Facile Synthesis of Natural Moracin Compounds using Pd(OAc)₂/P(^tBu)₃-HBF₄ as a Sonogashira Coupling Reagent

Jae Jun Lee, So-Ra Yun, and Jong-Gab Jun*

Department of Chemistry and Institute of Applied Chemistry, Hallym University, Chuncheon 200-702, Korea *E-mail: jgjun@hallym.ac.kr Received May 20, 2014, Accepted August 7, 2014

An efficient and practical synthesis of natural moracins, which have diverse range of biological properties including anticancer, antioxidant, and antibacterial activities, has been achieved using $Pd(OAc)_2/P('Bu)_3$ -HBF₄ as a Sonogashira coupling reagent which solved the unreactive problems in case of higher electron density of haloaryl compounds in the reaction. Lowering electron density of halophenol with acetylation and changing Sonogashira coupling reagent from $PdCl_2(PPh_3)_2$ to $Pd(OAc)_2/P('Bu)_3$ -HBF₄ smoothly produce the benzofuran structures in the syntheses of moracins M, N and S. The electron deficient halobenzaldehyde, however, easily forms the benzofuran using original Sonogashira conditions, and utilized for the first synthesis of moracin Y.

Key Words : Moracin, Sonogashira reaction, Benzofuran, Pd(OAc)₂/P('Bu)₃-HBF₄, Morus alba Linn

Introduction

Moracins are active components of mulberry (*Morus alba Linn.*), and isolated and identified of the structures as many as A through Z.¹ Most of them show important biological effects including antioxidant,² anti-inflammatory,³ anticancer,⁴ asthma,⁵ anxiolytic,⁶ analgesic,⁷ hyperlipidemic⁸ and antibacterial activities.⁹ Among them, moracin M (1) shows the most simple 2-arylbenzofuran structure and several synthetic reports with activity studies since the isolation from the heartwood of *Morus laevigata* in 1975.¹⁰ Direct lithiation methods developed by Widdowson¹¹ or Watanabe¹² for moracin M synthesis encountered problems, such as com-



Figure 1. Moracin M (1).

mercially not available starting materials or a low overall yield (7%), respectively. Recently, Cossio *et al.* reported an interesting strategy for moracin M synthesis which showed the benzofuran formation using one-pot reaction between phenol and α -bromoacetophenone in the presence of Al₂O₃.¹³ However, the benzofuran formation reaction showed only 26% yield with a low total yield. There were no reports for moracin M synthesis¹⁴ by using Sonogashira coupling reaction which is often used for benzofuran ring formation.¹⁵

Results and Discussion

We applied the Sonogashira coupling reaction of haloresorcinol **2** with aryl acetylene **3** using $PdCl_2(PPh_3)_2$ condition for moracin M synthesis. However, no reaction occurred despite every effort as shown in Scheme 1. Electron density of halophenol **2** in the reaction could be important. Halobenzaldehyde **4**, of which electron density was lowered by substitution of electron-withdrawing aldehyde comparing to **2**, was used once in our group^{15d} for XH-14 synthesis, and



Scheme 1. Sonogashira coupling reaction of aryl halide with aryl acetylene derivatives.



Figure 2. Tolman angle of Pd-acetylene complex.

successfully applied with Sonogashira reagent to yield the expected benzofuran 5. Acetylated haloresorcinol 6 for decreasing the electron density of 2, however, did not give the expected benzofuran using original Sonogashira condition, and we assumed that the chloride anion produced from the reaction could be used as a nucleophile causing deacetylation to increase the electron density again and prohibit the coupling reaction.

Fu¹⁶ introduced air-stable trialkylphosphonium salt and Kotschy¹⁷ used this catalyst with Pd(OAc)₂ for benzofuran ring formation. Fagnou also introduced the optimal metalligand combination using Pd(OAc)₂ with P('Bu)₃-HBF₄ to solve a cross-coupling problem.¹⁸ Using bulky phosphine ligands exhibiting a large cone angle (known as Tolman angle)¹⁹ has been known to accelerate the reductive elimination step in palladium catalysed coupling cycle (oxidative addition-transmetallation-reductive elimination) due to a decreased distance between the two R groups as shown in Figure 2. Changing the ligand from PPh₃ to P('Bu)₃ increase the cone angle between two PL₃ from 145° to 182° in Pd-acetylene complex, and decrease the angle between two R groups which accelerate the reductive elimination step in the reaction pathway.

