



# Intramuscular Single-dose Toxicity Test of *Bufonis venonum* Pharmacopuncture in Sprague-Dawley Rats

Kwang-Ho Lee<sup>1</sup>, Seung-Ho Sun<sup>2</sup>, Jun-Sang Yu<sup>3</sup>, Ki-Rok Kwon<sup>4\*</sup>

<sup>1</sup> Department of Acupuncture & Moxibustion Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>2</sup> Department of Internal Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>3</sup> Department of Sasang Constitutional Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>4</sup>Research Center of the Korean Pharmacopuncture Institute, Seoul, Korea

#### **Key Words**

*Bufonis venonum*, chan-su, pharmacopuncture, toxicity test

#### Abstract

**Objectives:** *Bufonis venonum* (BV) is the dried white secretions of the auricular and skin glands of the toads Bufo bufo gargarizans or Bufo melanosticus Schneider. This study was performed to evaluate the toxicity of intramuscularly-administered *Bufonis venonum* pharmacopuncture (BVP) and to calculate its approximate lethality through a single-dose test with Sprague-Dawley (SD) rats.

**Methods:** Twenty male and 20 female 6-week-old SD rats were injected intramuscularly with BVP or normal saline. The animals were divided into four groups with five female and five male rats per group: the control group injected with normal saline at 0.5 mL/animal, the low-dosage group injected with 0.125 mL/animal of BVP, the medium-dosage group injected with 0.25 mL/animal of BVP and the high-dosage group injected with 0.5 mL/animal of BVP. All injections were in the left thighs of the rats. After administration, we conducted clinical observations everyday and body weight measurements on days 3, 7 and 14 after the injection. We also carried out hematology, serum biochemistry, and histological observations on day 15 after treatment.

**Results:** No mortalities were observed in any experimental group. No significant changes in weight, hema-

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tology, serum biochemistry, and histological observations that could be attributed to the intramuscular injection of BVP were observed in any experimental group.

**Conclusion:** Lethal dose of BVP administered via intramuscular injection in SD rats is over 0.5 mL/animal.

# 1. Introduction

*Bufonis venonum* (BV), called Chan-Su in Chinese or Sum-So in Korean, is toad venom; in particular, it is the dried white secretions of the auricular and skin glands of the toads Bufo bufo gargarizans or Bufo melanosticus Schneider [1-3]. BV has detoxification, anti-inflammatory, cardiotonic, and pain-relief effects [1, 4], and studies on BV have reported local anesthetic actions and anti-cancer effects [5-8].

*Bufonis venonum* pharmacopuncture (BVP) is a pharmacopuncture that is produced by using various substances extracted from the toad venom. Choi *et al* [9] reported recently that BVP had therapeutic potential for treating neuropsychiatric disorders such as anxiety or depression disorder, but no side effects or toxicity of BVP have been reported so far. For that reason, we conducted an intramuscular single-dose toxicity test of BVP in Sprague-Dawley (SD) rats to determine the safety of its use safe and to estimating its appropriate dosage.

# 2. Materials and Methods

\*Corresponding Author

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Kirok Kwon. Korean Pharmacopuncture Institute, 4F, Association of Korean Oriental Medicine B/D, 26-27, Gayang-dong, Gangseo-gu, Seoul 157-200, Korea. Tel: +82-33-744-9304 Fax: +82-33-744-9305 E-mail: drkwon5031 (@daum.net

Twenty-four SD rats of each gender were obtained from Orientbio Inc. (Gyeong-gi, Korea) at 5 weeks of age and were used after a week of quarantine and acclimatization. The animals were housed in a room maintained at  $21.1 - 24.1^{\circ}$ C under a relative humidity of 40.7% - 64.5%. The room was illuminated with artificial lighting from 07:00 to 19:00 hours and had 10 - 15 air changes per hour. Three animals per cage were housed in suspended stainless-steel wire-mesh cages and were allowed sterilized tap water and commercial rodent chow (Teklad Certified Irradiated Global 18% Protein Rodent Diet 2918C, Harlan Laboratories, Inc., U.S.A.). This study protocol was approved by the institutional Animal Care Committee of Biotoxtech Co. (Oh Chang, Korea).

