

Verification of Heme Catalytic Cycle with 5-Aminosalicylic Acid and Its Application to Soil Remediation of Polycyclic Aromatic Hydrocarbons

Namhyun Chung¹, Kapsung Park², David K. Stevens³, Guyoung Kang^{2†}

¹College of Life Sciences and Biotechnology, Korea University, Seoul 136-713, Korea

²Department of Environmental Science, Hankyong University of Foreign Studies, Yongin 449-791, Korea

³Department of Civil & Environmental Engineering, Utah State University, Logan, UT 84321, USA

Abstract

Catalytic degradation of pentachlorophenol in soil by heme and hydrogen peroxide has been hypothesized to occur through nonspecific catalytic reactions similar to those involving ligninase. The present study examines the evidence for a heme catalytic mechanism for the oxidation of organic compounds. In the presence of hydrogen peroxide, heme is converted to the ferryl heme radical (Hm-Fe⁴⁺), which can oxidize organic compounds, such as 5-aminosalicylic acid (5-ASA). A second 5-ASA may later be oxidized by ferryl heme (Hm-Fe⁴⁺), which reverts to the ferric heme state (Hm-Fe³⁺) to complete the cycle. We believe that this catalytic cycle is involved in the degradation of hazardous pollutants, such as polycyclic aromatic hydrocarbons (PAHs). Remediation via heme catalytic reactions of PAHs in soil from a pole yard was evaluated, and about 96% of PAHs was found to disappear within 42 days after treatment with heme and hydrogen peroxide. In addition, benzo[a]pyrene and six other PAHs were undetectable among a total of 16 PAH compounds examined. Therefore, we propose heme catalysis as a novel technology for the remediation of hazardous compounds in contaminated soil.

Keywords: Heme, Hydrogen peroxide, Polycyclic aromatic hydrocarbons (PAHs), Remediation, 5-Aminosalicylic acid (5-ASA)

1. Introduction

Benzo[a]pyrene, a polycyclic aromatic hydrocarbon (PAH), is considered to be carcinogenic [1, 2]. It is classified by the United States Environmental Protection Agency (US EPA) as a class B2 carcinogenic compound with a mutagenic mode of action for inducing tumors [1, 2]. Additionally, the compound is regulated under the Korea Land Conservation Act of 2009. As a result, the interest in the biotic and abiotic processes for the destruction of hazardous materials in contaminated soil is renewed.

Creosote oil has been used for the preservation of railroad timbers for over 130 years in North America. Pentachlorophenol (PCP) and creosote oil were widely used before 1900 for railroad timbers, electric power poles, and bomb wood boxes [3, 4]. In Korea, waste timbers from railroads are usually recycled as landscaping material after 10–15 years of use [4]. Between 2005 and 2007, about 250,000 pieces of waste railroad timbers were recycled for use in playgrounds, trails, and timber houses. Kim et al. [5] reported that PAHs are major components of creosote oil (about 75%–80%). Therefore, it is expected that the soil around the recycled waste timbers would be heavily contaminated with PAHs [4–7]. Chung et al. [4] also reported that waste railroad tim-

bers contain high concentrations of PAHs, PCP, and benzene, toluene, ethylbenzene, and xylenes (BTEX). Sixteen PAH compounds were tested in waste timbers, and the maximum total PAH concentration was found to be 23,607 mg/kg, including a benzo[a]pyrene concentration of 153 mg/kg. These xenobiotic compounds are known to be toxic to microorganisms and carcinogenic to humans.

