0.34$, $p < 0.01$), and higher temperatures after 60 minutes ($r = 0.37$, $p < 0.001$), more skin blemishes ($r = 0.20$, $p < 0.05$) and greater marbling scores ($r = 0.22$, $p < 0.05$). Rigor mortis was more developed in pigs with higher concentrations of lactate ($r = -0.20$, $p < 0.05$) (Table 2).

As in our study, Edwards et al. (2010a, b) found that the concentration of blood lactate was negatively correlated with the initial pH value ($r = -0.25$, $p < 0.001$; $r = -0.32$, $p < 0.001$). Higher drip loss of meat was expected in pigs with increased concentrations of lactate (Edwards et al., 2010a, b), but no significant correlation was determined between these two parameters in this study. Although correlation coefficients between blood lactate and meat quality parameters (pH_{60min} and t_{60min}) were weak, they indicated that exsanguination blood lactate is a potential predictor of postmortem muscle metabolism and meat quality (Edwards et al., 2010b). As in our study, a higher degree of rigor mortis was found in pigs with higher levels of blood lactate (Warriss et al., 2003a). A positive correlation between blood lactate concentration and skin

damages score indicated that lactate was not only associated with meat quality, but also with the level of stress and procedures prior to slaughter. Rabaste et al. (2007) found that skin blemishes score tended to be higher in pigs after rough preslaughter treatment (pigs moved quickly with an electric prod) compared to gentle (pigs moved slowly with a plastic board or whip), while Edwards et al. (2010c) observed higher concentrations of blood lactate in pigs that experienced more frequent use of electric prods.

Higher levels of blood cortisol were associated with higher meat temperatures ($r = 0.23$, $p < 0.05$) and marbling scores ($r = 0.22$, $p < 0.1$). Warriss et al. (2003a) observed increased cortisol concentration in the blood of pigs having more developed rigor mortis.

Initial pH value was not significantly correlated with most meat quality parameters, except with temperature ($r = -0.16$, $p < 0.1$), L^* ($r = -0.19$, $p < 0.1$) and a^* ($r = -0.23$, $p < 0.05$) parameters of colour. In accordance with our results, Terlow and Rybarczyk (2008) found that a lower pH value after 40 minutes was associated with a higher temperature. Aaslyng and Barton Gade (2001) found that meat with lower pH value after 40 minutes had higher drip loss and became lighter. Other authors have found that drip loss increased when pH values after 45 minutes and 24 hours were lower (Hambrecht et al., 2003; Edwards et al., 2010a, b). Although a positive correlation can be found between initial and ultimate pH value, it is weak in most studies; $p = 0.14$ (Edwards et al., 2010b), 0.15 (Boler et al., 2008), 0.20 (Aaslyng et al., 2001), 0.33 (Borchers et al., 2007) and 0.44 (Klont et al., 2001). We found that relationship between initial and ultimate pH was not significant, probably due to the fact that these two parameters were under the influence of different factors. The extent of pH decline and the finally ultimate pH are primarily affected by amount of glycogen in muscles at the time of slaughter (Lonergan et al., 2008), while initial pH value depends on rate of postmortem glycolysis which is influenced by genetic (Barbut et al., 2007), pre-slaughter factors (Rosenvold and Andersen, 2003), postmortem carcass chilling (Tomovic et al., 2008) or combinations of all of these (Lonergan et al., 2008).

Several meat quality characteristics are correlated with pH value after 24 hours. In pigs with lower ultimate pH values, higher drip loss (Huff-Lonergan et al., 2002; Hambrecht et al., 2003; Edwards et al., 2010b) and lighter colour (Huff-Lonergan et al., 2002; Hambrecht et al., 2003; Boler et al., 2008; Edwards et al., 2010b) were measured. In carcasses with rapid development of rigor mortis, ultimate pH values were significantly lower (Warriss et al., 2003a). Contrary to those findings, in this study, ultimate pH value was positively correlated only with marbling, as was also observed by other authors (Przybylski et al., 2007; Boler et al., 2008; Czarniecka-Skubina et al., 2010). This can be explained by the fact that energy reserves in muscle fibres

are distributed among intramuscular fat and glycogen, and muscles with lower glycogen content have a higher intramuscular fat content. Therefore, higher marbling was accompanied by higher ultimate pH ($r = 0.27$, $p < 0.01$), lower drip loss ($r = -0.21$, $p < 0.05$) and darker colour expressed by L^* value ($r = -0.29$, $p < 0.01$) and sensory colour ($r = 0.30$, $p < 0.01$), indicating lower glycogen content in muscles. Boler et al. (2010) also found a significant correlation between sensory colour and marbling ($r = 0.29$, $p < 0.05$).

