# Food Consumption and Utilization Efficiency in *Samia ricini* Donovan Reared on *Ricinus communis*, *lin*. Leaves Supplemented with Cyanobacteria

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# **Abstract**

Food consumption and conversion efficiency of eri silkworm *Samia ricini* Donovan were studied during 4<sup>th</sup> and 5<sup>th</sup> larval instars by feeding castor leaves fortified with 100, 200, 300, 400 and 500 ppm concentrations of aqueous extracts of cyanobacteria *Anabaena variabilis*. The nutritional indices viz., ingesta, digesta, approximate digestibility (%), reference ratio and efficiency parameters like ECI and ECD were recorded which were significantly high at 400 ppm concentration treated batches of 4<sup>th</sup> instar larvae over control batches. The decline in nutritional efficiency parameters of 5<sup>th</sup> instar treated larvae might be due to higher utilization of the digested food for metabolic activities. Significant difference of ECI to cocoon % and non-significant difference of ECD to cocoon% and shell were observed between the treatments and control. Cyanabacteria feed supplement contains antibiotic and nutritions factors which has reflective effect on the biological parameters in eri silkworm and therefore has greater application in commercial eri silkworm rearing.

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### Introduction

Sericulture has been a traditional rural industry and instrumental in poverty alleviation in India. Qualitative and quantitative increase of cocoon production would certainly result into better economics for this industry and meet production needs. In order to increase the cocoon production, enrichment of mulberry leaves by supplementary compounds is an absolute important aspect (Etebari *et al.*, 2004). Nutritional supplements when added to normal food, not only tend to increase the nutritional value of the food but improves also the vital parameters of the silkworm also (Bajpeyi *et al.*, 1991).

Probiotics are the live microbial feed supplements which beneficially affect the host by improving the microbial balance (Yeung et al., 2002). The microbes, Lactobacilli and Bifidobacteria, are gram + ve bacteria producing lactic acid and extensively studied to improve cocoon characters in B.mori L (Masthan et al., 2010). Oral administration of mulberry leaf and castor leaf supplemented with single cell protein 'spirulina' (Blue green algae) as a feed to respective silkworms Bombyx mori L (Venkataramana et al., 2003) and Samia cynthia ricini Boisduval (Jayaprakash et al., 2005) was found to be effective in enhancing the larval and cocoon characters in mulberry silk worm. The antiviral protein of spirulina platensis

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exhibited 90% resistance to BmNPV of B.mori L (Babu et al., 2005).

Cyanobacteria (Blue green algae) are free living photosynthetic, and N<sub>2</sub> fixing bacteria found in fresh, marine water and terrestrial environments. Genus *Anabaena* having 40 common species store reserve food materials which can be utilized as a source of proteins, lipids, vitamins and secondary metabolites (Anupama, 2000 and Tan, 2007). The antibiotic and nutritious effects of *Anabaena variabilis* cyanobacteria (Sampath *et al.*, 2013) were evidenced by increased larval weights and cocoon yield of eri silkworm *S.ricini* Boisduval when they fed with castor leaves fortified with *A.variabilis* aqueous extracts. However, information on the synergistic action of combination of feed supplements is very scanty. Hence, the present investigation was undertaken to study the effect of cyanobacteria, *A.variabilis* as feed supplement on consumption, utilization and commercial indices of eri silkworm, *Samia ricini* Donovam.

## **Materials and Methods**

### Test insect and Host plant

Eri silkworm *S ricini* Donovan (Saturniidae: Lepidoptera) was selected for utilization studies because of their hardiness and less susceptibility to diseases. Leaves of primary host plant, *Ricinus communis* Lin (castor) were preferred as the source of food for the test insect which is next to host plant, Kessuru, *Heteropanax fragrans* (Roxb.) Seem, with respect to nutritional status of the host plant. Evaluation of food consumption and utilization in this insect was successfully undertaken as it is reared indoors like mulberry silkworm *B. mori L*.

