Synthesis and Potent Anti-leukemic Activity of Novel 5'-Norcarbocyclic C-nucleoside Phosphonic Acids

Seyeon Kim, Eunae Kim, Chang-Hyun Oh,[†] Kyung Ho Yoo,[‡] and Joon Hee Hong^{*}

BK21-Project Team, College of Pharmacy, Chosun University, Kwangju 501-759, Korea. *E-mail: hongjh@chosun.ac.kr [†]Center for Biomaterials, Korea Institute of Science and Technology, Seoul 130-650, Korea [‡]Chemical Kinomics Research Center, Korea Institute of Science and Technology, Seoul 130-650, Korea Received July 28, 2014, Accepted August 14, 2014

The first synthetic route to 5'-norcarbocyclic *C*-nucleoside [7-oxa-7,9-dideazadenosine (furo[3,2-*d*]pyrimidine) and 9-deazadenosine (pyrrolo[3,2-*d*]pyrimidine)] phosphonic acids from commercially available 1,3-dihydroxy cyclopentane was described. The key C-C bond formation from sugar to base precursor was performed using Knoevenagel-type condensation from a ketone derivative. Synthesized *C*-nucleoside phosphonic acids were tested for anti-HIV activity as well as anti-leukemic activity. Compound **26** showed significant anti-leukemic activity.

Key Words : Antiviral agent, Anti-leukemic agent, C-nucleoside, Knoevenagel reaction

Introduction

C-Nucleosides¹ such as pseudoisocytidine,² tiazofurin,³ selenazofurin,⁴ 9-deazaadenosine (1),⁵ and 4-amino-8- β -D-ribofuranosylpyrazolo[1,5-*a*]-1,3,5-triazine (2, D-APTR)⁶ have received considerable attention because of their chemical stability and anti-leukemic activity. The carbocyclic *C*-nucleoside⁷ is a unique class of nucleosides in which the heterocycle is connected to a sugar moiety by a C-C bond instead of the C-N bond of the natural nucleosides. Recently, Schneller *et al.* reported a novel synthetic route of carbocyclic *4*'-*epi*-formycin (3) *via* a procedure based on an asymmetric aldol/ring-closing metathesis strategy.⁸

Carbocyclic nucleosides⁹ are a group of compounds that are structurally similar to natural nucleosides in which the furanose oxygen is replaced by a methylene group. Replacement of the furanose ring oxygen by carbon is of particular interest because the resulting carbocyclic nucleosides possess greater metabolic stability to phosphorylase,¹⁰ which cleaves the glycosidic bond of nucleosides. The recent discovery of cpAP (**4**)¹¹ as an anti-HIV agent gives strong impetus to the search for novel nucleosides in this class of compounds (Figure 1).

A nucleoside 5'-phosphate is essentially a nucleoside monophosphate analogue.¹² However, the phosphonate has certain advantages over its phosphate counterpart because it is metabolically stable owing to its phosphorus-carbon bond, which is not susceptible to hydrolytic cleavage.¹³ The spatial location of the oxygen atom, particularly the β -position, with regard to the phosphorus atom in the nucleoside analogue plays a critical role in its antiviral activity. Increased antiviral activity conferred by this oxygen atom may be attributed to increased binding capacity of the phosphonate analogs to target enzymes.¹⁴ More importantly, the presence of a 5'-phosphonate allows the first phosphorylation step required



Figure 1. Design rationale of 5'-norcarbocyclic *C*-nucleoside phosphonic acid analogs

for nucleoside activation to be skipped, thereby bypassing this inefficient and often rate-limiting step in the conversion to 5'-triphosphate. This is frequently a limiting step in the phosphorylation sequence that ultimately leads to triphosphates.¹⁵ Like a nucleoside monophosphate, a nucleoside phosphonate can be further phosphorylated by cellular nucleotide kinases.¹⁶ The concept of nucleoside phosphonate has been applied to design chain terminators for anti-HIV chemotherapy and proved to be valid.

On the basis of the advantageous chemical stability and clinically significant biological properties of *C*-nucleosides and carbocyclic nucleoside phosphonic acids, we aimed to synthesize hybrid nucleosides in the form of 5'-norcarbocyclic *C*-nucleoside phosphonic acids. Herein, we report the synthesis of novel 5'-norcarbocyclic furo and pyrrolo[3,2-*d*]pyrimidine nucleoside phosphonic acids and their biological evaluation.

Novel 5'-Norcarbocyclic C-nucleoside analogs



periodinane, CH₂Cl₂; iii) NCCH₂CO₂Et, KOt-Bu, EtOH; iv) H₂, 10% Pd/C, MeOH; v) Dibal-H, ether.

Scheme 1. Synthesis of cyclopentyl enol intermediate 10.

Results and Discussion

For the synthesis of target carbocyclic *C*-nucleoside phosphonic acids, we utilized commercially available 1,3-dihydroxy cyclopropane **5** as a starting material (Scheme 1). Selective monosilylation of diol **5** produced cyclopentanol **6**, which was oxidized to the ketone **7** using Dess–Martin conditions.¹⁷ The cyclopentanone **7** was subjected to Knoevenagel-type condensation¹⁸ with ethyl cyanoacetate and potassium *tert*-butoxide in EtOH to give cyclopentylidene intermediate **8** with a 65% yield. The selective catalytic hydrogenation of **8** using 10% Pd/C under hydrogen produced compound **9** as a stereoisomeric mixture.

The intermediate **9** was reduced to the enol **10** by diisobutylaluminium hydride, which was *O*-alkylated with chloroacetonitrile using cesium carbonate to smoothly produce the enol ether **11** with a 52% two-step yield. Nitrile anion cyclization of **11** was performed with a 4-fold excess of LDA in anhydrous tetrahydrofuran at -70 °C to produce furan analogs **12** α (19%) and **12** β (21%).¹⁹ A complete nuclear Overhauser effect (NOE) NMR study enabled clear determination of the relative stereochemistry of **12** α and **12** β (Figure 2). For compound **12** β , the spectrum showed a strong NOE (1.5%) corresponding to H-1' \leftrightarrow H-4' coupling. On the basis of this finding, we concluded that the 4'-OTBDMS and the 1'-furan base of **12** β were located on the b face. In contrast, the spectra of **12** α showed only a weak H-1' \leftrightarrow H-4' NOE (1.0%); hence, it was defined as the 1,4'-*trans* isomer. The



Figure 2. NOE differences between the proximal hydrogens of 12α and 12β .

