

# Xenomonitoring of Different Filarial Nematodes Using Single and Multiplex PCR in Mosquitoes from Assiut Governorate, Egypt

Ahmed Kamal Dyab, Lamia Ahmed Galal\*, Abeer El-Sayed Mahmoud, Yasser Mokhtar

Department of Medical Parasitology, Faculty of Medicine, Assiut University, Assiut, Egypt

**Abstract:** *Wuchereria bancrofti*, *Dirofilaria immitis*, and *Dirofilaria repens* are filarial nematodes transmitted by mosquitoes belonging to *Culex*, *Aedes*, and *Anopheles* genera. Screening by vector dissection is a tiresome technique. We aimed to screen filarial parasites in their vectors by single and multiplex PCR and evaluate the usefulness of multiplex PCR as a rapid xenomonitoring and simultaneous differentiation tool, in area where 3 filarial parasites are coexisting. Female mosquitoes were collected from 7 localities in Assiut Governorate, were microscopically identified and divided into pools according to their species and collection site. Detection of *W. bancrofti*, *D. immitis*, and *D. repens* using single PCR was reached followed by multiplex PCR. Usefulness of multiplex PCR was evaluated by testing mosquito pools to know which genera and species are used by filarial parasites as a vector. An overall estimated rate of infection (ERI) in mosquitoes was 0.6%; the highest was *Culex* spp. (0.47%). *W. bancrofti*, *D. immitis*, and *D. repens* could be simultaneously and differentially detected in infected vectors by using multiplex PCR. Out of 100 mosquito pools, 8 were positive for *W. bancrofti* (ERI of 0.33%) and 3 pools each were positive for *D. immitis* and *D. repens* (ERI 0.12%). The technique showed 100% sensitivity and 98% specificity. El-Nikhila, El-Matiaa villages, and Sahel Seleem district in Assiut Governorate, Egypt are still endemic foci for filarial parasites. Multiplex PCR offers a reliable procedure for molecular xenomonitoring of filariasis within their respective vectors in endemic areas. Therefore, it is recommended for evaluation of mosquito infection after lymphatic filariasis eradication programs.

**Key words:** *Wuchereria bancrofti*, *Dirofilaria immitis*, *Dirofilaria repens*, xenomonitoring, mosquito, multiplex PCR, Egypt

## INTRODUCTION

Filariasis is one of the most common parasitic infections of the world. Approximately 106 million people are infected, with 1 billion people thought to be at risk of infection [1]. Lymphatic filariasis caused by *Wuchereria bancrofti* remains a public health problem in many tropical countries, including Nile delta and Assiut Governorate in Upper Egypt [2,3]. It is a serious tropical disease that can lead to chronic, disabling conditions such as lymphedema, elephantiasis, and genital deformities [4].

*Dirofilaria immitis* and *D. repens* are important filarioids of carnivores and are also accidental parasites of humans. An increase in annual numbers of reported cases in humans in areas

previously recognized as non-endemic [5], with 372 cases described between 1995 and 2000 [6], has led to human dirofilariasis to be classified as an emerging zoonosis [7]. Human dirofilariasis is manifested as subcutaneous or pulmonary nodules and in rare cases, eyes and periocular tissues can be involved, either caused by *D. immitis* or *D. repens* [8]. Although these lesions are typically benign but they may be misdiagnosed as more important diseases, such as lung tumors, needing prompt unnecessary diagnostic procedures with attendant cost, discomfort, and morbidity [9].

Xenomonitoring of *W. bancrofti*, *D. immitis*, and *D. repens* when based on individual dissection of female mosquito vectors (*Culex*, *Aedes*, and *Anopheles* sp.) [10] is a time-consuming and barely applicable technique [11]. Over the past decades, several PCR-based tools have been developed to detect filarial DNA in the definitive hosts and mosquito vectors [12]. Multiplex PCR is a variant of PCR techniques in which 2 or more target sequences can be amplified by including more than 1 pair of primers in the same reaction [13]. Multiplex PCR can offer

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\*Corresponding author (lamiazak@gmail.com)

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scope for simultaneous detection of the tested filarial parasites for rapid surveillance and monitoring of mosquitoes [14,15].

The present study aimed to screen the presence of filarial parasites in mosquitoes in certain endemic localities in Assiut Governorate, Egypt by single and multiplex PCRs, and to compare the results of these 2 techniques in order to evaluate the usefulness of multiplex PCR for simultaneous detection and differentiation between the different filarial parasites in various mosquito species as a rapid tool for epidemiological surveillance.