Barr and Aust [8] reported that lignin-degrading white fungi are able to degrade lignin as well as hazardous organic pollutants through the peroxidase catalytic cycle, which includes the protein heme. Heme, hemoglobin, and peroxidases (i.e., lignin peroxidase and horseradish peroxidase) react with hydrogen peroxide (H₂O₂) to degrade PAHs, PCP, crystal violet, and dibenzothiophene [7–11]. We were interested in the development of a soil-remediation technology based on the catalytic/degrading abilities of heme and H₂O₂ [7, 11]. Our previous work has demonstrated that PCP-contaminated soil was significantly remediated in the presence of heme and H₂O₂, possibly through a postulated catalytic reaction, under both laboratory conditions and at a pole yard field in Vancouver, WA, USA [11]. We have shown that



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>)

which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Received February 04, 2014 Accepted May 01, 2014

[†]Corresponding Author

E-mail: knaggy@hufs.ac.kr

Tel: +82-31-330-4269 Fax: +82-31-330-4529

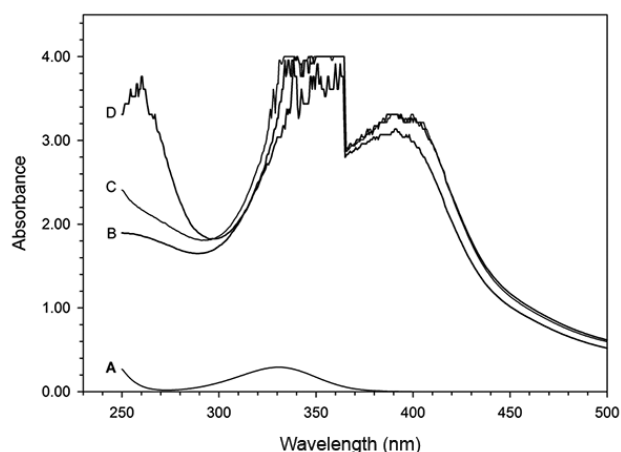


Fig. 1. Absorption spectra of 5-aminosalicylic acid (5-ASA) oxidation in the presence of 60- μM heme and 600- μM H_2O_2 . Curve A represents the absorption spectrum for 100- μM 5-ASA in 50-mM phosphate buffer (pH 7.0). Curve B represents the spectrum obtained for 60- μM heme and 600- μM H_2O_2 . Curve C represents the spectrum obtained for 60- μM heme and 100- μM 5-ASA (without H_2O_2 added). Curve D shows the changes in curve C after the addition of 600- μM H_2O_2 . The reaction temperature was 25°C and the reaction mixtures were scanned within 1 min.

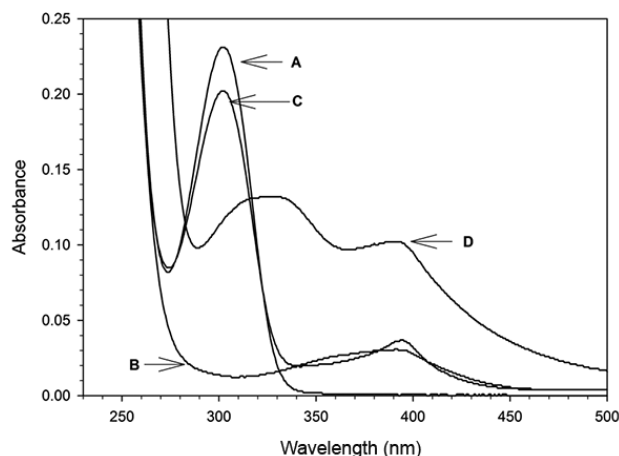


Fig. 2. Absorption spectra for 5-aminosalicylic acid (5-ASA) oxidation in the presence of 60- μM heme and 600- μM H_2O_2 after the removal of the heme from the reaction mixture. Heme was removed by 1 mL of 5% trichloroacetic acid and filtration prior to spectral analysis. Curve A is the absorption spectrum for 100- μM 5-ASA. Curve B is the spectrum for 60- μM heme and 600- μM H_2O_2 . Curve C is the spectrum for 60- μM heme and 100- μM 5-ASA (without H_2O_2 added). Curve D is the spectrum for 60- μM heme and 100- μM 5-ASA with 600- μM H_2O_2 added. All mixtures were reacted for 1 min at 25°C in a phosphate buffer solution.

about 80% of PCP was degraded within 4 hr in the presence of 8.5 g of heme and 47.5 g of $\text{H}_2\text{O}_2/\text{kg}$ soil.