Bull. Korean Chem. Soc. 2014, Vol. 35, No. 12 3503



Reagents: i) CICH₂CN, Cs₂CO₃, DMF; ii) LDA, -70 °C, THF; iii) formamidine acetate, EtOH; iv) BzCl, pyridine; v) TBAF, THF; vi) (EtO)₂POCH₂OTf, LiO-*t*-Bu, THF; vii) NH₃, MeOH, rt; viii) TMSBr, 2,6-lutidine, CH₃CN.

Scheme 2. Synthesis of 5'-norcarbocyclic 7-oxa-7,9-dideazaadenosine phosphonic acid.

aminonitrile 12β is a versatile intermediate for the synthesis of a number of furo[3,2-d]pyrimidine C-nucleosides. Treatment of 12β with formamidine acetate (HN=CHNH₂·HOAc) in refluxing ethanol provided 13 with a 69% yield. For the synthesis of desired nucleoside phosphonic acid, benzoylation of 13 with benzoyl chloride in dry pyridine yielded the di-N-benzoyl derivative 14 in an 85% yield.²⁰ The removal of the silvl protecting group of compound 14 by using tetra-n-butylammonium fluoride (TBAF) afforded the 5'-nornucleoside analogue 15, which was treated with diethylphosphonomethyl triflate²¹ using lithium *t*-butoxide to yield the nucleoside phosphonate analogue 16. The selective deprotection of the base moiety of 16 was achieved by using saturated methanolic ammonia at 25 °C to afford the corresponding 5'-nornucleoside phosphonate derivative 17. Hydrolysis of 17 by treatment with bromotrimethylsilane in CH₃CN in the presence of 2,6-lutidine gave a desired 5'norcarbocyclic furo[3,2-d]pyrimidine nucleoside phosphonic acid 18 (Scheme 2).22

For the synthesis of 5'-norcarbocyclic pyrrolo[3,2-d]pyrimidine nucleoside phosphonic acid, the common enol intermediate **10** was treated with aminoacetonitrile monosulfate (H₂NCH₂CN·H₂SO₄) in MeOH to produce enamine inter-



Reagents: i) aminoacetonitrile monosulfate, NaOAc, MeOH; ii) (a) ethyl chloroformate, DBU, CH_2Cl_2 ; (b) DBU, CH_2Cl_2 ; (c) K_2CO_3 , EtOH; iii) formamidine acetate, EtOH; iv) BzCl, pyridine; v) TBAF, THF; vi) (EtO)_2POCH_2OTf, LiO-t-Bu, THF; vii) NH_3, MeOH, rt; viii) TMSBr, 2,6-lutidine, CH_3CN.

Scheme 3. Synthesis of 5'-norcarbocyclic 9-deazaadenosine phosphonic acid.

mediate 19 with a 75% yield. Sequentially, N-protection of 19 with ethyl chloroformate, DBU-induced cyclization²³ and deformylation gave the pyrrolo derivatives 20α and 20β , respectively.²⁴ Their relative stereochemistries were also unambiguously determined by NOE studies. Treatment of 20β with formamidine acetate gave the protected 5'-norcarbocyclic-9-deazaadenosine analog 21 with a 62% yield. For the synthesis of 5'-norcarbocyclic-9-deazaadenosine nucleoside phosphonic acid, perbenzoylation of 21 with benzoyl chloride in dry pyridine yielded the tri-N-benzoyl derivative 22 in an 86% yield by using a procedure similar²⁰ to that described for 14, which was desilylated using TBAF to give 5'-norcarbocyclic nucleoside analog 23, which was phosphonated to produce 24 by the same conditions as those used to prepare 16. The debenzovlation of 24 was achieved by using saturated methanolic ammonia at 25 °C to afford the corresponding 5'-norcarbocyclic 9-deazaadenosine phosphonate 25. From this intermediate, the targeted 5'-norcarbocyclic pyrrolo[3,2-d]pyrimidine nucleoside phosphonic acid 26 was prepared using a hydrolysis procedure similar to that described for the preparation of 19 (Scheme 3).

It should be noted that the synthesized 5'-norcarbocyclic

Seyeon Kim et al.

 Table 1. The antiviral activity of the synthesized nucleoside analog compounds

Compound No.	Anti-HIV-1 (PBM) EC ₅₀ (µM)	Cytotoxicity (PBM) IC ₅₀ (µM)	Cytotoxicity (Vero) IC ₅₀ (µM)
18	21.6	42	25
26	9.7	20	8.4
AZT	0.007	>100	40
PMEA	0.42	>100	25

AZT: azidothymidine. PMEA: 9-[2-(phosphonomethoxy)ethyl]adenine. EC₅₀ (μ M): concentration (μ M) required to inhibit the replication of HIV-1 by 50%. IC₅₀ (μ M): concentration (μ M) required to inhibit the cell growth of unaffected cells by 50%

 Table 2. In vitro inhibitory activity for tumor growth of the synthesized nucleoside analog compounds

L-1210 ID ₅₀ (µg/mL)	P-815 ID ₅₀ (μg/mL)
7.6	8.3
1.7	0.8
< 0.01	< 0.01
	L-1210 ID ₅₀ (µg/mL) 7.6 1.7 < 0.01

 ID_{50} (µg/mL): dose (µg/mL) required to inhibit the *in vitro* tumor growth by 50%. in murine leukemic cell lines

nucleoside phosphonic acids **18** and **26** were novel compounds that were not previously reported in the literature. The antiviral activity of nucleoside phosphonic acids is mostly explained by their intracellular metabolism to their diphosphates, which is followed by incorporation into the viral genome and chain termination.²⁵ As shown in Table 1, the synthesized compounds **18** and **26** were tested against HIV-1 and showed moderate antiviral activity, as well as cytotoxicity, at concentrations up to 100 μ M. This result indicates that their antiviral activity might be a result of cytotoxicity. Anti-HIV activity was determined in human peripheral blood mononuclear (PBM) cells infected with HIV-1 strain *LAI*. The cytotoxicity of the compounds was evaluated in parallel with their antiviral activity based on the viability of mock-infected cells.²⁶

In addition, the final *C*-nucleosides **18** and **26** were screened *in vitro* inhibitory activity for tumor growth against mouse leukemia cell lines (L-1210, P-815), according to the technique described by Fischer,²⁷ with some modifications.²⁸ The results summarized in Table 2 indicate that pyrrolo[3,2-d]pyrimidine analog **26** was more active than **18**, but was much less active than 9-deazaadenosine (**1**) itself.