## MATERIALS AND METHODS

### Sampling areas and mosquito collection

The study was done in Assiut Governorate, Egypt, located about 375 km to the south of Cairo. It was carried out in 7 districts; El-Nikhila, El-Matiaa and Mankabad villages, Sahel Seleem, El-Badary, Dairout, and Manfalout districts (Fig. 1). A total of 2,500 adult female mosquitoes were collected (from March 2012 to December 2013) by using mechanical aspirators [16]. The collected mosquitoes were microscopically identified according to a previously described method [17]. Female mosquitoes were divided into 100 mosquito-pools according



**Fig. 1.** Map of Assiut Governorate, Egypt. Sites in red color represent areas of mosquito collection (<http://en.Wikipedia.Org/Wiki/Asyut>).

to species and collection site (each pool contained 25 mosquitoes). All pool specimens were labeled and maintained refrigerated until being used.

Two types of PCR assays were carried out for detection of filarial parasites (*W. bancrofti*, *D. immitis*, and *D. repens*) on all mosquito pools. Single PCR was firstly conducted followed by multiplex PCR. The same sets of primers used in single PCR were also applied in Multiplex PCR. The utility of Multiplex PCR in the simultaneous detection and differentiation of filarial parasites was evaluated by testing every pool of mosquito genera (*Culex*, *Anopheles*, and *Aedes*) to know which genera and species are used as a vector for filarial parasites in endemic localities, where they coexist.

### Extraction of filarial DNA from mosquitoes

Genomic DNA was isolated from all mosquito pools using Qiagen tissue kit (QIAamp DNA Minikit, Qiagen, Hilden, Germany) following the manufacturer's instructions.

### Single PCR Assay

A Perkin Elmer 480 Thermal Cycler (Perkin Elmer Cetus, Norwalk, Connecticut, USA) was used for the PCR amplification process. A pair of forward and reverse PCR primers were used for each of the 3 filarial parasites. The oligonucleotide primers used for the PCR were obtained from (Metabion International AG, Martinsried, Germany). PCR procedure for detection of *W. bancrofti* in mosquito vectors was executed according to Kithmini et al. [18], using forward pWb12 F (5'-CT-GAGTGAAATCAATGAACTGC-3') and reverse pWb12 R (5'-GTCCATCCGATGAAGTCCACC-3') primers. Rishniw et al. [19] described the PCR primers used for detection of *D. immitis* and *D. repens* in mosquito vectors, respectively. Forward DI COI-F1 (5'-AGTGTAGAGGGTCAGCCTGAGTTA-3') and reverse DI COI-R1 (5'-ACAGGCACTGACAATACCAAT-3') and *D. repens* used forward primer Dr ITS2-F (5'-CATTGATAGTTA-CATTCAAATAA-3') and reverse Dr ITS2-R (5'-GATTCATTTATTGCATTA-AGCAAGC-3'). Each amplification reaction was done in a final volume of 50  $\mu$ l containing 32.6  $\mu$ l double-distilled water, 5  $\mu$ l 10 $\times$ Taq buffer, 3  $\mu$ l MgCl<sub>2</sub> (25 mM), 2  $\mu$ l dNTP (5 mM), 1  $\mu$ l forward primer (10 pmol/ $\mu$ l), 1  $\mu$ l reverse primer (10 pmol/ $\mu$ l), 0.4  $\mu$ l Taq polymerase (5 U/ $\mu$ l), and 5  $\mu$ l template (mosquito DNA extract). Total 35 amplification cycles with template denaturation at 94°C (30 sec), primer annealing at 55°C (30 sec), primer extension at 72°C (30 sec), and an extra 30-sec extension as the final cycle were routinely used.

**Multiplex PCR Assay**

Multiplex PCR was done for all mosquito pools by using QIAamp DNA Minikit (Qiagen), following the manufacturer’s instructions. PCR products were detected by agarose gel electrophoresis. DNA was visualized by an ultraviolet transilluminator following staining the gel with 2% ethidium bromide. A 100-bp ladder molecular weight marker (100 bp DNA ladder, Promega, Madison, Wisconsin, USA) and a negative control (non-blood fed mosquitoes, which were obtained from Research Institute of Medical Entomology, Dokki, Egypt) were used in each PCR reaction.

