# Potential Zoonotic Trematodes Recovered in Stray Cats from Kuwait Municipality, Kuwait

Osama Mohamed ElShfei El-Azazy<sup>1,2,\*</sup>, Nadra-Elwgoud Mohamed Ibrahim Abdou<sup>1,3</sup>, Amal Iskander Khalil<sup>4</sup>, Maha Khaled Al-Batel<sup>1</sup>, Qais Abdulrazak Habeeb Majeed<sup>5</sup>, Adawia Abdul-Ruhman Henedi<sup>1</sup>, Laila Mohamed Azad Tahrani<sup>1</sup>

<sup>1</sup>Veterinary Laboratories, Public Authority of Agriculture Affairs and Fish Resources, Kuwait; <sup>2</sup>Department of Parasitology, Faculty of Veterinary Medicine, Zagazig University, Egypt; <sup>3</sup>Department of Medicine and Infectious Diseases, Faculty of Veterinary Medicine, Cairo University, Egypt; <sup>4</sup>Department of Zoology, Faculty of Science, Tanta University, Egypt; <sup>5</sup>Department of Science, College of Basic Education, Public Authority for Applied Education, Kuwait

**Abstract:** Stray cats are a common feature roaming the streets and alleys of Kuwait; they could be a source of parasites, including trematodes, that affect humans. A survey was conducted to identify feline trematodes and throw the light on their public health significance in Kuwait. Out of 240 stray cats trapped from different localities of Kuwait from June 2011 to May 2012, 59 (24.6%) were found to be infected with 14 species of trematodes. The most common were trematodes of the genus *Heterophyes*, particularly *H. heterophyes* and *H. dispar* that were found in respectively 15.8% and 10.8% of the cats examined. Other trematodes recorded, with lower prevalences, were *Heterophyes nocens* (2.9%), *Haplorchis taichui* (3.8%), *Stictodora sawakinensis* (2.1%), *Stellantchasmus falcatus* (1.6%), *Echinochasmus japonicus* (1.6%), and *Mesostephanus dottrensi* (1.3%). *Centrocestus cuspidatus, Galactosomum fregatae, Ascocotyle* sp., *Mesostephanus appendiculatus, Haplorchis yokogawai*, and *Pygidiopsis genata* showed the lowest prevalence (0.4%) and intensity. The majority of the trematodes are recorded for the first time in Kuwait and even in the Gulf region. The study reveals that stray cats are good indicators of fish-borne trematodes in the environment. As all trematodes recovered are zoonotic, their significance to public health should be considred.

Key words: Heterophyes heterophyes, Heterophyes dispar, trematode, heterophyid, zoonosis, stray cat, Kuwait

#### INTRODUCTION

In Kuwait, stray cats are a common feature roaming streets and alleys, living among houses, on farms, and near abattoirs and fish markets, and feeding on garbage, insects, and small animals as well as on meat and fish offal. Consequently, they may contract infection with parasites and pollute environment with them posing threat to humans and domestic animals.

Kuwait, a small country located in the desert geographical region with sparse vegetation, has a rich marine environment that includes a long 170 km shoreline, and open water of the Arabian Gulf where large quantities of fish and shrimp are caught. The fisheries sector in the country is important, and seafood is considered a significant source of protein for the people. Ac-

cording to the 2012 census, Kuwait produces 3,686,328 kg of fish including 30,482 kg from aquaculture, in addition it imports 8,803,178 kg of fish, mainly refregirated from Egypt, Iran, India, Pakistan, and Saudi Arabia as well as mainly frozen fish from Southeast Asia.

Although parasites of stray cats have been surveyed in many parts of the world [1-7] as well as in the Gulf region [8-10], little is known about the status of tremtode infections, particularly fish-borne zoonotic trematodes (FZTs), in Kuwait [11]. To assess the prevalence of FZTs and the role of cats as reservoir hosts for fish-borne parasites, a systematic study on parasites of stray cats in Kuwait was performed, with the identification of the recovered trematodes and the evaluation of their zoonotic importance.

#### • Received 15 September 2014, revised 26 March 2015, accepted 14 April 2015.

© 2015, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### **MATERIALS AND METHODS**

#### Cat sampling and study sites

During the period extended from June 2011 to May 2012, stray cats were trapped from various districts to cover the 6 ad-

<sup>\*</sup>Corresponding author (El\_Azazy@hotmail.com)

ministrative Governorates (Al-Asema, Hawalli, Farwanyiah, Mubarak Al-Kabeir, Al-Jahara, and Al-Ahmadi) of Kuwait (Fig. 1) using special live traps baited with fish. In order to determine the geographical distribution and demographics, the districts, from which cats were captured, were grouped into 2 main localities on the basis of the following criteria, e.g., the standard of living of inhabitants, density of population, level of municipal services, and accumulation of garbage. Locality 1 contained the districts which are inhabited by well-to do-Kuwaitis with good service levels and low population densities, while Locality 2 contains the districts which are inhabited by expatriates with low service levels and high population densities. Some districts of Locality 2, inhabited by laborers and fishermen, were characterized by the accumulation of garbage and large numbers of stary cats.

