

Power Absorption Measurements during NMR Experiments

N. Félix-González^{1,2*}, A. L. Urbano-Bojorge^{1,2}, C. Sánchez-L de Pablo^{1,2}, V. Ferro-Llanos^{1,2},
F. del Pozo-Guerrero^{1,2}, and J. J. Serrano-Olmedo^{1,2}

¹Centre for Biomedical Technology, Technical University of Madrid, Spain

²Biomedical Research Networking Center in Bioengineering Biomaterials and Nanomedicine (CIBER-BBN), Zaragoza, Spain

(Received 1 July 2013, Received in final form 11 December 2013, Accepted 20 December 2013)

The heating produced by the absorption of radiofrequency (RF) has been considered a secondary undesirable effect during MRI procedures. In this work, we have measured the power absorbed by distilled water, glycerol and egg-albumin during NMR and non-NMR experiments. The samples are dielectric and examples of different biological materials. The samples were irradiated using the same RF pulse sequence, whilst the magnetic field strength was the variable to be changed in the experiments. The measurements show a smooth increase of the thermal power as the magnetic field grows due to the magnetoresistive effect in the copper antenna, a coil around the probe, which is directly heating the sample. However, in the cases when the magnetic field was the adequate for the NMR to take place, some anomalies in the expected thermal powers were observed: the thermal power was higher in the cases of water and glycerol, and lower in the case of albumin. An ANOVA test demonstrated that the observed differences between the measured power and the expected power are significant.

Keywords : magnetoresistive effect, NMR, power release, RF power absorption, thermal power

1. Introduction

Resistive losses in biological tissues produce heat as an undesirable side effect of the RF radiation in MRI procedures. As the RF resonant pulse only has a very small associated electric field, the magnetically induced currents are the main cause of heating in biological tissue typically electrically conductive. However, in case of dielectric materials there are other many factors that determine the quantity of absorbed power such as the dielectric composition and the physical dimensions, shape and geometry of the sample in relation to the incident wavelength. In dielectric substances, the amount of energy absorbed could be neglected because a fraction of the incident energy would be transmitted through the material, while other fraction would be reflected back, so that, only a small amount of energy would be really absorbed. Actually, as the wavelength is much greater in relation to the sample, the fraction of energy deposited is small because the absorptive coupling is inadequate [1-4]. Several estimates

of power absorption in whole body during MRI experiments have been done taking into account the organism's size, anatomical features, duration of exposure, sensitivity of the involved tissues and many other complex variables, such as environmental factors. Their results show that the power deposition is a critical function of the frequency of the RF excitation pulse used [4-6]. Experimental measurements have demonstrated and confirm the dependency between the frequency of the pulse sequence and the specific absorption rate [7, 8].

Despite RF absorption is the main cause of tissue heating during MRI procedures, several works suggested measurable temperature increases by the effect of potent static magnetic fields [9, 10]. However, posterior works have demonstrated that temperature changes caused by powerful static magnetic fields, if any, are statistically insignificant [4, 10]. Heating produced by RF in MRI has been considered a secondary undesirable effect, but D.L. Parker suggested the use of MRI as hyperthermia therapy [11]. The researcher presented theoretical calculations about heating produced by the NMR effect, predicting a minimum increase of temperature, in the order of micro-degrees per hour. Although these calculations were discouraging, they left the possibility of finding a combi-

©The Korean Magnetism Society. All rights reserved.

*Corresponding author: Tel: +34615946196

Fax: +34615946196, e-mail: nazariofelix@hotmail.com

nation of pulse sequences for the use of NMR as thermal therapy.

Following this idea this study reports thermal power measurements during NMR procedures. Contrarily to the experiments mentioned in the literature, the RF pulse sequence was kept fixed and the static magnetic field was modified during the experiments. We have irradiated a sample of pure egg-albumin, pure glycerol and distilled water with a RF pulse sequence of 20 MHz at a variable magnetic field. The nuclei of Hydrogen have their NMR frequency at 20 MHz in a magnetic field of 0.46 T. We applied five different magnetic field strengths but the NMR only occurs for one of them. To obtain thermal powers from the observed time dependent temperature variations, we have performed a thermal calibration of our NMR equipment.