Sonogashira coupling of diacetylated bromoresorcinol **6a** with aryl acetylide **7** using Pd(OAc)₂/P(^{*i*}Bu)₃-HBF₄, CuI and

DIPA gave 63% yield of coupled product **8**, which was then deacetylated in basic condition to give the benzofuran **9** (Scheme 2). Demethylation of **9** using BBr₃ produced the moracin M in 25% yield as same as literature.¹³ The intermediate benzofuran **9** was utilized for the first synthesis of moracin N (**13**) which was isolated from mulberry²⁰ and showed tyrosinase inhibitory activity²¹ and better antifungal activity²² than moracin M. Propargylation of **9** with 3-chloro-3-methylbutyne and following reduction using Lindlar catalyst gave the prenyl ether **11**. Water-accelerated [3,3]-sigmatropic rearrangement²³ of **11** produced 5-propenyl-benzofuran **12** in 47% yield regioselectively, and followed demethylation of **12** using BBr₃ gave the moracin N in 21% yield.

Moracin S (19), which is the regioisomer of moracin N, was isolated from black mulberry^{1b} and showed similar antioxidant activity as vitamin E, ^{1b} however, no other reports on activity and synthesis are noticed. First total synthesis of moracin S is achieved using modified Sonogashira coupling reaction as shown in Scheme 3. Regioselective acetylation of **2a** with Ac₂O in acidic condition gave 40% yield of **6b** which was then propargylated to **14** in 65% yield and followed reduction with Lindlar catalyst produced **15** in 75% yield. Coupling reaction of **15** with aryl acetylide **7** using the modified Sonogashira reagent gave **16** in 56% yield and followed water-accelerated [3,3]-sigmatropic rearrangement produced the benzofuran **17** in one-step. Deacetylation of **17** with KOH and followed demethylation using BBr₃ produced the moracin S in 26% yield.

Moracin Y was extracted from mulberry leaves and only one bioactivity has been known.^{1a} Benzofuran **5a** containing



Scheme 2. Synthesis of moracins M and N.

Synthesis of Natural Moracins



Scheme 3. Synthesis of moracin S.



Scheme 4. Synthesis of moracin Y.

electron withdrawing aldehyde, which was prepared in Scheme 1 using original Sonogashira coupling condition, is utilized for the first synthesis of moracin Y (21) as shown in Scheme 4. Benzaldehyde 20 was iodinated with ICl and coupled with aryl acetylide 7a to give the benzofuran 5a, which was then deprotected with TBAF to produce the moracin Y with overall 23% yield in 3 steps.

In conclusion, efficient and practical syntheses of moracins M, N, and S have been achieved using $Pd(OAc)_2/P('Bu)_3$ -HBF₄ and the spectral data of the synthetic moracins N, S and Y are well matched with the literatures.^{20,1b,1a}

Experimental

All chemicals were purchased from Sigma-Aldrich Chemicals and were used without further purification unless noted otherwise. NMR spectra were recorded at Varian Mercury-300 MHz FT-NMR and 75 MHz for ¹³C, with the chemical shift (δ) reported in parts per million (ppm) relative to TMS and the coupling constants (*J*) quoted in Hz. CDCl₃ was used as a solvent and an internal standard. Mass spectra were recorded using a JMS-700 (JEOL) spectrometer. Melting points were measured on a MEL-TEMP II apparatus and were uncorrected. Thin-layer chromatography (TLC) was performed on DC-Plastikfolien 60, F_{254} (Merck, layer thickness 0.2 mm) plastic-backed silica gel plates and visualized by UV light (254 nm) or staining with *p*-anisaldehyde.

1,3-Diacetoxy-4-bromobenzene (6a). To a stirred solution 4-bromoresorcinol (**2a**) (0.50 g, 2.61 mmol) in THF (10 mL) was added triethylamine (0.74 mL, 5.31 mmol) under nitrogen atmosphere and stirred for 0.5 h at rt. AcCl (0.37 mL, 5.31 mmol) in THF (3 mL) was added slowly to this reaction mixture and stirred for 3 h at rt. The reaction mixture was extracted with EtOAc, washed with brine, dried over anhydrous MgSO₄, concentrated *in vacuo*, and purified by silica gel flash column chromatography (EtOAc/hexane = 1/10) to give a clean liquid. Yield: 0.69 g (98%); R_f 0.44 (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, CDCl₃) & 7.57 (1H, d, *J* = 8.4 Hz), 6.97 (1H, d, *J* = 0.9 Hz), 6.92 (1H, dd, *J* = 8.4, 0.9 Hz), 2.35 (3H, s), 2.29 (3H, s); ¹³C NMR (75 MHz, CDCl₃) & 168.51, 167.96, 150.05, 148.36, 133.20, 120.55, 117.49, 112.74, 21.17, 20.87.