The BVP (Lot No. KPI-2013-01) was manufactured in a pathogen-free facility at the Korean Pharmacopuncture Institute, Seoul, Korea by using BV purchased from Shandong, China. Then, the pharmacopuncture with a concentration of 0.1 mg BV/mL was filtered through 0.1-µm filter paper. Finally, the BVP was sterilized before being used for this experiment.

Twenty healthy male and 20 healthy female SD rats were selected from among the original 48 SD rats and were assigned to 1 of 4 groups with five male and five female SD rats per group: control (normal saline at 0.5 mL/animal), low-dosage (BVP at 0.125 mL/animal), medium-dosage (BVP at 0.25 mL/animal) and high-dosage (BVP at 0.5 mL/animal) groups. BVP or normal saline (Lot No. 12133, Choongwae Pharma Corp., Korea) was administered to the rats by intramuscular injection in the left thigh.

All animals were observed for clinical signs at 30 minutes, 1 hour and 2 hours immediately after the injection and once a day, starting the day after injection, for 14 days. The body weight of each rat was measured at the beginning of treatment and at 3 days, 7 days and 14 days after the injection.

On day 15 after treatment, the animals were fasted for 18 hours prior to necropsy and blood collection. Blood samples were drawn from the abdominal aorta under isoflurane anesthesia by using a syringe needle. Blood samples were collected into tubes containing ethylenediamine-tetraacetic acid (EDTA) and were analyzed to determine the red blood cell count (RBC), hemoglobin concentration (Hb), hematocrits (Ht), mean corpuscular cell volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular cell hemoglobin concentration (MCC), platelet count, white blood cell count (WBC), differential WBC count, reticulocyte (Reti) count, prothrombin time (PT) and active partial thromboplastin time (APTT) by using Hematology Systems (ADVIA 2120i, Siemens, Munich, Germany).

For the serum biochemistry analysis, blood samples were centrifuged at 3,000 rpm for 10 minutes and analyzed by using an auto-analyzer (7180, Hitachi, Tokyo, Japan). Serum biochemistry parameters, including alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), gamma glutamyl transpeptidase (GGT), blood urea nitrogen (BUN), creatinine, total bilirubin, total protein (TP), albumin, albumin/globulin ratio (A/G ratio), total cholesterol (T-Chol), triglycerides (TG), phosphate (P), glucose (Glu), calcium (Ca), chloride (Cl), sodium (Na) and potassium (K), were examined.

The tissues from the left thighs of the rats were routinely processed, embedded in paraffin and sectioned into 3- to  $5-\mu m$  pieces. The sections were stained with hematoxylin and eosin (H&E) for microscopic examination. All tissues taken from all animals were examined microscopically.

Data on animal weights and on their blood chemistry and hematology were tested by using a statistical analysis system (SAS, version 9.3, SAS Institute, Inc., Cary, NC, U.S.A.). The variance in the numerical data was checked by using the Bartlett test. If the variance was homogeneous, the data were subjected to a one-way analysis of variance (ANOVA). If either of the tests showed a significant difference among the groups, the data were analyzed using the multiple comparison procedure of the Dunnett test. If not, they were analyzed using the Kruskal-Wallis non-parametric ANOVA test (P < 0.05).

#### 3. Results

No treatment-related mortalities, clinical signs or weight changes occurred in either the control animals or the animals treated with any dose of BVP during the observation period (Tables 1, 2). On the hematological examination (Table 3), one female in the medium-dosage group showed a significant change; however, the change was not dose-dependent; the change seemed to have occurred sporadically. The blood chemistry tests (Table 4) showed no significant changes.

The necropsy examinations (Tables 5, 6) showed no abnormalities. Moreover, on the histopathological examination, one male in the control group and one female in the low-dosage group showed abnormal changes, but those changes were not dose-dependent. Thus, they were deemed not be important toxicological changes.

