

## Effects of Bacterial Inoculants and Cutting Height on Fermentation Quality of Barley Silage

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### ABSTRACT

This study was conducted to investigate the effects of bacterial inoculation (*Lactobacillus plantarum*) and cutting height on the chemical composition, fermentation characteristics and in vitro dry matter digestibility (IVDMD) in whole crop barley silage. Barley forage (Youngyang hybrid) was harvested at about 27% of dry matter (DM) level at two different cutting height (5 vs. 15 cm). And it was chopped to 5 cm length and treated with or without *L. plantarum*. Four replicates of each treatment were ensiled into 10 L mini silo (3 kg) for 100 days. After 100 days, bacterial inoculation decreased ( $p=0.001$ ) DM content, while increased cutting height increased ( $p=0.002$ ) DM in uninoculated silage. Crude protein (CP) concentration was decreased by increasing height in uninoculated silage (8.84 vs. 8.16) but increased in inoculated silage (8.19 vs. 8.99). Both neutral detergent fiber (NDF) ( $p<0.011$ ) and acid detergent fiber (ADF) ( $p<0.004$ ) were decreased by increasing cutting height of forage at harvest. The IVDMD and ammonia-N was increased ( $p=0.001$ ) by increasing cutting height and inoculation, respectively. Lactic acid bacteria (LAB) was increased ( $p=0.002$ ) in inoculated silage, but yeast count was decreased ( $p=0.026$ ) in uninoculated silages. It is concluded that increased cutting height of forage at harvest could be useful to make a fibrous portion with increase of dry matter digestibility of silages. (**Key words** : Barley, Cutting height, *Lactobacillus plantarum*, Silage)

### I . INTRODUCTION

The forage is indispensable resources for normal development of the rumen in ruminants. High quality forage is known as a key resource to improve the animal productivity (Kim et al., 2012). Oba and Allen (1999) reported that increased neutral detergent fiber (NDF) digestibility improved dry matter (DM) intake and 4% fat-corrected milk. Also high quality forage can replace some parts of concentrate from mixed feed (Seo et al., 2010).

In South Korea, more than 80% of demanded forage is producing in domestic, and the ensiling is the main procedure for forage preservation (McDonald et al., 1991). Barley is widely used to make silage in Korea for its high proportion of rumen-fermentable carbohydrates (Eun et al., 2004). Therefore, improving the nutritive quality and digestibility of barley silage has considerable importance for beef and dairy producers.

Usually extended wilting time lead to decreases of silage quality (Kim and Adesogan, 2006). In Korea, barley forage usually wilted for one or two days to adjust moisture content. However, it is hard to wilt quickly due to frequent rain during harvesting season. With this reason, increases of cutting height may increase the aeration at the bottom of harvested forages layed down at the field, which could help to increase silage quality by decreased wilting time. Moreover, increasing cutting height from the ground helps to improve fiber digestibility (Tolera and Sundstøl, 1999). Wu and Roth (2005) also reported that increase in cutting height of corn at harvest, crude protein (CP) concentration and NDF digestibility was improved in following silage.

Using bacterial inoculants is another approach to improve the silage fermentation and nutrient digestibility. Numerous precedence studies showed that bacterial inoculants are used in silage to stimulate lactic fermentation, decreasing pH and thereby improving silage preservation (Seale, 1986; Filya et

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al., 2000; Baah et al., 2011). Considerable studies were carried out to determine the effect of bacterial inoculants or cutting height separately, however, the effect of bacterial inoculants and cutting height together on silage quality was not studied.

Therefore, this study was carried out to determine the effect of applying inoculants in barley forage harvested at different cutting height on nutritive value and fermentation quality of barley silage.

