Efficient Synthesis of Novel 4'-Trifluoromethyl-5'-norcarbocyclic Purine Phosphonic Acid Analogs by Using the Ruppert-Prakash Reaction

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Novel 4'-trifluoromethyl-5'-norcarbocyclic purine phosphonic acid analogs were efficiently synthesized from commercially available 1,3-dihydroxy cyclopentane (5). Trifluoromethylation was successfully performed by using the Ruppert-Prakash reaction. The purine nucleosidic bases were efficiently coupled by using the Mitsunobu reaction. The synthesized adenosine phosphonic acids analogs 13 and 16 were screened for antiviral activity against human immunodeficiency virus-1 (HIV-1). Adenine derivative 13 exhibited significant anti-HIV-1 activity.

Key Words : Anti-HIV agents, 4'-Branched carbocyclic nucleoside, Phosphonic acid nucleosides

Introduction

4'-Branched-5'-norcarbocyclic phosphonic acid analogs, such as 4'-vinyl-cpAP $(1)^1$ and 4'-ethynyl-cpAP (2),² have encouraged the search for novel nucleosides as potential anti-human immunodeficiency virus (HIV) agents among this class of compounds. Molecular modeling studies demonstrated the presence of a hydrophobic 4'-pocket that could accommodate these substitutions and enhance anti-HIV activity.¹

Although monofluorinated³ and gem-difluorinated⁴ nucleosides have been widely studied, only a few trifluoromethylated⁵ nucleosides have been reported, which is probably because of the limitations of existing synthesis methods. The presence of a CF₃ group on the sugar moiety of nucleosides could confer many advantages including increased lipophilicity⁶ and improved chemical and/or enzymatic stability.⁷ In addition, the trifluoromethyl group can enhance the therapeutic properties of bioactive compounds.8 There has been increased interest in introducing a trifluoromethyl group into nucleosides in order to discover new nucleoside derivatives with high antiviral activities. Li et al. (2001) reported the first synthesis of 2'-C-\beta-trifluoromethyl pyrimidine ribonucleoside (3) with the Ruppert-Prakash reagent.⁹ Johnson and Kozak successfully synthesized a 4'-trifluoromethylated nucleoside analog (4) by introducing a CF₃ group into the C-4' position of ribose derivatives (Figure 1).¹⁰

Phosphonate has certain advantages over its phosphate counterpart, as it is metabolically stable because of its phosphorus-carbon bond, which is not susceptible to hydrolytic cleavage.¹¹ The spatial location of the oxygen atom, the β -position from the phosphorus atom in the nucleoside analog, is critical for antiviral activity. The increased antiviral activity conferred by the oxygen atom may be attributed to the increased binding capacity of the phosphonate analog to target enzymes.¹² Moreover, a phosphonate nucleoside does not require the first phosphorylation, which is a crucial step for the activation of nucleosides. This is frequently a limit-



Figure 1. Design rationale of 4'-trifluoromethyl-5'-norcarbocyclic phosphonic acid nucleoside analogs.

ing step in the phosphorylation sequence, which ultimately leads to triphosphates.¹³

Given that 4'-branched nucleoside analogs and 5'-norcarbocyclic nucleoside phosphonate have excellent biological activities, we aimed to synthesize a novel class of nucleosides, including 4'-trifluoromethyl-5'-norcarbocyclic phosphonic acid analogs, in order to identify more effective therapeutics against HIV and to provide analogs for probing the conformational preferences of enzymes associated with the nucleoside kinases of nucleosides and nucleotides.

As depicted in Scheme 1, the target compounds were readily prepared from 1,3-dihydroxy cyclopentane (**5**). Selective monosilylation of diol **5** produced alcohol derivative **6**, which was oxidized to the ketone 7 by using Dess-Martin conditions.¹⁴ Lavaire *et al.* (1996) reported fluoride-induced trifluoromethylation conditions by using the Ruppert-Prakash reagent, in which the *tert*-butyldimethylsilyl (TBDMS) protective group is retained.¹⁵ The ketone 7 was subjected to nucleophilic addition conditions¹⁶ with CF₃SiMe₃/*t*-butyl-





Reagents: i) TBDMSCI, imidazole, CH₂Cl₂, -10 °C; ii) Dess-Martin periodinane, CH₂Cl₂; iii) (a) CF₃SiMe₃, TBAF (cat), THF; (b) Na (cat), MeOH.