Conclusions

Based on the potent biological activity of *C*-nucleosides and 5'-norcarbocyclic nucleoside phosphonic acids, we have designed and successfully synthesized novel 5'-norcarbocyclic *C*-nucleoside phosphonic acids starting from 1,3dihydroxy-cyclopentane. The data presented herein indicate that synthesized nucleoside phosphonic acid analogs showed moderate cytotoxicity-derived anti-HIV activity and anti-

Novel 5'-Norcarbocyclic C-nucleoside analogs

leukemic activity. Further studies evaluating the anti-leukemic activity of these systems are in progress.

Experiments

Melting points were determined using a Mel-temp II laboratory device and are uncorrected. NMR spectra were recorded using a JEOL 300 Fourier transform spectrometer (JEOL, Tokyo, Japan). Chemical shifts are reported in parts per million (δ), and signals are reported as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and dd (doublet of doublets). UV spectra were recorded using a Beckman DU-7 spectrophotometer (Beckman, South Pasadena, CA, USA). Mass (MS) spectra were recorded in electrospray ionization (ESI) mode. Elemental analyses were performed using a Perkin-Elmer 2400 analyzer (Perkin-Elmer, Norwalk, CT, USA). Thin-layer chromatography (TLC) was performed using Uniplates (silica gel) purchased from Analtech Co. (7558, Newark, DE, USA). All reactions were carried out under nitrogen atmosphere unless otherwise specified. Anhydrous dichloromethane, benzene, and pyridine were obtained by distillation from CaH₂. Anhydrous THF was obtained by distillation from Na and benzophenone, immediately prior to use.

(*rel*)-(1*S* and 1*R*,3*S*)-3-(*t*-Butyldimethylsilanyloxy) cyclopentanol (6). TBDMSCI (2.29 g, 15.25 mmol) was added slowly to a solution of **5** (1.41 g, 13.87 mmol) and imidazole (1.41 g, 20.80 mmol) in CH₂Cl₂ (100 mL) at -10 °C and stirred for 7 h at the same temperature. Saturated NaHCO₃ solution (10 mL) was poured into the mixture and stirred for 1 h at room temperature. The solvent was evaporated under reduced pressure. The residue was dissolved in water (200 mL) and extracted with diethyl ether (200 mL). The organic layer was washed with brine, dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane, 1:10) to produce compound **6** (1.58 g, 53%) as an isomeric mixture: ¹H NMR (CDCl₃, 300 MHz) δ 3.27-3.20 (m, 2H), 1.98-1.52 (m, 6H), 0.89 (m, 9H), 0.02 (m, 6H).

(±)-3-(t-Butyldimethylsilanyloxy) cyclopentanone (7). Compound 6 (2.43 g, 11.25 mmol) was added to a solution of Dess-Martin periodinane (10.38 g, 24.5 mmol) in CH₂Cl₂ (100 mL) at 0 °C, and stirred for 24 h at room temperature under argon gas. The solvent was removed and the residue was triturated with diethyl ether (150 mL). Following filtration through a pad of silica gel, the organic solution was washed with a solution of sodium thiosulfate pentahydrate (13 g) in water (100 mL), ice-cold saturated NaHCO₃ (80 mL), and brine (80 mL) and dried over MgSO₄. The solvent was filtered, concentrated in vacuo, and purified by silica gel column chromatography (EtOAc/hexane, 1:10) to produce compound 7 (2.19 g, 91%) as a colorless syrup. ¹H NMR (CDCl₃, 300 MHz) δ 3.76-3.74 (m, 1H), 2.34-2.02 (m, 6H), 0.89 (s, 9H), 0.03 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 215.6, 63.5, 52.6, 37.1, 31.5, 25.7, 18.4, -4.7. Anal. calcd. for C₁₁H₂₂O₂Si: C, 61.63; H, 10.34; found: C, 61.72; H, 10.26; MS m/z 215 (M+H)⁺.

Bull. Korean Chem. Soc. 2014, Vol. 35, No. 12 3505

(±)-(*E&Z*)-Ethyl 2-cyano-2-[4-(*t*-butyldimethylsilanyloxy) cyclopentylidene] acetate (8). Potassium *tert*-butoxide (6.73 g, 60.0 mmol) was added to a solution of 7 (2.57 g, 12.0 mmol) and ethyl cyanoacetate (6.78 g, 60.0 mmol) in ethanol (80 mL) at 0 °C. The mixture was stirred for 5 h at room temperature and concentrated under reduced pressure. The mixture was quenched by water (12 mL) and further diluted with 150 mL of water. The mixture was then extracted with EtOAc (200 m) 2 times. The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and then concentrated under reduced pressure. The residue was purified by flash column chromatography (EtOAc/hexane, 1:15) to give crude ester derivative **8** (7.8 g, 65%) as a colorless oil. Without further purification, compound **8** was subjected to the next reaction.

(*rel*)-(1*S* and 1*R*,4*R*)-Ethyl-2-[4-(*t*-butyldimethylsilanyloxy)cyclopentyl]-2-isocyanoacetate (9). To a solution of 8 (1.2 g, 3.88 mmol) in methanol (10 mL), 10% Pd/C (120 mg) was added. The mixture was thoroughly deoxygenated, then saturated with hydrogen and stirred for 24 h. The charcoal was removed by filtration through a short Celite[®] pad, which was thoroughly washed with methanol. Evaporation of the solvent gave a crude product which was purified by column chromatography (EtOAc/hexane, 1:10) to yield 1.07 g (89%) of **9** as a colorless oil. ¹H NMR (CDCl₃, 300 MHz) δ 4.16 (q, *J* = 7.0 Hz, 2H), 3.41 (m, 1H), 3.26-3.28 (m, 1H), 2.24-2.25 (m, 1H), 1.75-1.35 (m, 6H), 1.31 (t, *J* = 7.0 Hz, 3H), 0.89 (s, 9H), 0.03 (s, 6H); Anal. calcd. for C₁₆H₂₉NO₃Si: C, 61.69; H, 9.38; N, 4.50; found: C, 61.82; H, 9.34; N, 4.54; MS *m/z* 312 (M+H)⁺.