## Laboratory investigations

The captured cats were transferred to the laboratory where date of examination, site of trapping, gender and age, were recorded. The age of cats was assessed by dentition, body size, and maturation of genital organs. The cats were grouped by age into adults (more than 6 months) and juveniles ( $\leq$ 6 months). After anesthetizing cats using Rompun 2% (Bayer, Leverkusen, Germany) intramuscular (1.5 ml/10 kg), they were killed humanely by intracardial injection with T61 (Schering-Plough Intervet, Elkhorn, Nebraska, USA), 1-4 ml according to age and weight.

#### Organ examination for trematodes

Extra-intestinal organs: Bile ducts and air passages were opened, and liver and lung tissues were dissected using fine scissors in a petri dish containing normal saline. Then, all the contents were examined under a stereomicroscope for detection of trematodes.

Intestinal tract: The gastrointestinal tracts were removed from cats at necropsy. The intestines were separated and opened in containers containing normal saline; the mucous membrane of each organ was scraped between the blades of a

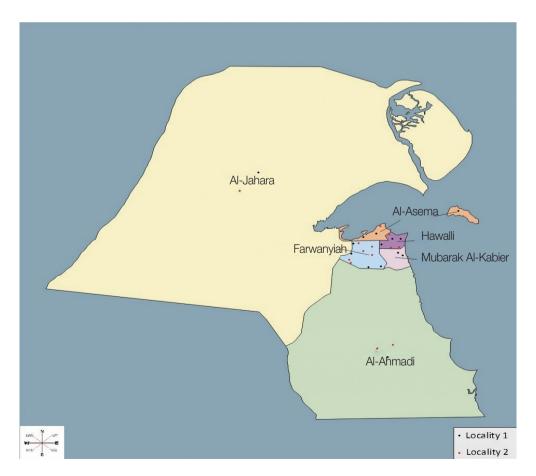


Fig. 1. Surveyed areas of the 6 governorates of Kuwait, grouped into 2 main localities.

forceps. The contents were then washed out over a 500  $\mu m$  sieve with tap water. The residues were systematically searched under a stereomicroscope for trematodes, and any specimens found were transferred to 70% ethanol for preservation until identification.

#### Preparation of specimens for identification

The worms were counted. For identification, they were stained with lactophenol cotton blue (LPCB) using the method of Henedi and El-Azazy [12]. Some specimens were stained with alum carmine, cleared in xylene and clove oil, and mounted in Canada balsam.

#### Trematode identification

The recovered parasites were identified using the keys/text-books of Soulsby [13] and Bray et al. [14,15]. In addition, helminth specimens were sent to John M. Kinsella, Helm-West Laboratory, Missoula, Montana, USA for confirmation, and voucher specimens were deposited in the US National Parasite Collection (accession no. 107983-107988). Measurement was done using a micrometer slide and a digital camera (Leica EC3, Wetzlar, Germany). All measurements are given in µm, as averages.

# Statistical analysis

Data were analyzed and statistical comparisons were performed using Statistix® (copyright® 1985-2003, analytical software, version 8, Tallahassee, Florida, USA). Summary data for the parasite intensity are expressed as arithmetic means ± SEM (standard error of mean). The mean intensity of infection as defined by Bush et al. [16] is the average intensity of a particular parasite species among the infected members of a particular host species. The prevalence and confidence intervals (CI) were calculated as described by Sokal and Rohlf [17]. The prevalence is the proportion of infected hosts among all hosts

examined. CI of prevalence indicates the accuracy of the estimation using CI 95% is advisable.

#### **RESULTS**

# Overall infection rate and species of trematodes recovered

A total of 240 stray cats were examined, 117 from Locality 1 and 123 from Locality 2 (Table 1). Cats were captured around houses of fishermen, fish markets, and trash containers. A total of 59 (24.6%) cats were found to be infected with FZTs, and 23 (39.0%) being infected with more than 2 different species. Total 14 species of FZTs were recovered in the small intestine of cats. Among them, the great majority (11 spp.) were members of the family Heterophyidae, 2 species (Mesostephanus dottrensi and M. appendiculatus) were those of the Cyathocotylidae, and only 1 species, Echinochasmus japonicus, belonged to the Echinostomatidae. Three species of the genus Heterophyes (H. heterophyes, H. dispar, and H. nocens) were detected in 15.8%, 10.8%, and 2.9% of stray cats examined, and their average numbers were 141, 83, and 203 worms per cat infected, respectively. Trematodes including Centrocestus cuspidatus, Galactosomum fregatae, Ascocotyle sp., M. appendiculatus, H. yokogawai, and Pygidiopsis genata showed the lowest prevalence (0.4%) and intensity (Table 2).