The objectives of the present work were to demonstrate the heme catalytic mechanism using 5-aminosalicylic acid (5-ASA) as a model organic chemical and to prove that PAHs in soil can be degraded by the catalytic mechanism, including heme and H_2O_2 . We assessed the interaction between heme (Hm-Fe^{3+}) and H_2O_2 in the presence of 5-ASA and the degradation of PAHs in contaminated soil under an optimized dose of heme and H_2O_2 .

2. Materials and Methods

2.1. Chemicals

Hemin, 5-ASA, 30% H_2O_2 , and trichloroacetic acid (TCA) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Reagent-grade chemicals for phosphate buffering were purchased from Duksan Pharmaceutical Co. (Ansan, Korea). The heme stock solution (388 μM) was prepared just before each experiment by dissolving 2.53 mg of hemin in 1 mL of 100-mM NaOH with gentle mixing, immediately adding 9 mL of 50-mM phosphate buffer (pH 7.0) and vortexing. The H_2O_2 solution was prepared to the desired concentration by diluting 30% H_2O_2 before each experiment. Five percent TCA was prepared and added to the reaction mixture for the precipitation of heme.

2.2. Spectral Characterization of Heme Catalytic Reactions

Spectra and kinetic measurements for the heme catalytic reaction were obtained using a UV/Vis recording spectrophotometer (UV-1650PC; Shimadzu, Tokyo, Japan). All catalytic reaction

mixtures contained 60- μM heme, 50-mM phosphate buffer, 100- μM 5-ASA, and 600- μM H_2O_2 in a 3-mL total volume using a 5-mL 6Q Quartz cuvette (light path 10 mm; Starna Scientific Ltd., Essex, UK) at 24°C \pm 2°C. The catalytic reactions were initiated by the addition of 20 μL of H_2O_2 (600 μM). The oxidation and reduction states of heme were determined by scanning between 250 and 700 nm. To study the heme catalytic mechanism, we assessed the ability of heme to catalyze the H_2O_2 -dependent oxidation of 5-ASA. The catalytic reactions were terminated by adding 1 mL of a solution containing 5% (w/v) TCA and vortexing for 10 sec. The coagulated heme was then removed from the reaction mixture by filtration using a 0.2- μm filter (Acrodose LC 13 PVDF; Gelman Sciences Inc., Ann Arbor, MI, USA) and scanned in a 1-mL QS Quartz cuvette (Hellma Analytics, Mullheim, Germany).

2.3. Degradation of PAHs in Soil by Heme Catalytic Reactions

The pan study was performed using soil from a pole yard in Vancouver, WA, USA. Five hundred grams of pole yard soil was placed in a stainless steel pan. First, 10 g of heme was dissolved with 40 mL of 0.05-M NaOH and diluted to 400 mL of the final volume with 50-mM phosphate buffer (pH 7.0) to make the feed solution. The heme feed solution was mixed with soil contaminated with PAHs; then, 54 g of $\text{H}_2\text{O}_2/\text{kg}$ soil was added and mixed. The PAH samples were extracted using a Soxhlet apparatus (US EPA Method 3540A), and the solvent extract was prepared for HPLC analyses using an acid-base partition cleanup (US EPA Method 3650). The solvent extracts were analyzed using HPLC (SIL-6B, Shimadzu) with a UV detector (LC 90 UV spectrophotometric; PerkinElmer, Waltham, MA, USA).