Scheme 1. Synthesis of trifluoromethylated cyclopentanol intermediate.



Figure 2. ¹⁹F-1H NOE differences between the proximal hydrogens of 8a and 8b.

ammonium fluoride (TBAF) followed by treatment with NaOMe in MeOH, and to give the cyclopentanols **8a** (34%) and **8b** (35%). The stereochemical assignment of **8a** and **8b** as α and β anomers, respectively, was determined by the ¹⁹F-¹H nuclear Overhauser effects (NOE) experiments. For **8b**, we observed strong NOE signals for ¹H when the ¹⁹F nuclei were irradiated (Figure 2).

The hydroxyl functional group of 8b was treated with diethylphosphonomethyl triflate¹⁷ by using lithium *t*-butoxide to yield the phosphonate analog 9 (Scheme 2). Removal of the silvl protective group of 9 by using TBAF produced the secondary alcohol 10. To synthesize the desired 5'-norcarbocyclic adenosine nucleoside analogs, the cyclopentanol 10 was treated with 6-chloropurine under Mitsunobu conditions¹⁸ [diethyl azodicarboxylate (DEAD) and PPh₃]. Slow addition of DEAD to a mixture of cyclopentanol 10, triphenylphosphine, and the 6-chloropurine in anhydrous tetrahydrofuran (THF) solvent produced a yellow solution, which was stirred for 2 h at -40 °C and further stirred overnight at room temperature to produce the protected 6-chloropurine analog 11 as a single N^9 -regioisomer (UV [MeOH] λ_{max} 264.0 nm).¹⁹ The chlorine group of **11** was then converted to an amine group with methanolic ammonia at 72 °C to produce the corresponding adenine phosphonate derivative 12. Hydrolysis of 12 by treatment with bromotrimethylsilane (TMSBr) in CH₃CN in the presence of 2,6-lutidine produced an adenine phosphonic acid derivative **13** (Scheme 2).²⁰

For the synthesis of guanine analogs, 2-fluoro-6-chloropurine²¹ was condensed with alcohol derivative **10** under similar coupling conditions as those used for the condensation of 6-chloropurine to produce the 2-fluoro-6-chloropurine analog **14** (61% yield). Bubbling ammonia into

OTBDMS OTBDMS FtO FtÓ 69% 8b 81% OH EtO FtÓ EtO 64% F₂Ĉ 10 EtÓ F₃Ĉ 11 62% NH₂ NH₂ 69% FtO EtÓ F₃Ĉ 13 12

Reagents: i) (EtO)₂POCH₂OTf, LiO-*t*-Bu, THF; ii) TBAF, THF; iii) 6chloropurine, DEAD, PPh₃, THF; iv) NH₃/MeOH, 70 °C; v) TMSBr, 2,6lutidine, CH₃CN.

Scheme 2. Synthesis of 5'-norcarbocyclic adenine phosphonic acid.



Reagents: i) 2-fluoro-6-chloropurine, DEAD, PPh₃, THF; ii) iv) NH₃/DME, 70 °C; iii) (a) TMSBr, 2,6-lutidine, CH₃CN; (b) NaOMe, HSCH₂CH₂OH, MeOH.

Scheme 3. Synthesis of 5'-norcarbocyclic guanine phosphonic acid.

compound 14 produced the 2-fluoro-6-aminopurine analog²² 15a and 2-amino-6-chloropurine analog 15b with 12% and 42% yields, respectively. The 2-amino-6-chloropurine derivative 15b was treated with TMSBr and 2,6-lutidine to produce the corresponding phosphonic acid, which was successively treated with sodium methoxide and 2-mer-captoethanol in MeOH resulting in the desired guanine phosphonic acid 16 (Scheme 3).²³

The synthesized nucleoside phosphonic acid analogs 13 and 16 were then evaluated for antiviral activity against

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Table 1. Anti-HIV	activity of synthesized compounds	

Compound No.	anti-HIV EC ₅₀ (µM) ^c	Cytotoxicity $CC_{50} (\mu M)^d$
13	8.4	90
16	39.2	98
\mathbf{AZT}^{a}	0.009	100
\mathbf{PMEA}^{b}	0.54	10