# Brief description of trematodes recovered (unit: µm) Heterophyes heterophyes

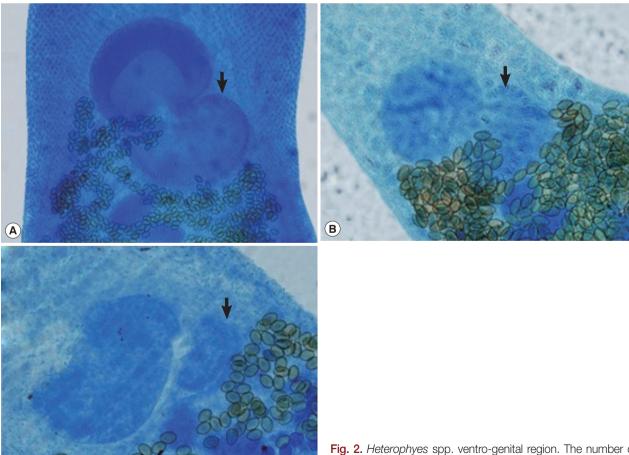
Pear-shaped,  $1,256\times379$  (n = 10). Oral sucker terminal,  $35\times61$ . Pre-pharynx clear, 60; pharynx,  $47\times45$ . Esophagus, 200 long. Ventral sucker separated from genital sucker, large, well developed, located at middle of the body between 2 ceca,  $146\times169$ . Genital sucker about the same size of ventral sucker,  $132\times157$ , located posterio-lateral to ventral sucker, armed with incomplete crown of 70-80 spines (Fig. 2). Ovary small,

Table 1. Demographic data (site, gender, age, and season) of stray cats examined in Kuwait

Locality	Season								
	Wet				Dry				
	Male		Female		Male		Female		
	Adult	Juvenile	Adult	Juvenile	Adult	Juvenile	Adult	Juvenile	
1	14	9	19	7	21	13	21	13	
2	20	9	41	10	10	10	14	9	
Total	34	18	60	17	31	23	35	22	

Table 2. Prevalence and mean intensity of trematodes in stray cats from Kuwait relative to age

	All cats (n=240)		Adult	(n = 160)	Young (n=80)		
	Positive (%)	Intensity ± SEM	Positive (%)	Intensity ± SEM	Positive (%)	Intensity ± SEM	
All trematodes	24.6	154±19	28.7	116±17	16.2	288±23	
Heterophyes heterophyes	15.8	141 ± 17	28.3	85±11	10.0	$351 \pm 24$	
Heterophyes nocens	2.9	$203 \pm 17$	3.8	187 ± 18	1.3	$301 \pm 0$	
Heterophyes dispar	10.8	83±12	13.8	70±11	5.0	156±14	
Haplorchis yokogawai	0.4	8±3	0.6	8±3	-	-	
Haplorchis taichui	3.8	4±2	5.0	4±2	1.3	2±0	
Stellantchasmus falcatus	1.6	5±2	2.5	5±2	-	-	
Centrocestus cuspidatus	0.4	3±0	0.6	3±0	-	-	
Ascocotyle sp.	0.4	1 ± 0	-	-	1.3	1 ± 0	
Pygidiopsis genata	0.4	1 ± 0	-	-	1.3	1 ± 0	
Stictodora sawakinensis	2.1	7±3	3.1	7±3	-	-	
Galactosomum fregatae	0.4	2±0	0.6	2±0	-	-	
Mesostephanus. dottrensi	1.3	4±2	0.6	6±0	2.5	3±1	
Mesostephanus appendiculatus	0.4	1 ± 0	-	-	1.3	1 ± 0	
Echinochasmus japonicus	1.6	5±2	2.5	5±2	-	-	



**Fig. 2.** Heterophyes spp. ventro-genital region. The number of the rodlets on the genotyl (arrow) is about 77 in *H. heterophyes* (A), 52 in *H. nocens* (B), and 30 in *H. dispar* (C). × 100.

compact, pretesticular,  $65 \times 69$ . Testes oval to round, horizontal, at caudal region, right,  $118 \times 98$ ; left,  $121 \times 97$ . Vitelline glands restricted in 2 patches anterior to testes. Eggs small, yellow  $25 \times 15$ .