**Statistical analysis**

Data was collected and entered by using the Excel program, and the statistical analysis was done by using SPSS 20 program. The rate of infection in mosquitoes was adjusted for pooled samples, calculating the Estimated Rate of Infection (ERI) using the following formula:  $ERI = 1 - (1 - x/m)^{1/k}$  [20], where ‘x’ is the number of positive pools; ‘m’ is the number of examined pools, and ‘k’ the average number of specimens in each pool. Sensitivity, specificity, positive predictive value, negative predictive value, and the accuracy rate of multiplex PCR as compared to single PCR were calculated.

**RESULTS**

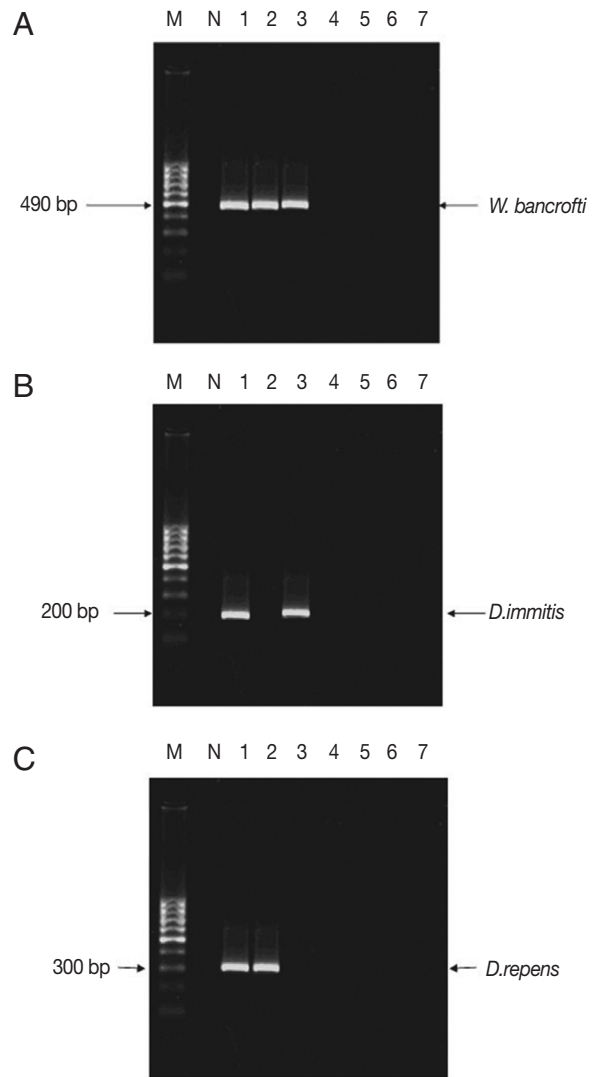
Mosquitoes collected from 7 localities in Assiut Governorate, Egypt were microscopically identified as *Culex pipiens molestus* (56%), *C. antennatus* (8%), *C. pusillus* (5%), *Anopheles*

**Table 1.** Comparison between the results of positive pools for filarial parasites in 100 collected mosquito pools detected by single and multiplex PCR