## II. MATERIALS AND METHODS

### 1. Preparation of silage

Barley forage (Youngyang hybrid) was grown at the animal research unit, Gyeongsang National University, South Korea and harvested at about 27% of DM level at two different cutting height (5 vs. 15 cm). Forage was chopped by conventional forage harvester (BHC-90, BUHEUNG Machinery Ind Co., Korea) to 5 cm theoretical cut and treated forage from both cutting height with or without inoculants. The inoculants (Chungmi-Lacto, CMBio) applied at 0.02 g/kg of fresh forage to supply  $1.2 \times 10^3$  cfu/g of *L. plantarum*. Four replicates of each treatment were ensiled into 10 L mini silo (3 kg) for 100 days at room temperature in an enclosed barn.

### 2. Laboratory analysis

At the day of silage making and at silo opening, fresh treated forage and silage (500 g), respectively, were subsampled for chemical analysis. A portion (20 g) of silage was macerated in laboratory blender along with 200 ml of distilled water to make silage extract, which was used for pH, ammonia-N and volatile fatty acid (VFA) determination, microbial enumeration and microbial DNA extraction. Samples were dried at 60°C for 48 h, ground to pass 1-mm screen of a grinder (Cutting Mill, SHINMYUNG ELECTRIC Co., Ltd, South Korea). Dry matter was determined in dry oven at 100°C for 24 h. The nitrogen and ether extract (EE) contents were determined by Kjeldahl procedure using N analyzer (B-324, 412, 435 and 719 S Titrino, BUCHI, Germany) and Soxhlet procedure (AOAC, 1990), respectively.

Concentrations of NDF and acid detergent fiber (ADF) were analyzed as described by Van Soest et al. (1991) using an AnKom Fiber Analyzer (A220, AnKom Technology). Alfa-amylase and sodium sulfite were used for NDF determination. *In vitro* DM digestibility (IVDMD) was measured by the method of Tilley and Terry (1963) using ANKOM DAISY<sup>II</sup> incubator (ANKOM Technology, USA). The pH was measured with electric pH meter (SevenEasy, Mettler Toledo, Switzerland) and ammonia-N was analyzed by colorimetry (Chaney and Marbach, 1962). The concentrations of VFA were measured using HPLC (L-2130, HITACHI) with an auto-sampler (L-2200, HITACHI), UV detector (L-2400, HITACHI) and a column (MetaCarb 87H, Varian) by the method of Muck and Dickerson (1988).

### 3. Microbiological count

The counts of Lactic acid bacteria, yeast and mold were carried out with using silage extract, considered it as the first dilution. The lactobacilli MRS agar medium (MRS; Difco, Detroit, MI, USA) used for the isolation and count of lactic acid bacteria, and potato dextrose agar medium (PDA; Difco, Detroit, MI, USA) used for the isolation and count of yeasts and molds. Lactobacilli MRS agar mediums was placed in a CO<sub>2</sub> incubator (Thermo Scientific, USA) at 39°C for 24 h and PDA mediums was incubated at 39°C for 24 h in normal incubator (Johnsam Corporation, Korea). Visible colonies were counted from the agar medium at appropriate dilutions and the number of colony forming units (CFU) was expressed per gram of silage.

### 4. DNA Extraction, primers and PCR conditions

Extraction of genomic DNA was conducted by using QIAamp DNA mini kit (Qiagen, USA) following the manufacturer's protocol and concentrations were measured by using a NanoDrop Spectrophotometer (ND-1000, USA). A Bio-Rad C1000 Touch<sup>TM</sup> Thermal cycler real-time PCR detection system (CFX96<sup>TM</sup> Real-Time system, Bio-Rad Laboratories, Inc., Hercules, CA, USA) was used to amplify DNA. Amplification conditions and primers for *L. plantarum* DNA were used as in Amanullah et al. (2014). The amplified fragments were subjected to electrophoresis

(Mupid<sup>®</sup>-2plus Submarine-type electrophoresis system, TaKaTa, Inc., Japan) on 1.5% agarose gel and visualized after staining with ethidium bromide under UV illumination (Molecular Imager Gel Doc<sup>TM</sup> XR+ Imaging System, Bio-Rad Laboratories, Inc., Hercules, CA, USA).