In this study, there was no significant correlation between temperature and other meat quality parameters. Contrary to that, meat temperature after 40 minutes was positively correlated with L^* value (Aaslyng et al., 2001) and drip loss (Aaslyng et al., 2001; Hambrecht et al., 2003), while Edwards et al. (2010b) found that increase in temperature reduced the amount of meat exudate. In addition, in the current study, pigs with rapid development of rigor mortis were observed to have a higher meat temperature, increased blood concentrations of cortisol and lactate, and lower ultimate pH value. Those changes in parameters indicated that stressful procedures prior to slaughter were accompanied by intensive metabolic processes in muscles and, consequently, more rapid development of rigor mortis (Warriss et al., 2003a).

The degree of skin blemishes was negatively correlated with L^* ($r = -0.31$, $p < 0.01$) and b^* ($r = -0.30$, $p < 0.01$) parameters, because during lairage the number of skin lesions increases (Nanni Costa et al., 2002; Perez et al., 2002; Guàrdia et al., 2009) and meat becomes darker (Jones et al., 1994; Faucitano, 1998; Warriss et al., 1998).

Drip loss was negatively correlated with sensory colour ($r = -0.20$, $p < 0.05$) and positively with L^* colour parameter ($r = 0.28$, $p < 0.01$). Other authors have come to the same conclusions (Hambrecht et al., 2003; Edwards et al., 2010b). Additionally, drip loss was lower in meat samples with higher marbling ($r = -0.21$, $p < 0.05$), as also determined by others (Przybylski et al., 2007; Czarniecka-Skubina et al., 2010).

In this study, sensory colour was negatively correlated with L^* ($r = -0.60$, $p < 0.001$) and b^* ($r = -0.25$, $p < 0.05$) parameters, and positively with a^* ($r = 0.20$, $p < 0.05$) colour parameter. Higher L^* values were correlated with higher b^* ($r = 0.60$, $p < 0.001$) values, and there was also a high correlation ($r = 0.72$, $p < 0.001$) between a^* and b^* colour parameters, as found previously (Edwards et al., 2010b).

The relationship among blood cortisol concentration and carcass quality parameters

Table 3 shows correlations between blood cortisol concentration and carcass quality parameters as well as correlations among carcass quality parameters.

Cortisol concentration was positively correlated with hot carcass weight ($r = 0.26$, $p < 0.1$) and with the amount of

Table 3. Relationship between blood cortisol concentration and carcass quality parameters as well as inter-relationships among carcass quality parameters

Parameters	Cor	HCW	FTB	FTS	M	Mar
Cor	1.0	0.26[†]	0.33*	0.25[†]	-0.29*	0.22[†]
HCW		1.0	0.56***	0.40***	-0.14	0.23*
FTB			1.0	0.74***	-0.39***	0.19[†]
FTS				1.0	-0.44***	0.15
M					1.0	-0.14
Mar						1.0

Cor, cortisol blood concentration; HCW, hot carcass weight; FTB, fat carcass thickness on the back; FTS, fat carcass thickness at the sacrum; M, meatiness (%); Mar, marbling.

[†] $p < 0.1$; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

fat in the body, expressed as carcass fat thickness on the back ($r = 0.33$, $p < 0.05$) and at the sacrum ($r = 0.25$, $p < 0.1$). Higher cortisol concentrations in the blood correlated with lower meatiness ($r = -0.29$, $p < 0.05$). In accordance with this, Foury et al. (2005; 2007) and Škrlep et al. (2009) have found that high concentrations of cortisol were associated with a greater thickness of subcutaneous fat tissue and lower meatiness. These results indicate that cortisol affects the metabolism of fats and proteins by stimulating the fat deposition at the expense of decreased synthesis and increased protein degradation (Mormede, 2007). In contrast to other authors (Foury et al., 2005; 2007), we found a positive correlation ($r = 0.22$, $p < 0.1$) between blood cortisol concentration and marbling, indicating that cortisol influenced deposition of adipose tissue not only under the skin and around organs, but also between and inside the muscle fibers. It is likely that the same mechanism controls deposition of fat regardless of location in the body, since Huff-Lonergan et al. (2002) have found a positive correlation ($r = 0.38$) between fat thickness on the back and marbling, which has been also observed in the present study ($r = 0.19$, $p < 0.1$).