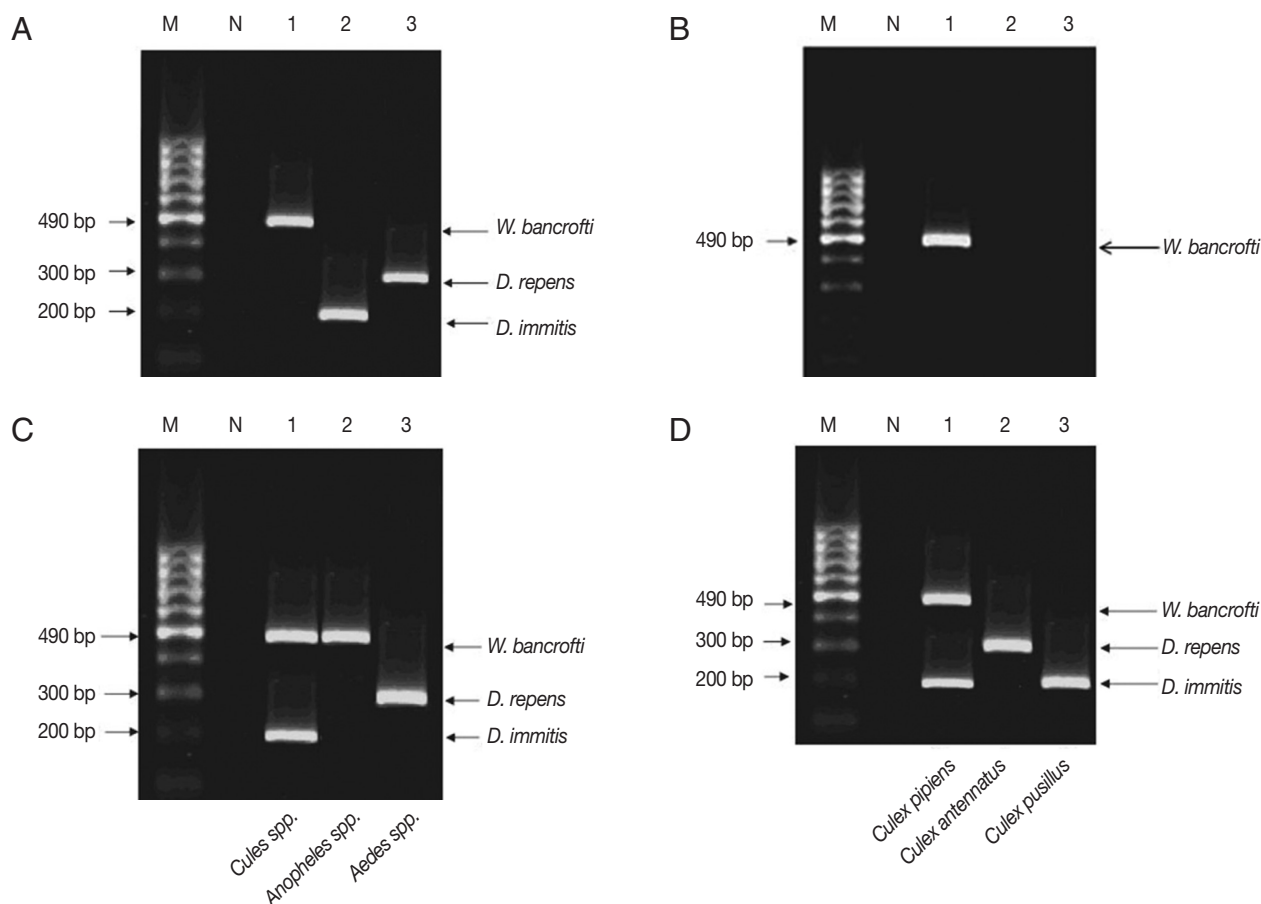
Filarial parasites	Locality	No. of positive pools by single PCR	No. of positive pools by multiplex PCR
<i>Wuchereria bancrofti</i>	El-Nikhila	3	4
	El-Matiaa	2	2
	Sahel Seleem	2	2
<i>Dirofilaria immitis</i>	El-Nikhila	2	2
	El-Matiaa	0	0
	Sahel Seleem	1	1
<i>Dirofilaria repens</i>	El-Nikhila	2	2
	El-Matiaa	1	1
	Sahel Seleem	0	0
<b>Total</b>		<b>13</b>	<b>14</b>

*pharoensis* (16%), and *Aedes caspius* (15%).

Preliminary, using single PCR technique could localize the presence of filarial nematodes only in 3 out of 7 localities. El-Nikhila was the only village where the 3 genera of filarial nematodes (*W. bancrofti*, 4%; *D. immitis*, 2%, and *D. repens*, 2%) coexist. Sahel Seleem district was harboring *W. bancrofti* (2%) and *D. immitis* (1%), while El-Matiaa village was harboring *W. bancrofti* (2%) and *D. repens* (1%). We could say that *W. bancrofti* is the most highly detected filarial nematode (Table 1;



**Fig. 2.** Distribution of filarial parasites in different Assiut localities using single PCR pattern. M, 100 bp DNA marker; N, negative control (non-blood fed mosquitoes); lane 1, El-Nikhila village; lane 2, El-Matiaa village; lane 3, Sahel Seleem district; lane 4, El-Badary district; lane 5, Mankabad village; lane 6, Dairout district; lane 7, Manfalout district. (A) *W. bancrofti* positive samples (lanes 1-3). (B) *D. immitis* positive samples (lanes 1, 2). (C) *D. repens* positive samples (lanes 1, 2).



**Fig. 3.** Multiplex PCR pattern on 2% ethidium bromide-stained agarose gel. Lane 1, *W. bancrofti*; lane 2, *D. immitis*; lane 3, *D. repens*. (A) Multiplex PCR confirmed the results of single PCR in El-Nikhila, El-Matkaa villages and Sahel Seleem district. (B) Multiplex PCR detected a positive sample of *W. Bancrofti* (lane 1) that could not be detected by single PCR in El-Nikhila village. (C) Distribution of filarial parasites among different mosquito genera using multiplex PCR. Lane 1, *Culex* spp.; lane 2, *Anopheles* spp.; lane 3, *Aedes* spp. (D) Distribution of filarial parasites among different *Culex* species in El-Nikhila village. Lane 1, *Culex pipiens*; lane 2, *Culex antennatus*; lane 3, *Culex pusillus*.