# 4. Discussion

BV has some toxic ingredients that can induce serious effects, including bradycardia, atrioventricular conduction block, ventricular tachycardia, ventricular fibrillation, and sudden death [10]. Bufadienolides, such as bufalin, cinobufagin and resibufogenin, which are major sources of BV, are known to increase vasoconstriction, vascular resistance and blood pressure probably by inhibiting Na, K-adenosine triphosphate (ATP) ase activity [11, 12], and these substances have recently been reported to have a strong surface anesthetic activity, cytotoxic effect and differentiation-apoptosis activity on murine leukemia human acute promyeocytic leukemia (HL-60) cells [13]. In addition, BV includes bufotenine, an indole alkaloid that produces effects such as aphrodisia and hallucination, and serotonin, which is involved in various psychiatric disorders such as depression, anxiety, chronic obsession syndrome and impulsivity [14-16]. Thus, appropriate dosage and careful use are very important for the safe use of BV [2].

For the above reasons, we conducted an intramuscular

Corr	Group / Dose	No. of		Days after dosing												Montolity		
Sex	(mL/animal)	animals	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	Mortality
	G1(0)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
Male	G2 (0.125)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
Male	G3 (0.25)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
	G4 (0.5)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
	G1 (0)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
Female	G2 (0.125)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
Female	G3 (0.25)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5
	G4 (0.5)	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/5

### Table 1 Summary of mortalities

### Table 2 Mean body weights

	Group / Dose	Mean		Days afte	er dosing		Gain (g)
Sex	(mL/animal)	S.D. N	0	3	7	14	0 — 14
	G1	Mean	191.5	219.1	255.3	315.4	123.9
	(0)	S.D.	4.8	6.7	10.3	17.9	13.6
	(0)	Ν	5	5	5	5	5
	G2	Mean	190.0	217.9	256.1	314.7	124.7
	(0.125)	S.D.	5.5	5.4	9.5	15.3	10.6
Male	(0.125)	Ν	5	5	5	5	5
wide	G3	Mean	188.8	214.0	250.2	309.6	120.8
	(0.25)	S.D.	3.0	4.3	6.3	9.0	7.1
	(0.23)	Ν	5	5	5	5	5
	G4	Mean	189.9	216.0	254.7	318.5	128.5
	(0.5)	S.D.	6.7	9.1	11.5	16.9	11.2
	(0.5)	Ν	5	5	5	5	5
	G1	Mean	157.3	173.0	192.0	220.5	63.2
	(0)	S.D.	8.3	11.7	15.5	18.8	12.0
	(0)	Ν	5	5	5	5	5
	G2	Mean	159.1	175.3	192.4	218.0	58.9
	(0.125)	S.D.	1.8	3.2	7.5	9.6	11.2
Female	(0.125)	Ν	5	5	5	5	5
remale	G3	Mean	157.0	173.3	192.7	220.6	63.6
		S.D.	8.5	7.8	8.5	12.5	4.5
	(0.25)	Ν	5	5	5	5	5
	G4	Mean	157.0	172.1	187.1	212.8	55.8
		S.D.	7.4	8.1	10.9	14.9	9.2
	(0.5)	Ν	5	5	5	5	5

S.D., standard deviation; N, number of animals.