2. Materials and Methods

A BRUKER electromagnet of variable field strength ranging from 0.23 T up to 1.9 T was used for the NMR experiments. The design and generation of the RF pulse sequence were done in a digital NMR console of a Fast Field Cycling Relaxometer (STELAR-SMARTracer). Due to restrictions in the space, it was impossible to use a thermal isolation around the probe to avoid temperature increase due to coil losses (Fig. 1). To determine the temperature increase, we used a LUXTRON m3300 fiber optic thermometry system. Since the LUXTRON probes are optical fibers, they are immune to electromagnetic interference and can measure temperatures in the presence of intense magnetic fields. Its accuracy is ± 0.2 °C within 20 °C of calibration point (manufacture's specifications). Fig. 1 shows the experimental set up.

2.1. Samples and performed experiments

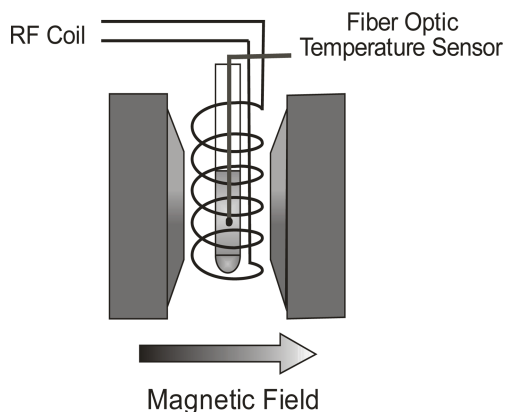


Fig. 1. Power absorption measurements.

The samples consisted of 150 μ L of distilled water, of pure glycerol and of natural egg-albumin in a 5 mm \varnothing NMR tube. All these substances were chosen due to its high dielectric coefficients in order to avoid induced currents inside the sample and because of their biological nature. The water is a standard reference; the glycerol is similar to many substances due to its OH terminations and tightly related to lipid substances. In the case of albumin, it is a protein similar to many abundant proteins in the human body and of high availability. The proton resonance was achieved by adjusting the magnetic field to 0.46 T, for a Larmor frequency of 20 MHz. Although the samples have the same NMR frequency at 0.46 T, their molecular structures are very different: whilst water and glycerol are compounds with a simple structure (glycerol formed by three hydroxyl groups) [12], albumin is a single polypeptide with 585 residues containing 17 pairs of disulfides bridges and one free cysteine [13].

Several magnetic field intensities were applied while keeping constant the RF excitation (at 20 MHz frequency, same power and sequencing), so that NMR only took place at 0.46 T, and the rest of values were: 0.23 T, 0.35 T, 0.58 T and 0.7 T. We have tuned the antenna to perfect impedance coupling with the RF pulse generator. The RF pulse sequence parameters were the same in all experiments to ensure that we have delivered the same RF power to the coil. The RF pulse sequence was repeatedly applied at intervals which were a fraction of the relaxation time T_1 . In all experiments, the samples were irradiated until the temperature reaches its maximum of stabilization, for about 30 minutes. Ten assays were performed for each sample and each magnetic field strength in order to reduce the uncertainty.