1-(2,4-Diacetoxyphenyl)-2-(3,5-dimethoxyphenyl)acetylene (8). To 6a (0.68 g, 2.53 mmol) were added Pd(OAc)₂ (30 mg, 0.12 mmol), CuI (30 mg, 0.12 mmol), P([']Bu)₃-HBF₄ (40 mg, 0.19 mmol) and 1-ethynyl-3,5-dimethoxybenzene 7 (0.52 g, 3.22 mmol) under nitrogen atmosphere and stirred for 3 d at 40 °C. The reaction mixture was dissolved with ether, filtered with Celite[®] 545, concentrated *in vacuo*, extracted with ether, washed with brine, dried over MgSO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give the product as white solid. Yield: 0.57 g (63%); R_f 0.29 (EtOAc/hexane = 1/3); mp 178-180 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.54 (1H, d, J = 8.4 Hz), 7.01 (1H, dd, J = 8.4, 2.4 Hz), 6.96 (1H, d, J = 2.4 Hz), 6.62 (2H, d, J = 2.7 Hz), 6.46 (1H, t, J = 2.7 Hz), 3.79 (6H, s), 2.36 (3H, s), 2.30 (3H, s); ¹³C NMR (75 MHz, CDCl₃) δ 168.50, 168.90, 151.84, 150.75, 133.15, 123.94, 119.17, 116.11, 114.72, 109.30, 101.79, 94.09, 83.10, 55.47, 21.14, 20.93.

2-(3,5-Dimethoxyphenyl)-6-hydroxybenzofuran (9). To a stirred solution of **8** (0.51 g, 1.44 mmol) in MeOH (5 mL) was added slowly KOH (0.28 g, 5.04 mmol) in H₂O (2 mL) under nitrogen atmosphere and refluxed for 5 h. The reaction was quenched with 1 N HCl, concentrated *in vacuo*, extracted with CH₂Cl₂, washed with brine, dried over Na₂SO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give a yellow solid. Yield: 0.21 g (54%); R_f 0.39 (EtOAc/hexane = 1/2); mp 218-221 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.38 (1H, d, *J* = 8.3 Hz), 7.00 (1H, d, *J* = 1.8 Hz), 6.95 (2H, d, *J* = 2.4 Hz), 6.92 (1H, s), 6.77 (1H, dd, *J* = 2.4, 8.3 Hz), 6.43 (1H, t, *J* = 1.8 Hz), 3.86 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 170.96, 161.03, 156.94, 153.14, 135.36, 133.21, 132.36, 109.72, 106.54, 103.67, 100.96 99.10, 55.66.

6-Hydroxy-2-(3,5-dihydroxyphenyl)benzofuran; Moracin M (1). Procedure followed as reported.¹³ Yield: (25%); R_f 0.53 (EtOAc/hexane = 1/1); mp 260-263 °C (lit.¹⁰ mp 259-262 °C; lit.;^{11a} ¹H NMR (300 MHz, methanol-*d*₄) δ 7.30 (1H, d, J = 8.4 Hz), 7.09 (1H, s), 6.95 (1H, d, J = 2.4 Hz), 6.68 (2H, d, J = 2.1 Hz); 6.55 (1H, dd, J = 8.4, 2.4 Hz), 6.22 (1H, t, J = 2.1 Hz); ¹³C NMR (75 MHz, methanol-*d*₄) δ 160.1, 157.4, 157.0, 156.3, 134.0, 123.2, 122.2, 113.4, 104.1, 103.7, 102.4, 98.6. HRMS (EI): m/z [M]⁺ calcd for C₁₄H₁₀O₄: 242.0579; found: 242.0580.

2-(3,5-Dimethoxyphenyl)-6-(1,1-dimethylprop-2-ynyloxy)benzofuran (10). To 9 (75 mg, 0.31 mmol) was added diisopropylethylamine (0.07 mL, 0.40 mmol) and stirred for 0.5 h at 0 °C. Cupper (II) chloride dihydrate (0.50 mg, 0.003 mmol) and 3-chloro-3-methylbutyne (0.04 mL, 0.34 mmol) was added slowly to this reaction mixture and stirred for 5 h at 0 °C. The reaction mixture was concentrated in vacuo, extracted with CH₂Cl₂, washed with brine, dried over MgSO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/8) to give a brown liquid. Yield: 81 mg (78%); $R_f 0.60$ (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, $CDCl_3$) δ 7.71 (1H, d, J = 9.3 Hz), 7.24 (1H, s), 6.87 (1H, d, J = 2.1 Hz), 6.62 (1H, dd, J = 9.3, 2.1 Hz), 6.40 (2H, d, J =1.5 Hz), 6.35 (1H, t, J = 1.5 Hz), 3.74 (6H, s), 2.66 (1H, s), 1.71 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 164.75, 162.52, 160.76, 136.37, 131.49, 113.56, 111.13, 110.99, 107.35, 106.31, 98.82, 95.54, 84.53, 75.16, 72.31, 55.36.