## 5. Statistical Analyses

This experiment was conducted in completely randomized design with a 2 × 2 factorial arrangement. The data were analyzed using GLM procedure of SAS (2002), with a model containing cutting height (5 vs. 15 cm), inoculant (no inoculant vs. inoculant) and interactions between them to analyze the data. The significance was declared at  $p < 0.05$ .

## III. RESULTS AND DISCUSSION

The chemical composition of the barley (Youngyang) forage before ensiling was shown in Table 1. The forage of 5 and 15 cm cutting heights had DM, CP, NDF and ADF contents of 26.5 vs. 28.0%; 9.0 vs. 8.1%; 54.6 vs. 55.5%; 36.5 vs. 34.9%, respectively. Barley forage cut at 5 cm had numerically higher CP and ADF concentrations than at 15 cm, but lower DM and NDF concentrations. The chemical compositions of barley forage used in this study were similar to the other studies (Kim et al., 2013; Amanullah et al., 2014). Generally, the lower part of the

Table 1. Chemical composition of barley silage (Youngyang) before ensiling (% DM)

	Cutting height <sup>1)</sup> , cm	
	5	15
DM <sup>2)</sup>	26.5	28.0
CP	9.00	8.12
EE	2.74	3.35
Curde ash	6.84	7.32
NDF	54.6	55.5
ADF	36.5	34.9
Hemicellulose	18.6	20.5

<sup>1)</sup> Forage were harvested at 5 and 15cm height from root, respectively.

<sup>2)</sup> DM, dry matter; CP, crude protein; EE, ether extract; NDF, neutral detergent fiber; ADF, acid detergent fiber.

forage is considered to be highly lignified and poorly digestible (Tolera and Sundstøl, 1999). Bernard et al. (2004) reported that increase in cutting height reduce the ADF concentration. The chemical composition of barley silage after 100 days of ensiling were described in Table 2. Bacterial inoculation decreased ( $p=0.001$ ) DM content, while increasing cutting height increased ( $p=0.002$ ) DM content only in uninoculated silage. Interaction effect ( $p=0.002$ ) of cutting height and inoculation was observed in silage DM content. Neither inoculant, nor cutting height does affected ( $p > 0.05$ ) CP content, however, interaction of inoculant and cutting height was observed ( $p < 0.001$ ). It was decreased by increasing cutting height in uninoculated silage (8.84 vs. 8.16), but increased in inoculated silage (8.19 vs 8.99). Both NDF ( $p=0.011$ ) and ADF ( $p=0.004$ ) in silage were decreased by increasing cutting height. These results are in agreement with several earlier studies, where it was also reported that fiber content (NDF and/or ADF) of silages decreased by increasing cutting height of forage at harvest (Neylon and Kung, 2003; Kennington et al., 2005; Kung et al., 2008). As the lower part of the forage plants is more fibrous (Tolera and Sundstøl, 1999), so leaving some portion of that part at harvest resulting to lower down the fiber content of silage.

*In vitro* DM digestibility and fermentation characteristics in barley silage (Youngyang) at 100 days of ensiling are presented in Table 3. The IVDMD was increased (52.9 vs. 47.7%;  $p=0.001$ ) with the increase of cutting height. Increasing cutting height at harvest helps to improve digestibility of forage or silage by reducing the lignified lower part of the plant (Wu and Roth et al., 2005). The fiber of lower part of the plants is usually lignified and less digestible (Tolera and Sundstøl, 1999). In this study, it was also observed a significant reduction in fiber content (ADF, NDF) in silages of higher cutting height. Kennington et al. (2005) also observed increased DM digestibility in higher cut silage upto 24 h of incubation. The pH was neither affected by inoculant, nor by cutting height ( $p > 0.05$ ) and was ranged from 4.75 to 4.78. Usually, bacterial inoculation with homolactic fermenters like *L. plantarum* helps to reduce silage pH, which effect was not observed here. The silage pH is related with organic acid production, especially of lactic acid by epiphytic and inoculated bacteria

Table 2. Chemical composition of barley silage (Youngyang) ensiled for 100 d (% , DM)