(*rel*)-(1*S* and 1*R*,4*R*)-2-[4-(*t*-Butyldimethylsilanyloxy) cyclopentyl]-3-hydroxyacrylonitrile (10). Diisobutylaluminium hydride (3.98 mL, 1 M in hexane) was added to a solution of 9 (1.24 g, 3.98 mmol) in anhydrous ether (8 mL) at -78 °C over 10 min. The resulting mixture was stirred for 10 min and quenched with MeOH (8 mL). The resulting white solid was filtered, and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane, 1:10) to give enol **10** (691 mg, 65%) as a colorless oil. ¹H NMR (CDCl₃, 300 MHz) δ 6.45 (s, 1H), 3.58-3.56 (m, 1H), 2.47-2.44 (m, 1H), 1.75-1.34 (m, 6H), 0.87-0.88 (m, 9H), 0.02-0.03 (m, 6H); Anal. calcd. for C₁₄H₂₅NO₂Si: C, 62.87; H, 9.42; N, 5.24; found: C, 62.74; H, 9.50, N, 5.19.

(*rel*)-(*E*&*Z*)-3-(Cyanomethoxy)-2-[(1*S* and 1*R*,4*R*)-4-(*t*butyldimethylsilanyloxy) cyclopentyl] acrylonitrile (11). To a solution of compound 10 (1.10 g, 4.14 mmol), cesium carbonate 5.39 g, 16.56 mmol) in anhydrous DMF (30 mL) was added chloroacetonitrile (2.48 g, 33.12 mmol), and the solution was stirred at room temperature for 24 h. The reaction mixture was poured water (150 mL) and extracted with *t*-butyl methyl ether (150 mL) three times. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (EtOAc/Benzene, 1:10) to give **11** (1.02 g, 81%) as an isomeric mixture. The mixture was subjected directly to the next step.

(rel)-3-Amino-4-[(1'R,4'R)-4'-(t-butyldimethylsilanyloxy) cyclopentyl] furan-2-carbonitrile (12a) and (rel)-3-amino-4-[(1'S,4'R)-4'-(t-butyldimethylsilanyloxy) cyclopentyl] furan-2-carbonitrile (12β) . To the solution of the carbonitrile 11 (1.6 g, 5.22 mmol) in anhydrous THF (20 mL), a solution of LDA (2.24 g, 21 mmol) in anhydrous THF (15 mL) was added dropwise at -70 °C and stirred for 2 h. The mixture was quenched with a saturated solution of ammonium chloride (150 mL) and stirred for 1 h at room temperature. The mixture was extracted with EtOAc (150 mL) 3 times. The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by flash column chromatography (EtOAc/Hexane, 3:1) to give 12a (303 mg, 19%) and 12 β (335 mg, 21%). Spectroscopic data for 12 α : ¹H NMR (CDCl₃, 300 MHz) δ 7.23 (s, 1H), 4.28 (br s, 2H, D₂O exchangeable), 3.31-3.29 (m, 1H), 2.82-2.83 (m, 1H), 2.02-1.37 (m, 6H), 0.88-0.89 (m, 9H), 0.02 (s, 6H); Anal. calcd. for C₁₆H₂₆N₂O₂Si: C, 62.70; H, 8.55; N, 9.14; found: C, 62.56; H, 8.50; N, 9.29; MS *m/z* 307 (M+H)⁺. Spectroscopic data for 12β: ¹H NMR (CDCl₃, 300 MHz) δ 7.25 (s, 1H), 4.30 (br, 2H, D₂O exchangeable), 3.34-3.31 (m, 1H), 2.81-2.79 (m, 1H), 2.00-1.35 (m, 6H), 0.88-0.89 (m, 9H), 0.03 (s, 6H); Anal. calcd. for C16H26N2O2Si: C, 62.70; H, 8.55; N, 9.14; found: C, 62.85; H, 8.63; N, 9.08; MS m/z 307 $(M+H)^{+}$.

(rel)-[(1S,4'R)-9-[4'-(t-Butyldimethylsilanyloxy) cyclopentyl] 7-oxa-7,9-dideazaadenosine (13). To a solution of 12β (650 mg, 2.12 mmol) in EtOH (20 mL), formamidine acetate (2.20 g, 21.2 mmol) was added and the reaction mixture was refluxed for 48 h. The solvent was concentrated under reduced pressure, and the residue was partitioned between dichloromethane and water. The organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CH₂Cl₂/MeOH, 10:1) to give **13** (487 mg, 69%). ¹H NMR (DMSO- d_6 , 300 MHz) δ 9.28 (br s, 2H, D₂O exchangeable), 8.63 (s, 1H), 8.51 (s, 1H), 3.29-3.30 (m, 1H), 2.82-2.83 (m, 1H), 2.07-2.08 (m, 1H), 1.94-1.47 (m, 5H), 0.88-0.89 (m, 9H), 0.03-0.04 (m, 6H); Anal. calcd. for C₁₇H₂₇N₃O₂Si (+1.0 MeOH): C, 59.19; H, 8.55, N, 11.50; found: C, 59.28; H, 8.48; N, 11.58; MS m/z $334 (M+H)^+$.

(*rel*)-[(1'S,4'R)-9-[4'-(*t*-Butyldimethylsilanyloxy) cyclopentyl] 4-*N*-dibenzoyl-7-oxa-7,9-dideazaadenosine (14). Benzoyl chloride (2.12 g, 15.12 mmol) was added dropwise to a stirred solution of compound 13 (1.26 g, 3.78 mmol) in dry pyridine (10 mL) at 0 °C and stirred overnight at rt. The reaction mixture was quenched by water (10 mL) and stirred for 1 h at 0 °C. The mixture was concentrated *in vacuo* and the residue was partitioned between H₂O (100 mL) and CH₂Cl₂ (100 mL). The aqueous layer was further extracted with CH₂Cl₂ 2 times. The combined organic layers were washed with cold saturated NaHCO₃ solution, then washed with brine, dried over anhydrous MgSO₄, and concentrated *in vacuo*. graphy (EtOAc/*n*-hexane, 1:8) to give **14** (1.74 g, 85%) as a form. ¹H NMR (CDCl₃, 300 MHz) δ 8.58 (s, 1H), 7.84-7.45 (m, 11H), 3.34-3.35 (m, 1H), 2.78-2.79 (m, 1H), 1.91-1.48 (m, 6H), 0.89-0.90 (m, 9H), 0.02-0.03 (m, 6H); Anal. calcd. for C₃₁H₃₅N₃O₄Si: C, 68.73; H, 6.51, N, 7.76; found: C, 68.87; H, 6.46; N, 7.88; MS *m*/*z* 542 (M+H)⁺.