3. Results and Discussion

3.1. Oxidation of 5-ASA by Heme Catalytic Reaction

UV/Vis spectra were obtained to study the interaction of heme with H_2O_2 in the presence of 5-ASA (Fig. 1). All catalytic reaction mixtures contained 60- μM heme, 50-mM phosphate buffer (pH 7.0), 100- μM 5-ASA, and 600- μM H_2O_2 in a total volume of 3 mL. The spectral characteristics were scanned between 250 and 700 nm. However, the maximum absorbance of heme (i.e., the Soret band) was not clearly observed because of the interference from the other reagents, such as H_2O_2 and 5-ASA. Thus, the experiments in Fig. 1 were repeated after the heme was precipitated by the addition of 1 mL of 5% TCA and removed by filtration [6]. Fig. 2 illustrates the UV/Vis absorption spectra for the mixtures of 5-ASA, heme, or H_2O_2 at pH 7.0. The absorption spectrum did not significantly change in the presence of either 5-ASA or 5-ASA/heme (Fig. 2(a) and (c)). In the presence of heme and H_2O_2 without 5-ASA, the peak at 300 nm was not observed and a weak peak appeared at 400 nm (curve B in Fig. 2). These results indicate that the peak at 300 nm can be attributed to 5-ASA while the weak peak at 400 nm is due to heme. In the spectrum for the mixture of all three reactants (heme, H_2O_2 , and 5-ASA), strong absorption peaks at 330 and 400 nm were observed, possibly because of the formation of oxidized 5-ASA (5-ASA_{ox}) by the heme catalytic oxidation reaction (curve D in Fig. 2). Thus, 5-ASA was oxidized only in the presence of both heme and H_2O_2 . It is known that 5-ASA_{ox} has an absorption peak at 400 nm that can be used to measure the change in its concentration [12, 13]. The formation of 5-ASA_{ox} was confirmed by measuring the change in absorbance at 400 nm. The oxidation of 5-ASA in the presence of heme was initiated by the addition of H_2O_2 . The reaction was stopped by the addition of 1 mL of 5% TCA solution and the precipitate was filtered to measure the absorbance at 400 nm. Fig. 3 shows that the absorbance at 400 nm increased over time and therefore that 5-ASA was oxidized by heme catalytic reactions.

3.2. Heme Catalytic Mechanism

The mechanism of the heme catalytic reaction has been postulated to be similar to that of peroxidase or hemoglobin [10, 13]. However, no clear experimental data exists to explain the mechanism. In our experiments, heme in the phosphate buffer (pH 7.0) had a maximum absorbance at 360 nm (Fig. 4, peak A). When H_2O_2 was added to the solution containing heme, the absorbance decreased from 1.450 to 1.191 (Fig 4, peaks A and B). This reaction suggests that ferric heme (Hm-Fe³⁺) was oxidized to ferryl heme (Hm-Fe⁴⁺) through the ferryl heme radical (Hm-Fe⁴⁺) upon reaction with H_2O_2 [14]. When 5-ASA was added to the reaction mixture, the absorbance increased from 1.191 to 1.307. This result suggests that the ferryl heme radical (Hm-Fe⁴⁺) oxidized 5-ASA to 5-ASA radical and was reduced back to ferric heme (Hm-Fe³⁺) with one more oxidation of 5-ASA or 5-ASA radical to complete the catalytic or redox cycle (Fig. 4, peak C), while 5-ASA as a substrate may be oxidized into 5-ASA_{ox}. In support of the data in Fig. 4, the duration of time for the three reactions was 2 min (Fig. 5). These results show that 5-ASA reacted with oxidized ferryl heme radicals or ferryl heme to reduce the oxidized hemes back to ferric heme. The heme catalytic reaction mechanism may therefore be defined as shown in Fig. 6, which is analogous to the catalytic cycle for the degradation of PCP [6, 7, 11, 13].