	No inoculant		Inoculant		SEM	Contrast <sup>2)</sup>		
	5 <sup>1)</sup>	15	5	15		Ino	Height	Ino*Height
DM <sup>3)</sup>	23.8	28.8	22.7	23.3	0.912	0.001	0.002	0.002
CP	8.84	8.16	8.19	8.99	0.222	0.827	0.080	0.000
EE	3.54	3.34	3.21	3.97	0.678	0.742	0.322	0.259
Curde ash	2.75	2.33	2.37	2.20	0.191	0.022	0.034	0.299
NDF	47.4	40.7	50.0	45.3	2.638	0.089	0.011	0.578
ADF	32.7	24.9	34.0a	27.0	3.084	0.832	0.004	0.838
Hemicellulose	16.3	15.8	15.2	18.3	2.707	0.701	0.274	0.271

<sup>1)</sup> Forage were harvested at 5 and 15 cm height from root, respectively.

<sup>2)</sup> Ino = inoculant effect; Height = cutting height effect; Ino\*Height = interaction effect between inoculant and cutting height.

<sup>3)</sup> DM, dry matter; CP, crude protein; EE, ether extract; NDF, neutral detergent fiber; ADF, acid detergent fiber.

Table 3. Fermentation indices of barley silage (Youngyang) ensiled for 100 d (% of DM or as stated)

	No inoculant		Inoculant		SEM	Contrast <sup>2)</sup>		
	5 <sup>1)</sup>	15	5	15		Ino	Height	Ino*Height
IVDMD <sup>3)</sup> , %	46.4	55.7	48.9	50.1	0.907	0.061	0.001	0.001
pH	4.75	4.78	4.76	4.76	0.072	0.984	0.815	0.621
Ammonia-N, %	0.28	0.22	0.32	0.33	0.022	0.001	0.265	0.042
Lactate, %	3.96	3.02	2.87	4.85	0.674	0.489	0.044	0.005
Acetate, %	4.32	6.76	4.70	4.33	0.827	0.135	0.144	0.022
Propionate, %	0.69	0.49	0.74	0.48	0.104	0.857	0.001	0.255
La:Ac ratio	0.91	0.47	0.54	1.12	0.177	0.059	0.246	0.001

<sup>1)</sup> Forage were harvested at 5 and 15 cm height from root, respectively.

<sup>2)</sup> Ino = inoculant effect; Height = cutting height effect; Ino\*Height = interaction effect between inoculant and cutting height.

<sup>3)</sup> IVDMD, *in vitro* dry matter digestibility.

(Zahiroddini et al., 2004). In this study, overall lactic acid production was not affected by bacterial inoculation. Therefore, the pH of resulting silage remained unaffected. Bacterial inoculation increased the concentration of ammonia-N expressed both in percent of DM and total N ( $p=0.001$ ). The increase of ammonia-N in silage usually indicates proteolysis in silage and considered as loss of protein (Ohshima and McDonald, 1978). However, in this study, CP content of silages was not affected by bacterial inoculation. Though, there was difference in ammonia-N between inoculated and uninoculated silages (0.33 vs. 0.25%;  $p=0.001$ ), but the amount was not significant to incur protein loss in silages. Cutting height affected lactate concentration (3.42 vs. 3.93%;  $p=0.044$ ), and also interacted with bacterial inoculation ( $p=0.005$ ). Propionate concentration was reduced in silage as the cutting height increased (0.72 vs. 0.49%;  $p=0.001$ ). These effects of cutting height on

barley silage are difficult to be discussed due to the limitation of available literature in this regard.

Microbial enumeration of LAB, yeast and mold in barley silage (Youngyang) at 100 days of ensiling are presented in Table 4. The LAB count was increased by both inoculation (7.11 vs. 6.47 log<sub>10</sub> cfu/g;  $p=0.002$ ) and higher cutting height (7.12 vs. 6.46 log<sub>10</sub> cfu/g;  $p=0.002$ ). This increase in LAB might be originated from *L. plantarum* inoculation in that group of silages. Yeast count was observed lower (6.01 vs. 6.83 log<sub>10</sub> cfu/g;  $p=0.026$ ) in uninoculated silage, but increased by increasing cutting height (6.84 vs. 6.01 log<sub>10</sub> cfu/g;  $p=0.017$ ). Numerically, the acetate concentration was higher in uninoculated silages that has greatest antimycotic effect among the short chain fatty acids in the resulting silages (Danner et al., 2003). Mold was neither affected by inoculation nor by cutting height ( $p>0.05$ ).