(*rel*)-(1'S,4'*R*)-9-(4'-Cyclopentyloxy) 4-*N*-dibenzoyl-7oxa-7,9-dideazaadenosine (15). To a solution of compound 14 (320 mg, 0.59 mmol) in THF (10 mL), tetrabutylammonium fluoride (TBAF; 0.65 mL, 1.0 M solution in THF) at 0 °C was added. The mixture was stirred for 5 h at room temperature and concentrated. The residue was purified by silica gel column chromatography (EtOAc/*n*-hexane, 1:1) to give compound 15 (176 mg, 70%). ¹H NMR (CDCl₃, 300 MHz) δ 8.50 (s, 1H), 7.91-7.34 (m, 11H), 3.32-3.30 (m, 1H), 2.81-2.79 (m, 1H), 2.06-2.07 (m, 1H), 1.91-1.42 (m, 5H); Anal. calcd. for C₂₅H₂₁N₃O₄: C, 70.25; H, 4.95, N, 9.83; found: C, 70.37; H, 4.88; N, 9.89; MS *m/z* 428 (M+H)⁺.

(rel)-(1'S,4'R)-Diethyl-9-[4'-(cyclopentyloxy) 4-N-dibenzoyl-7-oxa-7,9-dideazaadenosine] methylphosphonate (16). Both LiOt-Bu (2.996 mL of 0.5 M solution in THF, 1.498 mmol) and a solution of diethyl phosphonomethyltriflate (449 mg, 1.498 mmol) in 16.0 mL of THF were slowly added to a solution of the 15 analog (320 mg, 0.749 mmol) in 8.0 mL of THF at 0 °C and stirred overnight at room temperature under anhydrous conditions. The mixture was quenched by adding saturated NH₄Cl solution (5 mL) and further diluted with additional H₂O (100 mL). The aqueous layer was extracted with EtOAc (2×100 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography (nhexane/EtOAc, 1:2) to produce **16** (233 mg, 54%). ¹H NMR (CDCl₃, 300 MHz) & 8.52 (s, 1H), 7.93-7.39 (m, 11H), 4.12-4.09 (m, 4H), 3.86 (d, *J* = 8.0 Hz, 2H), 3.06-3.07 (m, 1H), 2.78-2.79 (m, 1H), 2.06-2.07 (m, 1H), 1.88-1.44 (m, 5H), 1.15-1.13 (m, 6H); Anal. calcd. for C₃₀H₃₂N₃O₇P: C, 62.39; H, 5.58; N, 7.28; found: C, 62.51; H, 5.49; N. 7.36; MS m/z $578 (M+H)^+$.

(*rel*)-(1'S,4'*R*)-Diethyl-9-[(4'-cyclopentyloxy) 7-oxa-7,9dideazaadenosine] methylphosphonate (17). A solution of 16 (280 mg, 0.485 mmol) in saturated methanolic ammonia (12 mL) was stirred overnight at room temperature for 6 h and the volatiles were evaporated. The residue was purified by silica gel column chromatography (MeOH/CH₂Cl₂, 1:8) to produce 17 (91 mg, 51%). ¹H (DMSO-*d*₆, 300 MHz) δ 9.30 (br s, 2H, D₂O exchangeable), 8.68 (s, 1H), 8.53 (s, 1H), 4.14-4.11 (m, 4H), 3.84 (d, *J* = 8.0 Hz, 2H), 3.12-3.13 (m, 1H), 2.78-2.79 (m, 1H), 2.01-1.43 (m, 6H), 1.12-1.13 (m, 6H); Anal. calcd. for C₁₆H₂₄N₃O₅P (+1.0 MeOH): C, 50.89; H, 7.03; N, 10.47; Found: C, 50.77; H, 6.93; N, 10.32; MS *m/z* 370 (M+H)⁺.

(*rel*)-(1'*S*,4'*R*)-9-[(4'-Cyclopentyloxy) 7-oxa-7,9-dideazaadenosine] methylphosphonic acid (18). TMSBr (786 mg, 5.14 mmol) was added to a solution of phosphonate 17 (190 mg, 0.514 mmol) in anhydrous CH₃CN (12 mL) and 2,6lutidine (1.10 g, 10.28 mmol). The mixture was heated over-

Novel 5'-Norcarbocyclic C-nucleoside analogs

night at 69 °C under nitrogen and then concentrated *in vacuo*. The residue was co-evaporated from concentrated aqueous ammonium hydroxide (NH₄OH; 2 × 20.6 mL) and purified by triturating with acetone (10.3 mL) twice and removing the acetone by evaporation. The residue was then purified by preparative reverse-phase column chromatography using C18 silica gel. Lyophilization of the appropriate fraction produced phosphonic acid salt **18** (98 mg, 58% yield) as a white salt (ammonium salt). ¹H NMR (D₂O, 300 MHz) δ 8.65 (s, 1H), 8.45 (s, 1H), 3.81 (d, *J* = 8.2 Hz, 2H), 3.17 (m, 1H), 2.77-2.78 (m, 1H), 2.03-1.41 (m, 6H); ¹³C NMR (D₂O, 75 MHz) δ 151.4, 147.6, 147.9, 138.1, 117.0, 85.6, 62.9 (*J* = 45.0 Hz), 50.2, 33.8, 31.2, 27.1; HPLC, *t*_R = 10.86 min; HRMS, [M–H]⁺ calcd. 312.0765, found 312.0766.