2-(3,5-Dimethoxyphenyl)-6-(1,1-dimethylprop-2-enyloxy)benzofuran (11). To **10** (80 mg, 0.24 mmol) were added 5% mol Pd-CaCO₃ (5 mg), quinoline (1 mL, 0.01 mmol) in EtOH (2 mL) under hydrogen atmosphere and stirred for 4 h at rt. The reaction mixture was dissolved with MeOH, filtered with Celite[®] 545, concentrated *in vacuo*, extracted with CH₂Cl₂, dried over Na₂SO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give a brown liquid. Yield: 77 mg (96%); R_f 0.71 (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, CDCl₃) δ 7.65 (1H, d, *J* = 8.4 Hz), 7.24 (1H, s), 6.53 (1H, d, *J* = 2.4 Hz), 6.44 (1H, dd, *J* = 8.4, 2.4 Hz), 6.40 (2H, d, *J* = 1.8 Hz), 6.35 (1H, d, *J* = 1.65 Hz), 5.20 (1H, d, *J* = 10.5 Hz), 3.76 (6H, s), 1.26 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 165.08, 163.87, 161.05, 159.25, 143.55, 136.78, 131.71, 114.50, 113.32, 111.65, 107.65, 106.54, 99.11, 81.12, 55.67, 27.63.

6-Hydroxy-2-(3,5-dimethoxyphenyl)-5-(3-methylbut-2enyl)benzofuran (12). 11 (70 mg, 0.23 mmol) was dissolved in EtOH-H₂O (4:1 v/v, 4 mL) in a 35-mL Ace pressure tube with PTFE bushes and an FETFE O-ring seal. The tube was then placed in an electric furnace at 80 °C for 8 h. When the tube had cooled to rt, the screw-cap was loosened. The reaction mixture was concentrated in vacuo, extracted with EtOAc-acetone (2:1), dried over MgSO₄ and purified by silica gel flash column chromatography (Acetone/hexane = 1/2) to give a yellow-green solid. Yield: 32 mg (47%); R_f 0.21 (EtOAc/hexane = 1/2); mp 232-234 °C; ¹H NMR (300 MHz, CDCl3) & 7.15 (1H, s), 6.91 (1H, s), 6.84 (1H, s), 6.59 (2H, d, J = 2.1 Hz), 6.31 (1H, t, J = 2.1 Hz), 1.61 (1H, s);¹³C NMR (75 MHz, CDCl₃) δ 162.53, 155.49, 155.00, 154.83, 131.11, 129.31, 125.44, 123.47, 122.82, 121.83, 105.43, 103.25, 99.80, 55.63, 27.00, 25.49, 17.64.

6-Hydroxy-2-(3,5-dihydroxyphenyl)-5-(3-methylbut-2envl)benzofuran (13); Moracin N. To a stirred solution of 12 (21 mg, 0.06 mmol) in CH_2Cl_2 (1 mL) was added slowly BBr₃ solution (0.25 mL, 0.25 mmol) (1 M in CH₂Cl₂) under nitrogen atmosphere and stirred for 1 h at -78 °C. Then the mixture was stirred for 19 h at 0 °C. MeOH was added to the reaction mixture and concentrated in vacuo, extracted with CH₂Cl₂, washed with brine, dried over Na₂SO₄ and evaporated to dryness. The crude was recrystallized from EtOAchexane to give the product as a brown solid.²⁰ Yield: 4 mg (21%); $R_f 0.57$ (EtOAc/hexane = 1/1); mp 220-221 °C (lit.^{20b} mp 187-188 °C); ¹H NMR (300 MHz, Methanol-d₄) δ 7.24 (1H, s), 7.05 (2H, s), 6.79 (2H, d, J = 2.1 Hz), 6.41 (1H, t, J)= 2.4 Hz), 5.27 (1H, t, *J* = 6.9 Hz), 3.77 (1H, d, *J* = 7.4 Hz), 1.78 (6H, s); ¹³C NMR (75 MHz, Methanol- d_4) δ 166.65, 156.96, 150.25, 150.13, 134.01, 131.09, 130.55, 129.55, 128.54, 127.37, 105.35, 104.55, 104.18, 100.32, 29.34, 26.13, 18.25. HRMS (EI) calcd for C₁₉H₁₈O₄ M⁺ 310.1205, found 310.1207.