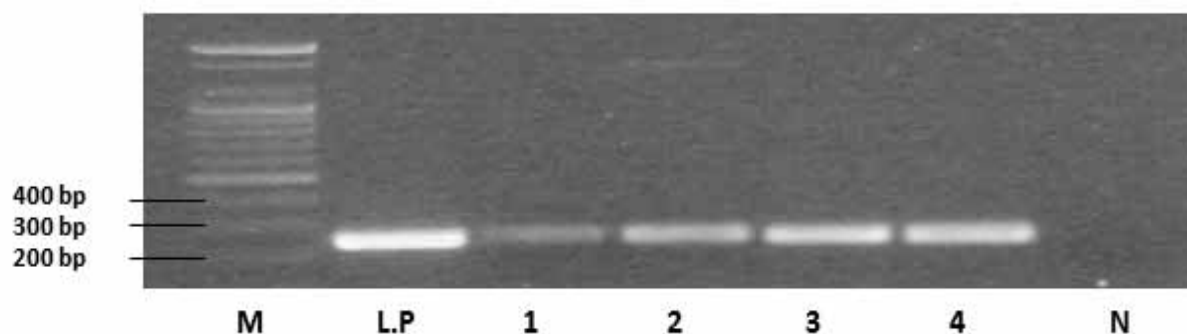
The PCR analysis for *L. plantarum* DNA followed by

Table 4. Microbial enumeration of lactic acid bacteria, yeast and mold in barley silage (Youngyang) ensiled for 100 d (log<sub>10</sub> cfu/g)

	No inoculant		Inoculant		SEM	Contrast <sup>2)</sup>		
	5 <sup>1)</sup>	15	5	15		Ino	Height	Ino*Height
Lactic acid bacteria	6.16	6.77	6.75	7.47	0.277	0.002	0.002	0.739
Yeast	5.79	6.23	6.22	7.44	0.568	0.026	0.017	0.263
Mold	4.30	4.07	4.33	4.98	0.452	0.145	0.382	0.155

<sup>1)</sup> Forage were harvested at 5 and 15cm height from root, respectively.

<sup>2)</sup> Ino = inoculant effect; Height = cutting height effect; Ino\*Height = interaction effect between inoculant and cutting height.



	Pure culture	No inoculant		Inoculant	
		5	15	5	15
Lanes	L.P	1	2	3	4
Band mass, ng	2049	20.85	94.58	348.2	379.2

Fig. 1. Gel electrophoresis analysis and band mass after PCR amplification of DNA from barley silage (Youngyang) fermented for 100 day without or with inoculants: M = DNA 100 bp size marker; L.P = Pure culture (*L. plantarum*,  $1.2 \times 10^3$  cfu/g); lane 1 to 4 = 5 and 15 cm cutting height without or with inoculant; N = negative control.

gel electrophoresis indicated the *L. plantarum* dominated fermentation in inoculated silages (Fig. 1). The band mass (ng) of DNA extracted from *L. plantarum* indicated that increased numerically by inoculation (364 vs. 58 ng) and cutting height (237 vs. 185 ng).

#### IV. CONCLUSION

It is concluded that increasing cutting height of forage at harvest is helpful to decrease fibrous portion but increase dry matter digestibility of silages. Bacterial inoculation did not bring any significant change in silage fermentation in this study. In some cases, the inoculated strain of bacteria may have some sort of antagonism and/or absence of synergism with the originally existed epiphytic bacteria on the plants. More detail study is needed in future to understand the effect of inoculation on silages of different

cutting height.

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