Fig. 2) with 0.33% ERI, while *D. immitis* and *D. repens* both had ERI of 0.12%. The overall estimated rate of infection of all collected mosquitoes with the 3 filarial parasites was 0.6%.

Afterwards, multiplex PCR had confirmed the single PCR results performed in the 3 localities (Fig. 3A), in fact multiplex PCR was superior to single PCR as it positively detected *W. bancrofti* in previously negative tested pool (Fig. 3B). The sensitivity of multiplex PCR as compared to single PCR was 100%, and the specificity was 98%. The positive predictive value was 93%, the negative predictive value was 100%, and the accuracy rate was 99% (Table 1).

Each pool of mosquito genera (*Culex*, *Anopheles*, and *Aedes*) was tested by multiplex PCR to know which filarial nematode is the vector in the 3 localities (Table 2; Fig. 3C). *Culex* spp. were found as the most common mosquito genus infected

with filarial nematodes in all localities, especially in El-Nikhila village, with the highest ERI of 0.47% (Table 2).

Since *Culex* spp. had the highest ERI among the 3 mosquito genera, and El-Nikhila village harbored the 3 filarial nematodes, multiplex PCR was applied to determine the species of *Culex* mosquitoes involved as the vector for these nematodes in El-Nikhila village. *C. pipiens* was the vector for all 3 filarial nematodes (ERI, 3.6%); *C. antennatus* was the vector for *D. repens* (ERI, 0.89%), whereas *C. pusillus* was the vector for *D. immitis* (ERI, 0.89%) (Table 3; Fig. 3D).

## DISCUSSION

*Culex* spp. were the most commonly collected mosquitoes (69%) throughout the period of the study. Southgate [21] re-

**Table 2.** Distribution of filarial parasites among mosquito genera performed by multiplex PCR in different Assiut localities and estimated rate of infection (ERI)

Locality	Mosquito vector	No. of Pools	<i>W. Bancrofti</i>		<i>D. immitis</i>		<i>D. repens</i>	
			No. of positive pools	ERI (%)	No. of positive pools	ERI (%)	No. of positive pools	ERI (%)
El Nikhila	<i>Culex</i> sp.	10	3	1.42	2	0.89	1	0.42
	<i>Anopheles</i> sp.	5	1	0.89	0	0.0	0	0.0
	<i>Aedes</i> sp.	5	0	0.0	0	0.0	1	0.89
El Matiaa	<i>Culex</i> sp.	10	2	0.89	0	0.0	0	0.0
	<i>Anopheles</i> sp.	5	0	0.0	0	0.0	1	0.89
	<i>Aedes</i> sp.	5	0	0.0	0	0.0	0	0.0
Sahel Seleem	<i>Culex</i> sp.	10	2	0.89	1	0.42	0	0.0
	<i>Anopheles</i> sp.	5	0	0.0	0	0.0	0	0.0
	<i>Aedes</i> sp.	5	0	0.0	0	0.0	0	0.0

**Table 3.** Distribution of filarial parasites among *Culex* spp. in El-Nikhila village performed by multiplex PCR and estimated rate of infection (ERI)

<i>Culex</i> spp.	No. of Pools	<i>W. Bancrofti</i>		<i>D. immitis</i>		<i>D. repens</i>	
		No. of positive pools	ERI (%)	No. of positive pools	ERI (%)	No. of positive pools	ERI (%)
<i>Culex pipiens</i>	5	2	2.0	1	0.89	0	0.0
<i>Culex antennatus</i>	5	0	0.0	0	0.0	1	0.89
<i>Culex pusillus</i>	5	0	0.0	1	0.89	0	0.0

ported that *C. pipiens* was widely distributed across Egypt; Khalil [22] found it as the most predominant mosquito species in Upper Egypt. In Assiut Governorate, Abdel-Aal [23], El-Nazer [24], and Mahmoud [25] found similar results.