# $\textbf{Table 3} \ \textbf{Mean hematology parameters}$

(wiale)
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Group/ Dose	Mean	RBC	LICD	LICT	]	RBC Indice			D-t:
(mL/animal)	S.D.	$(\times 10^{6}$	HGB (g/dL)	HCT (%)	MCV	MCH	MCHC	$(\times 10^{3})$	Reti (%)
(IIIL/ allillal)	Ν	cells/µL)	(g/uL)	(70)	(fL)	(pg)	(g/dL)	cells/ $\mu$ L)	(70)
G1	Mean	7.23	14.8	42.4	58.6	20.5	34.9	1235	4.52
(0)	S.D.	0.10	0.4	1.2	1.9	0.7	0.9	138	0.82
(0)	Ν	5	5	5	5	5	5	5	5
G2	Mean	7.56	15.3	44.2	58.4	20.2	34.5	1215	3.96
(0.125)	S.D.	0.25	0.4	1.7	1.2	0.4	0.5	92	0.58
(0.123)	Ν	5	5	5	5	5	5	5	5
G3	Mean	7.32	14.8	42.5	58.0	20.2	34.8	975	3.97
(0.25)	S.D.	0.30	0.6	1.7	0.8	0.4	0.6	239	0.43
(0.23)	Ν	5	5	5	5	5	5	5	5
G4	Mean	7.34	14.9	43.4	59.2	20.3	34.3	1083	4.82
(0.5)	S.D.	0.27	0.5	1.6	1.7	0.7	0.4	142	0.78
(0.5)	Ν	5	5	5	5	5	5	5	5
	Mean WBC		MDC D:f	formantial Car	unting (07)				
Group/ Dose	Mean	WBC $(\times 10^3)$		WBC Diff	ferential Cou	unting (%)		PT	APTT
Group/ Dose (mL/animal)	Mean S.D. N	WBC (× 10 <sup>3</sup> cells/µL)	NEU	WBC Diff	ferential Cou MONO	unting (%) EOS	BASO	PT (sec)	APTT (sec)
(mL/animal)	S.D.	$(\times 10^{3})$	NEU 18.7			<b>U</b>	BASO 0.1		
(mL/animal) G1	S.D. N	$(\times 10^3$ cells/µL)		LYM	MONO	EOS		(sec)	(sec)
(mL/animal)	S.D. N Mean	(× 10 <sup>3</sup> cells/μL) 8.29	18.7	LYM 78.4	MONO 1.4	EOS 0.6	0.1	(sec) 16.6	(sec) 12.9
(mL/animal) G1 (0)	S.D. N Mean S.D.	(× 10 <sup>3</sup> cells/µL) 8.29 2.35	18.7 4.2	LYM 78.4 3.9	MONO 1.4 0.3	EOS 0.6 0.2	0.1 0.1	(sec) 16.6 0.9	(sec) 12.9 2.6
(mL/animal) G1 (0) G2	S.D. N Mean S.D. N	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28	18.7 4.2 5	LYM 78.4 3.9 5	MONO 1.4 0.3 5	EOS 0.6 0.2 5	0.1 0.1 5	(sec) 16.6 0.9 5	(sec) 12.9 2.6 5
(mL/animal) G1 (0)	S.D. N Mean S.D. N Mean	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71	18.7 4.2 5 14.7	LYM 78.4 3.9 5 82.3	MONO 1.4 0.3 5 1.8	EOS 0.6 0.2 5 0.5	0.1 0.1 5 0.2	(sec) 16.6 0.9 5 17.6	(sec) 12.9 2.6 5 14.0
(mL/animal) G1 (0) G2 (0.125)	S.D. N Mean S.D. N Mean S.D. S.D. N Mean	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28	18.7 4.2 5 14.7 1.9	LYM 78.4 3.9 5 82.3 2.0	MONO 1.4 0.3 5 1.8 0.5	EOS 0.6 0.2 5 0.5 0.2	0.1 0.1 5 0.2 0.1	(sec) 16.6 0.9 5 17.6 1.2	(sec) 12.9 2.6 5 14.0 2.0
(mL/animal) G1 (0) G2 (0.125) G3	S.D. N Mean S.D. N Mean S.D. S.D. N	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28 5 7.62 2.28	18.7 4.2 5 14.7 1.9 5	LYM 78.4 3.9 5 82.3 2.0 5 79.8 6.7	MONO 1.4 0.3 5 1.8 0.5 5	EOS 0.6 0.2 5 0.5 0.5 0.2 5	0.1 0.1 5 0.2 0.1 5	(sec) 16.6 0.9 5 17.6 1.2 5	(sec) 12.9 2.6 5 14.0 2.0 5
(mL/animal) G1 (0) G2 (0.125)	S.D. N Mean S.D. N Mean S.D. S.D. N Mean	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28 5 5 7.62	18.7 4.2 5 14.7 1.9 5 17.4	LYM 78.4 3.9 5 82.3 2.0 5 79.8	MONO 1.4 0.3 5 1.8 0.5 5 1.6	EOS 0.6 0.2 5 0.5 0.2 5 0.2 5 0.4	0.1 0.1 5 0.2 0.1 5 0.2	(sec) 16.6 0.9 5 17.6 1.2 5 16.8	(sec) 12.9 2.6 5 14.0 2.0 5 13.8
(mL/animal) G1 (0) G2 (0.125) G3 (0.25)	S.D. N Mean S.D. N Mean S.D. N Mean S.D.	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28 5 7.62 2.28	18.7 4.2 5 14.7 1.9 5 17.4 6.9	LYM 78.4 3.9 5 82.3 2.0 5 79.8 6.7	MONO 1.4 0.3 5 1.8 0.5 5 1.6 0.4	EOS 0.6 0.2 5 0.5 0.2 5 0.2 5 0.4 0.2	0.1 0.1 5 0.2 0.1 5 0.2 0.1	(sec) 16.6 0.9 5 17.6 1.2 5 16.8 0.7	(sec) 12.9 2.6 5 14.0 2.0 5 13.8 1.9
(mL/animal) G1 (0) G2 (0.125) G3	S.D. N Mean S.D. N Mean S.D. N Mean S.D. N	(× 10 <sup>3</sup> cells/μL) 8.29 2.35 5 7.71 1.28 5 7.62 2.28 5	$     18.7 \\     4.2 \\     5 \\     14.7 \\     1.9 \\     5 \\     17.4 \\     6.9 \\     5 \\     5 $	LYM 78.4 3.9 5 82.3 2.0 5 79.8 6.7 5	MONO 1.4 0.3 5 1.8 0.5 5 1.6 0.4 5	EOS 0.6 0.2 5 0.5 0.2 5 0.4 0.2 5 5	0.1 0.1 5 0.2 0.1 5 0.2 0.1 5	(sec) 16.6 0.9 5 17.6 1.2 5 16.8 0.7 5	(sec) 12.9 2.6 5 14.0 2.0 5 13.8 1.9 5