(*rel*)-(*E*&*Z*)-3-(Cyanomethylamino)-2-[(1*S* and 1*R*,4*R*)-4-(*t*-butyldimethylsilanyloxy) cyclopentyl] acrylonitrile (19). Compound 10 (1.22 g, 4.56 mmol) was dissolved in MeOH (50 mL) followed by addition of aminoacetonitrile · monosulfate (2.81 g, 18.24 mmol) and sodium acetate (2.61 g, 31.92 mmol). The mixture was stirred for 6 h at room temperature, diluted with CHCl₃, and washed with water. The aqueous phase was back-washed with CHCl₃, the combined organic phase was dried, and the solvent was evaporated to give crude product 19 (1.04 g, 75%) as a mixture of E/Z diastereomers. The mixture was subjected directly to the next step.

(rel)-3-Amino-4-[(1'R,4'R)-4'-(t-butyldimethylsilanyloxy) cyclopentyl] 1H-pyrrole-2-carbonitrile (20a) and (rel)-3amino-4-[(1'S,4'R)-4'-(t-butyldimethylsilanyloxy) cyclo**pentyl**] 1*H*-pyrrole-2-carbonitrile (20β). To a solution of 19 (2.56 g, 8.38 mmol) in anhydrous CH_2Cl_2 (50 mL) was added 1,8-diazabicyclo[5,4,0] undec-7-ene (DBU, 2.5 mL, 16.78 mmol) and ethyl chloroformate (1.2 mL, 12.57 mmol) at 0 °C. The mixture was stirred for 2 h at room temperature. To the mixture, additional DBU (2.5 mL, 16.78 mmol) was added to induce cyclization, after which it was stirred at the same temperature for 20 h. The reaction mixture was diluted with CHCl₃ (150 mL) and extracted with an aqueous solution of citric acid (10%, 2×120 mL). The combined aqueous layer was back-washed with CHCl3. The combined organic layer was dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was dissolved in EtOH (50 mL), treated with potassium carbonate (1.15 g, 8.38 mmol) and stirred for 1 h at rt. The reaction mixture was quenched with water and extracted with CH₂Cl₂ two times. The combined organic layer was dried with anhydrous MgSO₄, filtered and concentrated. The residue was purified by flash column chromatography (EtOAc/n-hexane, 3:1) to give 20α (409 mg, 16%) and 20β (434 mg, 17%). Spectroscopic data for 20α : ¹H NMR (CDCl₃, 300 MHz) δ 7.87 (br s, 1H, D₂O exchangeable), 6.51 (s, 1H), 4.34 (br s, 2H, D₂O exchangeable), 3.37-3.38 (m, 1H), 2.82-2.83 (m, 1H), 2.02-1.50 (m, 6H), 0.88 (s, 9H), 0.02 (s, 6H); Anal. calcd. for C₁₆H₂₇N₃OSi: C, 62.91; H, 8.91; N, 13.75; found: C, 63.06; H, 8.85; N, 13.69; MS m/z 306 (M+H)⁺. Spectroscopic data for **20**: ¹H NMR (CDCl₃, 300 MHz) δ 7.91 (br s, 1H, D₂O exchangeable), 6.58 (s, 1H), 4.41 (br s, 2H, D₂O exchangeable), 3.31-3.32 (m, 1H), 2.79-2.80 (m, 1H), 1.98-1.47 (m, 6H), 0.89 (s, 9H), 0.03 (s, 6H); Anal. calcd. for $C_{16}H_{27}N_3OSi:$ C, 62.91; H, 8.91; N, 13.75; found: C, 62.83; H, 8.96; N, 13.87; MS *m*/z 306 (M+H)⁺.

(rel)-(1S,4'R)-9-[4'-(t-Butyldimethylsilanyloxy) cyclopentvl] 9-deazaadenosine (21). To a solution of 20b (450 mg, 1.47 mmol) in EtOH (12 mL), formamidine acetate (1.53 g, 14.7 mmol) was added and the reaction mixture was refluxed for 36 h. The solvent was concentrated under reduced pressure, and the residue was diluted with dichloromethane (100 mL) and extracted with water (2×100 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CH₂Cl₂/MeOH, 10:1) to give **21** (302 mg, 62%). ¹H NMR (DMSO- d_6 , 300 MHz) δ 12.53 (br s, 1H, D₂O exchangeable), 9.04 (br s, 2H, D₂O exchangeable), 8.45 (s, 1H), 7.77 (d, J = 3.0 Hz, 1H, singlet with D₂O), 3.36-3.37 (m, 1H), 2.80-2.81 (m, 1H), 1.99-1.47 (m, 6H), 0.87 (s, 9H), 0.01 (s, 6H); Anal. calcd. for C₁₇H₂₈N₄OSi (+1.0 MeOH): C, 59.35; H, 8.85, N, 15.38; found: C, 59.49; H, 8.78; N, 15.50; MS m/z $333 (M+H)^+$.

(rel)-(1'S,4'R)-9-[4'-(t-Butyldimethylsilanyloxy) cyclopentyl] 7-N-benzoyl-4-N-dibenzoyl-9-deazaadenosine (22). Benzoyl chloride (3.55 g, 25.26 mmol) was added dropwise to a stirred solution of compound 21 (1.40 g, 4.21 mmol) in dry pyridine (20 mL) at 0 °C and stirred overnight at room temperature. The reaction mixture was quenched by water (20 mL) and stirred for 1 h at 0 °C. The mixture was concentrated in vacuo and the residue was partitioned between H₂O (150 mL) and CH₂Cl₂ (150 mL). The aqueous layer was further extracted with CH₂Cl₂ 2 times. The combined organic layers were washed with cold saturated NaHCO3 solution, then washed with brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The residue was purified by flash column chromatography (EtOAc/n-hexane, 1:8) to give 22 (2.33 g, 86%). ¹H NMR (CDCl₃, 300 MHz) δ 8.68 (s, 1H), 7.97-7.29 (m, 16H), 3.32-3.33 (m, 1H), 2.80-2.81 (m, 1H), 1.97-1.43 (m, 6H), 0.89 (s, 9H), 0.02 (s, 6H); Anal. calcd. for C₃₈H₄₀N₄O₄Si: C, 70.78; H, 6.25, N, 8.69; found: C, 70.64; H, 6.36; N, 8.57; MS *m*/*z* 645 (M+H)⁺.