^{*a*}**AZT**: azidothymidine. ^{*b*}**PMEA**: 9-(2-[phosphonomethoxy]ethyl)adenine. ^{*c*}EC₅₀ (μ M): Concentration (μ M) required to inhibit the replication of HIV-1 by 50%. ^{*d*}CC₅₀ (μ M): Concentration (μ M) required to reduce the viability of unaffected cells by 50%

HIV-1. The antiviral activity of phosphonate nucleosides is due to their intracellular metabolism to diphosphates followed by incorporation into the viral genome and chain termination.²⁴ Anti-HIV activity was determined in human peripheral blood mononuclear (PBM) cells infected with HIV-1 strain LAI. PBM cells (1×10^5 cell/mL) were infected with HIV-1 at a multiplicity of infection of 0.02 and cultured in the presence of different concentrations of the test compounds. After 4 days of incubation at 37 °C, numbers of viable cells were determined by using the 3-(4,5-di-methylthiazole-2-yl)-2,5-diphenyltetrazolium bromide method. The cytotoxicity of the compounds was evaluated in parallel with their antiviral activity, based on the viability of mockinfected cells.²⁵ In particular, the adenine analog 13 showed significant anti-HIV-1 activity (Table 1), indicating diphosphorylation of the sugar moiety of the analog or some affinity of viral polymerases for its diphosphate. However, guanine nucleoside analog 16 showed weak anti-HIV activity at concentrations of up to $100 \,\mu$ M.

In summary, we have designed and successfully synthesized novel 4'-trifluoromethyl-5'-norcarbocyclic phosphonic acid nucleoside analogs starting from 1,3-dihydroxy cyclopentane (5). The adenine analog 13 exhibited significant antiviral activity against HIV-1 (EC₅₀ = 8.4μ M).

Experimental Section

Melting points (mp) were determined by using a Meltemp II laboratory device and are uncorrected. Nuclear magnetic resonance (NMR) spectra were recorded on a JEOL 300 Fourier transform spectrometer (JEOL, Tokyo, Japan). Chemical shifts are reported in parts per million (d) and signals are reported as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and dd (doublet of doublets). Ultraviolet (UV) spectra were obtained on a Beckman DU-7 spectrophotometer (Beckman, South Pasadena, CA, USA). Mass spectrometry (MS) spectra were collected in the electrospray ionization mode. The elemental analyses were performed by using a Perkin-Elmer 2400 analyzer (Perkin-Elmer, Norwalk, CT, USA). Thin layer chromatography was performed on Uniplates (silica gel) purchased from Analtech Co. (7558, Newark, DE, USA). All reactions were carried out under a nitrogen atmosphere unless otherwise specified. Dry dichloromethane, benzene, and pyridine were obtained

by distillation from CaH₂. Dry THF was obtained by distillation from Na and benzophenone immediately prior to use.

(*rel*)-(1*S* and 1*R*,3*S*)-3-(*t*-Butyldimethylsilanyloxy) cyclopentanol (6). TBDMSC1 (2.29 g, 15.25 mmol) was added slowly to a solution of **5** (1.41 g, 13.87 mmol) and imidazole (1.41 g, 20.80 mmol) in CH₂Cl₂ (100 mL) at -10 °C and stirred for 7 h at the same temperature. Saturated NaHCO₃ solution (10 mL) was poured into the mixture and stirred for 1 h at room temperature. The solvent was evaporated under reduced pressure. The residue was dissolved in water (200 mL) and extracted with diethyl ether (200 mL). The organic layer was washed with brine, dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane, 1:10) to produce compound **6** (1.58 g, 53%) as an isomeric mixture: ¹H NMR (CDCl₃, 300 MHz) δ 3.27-3.20 (m, 2H), 1.98-1.52 (m, 6H), 0.89 (m, 9H), 0.02 (m, 6H).