5-Acetoxy-2-bromophenol (6b). Sulfuric acid (1 drop) was added to acetic anhydride (0.5 mL, 5.28 mmol) in one neck flask at rt and then cooled to -20 °C. After stirring for 10 min at -20 °C, 4-bromo resorcinol (**2a**) (1 g, 5.28 mmol) in THF was added and the resulting mixture was stirred for 7 h at this temperature. After completion of the reaction, H₂O was added. The reaction mixture was extracted with EtOAc, washed with brine, dried over anhydrous MgSO₄, concentrated *in vacuo*, and purified by silica gel flash column

Synthesis of Natural Moracins

chromatography (EtOAc/hexane = 1/8) to give a clean liquid. Yield: 0.48 g (40%); R_f 0.31 (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, CDCl₃) δ 7.29 (1H, d, J = 8.4 Hz), 7.07 (1H, dd, J = 8.4, 3 Hz), 6.89 (1H, d, J = 3 Hz), 2.19 (3H, s); ¹³C NMR (75 MHz, CDCl₃) δ 166.31, 150.26, 149.16, 132.37, 118.65, 110.82, 108.14, 21.02.

4-Acetoxy-1-bromo-2-(1,1-dimethylprop-2-ynyloxy)benzene (14). 1,8-Diazabicycloundec-7-ene (0.32 mL, 2.07 mmol) was added to **6b** (0.41 g, 1.71 mmol) and stirred for 0.5 h at 0 °C. Cupper (II) chloride dihydrate (3 mg, 0.02 mmol) and 3-chloro-3-methylbutyne (0.21 mL, 1.81 mmol) was added slowly to this reaction mixture and stirred for 5 h at 0 °C. The reaction mixture was concentrated in vacuo, extracted with CH₂Cl₂, washed with brine, dried over MgSO₄ and purified by silica gel flash column chromatography (EtOAc/ hexane = 1/8) to give a brown liquid. Yield: 0.33 g (65%); R_f 0.75 (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, CDCl₃) δ 7.09 (1H, d, J = 8.4 Hz), 6.87 (1H, dd, J = 8.4, 3 Hz), 6.78 (1H, d, *J* = 3 Hz), 2. 12 (1H, s), 1.78 (3H, s), 1.19 (6H, s); 13 C NMR (75 MHz, CDCl₃) δ 168.40, 157.76, 149.69, 133.93, 119.40, 112.97, 110.23, 84.87, 74.82, 72.45, 29.56, 21.02.

4-Acetoxy-1-bromo-2-(1,1-dimethylprop-2-enyloxy)benzene (15). To **14** (0.30 g, 1.03 mmol) were added 5% mol Pd-CaCO₃ (15 mg) and quinoline (0.01 mL, 0.01 mmol) in EtOH (2 mL) under hydrogen atmosphere and stirred for 4 h at rt. The reaction mixture was dissolved with MeOH, filtered with Celite[®] 545, concentrated *in vacuo*, extracted with CH₂Cl₂, dried over Na₂SO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give a brown liquid. Yield: 0.29 g (75%); R_f 0.81 (EtOAc/hexane = 1/3); ¹H NMR (300 MHz, CDCl₃) δ 7.29 (1H, d, *J* = 8.7 Hz), 7.00 (1H, dd, *J* = 8.7, 3 Hz), 6.65 (1H, d, *J* = 3 Hz), 6.08 (1H, dd, *J* = 10.8, 10.8 Hz), 4.99 (2H, dd, *J* = 10.8, 10.8 Hz), 2.32 (3H, s), 1.28 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 168.40, 157.65, 149.88, 143.41, 134.11, 119.41, 114.42, 111.14, 107.74, 81.17, 27.07, 21.02.

1-(4-Acetoxy-2-(1,1-dimethylprop-2-enyloxy)phenyl)-2-(3,5-dimethoxyphenyl)acetylene (16). To a stirred solution of 15 (0.25 g, 0.83 mmol) in diisopropylamine (5 mL) were added slowly Pd(OAc)₂ (10 mg, 0.41 mmol), CuI (8 mg, 0.41 mmol), P('Bu)₃-HBF₄ (12 mg, 0.06 mmol) and 1ethynyl-3,5-dimethoxybenzene 7 (0.18 g, 1.08 mmol) under nitrogen atmosphere and stirred for 3 d at 40 °C. The reaction mixture was dissolved with ether, filtered with Celite® 545, concentrated in vacuo at 80 °C, extracted with ether, washed with brine, dried over MgSO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give a white solid. Yield: 0.18 g (56%); R_f 0.43 (EtOAc/ hexane = 1/3); mp 197-198 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.25 (1H, d, J = 8.9 Hz), 6.82 (1H, dd, J = 8.9, 2.8 Hz), 6.74 (1H, d, *J* = 2.8 Hz), 6.69 (2H, d, *J* = 2.3 Hz), 6.36 (1H, t, J = 2.3 Hz), 6.03 (1H, dd, J = 10.4, 10.4 Hz), 4.91 (2H, dd, *J* = 10.4, 10.4 Hz), 3.43 (6H, s), 2.24 (3H, s), 1.38 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 168.40, 163.97, 163.04, 149.72, 143.41, 134.76, 127.14, 117.37, 115.37, 114.42, 110.10, 108.02, 100.93, 98.38, 92.63, 82.43, 55.31, 27.53, 21.78.