From the hydrogen spectroscopy of the samples, see Fig. 2, we can be sure that our instrument “illuminates” (excites) their entire spectra. We have considered the chemical shifts in all the samples, but the most significant

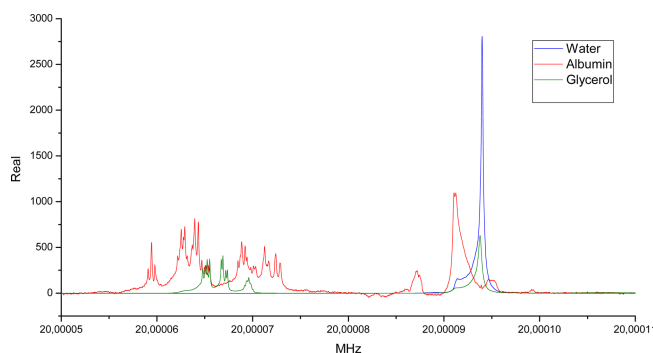


Fig. 2. (Color online) Hydrogen Spectroscopy of distilled water, albumin and glycerol samples.

peaks are contained in a bandwidth of very few hundreds of hertz, whilst the NMR console produces RF pulses with a bandwidth of 300 kHz. So, in the three cases, the radiation excites the whole spectrum when the pulse frequency is of 20 MHz at 0.46 T. In the same way, when the magnetic field intensity is set to 0.35 T and 0.58 T, the closest points to 0.46 T, both sides, and whose Larmor frequencies should be 15 MHz and 25 MHz respectively, the RF pulse of 20 MHz is unable to produce NMR in the samples. Actually we did not receive echoes at any other frequency than 20 MHz and a magnetic field intensity of 0.46 T.

2.2. Thermal Characterization of the System

To characterize the thermal behavior of the system, we used a resistor inside the sample probe as heater without RF excitation, so that the temperature variations were registered at constant pressure. The resistor was energized until the maximum stabilization temperature was reached. Then, it was extracted from the sample and the cooling curve was registered too. We assume that the thermal conductivity (thermal conductivity and heat transfer coefficient refer to the same concept) from inside the sample to the environment, C_D , is mainly determined by the air layer around the sample and the sample holder itself (quartz), much more than by the characteristics of the content, sample plus resistor during the heating or only sample during the cooling. On the contrary, given that C_D is, actually, quite low, the thermal capacity, C_T is much more dependent on the matter being heated or cooled, that is, depends on the thermal phase: heating ($C_{T\uparrow}$) or cooling ($C_{T\downarrow}$). The maximum increment of temperature obtained by power dissipation in the resistor was taken as $\Delta T_{R,\max}$, whereas the time constant τ_{\downarrow} of the cooling phase was obtained by fitting with the following equation:

$$\Delta T_R(t) = \Delta T_{R,\max} e^{(-t/\tau_{\downarrow})} \quad (1)$$

In the thermal model underneath, t depends on the thermal conductivity, C_D , and on the heat capacity, C_T as follows:

$$\tau_{\downarrow} = C_{T\downarrow} / C_D \quad (2)$$

and the thermal conductivity is given by:

$$C_D = P_R / \Delta T_{R,\max} \quad (3)$$

where the power P_R consumed by the resistor and dissipated as heat is exactly known.

It is convenient to use the thermal capacity instead of the thermal conductivity because it is larger sensibility to experimental artifacts, in our work. The thermal capacity can be calculated as follows:

$$C_{T\downarrow} = P_R \cdot \tau_{\downarrow} / \Delta T_{R,\max} \quad (4a)$$

Once the system has been thermally calibrated, the power supplied to the sample during NMR experiments, P_{NMR} , can be calculated from the measurement of the maximum increment of temperature, $\Delta T_{NMR,\max}$ by applying the following:

$$P_{NMR} = (C_{T\downarrow} / \tau_{\downarrow}) \cdot \Delta T_{NMR,\max} \quad (4b)$$

2.3. Electrical Power Characterization of the RF Coil

Since the emitting antenna is a coil in the middle of whose axis the sample is placed, an interfering thermal effect affects the experiments (Fig. 1). When the RF coil becomes hotter by Joule effect, the sample temperature rises as well. The other mechanism that would be present, is the heating produced by currents induced inside the samples, although it was avoided by choosing dielectric samples.