## Cyanobacteria culture

Cyanobacteria, *A. variabilis* strains selected as feed supplements and obtained from growing cultures of rice fields of Warangal district, Andhra Pradesh, India. These cultures were subcultured in BC-II media with or without nitrogenous source at 25±1°C temperature under continuous illumination (3000 lux) for 12 d. Then, the culture was washed with distilled water (Sampath *et al.*, 2013) to obtain pellets which were shade dried at room temperature.

### Rearing of Eri silkworm

Tray-cum shelf rearing method with 25 eri silkworm disease free

layings (DFLs) were carried out on castor leaves as suggested by Jayaprakash *et al.* (2006) in rearing chambers under controlled (temp. 25 - 26°C and relative humidity 85%) conditions.

### Consumption and utilization studies:

Feed utilization studies were confined to 4<sup>th</sup> and 5<sup>th</sup> instar eri silkworm larvae as 80% of total leaves was consumed in these instars. Since resumption to 4<sup>th</sup> instar, 60 larvae each in four replications (irrespective of sex) of a treatment were maintained separately from general rearing.

Agueous solution of A. variabilis at 100 ppm (T1), 200 ppm (T2), 300 ppm (T3), 400 ppm (T4), 500 ppm (T5) concentrations and control (T6) with distilled water were prepared daily for soaking of castor leaves for 20 min each treatment. Soaked leaves were shade dried and fed to worms daily four times at an interval of 6 h. Each treated castor leaf was cut into two halves one half was fed to the larvae while the other half was usied for the dry weight determination. Additional larval batch of each treatment was also maintained to determine the dry weight and for subsequent determination of daily increment in larval weight. The feeding duration of 4<sup>th</sup> and 5<sup>th</sup> instar larvae was also noted. The healthy larvae were counted daily, unequal, or dead and missing larvae were replaced with larvae of same age group from additional batches. Leaves and faeces were collected at 10 A.M daily and dried in an oven at 80°C to a constant weight. Thus, for each day of feeding period, dry matter ingested, digested, faeces produced and increase in larval weight was recorded daily.

The nutritional indices viz., ingesta, digesta, approximate digestibility (AD%), reference ratio (PR) and nutritional efficiency parameters viz., efficiency of converting the ingesta (ECI) and digesta (ECD) into larva, cocoon and shell weight were calculated. Besides, relative consumption and growth rates (RCR & RGR) were computed. The methods of calculation of various parameters (Mathur *et al.*, 2002 and Rath, 2011) were as follows:

- a) Ingesta = Dry weight of leaf fed Dry weight of left over leaf.
- b) Digesta = Dry weight of leaf ingested Dry weight of faeces.
- c) Approximate Digestibility (AD%) =  $\frac{Dry \ weight \ leaf \ ingesetd Dry \ weight \ of \ faeces}{Dry \ weight \ of \ food \ ingested} \times 100$

d) Reference Ratio = 
$$\frac{Dry\ weight\ food\ ingested}{Dry\ weight\ of\ litter} \times 100$$

e) 
$$ECI$$
 (%) =  $\frac{Dry\ weight\ gain\ of\ larvae}{Dry\ weight\ of\ ingeta} \times 100$ 

f) ECD (%) = 
$$\frac{Dry\ weight\ gain\ of\ larvae}{Dry\ weight\ of\ digesta} \times 100$$

g) ECI to Cocoon (%) = 
$$\frac{Dry\ weight\ gain\ of\ cocoon}{Dry\ weight\ of\ Ingesta} \times 100$$

h) EDC to cocoon (%) = 
$$\frac{Dry\ weight\ gain\ of\ Cocoon}{Dry\ weight\ of\ digesta} \times 100$$

i) ECI to shell (%) = 
$$\frac{Dry\ weight\ gain\ of\ Shell}{Dry\ weight\ of\ ingesta} \times 100$$

j) ECD to shell (%) = 
$$\frac{Dry\ weight\ gain\ of\ Shell}{Dry\ weight\ of\ digesta}$$

k) RCR =

Dry weight of food eaten

Duration of feeding(Days) × Meandry weight of the larvae during the feeding period

1) RGR =

Dry weight gain of the larvae

Duration of feeding(Days) × Meandry weight of the larvae during the feeding period

The experiments were repeated twice during summer and rainy seasons of 2013 year and pooled data were tabulated, compiled and

analyzed by using ANOVA (Analysis of Variance) test.