Bull. Korean Chem. Soc. 2014, Vol. 35, No. 12 3457

6-Acetoxy-2-(3,5-dimethoxyphenyl)-7-(3-methylbut-2enyl)benzofuran (17). 16 (0.17 g, 0.47 mmol) was dissolved in EtOH-H₂O (4:1 v/v, 4 mL) in a 35-mL Ace pressure tube with PTFE bushes and an FETFE O-ring seal. The tube was then placed in an electric furnace at 100 °C for 9 h. When the tube had cooled to rt, the screw-cap was loosened. The reaction mixture was concentrated in vacuo, extracted with EtOAc-acetone (2:1), dried over MgSO₄ and purified by silica gel flash column chromatography (Acetone/ hexane = 1/2) to give a green solid product. Yield: 55 mg (31%); $R_f 0.61$ (EtOAc/hexane = 1/2); mp 195-198 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.35 (1H, d, J = 8.7 Hz), 7.29 (1H, s), 7.06 (1H, d, J = 2.7 Hz), 6.37 (2H, d, J = 2.4 Hz), 6.30 (1H, t, J = 2.4 Hz), 5.24 (1H, m), 3.64 (6H, s), 3.51 (2H, d, J = 7.4 Hz), 2.35 (3H, s), 1.84 (6H, s); ¹³C NMR (75 MHz, CDCl₃) & 169.07, 161.33, 155.78, 154.53, 153.55, 130.71, 128.65, 125.21, 124.12, 121.25, 119.36, 112.32, 105.37, 105.11, 99.83, 55.67, 25.53, 20.11, 20.02, 18.36.

6-Hydroxy-2-(3,5-dimethoxyphenyl)-7-(3-methylbut-2enyl)benzofuran (18). To a stirred solution of 17 (55 mg, 0.14 mmol) in MeOH (5 mL) was added slowly KOH (16 mg, 0.29 mmol) in H₂O (2 mL) under nitrogen atmosphere and refluxed for 5 h. The reaction was quenched with 1N HCl, concentrated in vacuo, extracted with CH₂Cl₂, washed with brine, dried over Na₂SO₄ and purified by silica gel flash column chromatography (EtOAc/hexane = 1/6) to give a vellow solid. Yield: 25 mg (51%); $R_f 0.84$ (EtOAc/hexane = 1/1); mp 242-244 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.31 (1H, d, *J* = 8.9 Hz), 7.23 (1H, s), 6.82 (1H, d, *J* = 8.9 Hz), 6.75 (2H, d, *J* = 2.4 Hz), 6.41 (1H, t, *J* = 2.4 Hz), 5.22 (1H, m), 3.41 (6H, s), 3.35 (2H, d, J = 7.1 Hz), 1.72 (6H, s); ¹³C NMR (75 MHz, CDCl₃) δ 172.52, 156.22, 155.64, 150.65, 135.29, 128.51, 122.21, 121.42, 119.26, 117.32, 108.12, 105.21, 103.67, 97.51, 53.55, 25.60, 20.73, 17.47.

6-Hydroxy-2-(3,5-dihydroxyphenyl)-7-(3-methylbut-2enyl)benzofuran (19); Moracin S. To a stirred solution of 18 (20 mg, 0.059 mmol) in CH₂Cl₂(1 mL) was added slowly BBr₃ solution (0.23 mL, 0.23 mmol) (1 M in CH₂Cl₂) under nitrogen atmosphere and stirred for 1 h at -78 °C. Then the mixture was stirred for 19 h at 0 °C. MeOH was added to the reaction and concentrated in vacuo, extracted with CH₂Cl₂, washed with brine, dried over Na₂SO₄ and evaporated to dryness. The crude product was recrystallized from EtOAchexane to give the product as orange solid. Yield: 4.7 mg (26%); $R_f 0.63$ (EtOAc/hexane = 1/1); mp 217-220 °C.^{1b}; ¹H NMR (300 MHz, methanol- d_4) δ 7.27 (1H, d, J = 8.4 Hz), 7.12 (1H, s), 6.85 (1H, d, J = 8.4 Hz), 6.72 (2H, d, J = 2.1 Hz), 6.39 (1H, t, J = 2.1 Hz), 5.05 (1H, m), 3.66 (2H, d, J =7.2 Hz), 1.90 (6H, s); ¹³C NMR (75 MHz, methanol- d_4) δ 160.12, 156.11, 155.02, 152.21, 133.41, 133.11, 124.43, 123.71, 119.01, 113.34, 113.12, 104.01, 103.74, 103.18, 26.54, 24.01, 17.96. HRMS (EI) calcd for $C_{19}H_{18}O_4$ M⁺ 310.1205, found 310.1207.