(Female)

Group/ Dose	Mean	RBC	HGB	НСТ		RBC Indice	es	PLT	Reti
(mL/animal)	S.D.	$(\times 10^{6})$	(g/dL)	(%)	MCV	MCH	MCHC	$(\times 10^{3})$	(%)
(IIIL/ aIIIIIai)	Ν	cells/ $\mu$ L)	(g/uL)	(70)	(fL)	(pg)	(g/dL)	$cells/\mu L)$	(70)
Cl	Mean	7.72	15.5	42.8	55.4	20.0	36.1	1125	2.55
G1	S.D.	0.16	0.5	1.5	1.5	0.4	0.2	182	0.14
(0)	Ν	5	5	5	5	5	5	5	5
60	Mean	7.63	15.4	42.9	56.3	20.2	35.9	1027	2.25
G2 (0.125)	S.D.	0.25	0.4	1.4	1.9	0.6	0.5	104	0.50
(0.125)	Ν	5	5	5	5	5	5	5	5

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<u></u>	Mean	7.22	14.6*	40.6	56.3	20.2	35.9	1044	2.47
G3 (0.25)	S.D.	0.36	0.7	1.8	2.1	0.9	0.4	327	0.28
(0.23)	Ν	5	5	5	5	5	5	5	5
C 4	Mean	7.65	15.3	42.6	55.7	20.0	35.9	1078	2.03
G4 (0.5)	S.D.	0.34	0.4	1.0	2.1	1.0	0.4	90	0.37
(0.5)	Ν	5	5	5	5	5	5	5	5

Group/ Dose	Mean	WBC		WBC Diff	ferential Cou	nting (%)		PT	APTT
(mL/animal)	S.D. N	(× 10³ cells/µL)	NEU	LYM	MONO	EOS	BASO	(sec)	(sec)
G1	Mean	5.24	15.3	80.9	1.6	1.6	0.1	18.4	14.8
(0)	S.D.	1.25	5.7	6.8	0.8	0.4	0.1	0.3	1.4
	Ν	5	5	5	5	5	5	5	5
G2	Mean	4.83	10.7	85.8	1.5	1.0	0.2	17.8	13.4
(0.125)	S.D.	1.05	1.7	1.2	0.5	0.2	0.1	0.3	1.8
	Ν	5	5	5	5	5	5	5	5
G3	Mean	6.08	12.9	83.8	1.3	1.1	0.2	17.7	14.7
(0.25)	S.D.	3.63	6.0	6.5	0.4	0.5	0.1	1.2	0.6
	Ν	5	5	5	5	5	5	5	5
G4	Mean	6.05	12.1	84.4	1.5	1.1	0.2	17.8	15.3
(0.5)	S.D.	2.40	1.9	2.2	0.3	0.3	0.0	0.9	1.5
	Ν	5	5	5	5	5	5	5	5

Significantly different from control by Dunnett's *t*-test: P < 0.05.