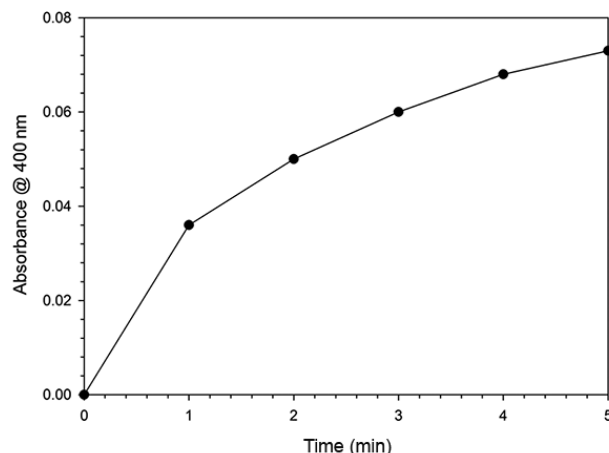


Fig. 3. Oxidation of 5-aminosalicylic acid (5-ASA) in the presence of 60- μM heme and 100- μM 5-ASA with 600- μM H_2O_2 added. Spectral analysis was conducted at 400 nm. Heme was removed by addition of 1 mL of 5% trichloroacetic acid and filtration prior to spectral analysis. Absorbance at 0 min was measured without H_2O_2 . Values are averages of triplicate analyses.

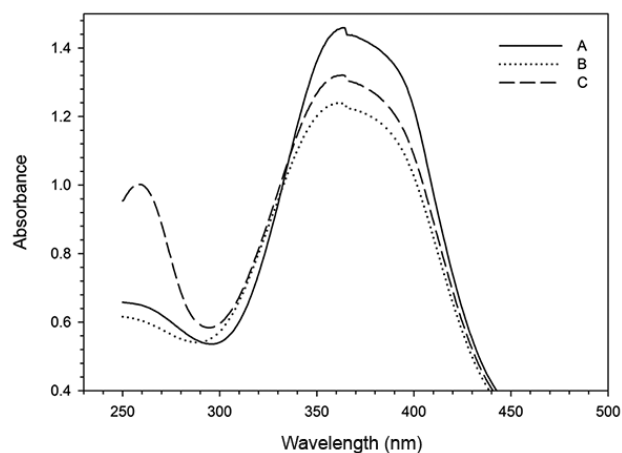


Fig. 4. Changes in the absorption spectra for (a) 30- μM heme, (b) 30- μM heme and H_2O_2 with 200- μM 5-aminosalicylic acid added, and (c) 30- μM heme and H_2O_2 .

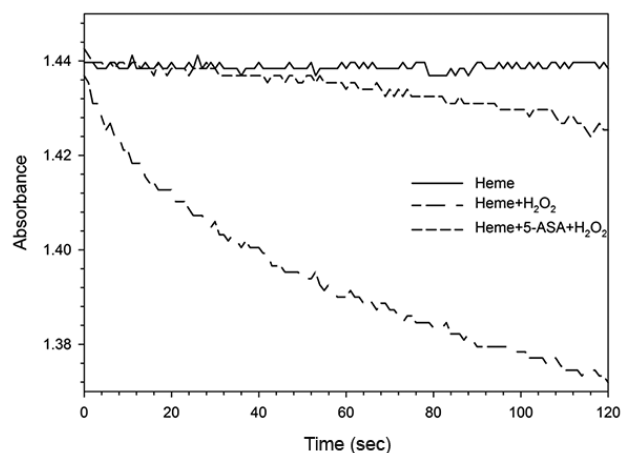


Fig. 5. Time-dependent observations of heme catalytic reactions for 30- μM heme, 200- μM 5-aminosalicylic acid (5-ASA), or 500- μM H_2O_2 . Absorbance was measured at 360 nm.

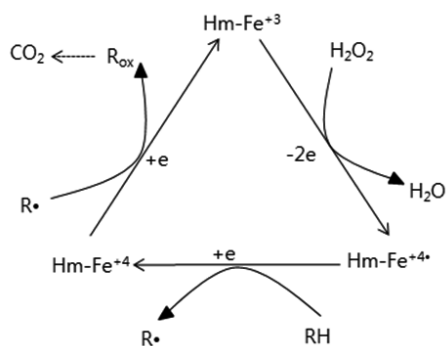


Fig. 6. Heme catalytic cycling for degradation of the organic pollutants.