2,4-Dihydroxy-5-iodobenzaldehyde (4a). To a stirred solution of 2,4-dihydroxybenzaldehyde (**20**) (1.50 g, 10.9 mmol) in AcOH (7.5 mL) was added slowly ICl (17.32 mL, 17.31 mmol) in CH_2Cl_2 under nitrogen atmosphere and

3458 Bull. Korean Chem. Soc. 2014, Vol. 35, No. 12

stirred for 5 h at rt. The reaction was quenched with sodium thiosulfate-H₂O (4:1) resulted solid product and the solid was filtered and washed with water resulted the product as brown solid. Yield: 2.87 g (51%); R_f 0.63 (EtOAc/hexane = 1/2); mp 167-170 °C; ¹H NMR (300 MHz, acetone- d_6) δ 9.67 (1H, s), 7.95 (1H, s), 6.36 (1H, s); ¹³C NMR (75 MHz, acetone- d_6) δ 193.02, 157.65, 156.77, 143.92, 117.75, 102.05, 70.61.

6-Hydroxy-2-(3-hydroxy-5-(tert-butyldimethylsilyl)oxyphenyl)benzofuran-5carbaldehyde (5a). To a stirred solution of 4a (0.1 g, 0.37 mmol) in DMF (2 mL) were added slowly PdCl₂(PPh₃)₂ (13 mg, 0.02 mmol), CuI (1.4 mg, 0.01 mmol), TEA (0.10 mL, 0.76 mmol) and ((5-ethynyl-1,3phenylene)bis(oxy))bis(tert-butyldimethylsilane) 7 (0.52 g, 3.22 mmol) under nitrogen atmosphere and stirred for 2 d at 70 °C. The reaction mixture was dissolved with ether, filtered with Celite[®] 545, concentrated in vacuo at 80 °C, extracted with ether, washed with brine, dried over MgSO₄. The crude product was recrystallized from acetone-hexane and brown solid. Yield: 73 mg (50%); Rf 0.38 (EtOAc/hexane = 1/3); mp 246-248 °C; ¹H NMR (300 MHz, acetone- d_6) δ 10.99 (1H, s), 9.79 (1H, s), 7.78 (1H, s), 7.03 (1H, s), 6.83 (1H, s), 6.68 (2H, d, *J* = 1.8 Hz), 6.17 (1H, t, *J* = 1.8 Hz), $0.78 (18H, s), 0.03 (12H, s); {}^{13}C NMR (75 MHz, acetone-d_6)$ δ 196.88, 160.36, 159.67, 157.40, 156.92, 131.60, 127.80, 123.05, 118.85, 108.24, 108.10, 105.37, 101.92, 98.87, 30.09, 25.52. - 4.70.

6-Hydroxy-2-(3,5-dihydroxyphenyl)benzofuran-5carbaldehyde (21); Moracin Y. To a stirred solution of 5a (34 mg, 0.07 mmol) in THF (1 mL) was added slowly TBAF (0.1 mL, 0.28 mmol) under nitrogen atmosphere and was stirred for 5 h at rt. After completion of the reaction, solvent was removed in vacuo, Acetone was added to the crude and filtered the solid TBAF. The filtrate was concentrated and addition of ether-MeOH to the crude offered the solid product moracin Y as yellow solid.^{1a} Yield: 17 mg (90%); R_f 0.32 (EtOAc/hexane = 1/1); mp 235-238 °C; ¹H NMR (300 MHz, methanol- d_4) δ 10.23 (1H, s), 7.72 (1H, s), 6.85 (1H, s), 6.73 (1H, s), 6.65 (2H, d, J = 2.1 Hz), 6.19 (1H, t, J = 2.1 Hz); ¹³C NMR (75 MHz, Methanol-d₄) δ 193.70, 169.51, 161.71, 159.56, 155.79, 131.88, 122.12, 121.50, 120.03, 103.21, 102.71, 101.08, 100.62. HRMS (EI) calcd for C₁₅H₁₀O₅ M⁺ 270.0528, found 270.0529.

Acknowledgments. This research was financially supported by the Ministry of Education, Science Technology (MEST) and Korea Institute for Advancement of Technology (KIAT) through the Human Resource Training Project for Regional Innovation (2012H1B8A2026036), and by Priority Research Centers Program through the National

Jae Jun Lee et al.

Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (NRF-2009-0094071).