El-Nikhila village had recorded the highest ERI (2%) with the 3 filarial parasites over El-Matiaa villages and Sahel Seleem district, and *Culex* spp. was the predominant vector. *Culex pipiens molestus* has the highest rate of infection (ERI, 2% and 0.89%) for *W. bancrofti* and *D. immitis*, respectively. Interestingly, the high *W. bancrofti* infection rate in *C. pipiens molestus* indicates the role of this species as a putative vector, as previously described [24-26]. Cancrini and Gabrielli [10] and Capelli et al. [27] added that *C. pipiens* is not only the most efficient natural vector for *W. bancrofti*, but also potential vectors of *D. immitis* and *D. repens*. It had to be in mind that Assiut Governorate villages had been enrolled as endemic foci in the international and national programs of elimination of lymphatic filariasis [2]. Although efforts were done all over the last years, the mass drug administration (MDA) program and intensive use of residual insecticides for mosquito control [28], vectors loaded with filarial parasites still exist. Abundance of vectors could be explained by environmental and biological features supporting vectors breeding in these localities. El-Nikhila, El-Matiaa villages, and Sahel Seleem district stretch along the banks of the Nile and have water canals traversing

their agricultural lands boosting the presence of stagnant water. Besides, poor quality of sanitation accompanied by inadequate disposal of humans and water waste lead to an alteration in water quality parameters with low dissolved oxygen, favorable temperature, and pH which may be contributing to successful *C. pipiens* mosquito breeding inside and/or near human habitations [29]. Moreover, this provided favorable conditions for the bacteria, alga, and protozoa, which are chief food sources for mosquito larvae [30]. In addition, dense vegetation covering throughout the periphery of water banks not only provides favorable oviposition sites but also protects mosquito larvae from predacious insects and fish [31].

Bahgat et al. [32] found *C. pipiens* to be primarily anthropophilic (96% human feeding) and to be fairly endophilic as most of the collected engorged females were from indoors. Appearance of insecticides "Chlorpyrifos" resistant strain among Assiut *C. pipiens* population might also decline the effort made in the elimination program and enhance the spread of vector-borne disease [33]. Previous documented factors in addition to earlier history of filariasis endemicity helped in making these localities foci for filarial parasites.

An overall ERI in mosquitoes was 0.6%, being higher in *Culex* spp. and *Anopheles pharoensis* (0.47% and 0.08%), respectively. Mosquito infection rate with filarial parasites in our study was higher than in previous studies performed in the

same localities [24-26], as they were based on mosquito dissection only. Therefore, we used the multiplex PCR. Multiplex PCR confirmed the results given by single PCR done for each parasite. Moreover, multiplex PCR was more accurate than single PCR in detection of filarial infections in mosquitoes with 100% sensitivity and 98% specificity. This may be explained by the combined use of Hot Start Taq DNA polymerase with a unique PCR buffer specifically developed for multiplex PCR reactions. The newly developed reaction buffer containing factor MP and a special multiplex PCR-enhancing synthetic factor that provides an efficient primer annealing and extension. This synthetic factor increases the local concentration of primers at the template DNA and stabilizes specifically bound primers [13]. This makes multiplex PCR as a good tool for simultaneous detection of *W. bancrofti*, *D. immitis*, and *D. repens* in mosquitoes. Similar results were previously reported by Latrofa et al. [15] in a study performed in Italy in which a duplex real-time PCR was used for the detection of and differentiation between *D. immitis* and *D. repens* in mosquitoes.

Data herein presented clearly indicate the usefulness of the molecular diagnosis assay for a broader field application in order to gain more information on the epidemiology of mosquitoes acting as vectors of filarial parasites. A continuous monitoring of infection rates for mosquito vector populations is pivotal to evaluate the success of an anti-filarial campaign in endemic areas. The advantages of using a highly sensitive molecular diagnostic tool for xenomonitoring was shown to be pivotal for assessing the efficacy and progress of eradication programs of human lymphatic filariasis based on MDA [34].

In conclusion, results presented submit 100% sensitivity and 98% specificity of multiplex PCR herein used in field-collected samples, clearly representing an alternative to classic microscopic methods and to single PCR-based assay for xenomonitoring of *W. bancrofti*, *D. immitis*, and *D. repens* in mosquitoes, mainly in areas where these species are endemic and/or occur sympatrically. Multiplex PCR could differentiate between *Dirofilaria* species in the same reaction, in addition to being easier and time and effort saving, and was more sensitive than single PCR in detection of mosquito filarial infection. We have reached that El-Nikhila, El-Matiaa villages, and Sahel Seleem district in Assiut Governorate, Egypt are still endemic areas of filarial parasites. More attention of health workers is recommended for control and elimination of filarial diseases in these areas.

## CONFLICT OF INTEREST

We have no conflict of interest related to this work.

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