3.3. Degradation of PAHs in Soil

The oxidized heme intermediate ferryl heme radical (Hm-Fe^{4•}) is formed by a two-electron oxidation upon interaction between H₂O₂ and heme. A one-electron oxidation results in the formation of Fe⁴⁺ from Fe³⁺ and another one-electron oxidation is due to the oxidation of the heme porphyrin ring. It has been suggested that the ferryl heme radical (Hm-Fe^{4•}) is short-lived and that it rapidly decomposes to ferric heme upon reaction with a substrate by two successive one electron oxidations. A few studies have shown that heme is capable of oxidizing a wide variety of substrates [6, 7, 11, 13, 15, 16]. Specifically, Chen et al. [7, 11] reported significant degradation of PCP in soil in the presence of heme and H₂O₂. To extend the use of the heme catalytic reaction, we investigated the degradation of PAHs, which are commonly found in soil around wood preserved with creosote oil. The PAH-contaminated soil came from a pole yard in Washington, USA.

Table 1. PAH degradation by heme and hydrogen peroxide during the pan study of contaminated pole yard soil (Washington, USA)

PAH compound	Operation time in the pan study		
	Day 0	Day 14	Day 42
Naphthalene	49.32	34.64	ND
Acenaphthylene	16.64	4.88	ND
Acenaphthene	102.78	79.72	ND
Fluorene	3.08	1.62	ND
Phenanthrene	11.30	0.10	ND
Anthracene	5.24	0.58	ND
Fluoranthene	111.84	13.70	0.44
Pyrene	73.40	80.68	3.70
Benzo[a]anthracene	24.62	13.32	0.58
Chrysene	28.28	21.84	0.40
Benzo[b]fluoranthene	12.28	3.78	0.32
Benzo[k]fluoranthene	4.66	1.92	3.00
Benzo[a]pyrene	5.38	2.80	ND
Dibenzo[a,h]anthracene	3.00	0.18	1.28
Benzo[g,h,i]perylene	8.70	1.70	0.08
Indeno[1,2,3 cd]pyrene	3.26	1.22	0.24
Total PAHs	463.78	262.68	10.04

Values are average of triplicate analyses (unit: mg/kg soil). PAH: polycyclic aromatic hydrocarbon, ND: not detected.

This contaminated soil was a sandy loam and contained 464 mg of PAH/kg soil. Five hundred grams of the PAH-contaminated soil was placed into a stainless pan and mixed with 400 mL of heme feed solution containing 10 g of heme as described in the Materials and Methods section. Finally, 159 mL of 30% H₂O₂ was diluted with distilled water to make a final volume of 250 mL and mixed well by spraying into the soil in the pan. The water content in this soil was maintained at 50% for 42 days. The PAH concentrations in the soil decreased from 464 to 263 mg/kg of soil at day 14 and to 10 mg/kg of soil at day 42, as shown in Table 1. Thus, the extent of the PAH degradation was about 96% by day 42. Approximately 40% of the toxic components of the PAHs (e.g., benzo[a]pyrene) were no longer detectable by day 42. Therefore, with the combination of heme and H₂O₂, PAHs in contaminated soil can be effectively and economically remediated. This technology can be considered to be an innovative technology. We believe that the remediated soil is environmentally safe because heme cannot perform any catalytic activity in the absence of H₂O₂.