# Heterophyes nocens

Oval to elongate body,  $1,157 \times 390$  (n = 10). Oral sucker terminal,  $44 \times 50$ . Pre-pharynx short, 64 long; followed by small pharynx,  $37 \times 37$ . Esophagus, 157 long. Ventral sucker large,  $150 \times 170$ . Crescent genital sucker posterio-lateral to ventral sucker carries 50-62 spines (Fig. 2),  $110 \times 120$ . Seminal vesicle bilobed, between genital sucker and ovary. Ovary globular,  $62 \times 81$ . Testes horizontal at posterior part of the body, right,  $95 \times 77$ ; left,  $98 \times 81$ . Eggs numerous,  $25 \times 15$ .

#### Heterophyes dispar

Elongate to pear-shaped body,  $943 \times 318$  (n = 10). Oral sucker terminal to subterminal, small,  $54 \times 67$ . pre-pharynx, 64 long; pharynx,  $48 \times 45$ . Esophagus, 159 long. Ventral sucker large,  $126 \times 150$ . Genital sucker small, armed with 15-35 spines (Fig. 2), at the lower left side of ventral sucker,  $81 \times 57$ . Ovary round,  $73 \times 64$ . Testes obligue or opposite, right,  $104 \times 73$ ; left,  $115 \times 70$ . Seminal receptacles at lower right side of the ovary. Eggs numerous,  $25 \times 10$ .

#### Stictodora sawakinensis

Body elongate, pyriform  $870 \times 208$  (n = 10). Oral sucker round, subterminal,  $52 \times 52$ . Pre-pharynx, 56 long; Pharynx,  $40 \times 41$ . Esophagus, 46 long. Ventral sucker and gonotyl enclosed in ventrogenital sac. Ventrogenital sac looks like egg plant, divided in 2 portions, smaller  $46 \times 55$  armed with 6-10 longitudinal rows of spines which trifid or single tipped, vary in number (14-28), large portion, saccular,  $99 \times 65$ . Seminal vesicle, bilobed or tripartite, posterior to ventrogenital sac. Testes obliquely, oval to round in shape located at second third of the body. Upper testis,  $95 \times 50$ ; lower,  $49 \times 52$ . Ovary between testes,  $39 \times 50$ . Vitellaria post-testicular extending outside cecal field. Eggs oval, dark yellow,  $20 \times 10$ .

#### Ascocotyle (Phagicola) longa

Body pyriform,  $889 \times 265$ , (n = 1). A small prolongation, 10, is seen at the top of oral sucker. Oral sucker,  $51 \times 51$ , surrounded by single crown of 16 well-developed spines. At the back of oral sucker, dorsal to prephrynx, extends a tapered muscular extensible prolongation. Pre-pharynx, 204 long. Pharynx mus-

cular, well developed,  $80 \times 50$ , esophagus short, 20. Ventral sucker spherical,  $80 \times 80$ . Testes equal, opposite in position,  $80 \times 50$ . Ovary lies posterior ventral sucker, obscured by numerous eggs. Vitelline glands follicular, in latero-posterior region. Eggs,  $50 \times 30$ , fill area between ventral sucker and testes.

#### Haplorchis yokogawai

Oval to pyriform body,  $662 \times 295$  (n = 10). Oral sucker terminal,  $45 \times 53$ . Pre-pharynx short, 21 long. Pharynx oval to spherical,  $29 \times 34$ . Esophagus, 120 long. Ventral sucker small,  $66 \times 82$  with apex composing of large ventral lobe armed with numerous tiny spines and larger pair of sclerites in addition to 3 smaller lobes armed with tiny spines (reduced cavity, armed with small spines). Gonotyl absent. Ventrogenital sac unarmed,  $142 \times 119$ . Ovary round,  $47 \times 49$  posterior to ventral sucker at left side. Testis single,  $79 \times 71$  at posterior part of the body. Eggs numerous, uncountable, distributed around and posterior to ventral sucker,  $20 \times 10$ .

#### Haplorchis taichui

Body elongated,  $562 \times 197$  (n=4). Oral sucker terminal,  $32 \times 50$ . Pre-pharynx, 10 long. Pharynx,  $27 \times 30$ . Esophagus, 85 long. Ventral sucker median,  $60 \times 55$  possesses reduced cavity, armed with 1 or 2 ventral crescent shaped groups of 12-16 sclerites. Seminal vesicle between ventral sucker and ovary. Ovary globular,  $53 \times 53$ . Testis single, fan shaped, lobulated, at posterior part of the body,  $46 \times 36$ . Vitteline glands at posterior part of the body, masked with large amount of yellow eggs which are  $20 \times 10$ .