S.D., standard deviation; N, number of animals; RBC, red blood cell; HGB, hemoglobin; HCT, hematocrit; MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; PLT, platelet; Reti, reticulocytes; WBC, white blood cell; NEU, neutrophils; LYM, lymphocytes; MONO, monocytes; EOS, Eosinophils; BASO, basophils; PT, prothrombin time; APTT, activated partial thromboplastin time.

Table 4 Mean clinical chemistry

(Male)

Glu BUN T-Chol Mean Crea T-Bili Group/ Dose ALT AST ALP GGT S.D. (mg/ (mg/ (mg/ (mg/ (mg/ (mL/animal) (U/L)(U/L)(U/L)(U/L)Ν dL) dL) dL) dL) dL) Mean 30.0 92.1 937.3 0.41 113 12.2 0.41 0.03 70 G1 S.D. 2.2 12.2 176.7 0.08 15 1.1 0.01 0.03 6 (0)Ν 5 5 5 5 5 5 5 5 5 Mean 32.1 107.3 851.6 0.39 116 11.8 0.39 0.02 66 G2 174.7 S.D. 4.222.0 0.05 12 1.0 0.02 0.01 15 (0.125)Ν 5 5 5 5 5 5 5 5 5 Mean 32.3 105.6 917.2 0.41 109 12.8 0.40 0.01 68 G3 S.D. 4.522.3 117.0 0.14 18 2.3 0.04 0.01 19 (0.25)Ν 5 5 5 5 5 5 5 5 5 Mean 28.5 118.6 777.7 0.45 117 12.2 0.40 0.02 69 G4 18 S.D. 3.2 15.3 169.7 0.12 8 0.8 0.02 0.01 (0.5)Ν 5 5 5 5 5 5 5 5 5

(Continued)

Group/ Dose (mL/animal)	Mean S.D. N	TG (mg/ dL)	TP (g/dL)	Alb (g/dL)	A/G ratio	P (mg/ dL)	Ca (mg/ dL)	Na (mmol /L)	K (mmol /L)	Cl (mmol /L)
G1	Mean	37	5.4	2.3	0.76	8.70	9.7	140	4.8	105
(0)	S.D.	16	0.2	0.1	0.03	0.66	0.3	0	0.6	2
(0)	Ν	5	5	5	5	5	5	5	5	5
Ca	Mean	40	5.4	2.3	0.76	8.69	10.0	141	4.7	105
G2 (0.125)	S.D.	24	0.3	0.1	0.05	0.57	0.4	1	0.3	1
(0.125)	Ν	5	5	5	5	5	5	5	5	5
62	Mean	33	5.3	2.3	0.76	8.55	9.9	140	4.6	105
G3 (0.25)	S.D.	16	0.0	0.1	0.04	0.38	0.2	1	0.3	1
(0.25)	Ν	5	5	5	5	5	5	5	5	5
64	Mean	37	5.2	2.3	0.80	8.65	9.7	140	4.9	105
G4 (0.5)	S.D.	13	0.1	0.1	0.09	0.28	0.2	1	0.2	1
(0.5)	Ν	5	5	5	5	5	5	5	5	5