(±)-3-(t-Butyldimethylsilanyloxy) Cyclopentanone (7). Compound 6 (2.43 g, 11.25 mmol) was added to a solution of Dess-Martin periodinane (10.38 g, 24.5 mmol) in CH₂Cl₂ (100 mL) at 0 °C, and stirred for 24 h at room temperature under argon gas. The solvent was removed and the residue was triturated with diethyl ether (150 mL). Following filtration through a pad of silica gel, the organic solution was washed with a solution of sodium thiosulfate pentahydrate (13 g) in water (100 mL), ice-cold saturated NaHCO₃ (80 mL), and brine (80 mL) and dried over MgSO₄. The solvent was filtered, concentrated under vacuum, and purified by silica gel column chromatography (EtOAc/hexane, 1:10) to produce compound 7 (2.19 g, 91%) as a colorless syrup: ¹H NMR (CDCl₃, 300 MHz) δ 3.76-3.74 (m, 1H), 2.34-2.02 (m, 6H), 0.89 (s, 9H), 0.03 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 215.6, 63.5, 52.6, 37.1, 31.5, 25.7, 18.4, -4.7. Anal. Calcd. for C₁₁H₂₂O₂Si: C, 61.63; H, 10.34; Found: C, 61.72; H, 10.26; MS m/z 215 (M+H)⁺.

(rel)-(1R,4S)-1-(t-Butyldimethylsilanyloxy)-4-(trifluoromethyl)cyclopentanyl-4-ol (8a) and (rel)-(1R,4R)-1-(t-butyldimethylsilanyloxy)-4-(trifluoromethyl)cyclopentanyl-4-ol (8b). A catalytic amount of TBAF (0.37 mL, 1 M solution in THF) was added into mixture 7 (1.54 g, 7.2 mmol) with trifluoromethylsilane (25 mL, 12.5 mmol, 0.5 M solution in THF) in dry THF (10 mL) at -78 °C under argon gas. The reaction mixture was allowed to warm to room temperature. The solution became yellow immediately upon the addition of TBAF and eventually became dark brown. After the reaction mixture was stirred for 15 h at room temperature, it was washed with saturated NH₄Cl solution. The aqueous layer was extracted with $Et_2O(2 \times 20 \text{ mL})$ and the combined organic layer was dried with magnesium sulfate, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography. Without further purification, the crude residue was dissolved in methanol (10 mL) and a catalytic amount (0.1 eq) of metallic Na (17 mg) was added at 0 °C. The reaction was completed within 1 h, the reaction mixture was treated with a saturated NH₄Cl solution, and the aqueous layer was extracted with Et₂O. The combined organic layer was dried over Na₂SO₄ and filtered. The solvent was concentrated under high vacuum and the residue was purified by silica gel column chromatography (EtOAc/hexane, 1:8) to produce **8a** (695 mg, 34%) and **8b** (760 mg, 35%) as oils. Compound **8a**: ¹H NMR (CDCl₃, 300 MHz) δ 3.25-3.23 (m, 1H), 1.90-1.56 (m, 6H), 0.90 (s, 9H), 0.02 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 140.4 (q, *J* = 283.7 Hz), 80.2 (q, *J* = 30.4 Hz), 68.5, 35.2, 29.8, 25.5, 19.3, 18.4, -5. Anal. Calcd. for C₁₂H₂₃F₃O₂Si: C, 50.68; H, 8.15; Found: C, 50.77; H, 8.21; MS *m/z* 285 (M+H)⁺.

Compound 8b: ¹H NMR (CDCl₃, 300 MHz) δ 3.37-3.34 (m, 1H), 1.91-1.55 (m, 6H), 0.89 (s, 1H), 0.02 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 139.7 (q, *J* = 282.8 Hz), 79.5 (q, *J* = 31.8 Hz), 66.8, 36.2, 30.2, 25.7, 19.5, 18.1, -4.4. Anal. Calcd. for C₁₂H₂₃F₃O₂Si: C, 50.68; H, 8.15; Found: C, 50.56; H, 8.08; MS *m*/*z* 285 (M+H)⁺.