### **Results and Discussion**

The food consumption, utilization efficiencies and growth rates of 4<sup>th</sup> and 5<sup>th</sup> instars are presented in Table 1 and 2, respectively. The amount of ingesta and digesta varied significantly in 4<sup>th</sup> and 5<sup>th</sup> instar larvae fed with castor, R. *communis*, leaves treated with different concentrations of *A.variabilis* and in different seasons. The amount of food consumed and digested was significantly high in both the batches fed on castor leaves treated with 400 ppm (T4) of cyanobacteria during summer and rainy seasons. Descending order of values in ingestion and digestion capacity of 4<sup>th</sup> instar larvae was observed at 300 (T3), 200 (T2), 500 (T5), 100 (T1) ppm concentration of *A.variabilis* whereas in 5<sup>th</sup> instar it was at T5, T3, T2 and T1 concentrations in both the seasons. Significantly low ingesta and digesta was recorded in control (T6) treatments of both larval

**Table 1.** Food intake and utilization budget of *S. ricini* during fourth instar larvae fed on castor leaves fortified with *A. variabilis* 

Season	Treatment	Ingesta	Digesta	AD (%)	Reference Ratio (RR)	ECI (%)	ECD (%)	RCR	RGR
Summer (May – June)	T1	141.41	55.16	39.05	1.64	40.50	59.86	0.675	0.170
	T2	143.11	60.83	42.60	1.69	41.93	58.77	0.844	0.251
	Т3	155.65	85.40	45.35	2.20	46.56	68.78	1.251	0.575
	T4	163.35	97.00	59.00	2.47	55.10	82.23	1.275	0.775
	T5	140.47	48.55	34.56	1.46	42.81	54.25	0.787	0.201
	T6 (Control)	136.88	52.40	38.28	1.61	33.30	44.44	0.355	0.171
Rainy (July – Aug)	T1	137.60	50.67	36.83	1.58	38.52	48.50	0.555	0.140
	T2	159.25	66.36	42.14	1.70	45.45	60.00	0.915	0.350
	Т3	175.75	73.24	41.67	1.80	58.30	74.35	1.345	0.775
	T4	183.75	109.12	58.23	2.38	61.95	80.10	1.892	0.925
	T5	160.65	56.66	35.68	1.55	53.91	68.35	1.112	0.665
	T6 (Control)	127.25	47.60	37.40	1.60	33.48	42.08	0.434	0.135
	F. Test	HS	HS	HS	NS	HS	HS	NS	NS
	CD at 5%	4.89	11.13	7.77	0.220	12.75	15.60	8.050	0.350
	CD at 1%	9.60	17.20	11.23	0.816	19.23	22.31	14.250	1.315

AD = Approximate Digestibility: RCR = Relative Consumption Rate: ECI = Efficiency of Converting the Ingesta: RGR = Relative Growth Rate: HS = Highly Significant:

ECD = Efficiency of converting the Digesta: NS = Non Significant

Table 2. Food intake and utilization budget of S. ricini during fifth instar larvae fed on castor leaves fortified with A.variabilis