#### Stellantchasmus falcatus

Small pyriform body,  $528 \times 180$  (n = 10). Oral sucker terminal,  $30 \times 39$ . Pre-pharynx short, 26. Pharynx globular,  $23 \times 29$ . Esophagus, 106 long. Ventral sucker semicircle,  $30 \times 31$ , slightly deviated to right side between 2 caeca, armed with minute spines encircling mouth and forming 2 denses. Seminal vesicle, elongated sac-like, situated at the opposite side of ventral sucker,  $66 \times 24$ . Ovary submedian, lies between ventral sucker and right testis,  $40 \times 40$ . Testes horizontal, right,  $65 \times 41$ ; left,  $64 \times 44$ . Vitellaria follicular. Eggs,  $20 \times 10$ .

#### Centrocestus cuspidatus

Body very small  $468 \times 186$ , with unique conical flask shape, with a spiny neck, (n=3). Oral sucker sub-terminal,  $32 \times 57$ , with dorsal lip armed with 2 circles of alternating spines. Inner

circle contains 9-21 spines, outer contains 10-30 spines. Prepharynx, 41 long. Pharynx,  $30 \times 40$ . Esophagus short, 59 long. Ventral sucker,  $40 \times 60$ . Seminal vesicle bipartite. Ovary,  $60 \times 90$ . Testes opposite, right,  $70 \times 30$ ; left,  $80 \times 90$ . Eggs large,  $35 \times 22$ , few in number (5-11).

#### Galactosomum fregatae

Body elongated, rounded at both extremities, with slight constriction behind ventrogenital complex, covered with spines and scattered pigment granules  $2.877 \times 475$  (n = 2). Oral sucker subterminal, large; 90×100. Pre-pharynx, 325 long. Pharynx, 100 × 90. Esophagus very short, 87 long. Ventrogenital complex at the beginning of second third of body, it's composed of ventral genital sac that's connected to lobulated seminal vesicle by prostatic ejaculatory duct. Ventral genital sac looks like a rose, contains ventral sucker and genital atrium. Ventral sucker, 143×155 with 2 separate groups of spines, first group at right with large spines arranged alternatively in horse shoe manner, while second group of spines opposite to first one and composed of 2 sets of spines, first set tiny, arranged in crescent order which is connected to another set of small spines arranged in small patch. Genital atrium, 124×166. Seminal vesicle strongly developed lying opposite to acetabulum, 430×160, composed of 2 chambered separated by a distinct constriction, upper oval to round while lower elongated, sac like. Testes diagonal, upper, 100×170; lower, 170×120. Ovary round, post-acetabulam, 120×110. Seminal receptacle large, 120×90 post ovarian. Eggs numerous,  $20 \times 10$ .

# Pygidiopsis genata

Body looks like conical flask, with maximum width at the ovary level,  $550 \times 240$  (n=1). Oral sucker large, terminal, without appendage,  $40 \times 50$ . Pre-pharynx, 36. Pharynx muscular,  $30 \times 40$ . Ceca reach ovary and turn somewhat dorsomedially at terminal portion. Ventral sucker median, round, globular,  $60 \times 40$ . Genital sucker crescent-like, lobulated, spineless, attached to ventral sucker,  $60 \times 30$ . Seminal vesicle bipartite, post acetabulum,  $50 \times 28$ . Ovary round, post to seminal vesicle,  $50 \times 40$ . Seminal receptacles attached to ovary,  $40 \times 48$ . Testes oval, opposite, right,  $30 \times 60$ ; left,  $40 \times 40$ . Eggs scattered on area posterior to ventral sucker and anterior to testes,  $20 \times 10$ .

#### Mesostephanus dottrensi

Pyriform in shape, with small dorsoterminal appendage at end of body,  $1,000 \times 398$  (n = 6). Oral sucker,  $47 \times 57$ . Pre-

pharynx absent; pharynx,  $41 \times 49$ . Esophagus, 62 long. Intestinal ceca reaching close to posterior extremity. Between 2 ceca; 2 columns, each of 6 small dense patches. Acetabulum,  $47 \times 52$ . A hold fast organ occupies the region behind ventral sucker. Two large testes, subspherical or oval, tandem, at posterior half of the body. Upper,  $102 \times 100$ ; lower,  $107 \times 100$ . Ovary pyramidal, between testes,  $140 \times 140$ . Cirrus sac long, well developed,  $270 \times 65$ ; genital pore terminal. Vitellaria well-developed; in horse-shoe manner, composed of large follicles confined around gonads, but not confluent posteriorly. Vaginal sphincter ellipsoidal. Coffee bean-shaped eggs large; yellowish in color, few in number (1-3 eggs)  $74 \times 51$ .