The coil self-heating is strongly conditioned by the magnetoresistive effect. The magnetic field alters the trajectories of the charge carriers inside the conductor, making their paths longer, and, consequently, enlarging the electric resistance and the electric power consumed in the coil [14]. The current actually running through the coil also depends on the impedance circuit matching the antenna and the transmitter/receiver console of the NMR equipment, which must be as good as possible [15]. This matching circuit consists of a couple of variable capacitor (C_s , C_p) mounted as shown in Fig. 3, where the auto-inductance (L) of the coil is in series to its internal series ohmic resistance (R). The capacitors must be adjusted before the NMR measurement in order to tune the antenna to the Larmor frequency.

In the NMR tuning condition, the measured impedance of the RF coil was of 41.80-j21.50 W at 20 MHz (wo) and at null magnetic field. Therefore, the impedance

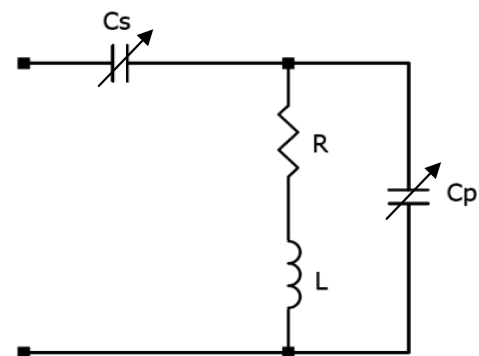


Fig. 3. Electric diagram of the RF coil and the matching capacitors.

modulus $|Z|$ and phase θ_z of the tuned antenna depends on the serial resistance R , and can be expressed as:

$$|Z|^2 = \frac{1}{\omega_o^4 \cdot C_p^4 \cdot R^2} + \frac{1}{\omega_o^2} \left(\frac{1}{C_s} + \frac{1}{C_p} \right)^2 = \frac{23.1}{R^2} + 462.2 \quad (5)$$

$$\theta_z = \tan^{-1} \left(R \cdot \omega_o \cdot C_p \cdot \left(1 + \frac{C_p}{C_s} \right) \right) = \tan^{-1} (R \cdot 4.473) \quad (6)$$

The resistance of the wire coil at null field when taking into account the skin effect at 20 MHz is $R_0 = R(B = 0) = 0.115 \Omega$. We assume that the skin effect and the magneto-resistive effect are independent each other. The variation on the series resistor associated to the wire and due to the magneto-resistive effect is defined by the Kohler rule [16]. For room temperature and copper single-wire conductor transverse to the magnetic field B it can be expressed as:

$$\frac{R(B) - R_0}{R_0} = f(B) = \frac{\Delta\rho(B)}{\rho_0} \rightarrow R(B) = R_0 \cdot (1 + f(B)) \quad (7)$$

where the function $f(B)$ is known for a straight conductor, whilst the coil wire is not straight but curled.

The differential wire resistance dR for a differential length of coil, dl , can be calculated as follow (see Fig. 4 for the geometric details):

$$\begin{aligned} dR &= \frac{\rho(l)}{\pi \cdot a^2} dl = \frac{R_0}{2\pi \cdot A} \left(1 + \frac{\Delta\rho(l)}{\rho_0} \right) \cdot dl \\ &= \frac{R_0}{2\pi \cdot A} \left(1 + f \left(B \cdot \left| \sin \left(\frac{l}{A} \right) \right| \right) \right) \cdot dl \end{aligned} \quad (8)$$

where a is the wire radius, A is the radius of the spire and l/A is θ .