Season	Treatment	Ingesta	Digesta	AD (%)	Reference Ratio (RR)	ECI (%)	ECD (%)	RCR	RGR
Summer (May – June)	T1	400.38	100.38	20.08	1.25	23.53	43.32	0.780	0.082
	T2	462.60	127.00	25.10	1.37	24.20	45.20	0.885	0.112
	Т3	463.50	148.60	32.20	1.45	25.35	47.32	1.125	0.427
	T4	594.40	199.40	33.55	1.51	33.52	58.67	1.950	0.955
	T5	515.35	150.35	28.17	1.39	21.94	44.57	1.775	0.684
	T6 (Control)	394.44	77.44	19.63	1.21	18.76	37.61	0.675	0.045
Rainy (July – Aug)	T1	495.68	107.68	21.32	1.28	26.46	38.37	0.906	0.100
	T2	510.80	137.23	26.88	1.37	29.80	46.25	0.995	0.104
	Т3	557.30	178.67	32.10	1.47	28.88	47.22	1.059	0.513
	T4	625.35	233.27	38.47	1.59	34.40	61.30	2.010	0.911
	T5	568.30	144.25	25.38	1.34	28.96	50.77	1.553	0.735
	T6 (Control)	404.10	81.15	20.04	1.25	20.29	32.23	0.553	0.075
	F. Test	HS	HS	HS	NS	HS	HS	NS	NS
	CD at 5%	6.73	53.28	12.75	0.250	10.76	26.26	4.750	0.985
	CD at 1%	9.44	60.15	18.50	0.733	16.20	33.14	6.350	3.885

AD = Approximate Digestibility: RCR = Relative Consumption Rate: ECI = Efficiency of Converting the Ingesta: RGR = Relative Growth Rate: ECD = Efficiency of converting the Digesta:

HS = Highly Significant:

NS = Non Significant

instars. The findings of Horie *et al.* (1976), Jaksheva and Genova (1991), Anantha Raman *et al.* (1993), Mathur *et al.* (2002) and Gokulamma and Reddy (2005) had supported the present findings by revealing the higher ingesta and digesta values for 4<sup>th</sup> and 5<sup>th</sup> instars of *B. mori L.* The higher ingesta in 5<sup>th</sup> instar is necessary as the larvae has to maintain metabolic demand during the transformation stages from larvae to pupa, pupa to moth. Similar results were also reported by Poonia (1978), Reddy and Alfred (1979) in eri silkworm *Philosamia ricini* and Rana *et al.* (1987) in Oak tasar silkworm, Antheraea proylei Jolly. Larval weight of eri silkworm has increased substantially (Tomy Philip *et al.*, 2009) when the larvae from 3<sup>rd</sup> instar to spinning stage were fed on castor foliage fortified with leaf

Sampath *et al.* (2013) observed increase in larval weight (59.95%), pupation rate (31.83%) and cocoon yield (146.86%) of *S.C ricini* Boisduval fed on castor leaves foliated with 500 ppm of *A. variabilis* aqueous extracts. Higher cocoon yield of *S.C. ricini* might be attributed to reduction in larval mortality effected by antibacterial and antiviral

extracts of Stachytarpheta indica (L) Vahl.

activities of cyanobacteria, A.variabilis (Bloor and England, 1989).

The approximate digestibility (AD) defines the quantity of food that was actually digested. AD gradually increased in the treated batches 1 to 4<sup>th</sup> instar when compared to 5<sup>th</sup> instar larvae during summer and rainy seasons. significant difference of AD% between control and other treatments batches was noticed (Table 1 &2). This is probably dur to feeding of larvae on supplemented nutrients along with castor leaves. The digestibility is affected by high content of crued fiber deficiency of nutriunts and water in food (Waldbauer, 1964). Similar results of AD as per the age of worms were also reported by Poonia (1978) and Rana *et al.* (1987).

Reference ratio (RR) expresses ingesta required per unit excreta production. High value of RR indicates high rate of digestion and absorption of food. It also expresses the retention efficiency of food. comparatively higher RR (Table 1) was recorded in 400 ppm (T4) treatment(2.47)(2.38) batches than control (1.61) and (1.60) batches of 4<sup>th</sup> instar larvae recpectively during the both seasons. It indicates that 4<sup>th</sup> instar larva has to ingest more to produce one gram of excreta.