(*rel*)-(1'S,4'*R*)-9-[(4'-Cyclopentyloxy) 7-*N*-benzoyl-4-*N*dibenzoyl-9-deazaadenosine (23). To a solution of compound 22 (860 mg, 1.33 mmol) in THF (12 mL), tetrabutylammonium fluoride (TBAF, 1.59 mL, 1.0 M solution in THF) at 0 °C was added. The mixture was stirred for 6 h at room temperature and concentrated. The residue was purified by silica gel column chromatography (EtOAc/n-hexane, 1:1) to give compound 23 (535 mg, 76%). ¹H NMR (CDCl₃, 300 MHz) δ 8.76 (s, 1H), 8.08-7.34 (m, 16H), 3.35-3.36 (m, 1H), 2.81-2.82 (m, 1H), 2.01-2.02 (m, 1H), 1.97-1.46 (m, 5H); Anal. calcd. for C₃₂H₂₆N₄O₄: C, 72.44; H, 4.94, N, 10.56; found: C, 72.56; H, 4.85; N, 10.69; MS *m/z* 531 (M+H)⁺.

(*rel*)-(1'*S*,4'*R*)-Diethyl-9-[(4'-cyclopentyloxy]) 7-*N*-benzoyl-4-*N*-dibenzoyl-9-deazaadenosine] methylphosphonate (24). Phosphonation of the alcohol 23 was accomplished by similar reaction procedures as were used for 16 to produce **24** with a 60% yield; ¹H NMR (CDCl₃, 300 MHz) δ 8.78 (s, 1H), 8.08-7.34 (m, 16H), 4.10-4.08 (m, 4H), 3.79 (d, *J* = 8.0 Hz, 2H), 3.24-3.25 (m, 1H), 2.79-2.80 (m, 1H), 2.02-1.40 (m, 6H), 1.15-1.13 (m, 6H); Anal. calcd. for C₃₇H₃₇N₄O₇P: C, 65.29; H, 5.48; N, 8.23; found: C, 65.44; H, 5.41; N. 8.15; MS *m/z* 681 (M+H)⁺.

(*rel*)-(1'*S*,4'*R*)-Diethyl-9-[(4'-cyclopentyloxy) 9-deazaadenosine] methylphosphonate (25). Ammonolysis of the tri-*N*-benzoyl protection groups of 24 was performed by similar reaction conditions as those used for 17 to produce 25 with a 49% yield; ¹H (DMSO-*d*₆, 300 MHz) δ 12.18 (br s, 1H, D₂O exchangeable), 9.01 (br s, 2H, D₂O exchangeable), 8.52 (s, 1H), 7.89 (d, *J* = 2.9 Hz, 1H, singlet with D₂O), 4.11-4.09 (m, 4H), 3.81 (d, *J* = 8.2 Hz, 2H), 3.31-3.32 (m, 1H), 2.84-2.85 (m, 1H), 2.03-1.46 (m, 6H), 1.13-1.10 (m, 6H); Anal. calcd. for C₁₆H₂₅N₄O₄P (+1.0 MeOH): C, 51.02; H, 7.30; N, 13.99; Found: C, 50.91; H, 7.23; N, 13.85; MS *m*/z 369 (M+H)⁺.

(*rel*)-(1'*S*,4'*R*)-9-(4'-Cyclopentyloxy) 9-deazaadenosine] methylphosphonic acid (26). Deprotection of diethylphosphonate 25 to phosphonic acid 26 was performed by similar reaction conditions as were used for 18 with a 51% yield (ammonium salt). UV (H₂O) λ_{max} 268.0 nm; ¹H NMR (D₂O, 300 MHz) δ 8.55 (s, 1H), 7.82 (s, 1H), 3.76 (d, *J* = 8.1 Hz, 2H), 2.86-2.87 (m, 1H), 2.79-2.80 (m, 1H), 2.01-1.45 (m, 6H); ¹³C NMR (D₂O, 75 MHz) δ 154.6, 145.3, 134.7, 129.4, 118.7, 84.8, 62.7 (*J* = 40.0 Hz), 48.5, 36.7, 32.6, 26.9; HPLC, *t*_R = 10.71 min; HRMS, [M–H]⁺ calcd.: 311.0657; found: 311.0658.

References

- (a) Shaban, M. A. E.; Nasr, A. Z. Adv. Heterocycl. Chem. 1997, 68, 223-432.
 (b) Shaban, M. A. E. Adv. Heterocycl. Chem. 1997, 70, 163-337.
 (c) Liang, C.; Ma, T.; Cooperwood, J. S.; Du, J.; Chu, C. K. Carbohydr. Res. 1997, 303, 33-38.
 (d) Zhou, J.; Yang, M.; Akdag, A.; Wang, H.; Schneller, S. W. Tetrahedron 2008, 64, 433-438.
- Buchenal, J. H.; Ciovacco, K.; Kalaher, K.; O'Toole, T.; Kiefner, R.; Dowing, M. D.; Chu, C. K.; Watanabe, K. A.; Wempen, I.; Fox, J. J. *Cancer Res.* **1976**, *36*, 1520-1523.
- (a) Fuertes, M.; Garcia-Lopez, M. T.; Garcia-Munoz, G.; Stud, M. J. Org. Chem. 1976, 41, 4074-4077. (b) Srivastava, P. C.; Pickering, M. V.; Allen, L. B.; Streeter, D. C.; Campbell, M. T.; Witkowski, J. T.; Sidwell, R. W.; Robins, R. K. J. Med. Chem. 1977, 20, 256-262.
- 4. Srivastava, P. C.; Robins, R. K. J. Med. Chem. 1983, 26, 445-448.
- (a) Chu, C. Y.; Zuckerman, L. B.; Sato, S.; Crabtree, G. W.; Bogden, A. E.; Lim, M. I.; Klein, R. S. *Biochem. Pharmacol.* **1984**, *33*, 1229-1234. (b) Zimmerman, T. P.; Deeprose, R. D.; Wolberg, G.; Stopford, C. R.; Duncan, G. S.; Miller, W. H.; Miller, R. L.; Lim, M. I.; Ren, W. Y.; Klein, R. S. *Biochem. Pharmacol.* **1983**, *32*, 1211-1217.
- (a) Tam, S. Y.-K.; Klein, R. S.; Wempen, I.; Fox, J. J. J. Org. Chem. 1979, 44, 4547-4553. (b) Tam, S. Y.-K.; Hwang, J. S.; De Las Heras, F. G.; Klein, R. S.; Fox, J. J. J. Heterocycl. Chem. 1976, 13, 1305-1308.
- (a) Fissekis, J. D.; Creegan, B. M. J. Org. Chem. 1967, 32, 3595-3603. (b) Katagiri, N.; Haneda, T.; Kaneko, C. Chem. Pharm.