#### Mesostephanus appendiculatus

Body,  $680 \times 410$  (n=1). Oral sucker, 54 long. Pre-pharynx very short. Pharynx,  $50 \times 36$ ; esophagus length 25. Ventral sucker,  $60 \times 60$ . Tribocytic organ well developed. Ovary intertesticular,  $60 \times 50$ . Testes oblique, in posterior part of the body, right,  $116 \times 70$ ; left,  $118 \times 75$ . Cirrus sac and cirrus well developed. Vitellaria follicular arranged in semi circle at posterior half of the body. Eggs,  $107 \times 60$ .

#### Echinochasmus japonicus

Body ovoid or plump,  $1,200\times39$  (n=6). Anterior part of the body armed with collar of 24 spines (12 on each side) alternating in position. Head crown,  $75\times90$ . Oral sucker spherical,  $40\times50$ . Pre-pharynx between 2 sets of spines, 51 long. Pharynx,  $70\times110$ . Esophagus, 113 long. Caecum blind, reaching end of posterior extremity of trematode. Ventral sucker large,  $150\times110$ . Ovary posterior to ventral sucker,  $130\times160$ . Cirrus sac large, elongated to oval, between the intestinal bifurcation and ventral sucker. Seminal receptacle large, sac-like between ovary and upper testis. Testes tandem, posterior at one-third body, upper, transversely oval,  $110\times120$ ; lower, rounded,  $160\times180$ . Ootype obvious between ovary and upper testis. Vitelline glands extend on from acetabular level to posterior body extremity. Eggs few (1-2) and measure  $90\times50$ .

## Effect of demographic factors on parasitism

Older cats had a significantly higher infection rate, 46 adult cats had a significantly higher intensity and infection rate with trematodes than juveniles did (P=0.03). In addition, trematodes showed significantly higher prevalence and intensity in cats which were trapped in Locality 2 than in Locality 1 (P=0.03). These data were very much influenced by prevalence

		Se	Sex		Age		Season		Locality	
		Male	Female	Adult	Juvenile	Wet	Dry	1	2	Total
Prevalence	е	21.7	26.9	28.7*	16.2	29.5	18.9	13.71	34.9ª	24.6
Range	Low	14.27	19.57	8.94	21.87	21.8	12.1	8.0	26.5	19.3
	High	30.76	35.21	26.19	36.43	38.1	27.5	21.3	44.1	30.5

Table 3. Prevalence and 95% confidence intervals (C.I.) of parasites in cats (240) of different age groups, sexes, and seasons

es of *Heterophyes* spp., which constituted the majority of the trematode community. *Heterophyes* spp. were influenced by age and site but not by season and gender; significantly higher (P=0.00) prevalences were observed in adults and in Locality 2 when compared to juveniles and Locality 1, respectively (Table 3). The prevalence of trematode infection was higher (29.5%) in the wet season than in the dry season (18.9%), while the intensity of infection was higher in the dry season (mean = 279.1 burdens) than in the wet season (mean = 84.9 burdens). However, these differences were not statistically significant (P>0.05).

#### **DISCUSSION**

All trematodes recorded in this study are transmitted by eating raw or undercooked fish, and most of them have been reported to cause health problems in humans [18,19]. The role of stray cats as reservoir hosts of fish-borne trematodes is highlighted in this study; however, their role in sustaining transmission of them is questionable. Little is known about the epidemiology of fish-borne trematodiasis in Kuwait.

It is known that cats and other fish-eating mammals and birds act as final hosts for fish-borne trematodes. However, few studies have been done on the role of reservoir hosts, including cats, in the epidemiology of FZTs; in some endemic areas in Southeast Asian countries, it has been suggested that a minor role is played [18,19]. However, in Vietnam, cats, dogs, and pigs have been found to play major roles in the epidemiology of FZTs in aquaculture [20].

In the present study, although it is evident that cats were aquired infection with trematodes by eating raw fish, it is not possible to know the source of infection, i.e., via local or imported fish. This point needs more investigation. Locally, no systematic studies have been conducted to investigate infection of fish with metacercariae. However, on 2 occasions, *Stictodora fuscata* and *S. tridactyla* were reported after experimental feeding of kittens with metacercaria-infected fish species, respectively, *Liza macrolepis* (mullet) and *Aphanius dispar* (killfish),

caught from Kuwait Bay [21,22]. On other hand, metacercariae of fish-born trematodes have been detected in fish in some countries nearby Kuwait [23-27].

Kuwait imports fish from many countries where FZTs are endemic, e.g., Egypt [25], Iran [28], India [29,30], and Southeast Asia [18,19]. It is worth mentioning that metacercariae in fish tissues can survive and be infective for 10-18 days at -12°C and for 3-7 days at -20°C [31]. Importation of fish from a wide variety of geographical and ecological regions can explain the broad spectrum of FZTs diversity and recording a large number of species in this study.