Table 3. Conversion Efficiency Parameters to Cocoon and Shell

Season	Treatment	ECI to Cocoon (%)	ECD to Cocoon (%)	ECI to Shell (%)	ECD to Shell (%)
	T1	19.37	33.54	2.65	6.94
	T2	20.95	32.61	2.93	8.05
Summer	Т3	20.71	31.76	2.86	8.00
(May – June)	T4	24.83	35.53	4.55	9.35
	T5	21.21	32.45	3.65	8.00
	T6 (Control)	18.52	27.35	2.15	5.11
	T1	18.38	30.83	2.24	5.86
	T2	22.36	35.80	3.27	8.98
Painy	Т3	21.00	32.60	2.94	8.25
Rainy (July – Aug)	T4	25.65	37.80	4.92	10.11
	T5	21.21	33.55	3.85	8.46
	T6 (Control)	17.55	25.65	2.02	4.66
	F. Test	HS	NS	NS	NS
	CD at 5%	3.86	6.83	0.398	2.32
	CD at 1%	5.47	9.78	1.210	4.16

HS = Highly Significant: NS = Non Significant

Gokulamma and Reddy (2005) reported that RR and AD (%) values were high during early instars and reduced with advancement of instars of *B.mori L*. (Table 1 and 2)

Efficiency conversion of ingested food (ECI) is an overall measure of the ability of larvae to utilize the undigested food. The ECI recorded was high in 4<sup>th</sup> instar and lowest in the 5<sup>th</sup> instar of treated lots. Fourth instar larvae of treated lots performed better with respect to efficiency of conversion of digested food (ECD) recording 82.23 and 80.10% during summer and rainy seasons followed by 5<sup>th</sup> instar (58.67 and 61.30%) at 400 ppm concentrations. The higher ECD values might be due to higher absorption ability and associated with higher digestibility. The low ECD in the last instar is due to higher utilization of the digested food for metabolic activities. Our findings are in the confirmity with those of Rana *et al.* (1987), Mathur *et al.* (2002) and Manimeghalai and Aruna (2010) who observed that AD and ECD values are higher in young age worms than in late age worms of *A.proylei* and *B.mori L.* 

There was consistent increase in the efficiency of conversion of ingested (ECI) and digested (ECD) food into the body substance in 4<sup>th</sup> instar when compared to decline of both efficiencies in 5<sup>th</sup> instar. This

decline might be associated with energy consuming physiological activities associated with the approach of larval maturity. Similar patterns were also observed by Rana *et al.* (1987) and Barah (2007) while dealing with the consumption and utilization studies of *A. proylei* and *A.assamensis*. In the present work, higher ECI and ECD values of 4<sup>th</sup> and 5<sup>th</sup> instar larvae were noticed during rainy season followed by summer. Seasonal and feed supplement influence on the nutritive indices that clearly indicate the significance of leaf moisture both in palatability and assimilation of nutritive components of the leaf

Relative consumption rate (RCR) and Relative growth rate (RGR) of 4<sup>th</sup> (1.892 and 0.925) and 5<sup>th</sup> (2.010 and 0.911) instar larvae were comparatively higher at 400 ppm (T4) of *A. variabilis* treated leaves when compared to control and other treatments in rainy season (Table 1&2). The growth rate was the highest in rainy season followed by summer season at 400 ppm. Growth rate directly influences the speed of development of larvae which in turn depends on the quality of leaf and physiological stages of the larvae. The present findings are in close conformity of the findings of Gokulamma and Reddy (2005) and Manimegalai and Aruna (2010).

The efficiency of conversion of ingesta and digesta to cocoon and shell are given in Table 3. Significant difference of ECI to cocoon% was observed between different treatments and control. Higher ECD to cocoon % was recorded in T4 (37.80% in rainy and 35.53% in summer season) and was found non-significant. No Significant difference of ECI and ECD to shell % was observed between all the treatments especially at 400 ppm in comparison to control (Table 3). Maachi and Katagiri (1991) inferred that ECI to cocoon % and shell % were ultimate nutritional indices in terms of productivity to evaluate the production efficiency of breed or hybrid castor leaves supplemented with aqueous extracts of "Spirulina" (Jayaprakash et al., 2005) leaf extracts of S.indica (Tomy Philip et al., 2009) and A. variabilis (Sampath et al., 2013) found to be effective in increasing cocoon weight, shell weight and silk ratio of eri silkworm. "Serifeed" supplemented mulberry feed (Narayana Swamy and Ananthanarayana 2006) improved the commercial characters of B.mori L. However, bioassay studies of mulberry foliage from cyanobacteria biofertilizer + NPK 50% treated plots (Dasappa et al., 2006) revealed no significant difference in mulberry silkworm growth and cocoon characters was observed.