Bull. 1986, 34, 4875-4878. (c) Katagiri, N.; Haneda, T.; Hayasaka,
E.; Watanabe, N.; Kaneko, C. J. Org. Chem. 1988, 53, 226-227.
(d) Lee, C. H.; Kim, J. Y.; Kim, W. J.; Kim, Y. H. Heterocycles
1990, 31, 211-214. (e) Katagiri, N.; Tomura, M.; Haneda, T.;
Kaneko, C. J. Chem. Soc., Chem. Commun. 1987, 19, 1422-1423.
(f) Coockson, R. C.; Dufield, P. J.; Scopes, D. I. C. J. Chem. Soc., Perkin Trans. 1 1986, 393-398. (g) Takahashi, T.; Kotsubo, H.;
Koizumi, T. Tetrahedron: Asymmetry 1991, 2, 1035-1040. (h) Zhou,
J.; Yang, M.; Schnell, S. W. Tetrahedron Lett. 2004, 45, 8233-8234.

- Zhou, J.; Yang, M.; Akdag, A.; Wang, H.; Schneller, S. W. *Tetra*hedron 2008, 64, 433-438.
- (a) Borthwick, A. D.; Biggadike, K. *Tetrahedron* 1992, 48, 571-623.
 (b) Huryn, D. M.; Okabe, M. *Chem. Rev.* 1992, 92, 1745-1768.
 (c) Agrofoglio, L.; Suhas, E.; Farese, A.; Condom, R.; Challand, S.; Earl, R. A.; Guedj, R. *Tetrahedron* 1994, 50, 10611-10670.
 (d) Crimmins, M. T. *Tetrahedron* 1998, 54, 9229-9272.
 (e) Ariona, O.; Gómez, A. M.; López, J. C.; Plumet, J. *Chem. Rev.* 2007, 107, 1919-2036.
 (f) Jeong, L. S.; Lee, J. A. *Antiviral Chem. Chemother.* 2004, 15, 235-250.
 (g) Boutureira, O.; Matheu, M. I.; Díazm, Y.; Castillón, S. *Chem. Soc. Rev.* 2013, 42, 5056-5072.
- 10. Stoeckler, J. D.; Cambor, C.; Parks, R. E., Jr. *Biochemistry* 1980, 19, 102-107.
- Boojamra, C. G; Parrish, J. P.; Sperandio, D.; Gao, Y.; Petrakovsky, O. V.; Lee, S. K.; Markevich, D. Y.; Vela, J. E.; Laflamme, G; Chen, J. M.; Ray, A. S.; Barron, A. C.; Sparacino, M. L.; Desai, M. C.; Kim, C. U.; Cihlar, T.; Mackman, R. L. *Bioorg. Med. Chem.* 2009, *17*, 1739-1746.
- Koh, Y. H.; Shim, J. H.; Wu, J. Z.; Zhong, W.; Hong, Z.; Girardet, J. L. J. Med. Chem. 2005, 48, 2867-2875.
- Wu, T.; Froeyen, M.; Kempeneers, V.; Pannecouque, C.; Wang, J.; Busson, R.; De Clercq, E.; Herdewijn, P. J. Am. Chem. Soc. 2005, 127, 5056-5065.
- Kim, C. U.; Luh, B. Y.; Misco, P. F.; Bronson, J. J.; Hitchcock, M. J.; Ghazzouli, I.; Martin, J. C. J. Med. Chem. 1990, 33, 1207-1213.
- Kim, C. U.; Luh, B. Y.; Martin, J. C. J. Org. Chem. 1991, 56, 2642-2647.
- (a) De Clercq, E. *Biochem. Pharmacol.* 2011, *82*, 99-109. (b) Balzarini, J.; Hao, Z.; Herdewijn, P.; Johns, D. G; De Clercq, E. *Proc. Natl. Acad. Sci. USA* 1991, *88*, 1499-1503.
- 17. Dess, D. B.; Martin, J. C. J. Am. Chem. Soc. 1991, 113, 7277-7287.
- Chun, B. K.; Song, G. Y.; Chu, C. K. J. Org. Chem. 2001, 66, 4852-4858.
- Bhattacharya, B. K.; Otter, B. A.; Berens, R. L.; Klein, R. S. Nucleosides Nucleotides 1990, 9, 1021-1043.
- Rao, K. V. B.; Ren, W.-Y.; Burchenal, J. H.; Klein, R. S. *Nucleosides Nucleotides* 1986, *5*, 539-569.
- (a) Phillion, D. P.; Andrew, S. S. *Tetrahedron Lett.* **1986**, *27*, 1477-1480.
 (b) Xu, Y.; Flavin, M. T.; Xu, Z.-Q. J. Org. Chem. **1996**, *61*, 7697-7701.
- Hocková, D.; Holý, A.; Masojídková, M.; Keough, D. T.; De Jersey, J.; Guddat, L. W. *Bioorg. Med. Chem.* 2009, *17*, 6218-6232.
- 23. Lim, M. I.; Klein, R. S. Tetrahedron Lett. 1981, 22, 25-28.
- 24. Kamath, V. P.; Ananth, S.; Bantia, S.; Morris, P. E. J. Med. Chem. 2004, 47, 1322-1324.
- Holy, A.; Votruba, I.; Merta, A.; Cerny, J.; Vesely, J.; Vlach, J.; Sediva, K.; Rosenberg, I.; Otmar, M.; Hrebabecky, H.; Travniekb, M.; Vonkac, V.; Snoeck, R.; De Clercq, E. *Antiviral Res.* 1990, *13*, 295-311.
- Pauwels, R.; Balzarini, J.; Baba, M.; Snoeck, R.; Schols, D.; Herdewijn, P.; Desmyter, J.; De Clercq, E. J. Virol. Methods 1988, 20, 309-321.
- 27. Fischer, G. A. Ann. N.Y. Acad. Sci. 1958, 76, 673-680.
- Burchenal, J. H.; Chou, T.-C.; Lokye, L.; Smith, R. S.; Watanabe, K. A.; Su, T.-L.; Fox, J. J. *Cancer Res.* **1982**, *42*, 2598-2600.