Heterophyes spp., particularly *H. heterophyes*, were the most prevalent trematodes and the most abundant helminth parasites in the present study. In previous studies in Kuwait, *H. heterophyes* was the only trematode recorded from dogs [32] and cats [11]. In addition, it has been recorded from humans and animals in other Gulf countries, e.g., Iran [28], United Arab Emirates [9], and Iraq [33]. Seo [34] and Chai et al. [35] reported *H. heterophyes* infections in Korean workers who returned from Saudi Arabia, Bahrain, and Kuwait. This could indicate that heterophyiasis is endemic in the Gulf region.

The significantly higher prevalence observed in adult cats, is believed to be caused by longer exposure and accumulation of parasites [36]. This could be also related to the competition for food, where adults have a greater capacity to obtain fish offal at garbage containers near fish markets in comparison to younger cats. Higher prevalence and intensity rates of trematodes were observed in cats captured in Locality 2 where expatriate laborers clean fish by themselves and throw the offal in garbage to be accessed by stray cats. Another reason for the higher prevalence rates at Locality 2 could be that the inhabitants prefer the consumption of imported fish, which could have higher numbers of FZTs.

The great majority of FZTs are recorded for the first time in Kuwait, and even in the Gulf region. This study proved that stray cats are good indicators of FZTs in the environment of Kuwait. The paper draws the attention of food safety services,

<sup>&</sup>lt;sup>a</sup>Statistically significant (P<0.05).

fishery officials, marketing agencies, and public and veterinary authorities in Kuwait to the risk, which may be posed by consumption of raw or under cooked seafood.

#### **ACKNOWLEDGMENTS**

The authors are grateful to the Kuwait Foundation for Advancement of Sciences for financial support as well as to John M. Kinsella, Helm-West Laboratory, Missoula, Montana, USA, for his kind help in verifying the identification of trematodes.

#### **CONFLICT OF INTEREST**

We have no conflict of interest related to this work.

#### **REFERENCES**

- McGlade TR, Robertson ID, Elliot AD, Read C, Thompson RCA. Gastrointestinal parasites of domestic cats in Perth, Western Australia. Vet Parasitol 2003; 117: 251-262.
- Carleton, RE, Tolbert MK. Prevalence of *Dirofilaria immitis* and gastrointestinal helminths in cats euthanized at animal control agencies in northwest Georgia. Vet Parasitol 2004; 119: 319-326.
- Labarthe N, Serrao ML, Ferreira AMR, Almeida NKO, Guerrero J. A survey of gastrointestinal helminths in cats of the metropolitan region of Rio de Janeiro, Brazil. Vet Parasitol 2004; 123: 133-139.
- 4. Sohn WM, Chai JY. Infection status with helminthes in feral cats purchased from a market in Busan, Republic of Korea. Korean J Parasitol 2005; 43: 93-100.
- Borthakur SK, Mukharjee SK. Gastrointestinal helminthes in stray cats (*Felis catus*) from Aizawl, Mizoram, India. Southeast Asian J Trop Med Public Health 2011; 42: 255-258.
- Khalafalla RE. A survey study on gastrointestinal parasites of stray cats in Northern region in Nile Delta, Egypt. PLoS One 2011; 6: e20283.
- 7. Chai JY, Bahk YY, Sohn WM. Trematodes recovered in the small intestine of stray cats in the Republic of Korea. Korean J Parasitol 2013; 51: 99-106.
- 8. Changizi E, Mobedi I, Salimi-Bejestani MR, Razaei-Doust A. Gastrointestinal helminthic parasites in stray cats (*Felis catus*) from North of Iran. Iranian J Parasitol 2007; 2: 25-29.
- 9. Schuster RK, Thomas K, Sivakumar S, O' Donovan D. The parasite fauna of stray domestic cats (*Felis catus*) in Dubai, United Arab Emirates. Parasitol Res 2009; 105: 125-134.
- Abu-Madi MA, Behnke JM, Prabhaker KS, Al-Ibrahim R, Lewis JW. Intestinal helminthes of feral cat populations from urban and suburban districts of Qatar. Vet Parasitol 2010; 168: 284-292.