4. Conclusions

The present study focused on the heme catalytic reaction in the presence of H₂O₂ using substrates, such as 5-ASA. The study also evaluated the practical use of the catalytic reaction in degrading the PAHs present in the soil obtained from a pole yard. We concluded that the reactions in the presence of heme and H₂O₂ can be explained as a possible catalytic cycle that include ferric heme (Hm-Fe³⁺), ferryl heme radical (Hm-Fe^{4•}), and ferryl heme (Hm-Fe⁴⁺). Even though PAH compounds have been shown to be toxic, carcinogenic, and recalcitrant to microbial degradation, about 96% of the PAH content in the pole yard soil was degraded by heme and H₂O₂. Moreover, benzo[a]pyrene and six other PAH compounds, out of 16 compounds tested, were undetectable in the soil at day 42. In conclusion, because of its environmental safety the heme catalytic degradation of PAHs can be used as a novel technology for the remediation of hazardous organic compounds.

Acknowledgments

This subject is supported by the Korea Ministry of Environment under the GAIA Program.

References

- Mumtaz M, George J. Toxicological profile for polycyclic aromatic hydrocarbons (PAHs). Atlanta: Agency for Toxic Substances and Disease Registry; 1995.
- Menzie CA, Potocki BB, Santodonato J. Exposure to carcinogenic PAHs in the environment. *Environ. Sci. Technol.* 1992;26:1278-1284.
- Lee JC, Choi S. Solvent extraction of pentachlorophenol (PCP) from PCP-treated wood. *Korean Chem. Eng. Res.* 2006;44:227-233.
- Chung D, Yoon JI, Kim MS, et al. Hazardous characteristics of waste timber from rail road. *J. Korea Soc. Waste Manag.* 2012;29:59-69.
- Kim H, Kang S, Han K, Du S, Kang S. Waste rail timber used survey and management. Gwacheon: Ministry of Environ-

- mental; 2008.
6. Kang G, Jung J, Park K, Stevens DK. Mineralization of hazardous chemicals by heme reaction. In: Teddler DW, Pohland FG, eds. *Emerging technologies in hazardous waste management 7*. Heidelberg: Springer; 1995. p. 69-77.
 7. Chen ST, Stevens DK, Kang G. Pentachlorophenol and crystal violet degradation in water and soil using heme and hydrogen peroxide. *Water Res.* 1995;33:3657-3665.
 8. Barr DP, Aust SD. Mechanisms white rot fungi use to degrade pollutants. *Environ. Sci. Technol.* 1994;28:78A-87A.
 9. Lamar RT, Evans JW. Solid-phase treatment of a pentachlorophenol-contaminated soil using lignin-degrading fungi. *Environ. Sci. Technol.* 1993;27:2566-2571.
 10. Stachyra T, Guillochon D, Pulvin S, Thomas D. Hemoglobin, horseradish peroxidase, and heme-bovine serum albumin as biocatalyst for the oxidation of dibenzothiophene. *Appl. Biochem. Biotechnol.* 1996;59:231-244.
 11. Chen ST, Stevens DK, Kang G, Hsieh M. Treating soil PCP at optimal conditions using heme and peroxide. *J. Environ. Eng.* 2006;132:704-708.
 12. Yamada T, Volkmer C, Grisham MB. The effects of sulfasalazine metabolites on hemoglobin-catalyzed lipid peroxidation. *Free Radic. Biol. Med.* 1991;10:41-49.
 13. Kang G, Park K. Heme and hydrogen peroxide catalytic reaction mechanism and mineralization. *J. Korean Soc. Environ. Eng.* 1996;18:399-405.
 14. Yoshida T, Migita CT. Mechanism of heme degradation by heme oxygenase. *J. Inorg. Biochem.* 2000;82:33-41.
 15. Wood AW, Levin W, Lu AY, et al. Metabolism of benzo(a)pyrene and benzo(a)pyrene derivatives to mutagenic products by highly purified hepatic microsomal enzymes. *J. Biol. Chem.* 1976;251: 4882-4890.
 16. Pinsky C, Boss R. Pyridine and other coal tar constituents as free radical-generating environmental neurotoxicants. *Mol. Cell. Biochem.* 1988;84:217-222.