(rel)-(1R,4R)-Diethyl 1-(t-butyldimethylsilanyloxy) 4-[(trifluoromethyl) cyclopentanyloxy] methylphosphonate (9). Both LiOt-Bu (3.172 mL of 0.5 M solution in THF, 1.586 mmol) and a solution of diethyl phosphonomethyltriflate (475 mg, 1.586 mmol) in 12.0 mL of THF were slowly added to a solution of the **8b** analog (225 mg, 0.793 mmol) in 6.0 mL of THF at 0 °C and stirred overnight at room temperature under anhydrous conditions. The mixture was quenched by adding saturated NH₄Cl solution (5 mL) and further diluted with additional H₂O (100 mL). The aqueous layer was extracted with EtOAc (3×100 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO4, and concentrated in vacuo. The residue was purified by silica gel column chromatography (Hexane/ EtOAc, 4:1) to produce 9 (237 mg, 69%): ¹H NMR (CDCl₃, 300 MHz) $\delta 4.38 \text{ (m, 4H)}$, 3.94 (d, J = 8.0 Hz, 2H), 3.26 (m, 4H)1H), 1.89-1.49 (m, 6H), 1.13 (m, 6H), 0.89 (s, 9H), 0.04 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 138.4 (q, J = 281.4 Hz), 85.3 (q, J = 28.2 Hz), 70.1, 62.8, 62.3, 32.1, 30.5, 25.3, 18.6, 17.4, 13.7, -4.6. Anal. Calcd. for C₁₇H₃₄F₃O₅PSi: C, 46.99; H, 7.89; Found: C, 47.11; H, 7.95; MS *m/z* 435 (M+H)⁺.

(*rel*)-(1*R*,4*R*)-Diethyl [4-(trifluoromethyl) cyclopentanyloxy] methylphosphonate (10). TBAF (0.60 mL, 1.0 M solution in THF) was added to a solution of 9 (235 mg, 0.54 mmol) in THF (8 mL) at 0 °C. The mixture was stirred overnight at room temperature and concentrated. The residue was purified by silica gel column chromatography (EtOAc/hexane/MeOH, 1:4:0.02) to produce 10 (140 mg, 81%) as an oil: ¹H NMR (DMSO-*d*₆, 300 MHz) δ 4.41-4.38 (m, 4H), 3.87 (d, *J* = 8.1 Hz, 2H), 3.31 (m, 1H), 1.94-1.52 (m, 6H), 1.11 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 137.6 (q, *J* = 281.8 Hz), 84.3 (q, *J* = 29.7 Hz), 70.9, 62.4, 61.8, 31.3, 29.5, 17.3, 13.2. Anal. Calcd. for C₁₁H₂₀F₃O₅P: C, 41.26; H, 6.29; Found: C, 41.19; H, 6.34; MS *m/z* 321 (M+H)⁺.

(*rel*)-(1'*S*,4'*R*)-Diethyl [9-(4'-trifluoromethylcyclopentanyloxy-1'-yl)-6-chloropurine] Methylphosphonate (11). DEAD (245 mg, 1.412 mmol) was added dropwise at -40 °C to a solution containing compound 10 (226 mg, 0.706 mmol), triphenylphosphine (555 mg, 2.118 mmol), and 6chloropurine (218 mg, 1.412 mmol) in anhydrous THF (15.0 mL) for 10 min under nitrogen. The reaction mixture was Seyeon Kim et al.

stirred for 4 h at the same temperature under argon and further stirred overnight at room temperature. The solvent was concentrated *in vacuo* and the residue was purified by silica gel column chromatography (EtOAc/hexane, 3:1) to produce compound **11** (206 mg, 64%): mp 171-173 °C; UV (MeOH) λ_{max} 263.0 nm; ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.80 (s, 1H), 8.33 (s, 1H), 4.35 (m, 1H), 3.84 (d, *J* = 8.2 Hz, 2H), 3.75 (m, 1H), 2.21-1.52 (m, 6H), 1.12 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 151.7, 151.6, 151.4, 145.2, 137.6 (q, *J* = 282.8 Hz), 132.5, 85.3 (q, *J* = 28.6 Hz), 63.0, 62.5, 61.9, 52.5, 26.7, 23.5, 17.5, 15.2. Anal. Calcd. for C₁₆H₂₁ClF₃N₄O₄P (+1.0 MeOH): C, 41.83; H, 5.16; N, 11.48; Found: C, 41.74; H, 5.25; N, 11.53; MS *m/z* 457 (M+H)⁺.