Given that the antenna is made of copper, and that the function $f(B)$ is linear on B in the region of interest [16],

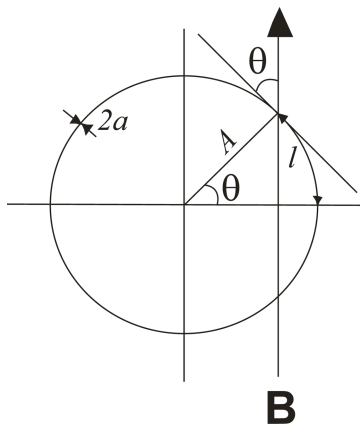


Fig. 4. Schematic diagram for determining the RF coil resistance.

the problem of integration of R gets reduced to the integration of the absolute value of the sine in a half of spire. The magneto-resistive effect leads to a dependency of the coil resistive component on the magnetic field as follows:

$$R(B) = R_0 \left(1 + \frac{2}{\pi} \cdot f(B) \right) \quad (9)$$

Therefore, the electric power dissipated in the coil at a variable magnetic field of B intensity, $P(B)_{coil}$ can be expressed as:

$$P(B)_{coil} = \frac{V^2 \cos(\theta_z)}{2 \cdot |Z|} \quad (10a)$$

$$P(B) = k \cdot \frac{\cos(\tan^{-1}(R(B) \cdot 4.473))}{\sqrt{\frac{23.1}{R(B)^2} + 462.25}} \propto P(B)_{coil} \quad (10b)$$

where $P(B)$ is the power actually reaching the sample from the coil, and k is an unknown constant representing the fact than only a fraction of $P(B)_{coil}$ reaches the sample to heat it. This constant does not depend on the field strength neither on the radiofrequency. It is only determined by the geometrical aspects of the system, so becoming a fitting constant as shown afterwards.

3. Results

The relaxation times at 0.46 T were of 2.6 s for distilled water, 1.1 s for egg-albumin and 0.9 s for glycerol. The observed temperatures behave as exemplified by Fig. 5. It shows that the temperature stabilizes after about 30 minutes of continuous irradiation. The plot shows the measured data at different magnetic field strengths obtained with the fiber optic thermometry system and the fitted curve to obtain maximum temperature increment $\Delta T_{NMR,max}$ and the time constant T_{\uparrow} . It can be observed in the measured temperature the accuracy of the temperature sensor. The maximum increases reflect the temperature dependence with the magnetic field by the magneto-resistive effect, as expected.

We processed the measured heating curves to calculate the thermal power that heats the sample by taking into account the thermal calibration mentioned before (4b). When analyzing the components of the obtained powers, we need, first, to neglect the absorption of the RF power in the sample because of the small size of the sample ($\varnothing 5 \text{ mm} \times 9.5 \text{ mm}$) in relation to the incident wave length (15 m) and because of their dielectric nature. Moreover, at the used frequency, the power loss due to electric coupling is also neglectable. So, changes in the thermal power mea-

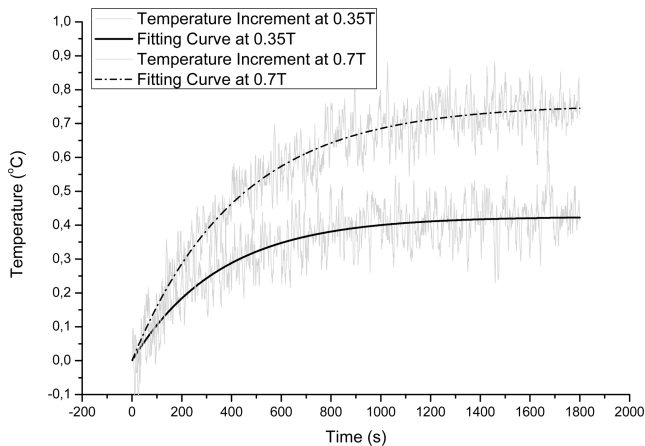


Fig. 5. Example of temperature rise in the sample during power absorption experiments.

measured at the sample must be principally caused by direct heating from the copper antenna, or by internal heating due to the NMR procedure. In absence of NMR (four of the five essayed points of magnetic field strength) we assumed that the thermal power calculated for a sample must be the power coming from the coil, which is known except for the constant k and which is only dependent on the magneto-resistive effect. Therefore, we set the value of k in (10) to match the calculated power curve with the measured thermal powers (out of the NMR condition). The results for the thermal powers corresponding to the egg-albumin, the glycerol and the distilled water samples are shown in Fig. 6. The thermal power heating the samples, triangles, increases as the magnetic field increases,