References

- (a) Yang, Y.; Gong, T.; Liu, C.; Chen, R.-Y. *Chem. Pharm. Bull.* 2010, *58*, 257-260. (b) Kapache, G D. W. F.; Fozing, C. D.; Donfack, J. H.; Fotso, G. W.; Amadou, D.; Tchana, A. N.; Bezabih, M.; Moundipa, P. F.; Ngadjui, B. T.; Abegaz, B. M. *Phytochemistry* 2009, *70*, 216-221 and references cited therein.
- Naowaboot, J.; Pannangpetch, P.; Kukongviriyapan, V.; Kongyingyoes, B.; Kukongviriyapan, U. *Plant Food Human Nut.* 2009, 64, 116-121.
- 3. Ahn, K.-S.; Sim, W.-S.; Kim, I.-H. Planta Med. 1996, 62, 7-9.
- Khyade, V. B.; Khyade, V. V.; Khyade, S. V. IOSR J. Environ. Sci. Toxicol. Food Tech. 2013, 4, 96-104.
- Chen, S.-K.; Zhao, P.; Shao, Y.-X..; Li, Z.; Zhang, C.; Liu, P.; He, X.; Luo, H.-B.; Hu, X. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 3261-3264.
- Yadav, A. V.; Kawale, L. A.; Nade, V. S. Indian J. Pharm. 2008, 40, 32-36.
- 7. Aditya, R. S. J.; Ramesh, C. K.; Basavaraj, P.; Jamuna, K. S. *RJPBCS* **2013**, *4*, 822.
- Zeni, A. L. B.; Dall'Molin, M. Rev. Bras. Farmacogn. Braz. J. Pharm. 2010, 20, 130-133.
- Yang, Z.-G.; Matsuzaki, K.; Takamatsu, S.; Kitanaka, S. *Molecules* 2011, 16, 6010-6022.
- Deshpande, V. H.; Spinivasan, R.; Rama Rao, A. V. Indian J. Chem. 1975, 13, 453-457.
- (a) Clough, J. M.; Mann, I. S.; Widdowson, D. A. *Tetrahedron Lett.* **1987**, *28*, 2645-2648. (b) Mann, I. S.; Widdowson, D. A. Clough, J. M. *Tetrahedron* **1991**, *47*, 7981-7990.
- Watanabe, M.; Kawanishi, K.; Furukawa, S. Chem. Pharm. Bull. 1991, 39, 579-583.
- 13. Arias, L.; Vara, Y.; Cossio, F. P. J. Org. Chem. 2012, 77, 266-275.
- 14. (a) Kinoshita, T.; Ichinose, K. *Heterocycles* 2005, *65*, 1641-1654.
 (b) Celaje, J. A.; Zhang, D.; Guerrero, A. M.; Selke, M. *Org. Lett.* 2011, *13*, 4846-4849.
- (a) Hiroya, K.; Suzuki, N.; Yasuhara, A.; Egawa, Y.; Kasano, A.; Sakamoto, T. *J. Chem. Soc. Perkin Trans. 1* **2000**, 4339-4346. (b) Yue, D.; Yao, T.; Larock, R. *J. Org. Chem.* **2005**, *70*, 10292-10296.
 (c) Carril, M.; Correa, A.; Bolm, C. *Angew. Chem. Int. Ed.* **2008**, *47*, 4862-4865. (d) Bang, H. B.; Han, S. Y.; Choi, D. H.; Hwang, J. W.; Jun, J.-G. *ARKIVOC* **2009**, *(ii)*, 112-125.
- 16. Netherton, M. R.; Fu, G. C. Org. Lett. 2001, 3, 4295-4298.
- 17. Novak, Z.; Timari, G.; Kotschy, A. *Tetrahedron* **2003**, *59*, 7509-7513.
- Campeau, L.-C.; Rousseaux, S.; Fagnou, K. J. Am. Chem. Soc. 2005, 127, 18020-18021.
- 19. Tolman, C. A. Chem. Rev. 1977, 77, 313-348.
- (a) Matsuyama, S.; Kuwahara, Y.; Suzuki, T. Agric. Biol. Chem. 1991, 55, 1409-1410. (b) Matsuyama, S.; Kuwahara, Y.; Nakamura, S.; Suzuki, T. Agric. Biol. Chem. 1991, 55, 1339-1341.
- 21. Yang, Z.; Wang, Y.; Wang, Y.; Zang, Y. Food Chem. 2012, 131, 617-625.
- Royer, M.; Rodrigues, A. M. S.; Herbette, G.; Beauchene, J.; Chevalier, M.; Herault, B.; Thibaut, B.; Stien, D. Int. Biodeterior: Biodegrad. 2012, 70, 55-59.
- 23. Jeon, J.-H.; Kim, M. R.; Jun, J.-G. Synthesis 2011, 43, 370-376.