(rel)-(1'S,4'R)-Diethyl [9-(4'-trifluoromethylcyclopentanyloxy-1'-yl)-adenine] methylphosphonate (12). A solution of 11 (220 mg, 0.482 mmol) in saturated methanolic ammonia (12 mL) was stirred overnight at 72 °C in a steel bomb and the volatiles were evaporated. The residue was purified by silica gel column chromatography (MeOH/ CH_2Cl_2 , 1:8) to produce 12 (130 mg, 62%) as a white solid: mp 163-165 °C; UV (MeOH) λ_{max} 261.0 nm; ¹H (DMSO-*d*₆, 300 MHz) & 8.28 (s, 1H), 8.13 (s, 1H), 4.37-4.34 (m, 1H), 3.87 (d, J = 8.1 Hz, 2H), 3.72 (m, 1H), 2.19-1.49 (m, 6H), 1.11 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 155.5, 152.7, 150.5, 141.4, 137.3 (q, J = 283.2 Hz), 120.1, 86.1 (q, J = 30.2 Hz), 62.6, 61.8, 52.2, 26.7, 22.5, 17.5, 13.9. Anal. Calcd. for C₁₆H₂₃F₃N₅O₄P (+1.0 MeOH): C, 43.52; H, 5.80; N, 14.93; Found: C, 43.65; H, 5.73; N, 14.85; MS m/z 438 $(M+H)^{+}$.

(rel)-(1'S,4'R)-[9-(4'-Trifluoromethylcyclopentanyloxy-1'-yl) adenine] 4'-Methylphosphonic Acid (13). TMSBr (0.619 mL, 4.69 mmol) was added to a solution of phosphonate 12 (205 mg, 0.469 mmol) in anhydrous CH₃CN (10 mL) and 2,6-lutidine (1.09 mL, 9.38 mmol). The mixture was heated overnight at 72 °C under nitrogen and then concentrated under vacuum. The residue was co-evaporated from concentrated aqueous ammonium hydroxide (NH₄OH; 2×20 mL) and purified by triturating with acetone (10 mL) twice and removing the acetone by evaporation. The residue was then purified by preparative reverse-phase column chromatography using C18 silica gel. Lyophilization of the appropriate fraction produced phosphonic acid salt 13 (128 mg, 69% yield) as a white salt (ammonium salt): UV (H₂O) λ_{max} 262.0 nm; ¹H NMR (D₂O, 300 MHz) δ 8.38 (s, 1H), 8.19 (s, 1H), 3.81 (d, J = 8.2 Hz, 2H), 3.72 (m, 1H), 2.15-1.51 (m, 6H); ¹³C NMR (D₂O, 75 MHz) 155.7, 152.9, 150.8, 141.6, 138.5 (q, J = 281.8 Hz), 120.5, 85.9 (q, J = 29.4 Hz), 63.2, 52.2, 26.7, 22.5, 18.0; High-performace liquid chromatography (HPLC), $t_{\rm R} = 10.26$ min; High-resolution mass spectrometry (HRMS) $[M - H]^+$ calcd. 380.0692, found 380.0693.

(*rel*)-(1'*S*,4'*R*)-Diethyl [9-(4'-Trifluoromethylcyclopentanyloxy-1'-yl)-2-fluoro-6-chloropurine] Methylphosphonate (14). Coupling of 10 with 2-fluoro-6-chloropurine was accomplished by using a similar Mitsunobu reaction as described for the synthesis of 11: yield 61%; mp 176-178 °C; UV (MeOH) λ_{max} 270.5 nm; ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.45 (s, 1H), 4.21-4.15 (m, 4H), 3.81 (d, *J* = 10.5 Hz, 2H), 3.69 (m, 1H), 2.15-1.50 (m, 6H), 1.14 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 157.6 (d, *J* = 220.5 Hz), 154.1, 145.2, 137.3 (q, *J* = 281.5 Hz), 136.5, 121.3, 86.7 (q, *J* = 30.4 Hz), 62.7, 62.4, 61.5, 52.2, 26.7, 23.1, 18.2, 13.5. Anal. Calcd. for C₁₆H₂₀ClF₄N₄O₄P (+0.5 MeOH): C, 40.43; H, 4.52; N, 11.43; Found: C, 40.50; H, 4.48; N, 11.49; MS *m*/z 475 (M+H)⁺.