This seems to be the first report on consumption and utilization of cyanobacteria, *A.variabilis* feed supplement by eri silkworm as review of literature reveals that no such studies have been conducted so far. Earlier, Sampath *et al.* (2013) observed the positive effects of *A.variabilis* feed supplemention on larval weight and cocoon yield of eri silkworm. From the present investigation, it is quite apparent that 400 ppm of feed supplement cyanobacteria *A.variabilis* brings in a positive effect on growth indices viz., ingesta, digesta, efficiency of conversion of ingested (ECI) and digested (ECD) food, relative consumption (RCR) and growth (RGR) rate of *S. ricini* Donayan.

It can be assumed that the presence of antibiotic and nutritional factors viz., lipids, proteins, vitamins and secondary metabolites in cyano bacteria *A.variabilis* induced eri silkworm to ingest more, digest the food in a better way and to convert the utilized food efficiently into body matter. So, this supplementation could be prescribed to farmers to get more quantity of eri silk.

### References

Anantha Raman KV, Benchamin KV, Magadum SB, Ramadevi OK., and Datta RK (1993) Studies on the nutritional efficiency in silkworm *Bombyx mori L*. Indian J. Seric. 32(1): 43-49.

Anupama PR (2000) Value added food single cell protein, Biotechnol. Adv. 18: 459-479.

Babu SM, Swamy, G and Chandramohan N (2005) Identification of an antiviral principle in *Spirulina plantensis* against *Bombyx mori L*. Nuclear polyhedrosis virus (BmNPV). Ind. J. Biotechnol. 4: 384-388.

Bajpeyi CM, Singh RN and Thangavelu K (1991) Supplementary nutrients to increase silk production. Indian Silk 30(7): 41-42.

Barah A (2007) Utilisation of Soalu (*Litsea monopetala* Roxb.) leaves by muga silkworm (*Antheraea assamensis* Helfer). Indian J.Entomol. 69(4): 392-395.

Bloor S and England RR (1989) Antibiotic production by the cyanobacterium *Nostoc muscorum*. J. Appl. Phycol. 1: 367-372.

Dasappa, Rama Rao DM and Ramaswamy SN (2006) Efficacy of cyanobacterial biofertilizer (CBB) on leaf yield and quality of mulberry and its impact on silk worm cocoon characters. Int. J. Indust. Entomol. 13(1): 15-22.

Etebari K, Kaliwal B and Matindoost L (2004) Supplementation of mulberry leaves in sericulture, Theoretical and applied aspects. Int. J. Indust, Entomol. 9: 14-28.

Gokulamma K and Reddy YS (2005) Role of nutrition and environment on the consumption, growth and utilization indices of selected silkworm races of *Bombyx mori L*. Indian J. Seric. 44(2): 165-170.

Horie Y, Inokuchi T, Watanable K, Nakasone S and Yanagawa H (1976) Quantitative studies on food utilization by the silkworm *B.mori L*. through its life cycle. I.Economy of dry matter, energy and carbon. Bull. Seric. Exp. Stn. Jpn. 26(6): 413-442.

Jaksheva V and Genova E (1991) Daily ingestion and utilization of various kinds of nutrients in mulberry leaves by the silkworm *B.mori L*, sericologia. 31(1): 167-172.

Jayaprakash P. Singh RSJ, Rao BVS and Suryanarayana N (2006) Rearing performance of Eri silkworm *Samia ricini* Donovan on castor and tapioca in non-traditional areas. In: Natl. Workshop on eri food plants, 11-12, October, 2006. Central Muga Eri Research and Training Institute, Jorhat Assam, India. pp. 38-46.