Carcass quality parameters were inter-correlated, where hot carcass weight was associated with fat thickness on the back ($r = 0.56$, $p < 0.001$) and at the sacrum ($r = 0.40$, $p < 0.001$), as well as with marbling ($r = 0.23$, $p < 0.05$). Fat thickness on the back and at the sacrum was negatively correlated with meat yield ($r = -0.39$, $p < 0.001$, and $r = -0.44$, $p < 0.001$). Similarly to our results, Huff-Lonergan et al. (2002) and Škrlep et al. (2009) found that increasing of hot carcass weight increased subcutaneous fat thickness and slightly reduced carcass meatiness.

The effect of blood lactate concentration on pH and temperature at 60 minutes postmortem

Figures 1 and 2 represent pH values and temperatures measured 60 minutes after slaughter in relation to the blood lactate concentration (below and above 12 mmol/L). In the group of pigs with the lower lactate concentration, a

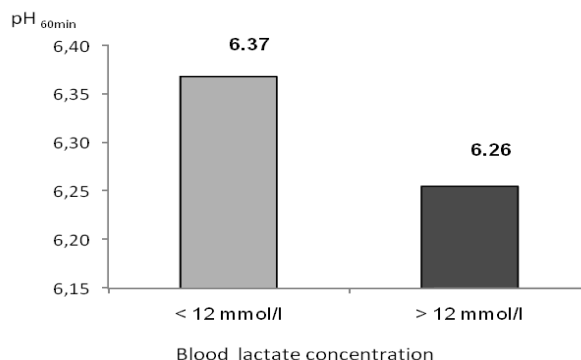


Figure 1. In the group of pigs with the lower lactate concentration, a significantly higher ($p < 0.01$) average pH value (6.37) was determined compared to the group of pigs with the higher lactate concentration (6.26).

significantly higher ($p < 0.01$) average pH value (6.37) was determined compared to the group of pigs with the higher lactate concentration (6.26). Furthermore, in the group of pigs with blood lactate concentration above 12 mmol/L, meat temperature (38.92°C) was, on average, higher ($p < 0.001$) than in the group of pigs with lower blood lactate concentration (38.40°C). Drip loss did not differ between two groups (data not shown). Edwards et al. (2010b) compared the effect of low (up to 10 mmol/L) and high (more than 10 mmol/L) lactate concentration on drip loss and found a higher drip loss in the group with a higher lactate concentration. These results support the statement that lactate can be used as a predictor of meat quality (Edwards et al., 2010a) and the concentration of 12 mmol/L may be taken as a critical value above which the most important meat quality parameters deteriorate.

CONCLUSION

Lairage time affected stress and meat quality parameters

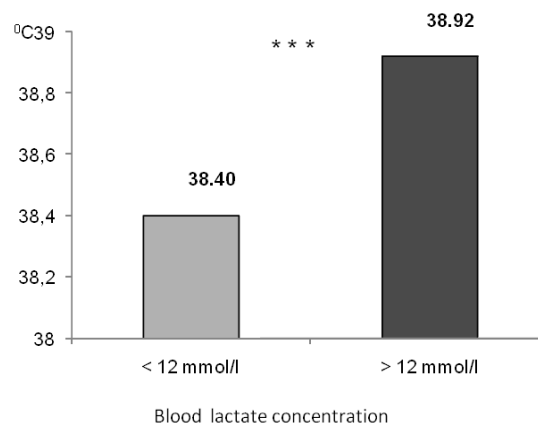


Figure 2. In the group of pigs with blood lactate concentration above 12 mmol/L, meat temperature (38.92°C) was, on average, higher ($p < 0.001$) than in the group of pigs with lower blood lactate concentration (38.40°C).

and after longer lairage, blood lactate and degree of injuries increased, meat became darker, while drip loss decreased. Higher lactate was associated with lower initial pH value, higher temperature and skin blemishes score and more developed rigor mortis, suggesting that lactate could be predictor of both meat quality and the level of preslaughter stress. Cortisol affected carcass quality, so higher levels of cortisol were associated with increased hot carcass weight, carcass fat thickness on the back and at the sacrum and marbling, but also with decreased meatiness. The most important meat quality parameters (pH and temperature after 60 minutes) deteriorated when blood lactate concentration was above 12 mmol/L.

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