### (Female)

Group/ Dose (mL/animal)	Mean S.D.	ALT (U/L)	AST (U/L)	ALP (U/L)	GGT (U/L)	Glu (mg/	BUN (mg/	Crea (mg/	T-Bili (mg/	T-Chol (mg/
(IIIL/ aIIIIIaI)	Ν	(0/L)	(0/L)	(0/L)	(0/L)	dL)	dL)	dL)	dL)	dL)
61	Mean	26.8	90.3	572.4	0.65	116	13.2	0.42	0.01	77
G1	S.D.	1.6	10.1	196.5	0.21	3	1.0	0.02	0.01	13
(0)	Ν	5	5	5	5	5	5	5	5	5
<u></u>	Mean	24.1	90.3	456.9	0.52	117	13.7	0.41	0.01	76
G2	S.D.	3.7	20.5	90.9	0.22	7	1.5	0.03	0.01	10
(0.125)	Ν	5	5	5	5	5	5	5	5	5
	Mean	22.1	94.8	531.7	0.60	118	11.8	0.43	0.01	89
G3	S.D.	3.8	27.1	101.4	0.11	4	1.3	0.02	0.00	15
(0.25)	Ν	5	5	5	5	5	5	5	5	5
_	Mean	24.6	93.3	543.8	0.52	118	14.7	0.45	0.01	92
G4	S.D.	3.1	12.1	150.3	0.17	5	3.2	0.04	0.01	8
(0.5)	Ν	5	5	5	5	5	5	5	5	5
0 /D	Mean	TG	(III)	4.11		Р	Са	Na	K	Cl
Group / Dose	S.D.	(mg/	TP	Alb	A/G	(mg/	(mg/	(mmol	(mmol	(mmol
(mL/animal)	Ν	dL)	(g/dL)	(g/dL)	ratio	dL)	dL)	/L)	/L)	/L)
61	Mean	16	5.8	2.6	0.83	7.11	9.9	140	4.6	106
G1	S.D.	7	0.3	0.1	0.04	0.50	0.2	1	0.1	2
(0)	Ν	5	5	5	5	5	5	5	5	5
	Mean	13	5.8	2.7	0.87	7.49	10.0	140	4.5	106
G2	S.D.	3	0.2	0.1	0.04	0.32	0.1	1	0.3	2
(0.125)	Ν	5	5	5	5	5	5	5	5	5

(Continued)

	Mean	16	5.7	2.6	0.83	7.57	9.8	140	4.5	106
G3 (0.25)	S.D.	6	0.5	0.3	0.05	0.64	0.5	1	0.5	2
(0.23)	Ν	5	5	5	5	5	5	5	5	5
C 4	Mean	18	5.6	2.5	0.83	7.83	9.8	140	4.5	107
G4 (0.5)	S.D.	2	0.1	0.1	0.04	0.27	0.2	1	0.3	1
(0.5)	Ν	5	5	5	5	5	5	5	5	5

S.D., standard deviation; N, number of animals; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; GGT, gamma glutamyl transpeptidase; Glu, glucose; BUN, blood urea nitrogen; Crea, creatinine; T-Bili, total bilirubin; T-Chol, total cholesterol; TG, triglycerides; TP, total protein; Alb, albumin; A/G ratio, albumin/globulin ratio; P, phosphorus; Ca, calcium; Na, sodium; K, potassium; Cl, chloride.

#### Table 5 Summary of necropsy findings

Sex		Ma	ale		Female						
Group	G1	G2	G3	G4	G1	G2	G3	G4			
Dose (mL/animal)	0	0.125	0.25	0.5	0	0.125	0.25	0.5			
No. of animals	5	5	5	5	5	5	5	5			
Unremarkable findings	5	5	5	5	5	5	5	5			
No. of rats examined	5	5	5	5	5	5	5	5			

External surface and all organs in the body cavity were unremarkable.

#### Table 6 Summary of histopathological findings

	Sex			Ma	ale				Female		
Organ /	Group		G1	G2	G3	G4		G1	G2	G3	G4
Organ / Findings	Dose (mL/animal)		0	0.125	0.25	0.5		0	0.125	0.25	0.5
Fillulligs	No. of animals		5	5	5	5		5	5	5	5
Injection site	-Cell infiltration, inflammatory, focal	±	1	0	0	0	±	0	1	0	0
Injection site	-Cell infiltration, macrophages, focal						±	0	1	0	0

Grade- ±, minimal.

single-dose toxicity test of BVP in SD rats to determine an appropriate dosage for its safe use. The results showed no treatment-related abnormalities for any of the used doses of BVP. The dose used for the high-dosage group was 0.5 mL/animal, and no dangerous signs were observed. Thus, we may conclude that 0.5 mL/animal of BVP is a safe dose in both male and female SD rats.