- 11. Abdul-Salam J, Baker K. Prevalence of intestinal helminths in stray cats in Kuwait. Pakistan Vet J 1990; 10: 17-21.
- 12. Henedi AAM, El- Azazy OME. A simple technique for staining of platyhelminthes with the lactophenol cotton blue stain. J Egypt Soc Parasitol 2013; 43: 419-423.
- 13. Soulsby EJL. Helminths, Arthropods and Protozoa of Domesticated animals, 7th ed. London, UK. Bailliere Tindall. 1982, pp 8-87
- Bray RA, Gibson DI, Jones A. Keys to the Trematoda. Vol. 1. London, UK. CAB International and Natural History Museum. 2002, pp 1-521.
- 15. Bray RA, Gibson DI, Jones A. Keys to the Trematoda. Vol. 3. London, UK. CAB International and Natural History Museum. 2008, pp 1-824.
- Bush AO, Lafferty KO, Lotz JM, Shostak AW. Parasitology meets ecology on its own terms: Margolis et al. revisited. J Parasitol 1997; 83: 575-583.
- 17. Sokal RR, Rohlf FJ. Biometry: the principles and practice of statistics in biological research. 4th ed. New York, USA. WH Fereeman and Co. 1994, pp 1-937.
- 18. Chai JY, Murrell KD, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. Int J Parasitol 2005; 35: 1233-1254.
- 19. Chai JY, Lee SH. Food-borne intestinal trematode infections in the Republic of Korea. Parasitol Int 2002; 51: 129-154.
- 20. Lan-Anh NT, Phuong NT, Murrell KD, Johansen MV, Dalsgaard A, Thu LT, Kim-Chi TT, Thamsborg SM. Animal reservoir hosts and fish-borne zoonotic trematode infections on fish farms, Vietnam. Emerg Infect Dis 2009; 15: 540-546.
- 21. Abdul-Salam J, Nair SBH, Ashkanani H. Surface ultrastructure of *Stictodora fuscatum* (Trematoda: Heterophyidae) from Kuwait Bay. Parasitol Día 2000; 24: 55-59 (online).
- 22. Abdul-Salam J, Sreelatha BS, Ashkanani H. Surface ultrastructure of *Stictodora tridactyla* (Trematoda: Heterophyidae) from Kuwait Bay. Parasitol Int 2000; 49: 1-7.
- Farahnak A, Massoud J. Medically important metacercariae (larva trematodes) in Khuzestan fishes, Iran. Acta Med Iranica 1999; 37: 59-62.
- 24. Ali AH. Pathological effects of helminths partasitic on some local fishes. M. Sc. thesis, College of Agriculture, University of Basrah, Iraq. 2001, pp 1-174 (in Arabic).
- 25. El-Seify MA, Desouky ARY, Bazh EK. Epidemiological studies on some fish-borne parasites. Egypt Vet Med Soc Parasitol J 2005; 2: 570-592.
- 26. Kardousha MM. Helminth parasite larvae collected from Arabian Gulf fish. IV. Description of 4 larvae including two metacercariae, one didymozoid and one acanthocephalan. Arab Gulf J Sci Res 2005; 23: 23-27.
- 27. Karimian E, Ghorbani R, Hajimoradlou A. First occurrence and intensity of *Posthodiplostomum cuticola* (Nordmann, 1832) (Digenea; Diplostomatidae) metacercariae in monkey goby (*Neogobius pallasi* berg, 1916) in the Zarringol stream, Golestan Province, Iran. Global Veterinaria 2013; 10: 505-510.
- 28. Massoud J, Jalali H, Reza M. Studies on trematodes of the family

- Heterophyidae (Odhner, 1914) in Iran. 1. Preliminary epidemiological surveys in man and carnivores in Khuzestan. J Helminthol 1981; 55: 255-260.
- 29. Rajavelu G, Raja EE. On helminth parasites in domestic cat in Madras. Cheiron 1988; 17: 11-14.
- 30. Jacob L, Pillaik M. Incidence of parasitic infection in dogs in Thrissur, Kerala. J Vet Anim Sci 1991; 22: 149-150.
- 31. Fan PC. Viability of metacercariae of *Clonorchis sinensis* in frozen or salted fresh water fish. Int J Parasitol 1988; 28: 603-605.
- 32. Abdul-Salam J. Intestinal helminth parasites of stray dogs in Kuwait. Arab Gulf J Sci Res 1986; 4: 659-663.
- 33. Mohammad KM. The parasitic fauna and the food habits of the wild jungle cat *Felis chausfuraxde* Winton, 1898, in Iraq. Bull Iraq Nat Hist Mus 2008; 10: 65-78.
- 34. Seo BS. Biology and clinical aspects of Heterophyidae. Human Sci 1979; 3: 784-791 (in Korean).
- 35. Chai JY, Seo BS, Lee SH, Hong SJ, Sohn WM. Human infections by *Heterophyes heterophyes* and *H. dispar* imported from Saudi Arabia. Korean J Parasitol 1986; 24: 82-88.
- 36. Dung DT, De NV, Waikagul J, Dalsgaard A, Chai JY, Sohn WM, Murrell D. Fishborne zoonotic intestinal trematodes, Vietnam. Emerg Infect Dis 2007; 13: 1828-1833.