Jayaprakash P, Singh RSJ, Rao BVS, Vijay Kumar M, Sasikala A and Suryanarayana N (2005) Effect of spirulina on the larval and economic characters of silkworm, *Samia cynthia ricini* boisduval, Proc. XX Cong. International Sericultural Commission, 15-18<sup>th</sup> December, 2005, Bangalore, India. pp. 254-257.

Maachi H and Katagiri K (1991) Varietal differences in nutritive values of mulberry leaves for rearing silkworm, JARQ. 25: 205-208.

Manimegalai S and Aruna GR (2010) Nutritional efficiency of selected hybrids and a cross breed of silkworm, *Bombyx mori L*. Indian J.

- Seric. 49 (2): 125-133.
- Masthan K, Raj Kumar T, Usharani CV and Narasimhamurthy CV (2010) Use of *Lactobacillus acidophilus* as a probiotics to improve cocoon production of mulberry silkworm *Bombyx mori L.* J. Curr. Sci. 15(2): 445-449.
- Mathur VB, Rahmathulla VK and Vijayabhaskar O (2002) Consumption and conversion efficiency of food in new elite bivoltine hybrid silkworm, *Bombyx mori L*. under restricted feeding levels. Int. J. Indust. Entomol. 5(2): 213-216.
- Narayana swamy M and Ananthanarayana SR (2006) Biological role of feed supplement "Serifeed" on nutritional parameters, cocoon characters and cocoon yield in silkworm, *Bombyx mori L*, (Lepidoptera: Bombycidae), Indian J. Seric. 45(2): 110-115.
- Poonia FS (1978) Studies on food utilization and rate of growth during the developmental stages of Eri silkworm *Philosamia ricini* Hutt. Indian. J. Seric. 17: 48-60.
- Rana B, Prasad B and Nigam MP (1987) Consumption and utilization of food by oak tasar silkworm *Antheraea proylei* Jolly (Lepidoptera: Saturniidae). Sericologia, 27(1): 11-19.
- Rath SS (2011) Food utilization efficiency in *Antheraea mylitta* fed on *Terminalia ariuna* leaves. Sericologia. 51(1): 91-99.
- Reddy MV and Alfred JRB (1979) Utilization of castor *Ricinus communis* Linn, leaves by the last instar larvae of the silkworm

- *Philosamia ricini* Hutt. (Lepidoptera: Saturniidae). Indian Biol. 2:35-40
- Sampath A, Ramesh Babu M, Sujatha K, Singh RSJ and Rao DB, (2013) Beneficial effect of cyanobacteria *Anabaena variabilis* on quantitative traits of Eri silkworm *Samia cynthia ricini*, Boisduval, Asian J. Agric. Sci. 5(3): 36-39.
- Tan LT (2007) Bioactive natural products from marine cyanobacteria for drug discovery, Phytochem. 68: 954-979.
- TomyPhilip, Somaprakash DS and Qadri SMH (2009) Effect of fortification of castor (*Ricinus communis L.*) leaves with plant extracts on the biological performance of eri silkworm. Indian. J. Seric. 48(2): 191-193.
- Venkataramana P, Rao TVSS, Reddy PS and Suryanarayana N (2003) Effect of spirulina on the larval and cocoon characters of the silkworm *Bombyx mori L*, proc. Nat. Acad., Sci. India. 73B(1): 89-94.
- Waldbauer GP (1964) The consumption, digestion and utilization of Solanaceous and non-solanaceous plants by the larvae of the tobacco horn worm *Protoparea sexta* (Johan) (Lepidoptera: Sphingidae), Entomol. Exp. Appl. 7:253-269.
- Yeung PSM, Sanders CL, Kitts R, Cano and Tong PS (2002) Species specific identification of commercial probiotic strains. J.Dairy. Sci. Association. 85: 1039-1051.