(*rel*)-(1'S,4'R)-Diethyl [9-(4'-Trifluoromethylcyclopentanyloxy-1'-yl) 2-Fluoro-6-aminopurine] Methylphosphonate (15a) and (*rel*)-(1'S,4'R)-Diethyl [9-(4'-Trifluoromethylcyclopentanyloxy-1'-yl) 2-Amino-6-chloropurine] Methylphosphonate (15b). Dry ammonia gas was bubbled into a solution of 14 (310 mg, 0.65 mmol) in dimethoxyethane (15 mL) while stirring at room temperature overnight. The salts were removed by filtration, and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (MeOH/CH₂Cl₂, 1:10) to produce 15a (35 mg, 12%) and 15b (128 mg, 42%).

Data for 15a: UV (MeOH) λ_{max} 261.0 nm; ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.21 (s, 1H), 4.15-4.09 (m, 4H), 3.84 (d, *J* = 10.2 Hz, 2H), 3.70 (m, 1H), 2.18-1.56 (m, 6H), 1.25-1.20 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 160.9 (d, *J* = 254.3 Hz), 155.2, 152.8, 142.0, 138.4 (q, *J* = 281.4 Hz), 120.2, 85.4 (q, *J* = 31.8 Hz), 63.2, 62.7, 61.7, 52.7, 26.8, 24.5, 19.2, 13.7. Anal. Calcd. for C₁₆H₂₂F₄N₅O₄P (+1.0 MeOH): C, 41.91; H, 5.38; N, 14.37; Found: C, 41.88; H, 5.43; N, 14.42; MS *m*/z 456 (M+H)⁺.

Data for 15b: UV (MeOH) λ_{max} 309.0 nm; ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.13 (s, 1H), 4.17-4.14 (m, 4H), 3.84 (d, *J* = 8.0 Hz, 2H), 3.73 (m, 1H), 2.17-1.57 (m, 6H), 1.24-1.20 (m, 6H); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 158.4, 154.1, 150.7, 142.7, 136.9 (q, *J* = 281.3 Hz), 125.2, 86.3 (q, *J* = 29.5 Hz), 62.6, 62.1, 61.5, 53.7, 27.6, 23.2, 13.7. Anal. Calcd. for C₁₆H₂₂ClF₃N₅O₄P (+ 1.0 MeOH): C, 40.58; H, 5.21; N, 13.92; Found: C, 40.62; H, 5.28; N, 13.87; MS *m*/*z* 472 (M+H)⁺.

(rel)-(1'S,4'R)-[9-(4'-Trifluoromethylcyclopentanyloxy-1'-yl) Guanine] 4'-Methylphosphonic Acid (16). Dry acetonitrile (15 mL), 2,6-lutidine (1.01 mL, 8.72 mmol), and TMSBr (0.57 mL, 4.36 mmol) were added to a solution of 15b (205 mg, 0.436 mmol) at room temperature. After stirring this mixture for 32 h, the solvent was removed and co-evaporaed three times with MeOH. The residue was dissolved in MeOH (16.0 mL), and 2-mercaptoethanol (170 mg, 2.18 mmol) and sodium methoxide (117 mg, 2.18 mmol) were added to the mixture. The mixture was refluxed for 24 h under nitrogen, cooled, neutralized with glacial acetic acid, and evaporated to dryness under vacuum. The obtained residue was co-evaporated from concentrated $NH_4OH (2 \times 16 \text{ mL})$ and the resultant solid was triturated with acetone (2×16 mL). After evaporating the acetone, the residue was purified by preparative column chromatography by using reverse-phase C18 silica gel and eluting with water. Lyophilization of the appropriate fraction produced compound 16 (92 mg, 51%) in the form of a yellowish salt (ammonium salt): mp 180-182 °C; UV (H₂O) λ_{max} 255.5 nm; ¹H NMR (D₂O, 300 MHz) δ 7.84 (s, 1H), 3.86 (d, J = 8.3

Hz, 1H), 3.69 (m, 1H), 2.19-1.60 (m, 6H); ¹³C NMR (D₂O, 75 MHz) δ 157.5, 154.4, 152.1, 137.2, 136.4 (q, *J* = 280.8 Hz), 119.1, 86.3 (q, *J* = 30.4 Hz), 63.4, 44.6, 24.1, 18.4; HPLC *t*_R = 9.37 min; HRMS [M-H]⁺ req. 396.0754, found 396.0755.

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