# 5. Conclusion

This study showed that the lethal dose of BVP was over 0.5 mL/animal in both male and female SD rats.

# **Conflict of interest**

The authors declare that there are no conflict of interest.

## References

- 1. Chen KK, Kovarikova A. Pharmacology and toxicology of toad venom. J Pharm Sci. 1967;56(12):1535-41.
- Lee HJ, Koung FP, Kwon KR, Kang DI, Cohen L, Yang PY, *et al.* Comparative analysis of the *Bufonis venenum* by using TLC, HPLC, and LC-MS for different extraction methods. J Pharmacopuncture. 2012;15(4):52-65.

- 3. Xie JT, Maleckar SA, Yuan CS. Beneficial and adverse effects of toad venom, a traditional oriental medicine. Orient Pharm Exp Med. 2002;2(1):28-35.
- 4. Professors of herbology in colleges of oriental medicine. [Herbology]. Seoul: Yeonglimsa; 2006. p. 569-70. Korean.
- 5. Kim YH, Jeong SH, Kim JH, Choi JM, Ji JH, Kang JK, *et al.* [Pharmacological effects of extract of *Bufonis vene-num*]. Biomol Ther. 2001;9(1):51-4. Korean.
- Park TY, Park C, Yoon HJ, Choi YH, Ko WS. [Growth arrest by *Bufonis venenum* is associated with inhibition of Cdc2 and Cdc25C, and induction of p21WAF1/CIP1 in T24 human bladder carcinoma cells]. Korean J Orient Physiol Pathol. 2004;18(5):1449-55. Korean.
- Kang AM, Kim BR, Kim SU, Lim SW. [Screening of the Bufonis venenum on Hep G2 cells]. J Korean Orient Med. 2008;29(4):171-9. Korean.
- 8. Lee SH, Choi DY, Baek YH, Lee JD. [A bibliographic studies on the *Bufonis venenum* for clinical treatment: important to toxicity and processing]. J Korean Acupuncture & Moxibustion Soc. 2009;26(1):121-33. Koran.
- 9. Choi MJ, Kim KN, Lee JE, Suh JW, Kim SC, Kwon KR, *et al.* Effects of sumsu (*Bufonis venenum*) pharmacopuncture treatment on depression in mice. J Pharmacopuncture. 2014;17(2):27-33.
- 10. Gowda RM, Cohen RA, Khan IA. Toad venom poisoning: resemblance to digoxin toxicity and therapeutic implications. Heart. 2003;89(4):e14.
- Pamnani MB, Chen S, Bryant HJ, Schooley JF Jr, Eliades DC, Yuan CM, *et al*. Effects of three sodium-potassium adenosine triphosphatase inhibitors. Hypertension. 1991;18(3):316-24.
- 12. Bick RJ, Poindexter BJ, Sweney RR, Dasgupta A. Effects of chan su, a traditional Chinese medicine, on the calcium transients of isolated cardiomyocytes: cardiotoxicity due to more than Na, K-ATPase blocking. Life Sci. 2002;72(6):699-709.
- Kamano Y, Kotake A, Hashima H, Inoue M, Morita H, Takeya K, *et al.* Structure-cytotoxic activity relationship for the toad poison bufadienolides. Bioorg Med Chem. 1998;6(7):1103-15.
- Barry TL, Petzinger G, Zito SW. GC/MS comparison of the west indian aphrodisiac "love stone" to the Chinese medication "chan su": bufotenine and related bufodienolides. J Forensic Sci. 1996;41(6):1068-73.
- 15. Fuller RW, Snoddy HD, Perry KW. Tissue distribution, metabolism and effects of bufotenine admistered to rats. Neuropharmacology. 1995;34(7):799-804.
- Gauthier C, Hassler C, Mattar L, Launay JM, Callebert J, Steiger H, *et al.* Symptoms of depression and anxiety in anorexia nervosa: links with plasma tryptophan and serotonin metabolism. Psychoneuroendocrinology. 20 14;39:170-8.