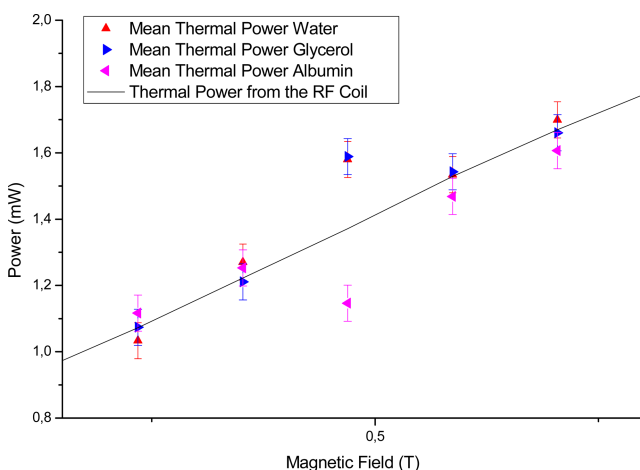


Fig. 6. (Color online) Thermal power actually heating the sample and thermal power dissipated by the RF coil that heats the sample. The measures were made at different magnetic field strengths under the same RF irradiation conditions. The samples were distilled water, glycerol and egg-albumin.

Table 1. ANOVA test representing the probability that all experiments belong to the curve described by the Kohler Rule

Sample	ANOVA	
	All experiments	Without 0.46T experiments
Water	0.02162	0.67876
Glycerol	0.00289	0.97208
Albumin	0.00868	0.61513

in concordance to the predicted magneto-resistive effect, black line.

It is noticed that certain anomalies are presented at the point of 0.46 T, that is, at NMR condition. The measured heating powers differ from the expected at 0.46 T, whilst for the rest of used magnetic strengths the measured powers follow the trend derived from the coil heating. We performed an ANOVA analysis on the data, to question if the NMR points do or do not belong to the calculated power curve, that is, to question if there are or not more thermal effects affecting the sample at the NMR condition, or, contrarily, the observed large differences are only statistical deviations. That is, if the glycerol and the water samples are significantly heated more than their expected value, whereas albumin samples are heated less than the expected value.

In Table 1, the p-value in the ANOVA tests when checking if all the powers belong to the same curve, per each sample type, is clearly under 0.05 (first column), so neglecting the hypothesis: one point at least is not part of the group. When using the other points without including the 0.46 T one, the test does not reject the same hypothesis (second column). Although it is not shown, when not including some other point, but including the 0.46 T, again the p-value is always under 0.05: some point still do not belong to the group formed by the rest, that is, does not follow the trend of heating as determined by the coil heating. Therefore, these results indicate that distilled water and glycerol molecules have similar behaviors, in some way they absorb more energy when they are in NMR conditions. On the other hand, the albumin sample seems to be not as warm as it should be, that is, the albumin sample is not absorbing energy by the NMR phenomenon, even it could be releasing energy.

4. Discussion

We have developed a method to measure tiny thermal powers when performing NMR experiments in small samples in the presence of high magnetic fields, with and without nuclear magnetic resonance condition. In this first work, we have tried three different types of samples, related to

abundant typical organic substances: distilled water, glycerol and egg-albumin. Because of their nature, we can neglect any other cause as heat source that the antenna, a copper coil around the sample that heats itself by Joule effect; and the NMR related phenomena to be measured. To fix the amount of thermal power delivered to the sample from the electric losses in the antenna, whatever the experiment, we use the same RF excitation (frequency and pulse sequencing) while changing the magnetic field strength, so warranting that the NMR takes place at one unique magnetic field value, at which the NMR condition is met. Since we measure temperatures, we have needed to calibrate the NMR equipment to calculate the thermal powers that actually were responsible for the measured temperature changes. The temperature changes out of the NMR condition can be well explained in terms of heating from the antenna selfheating. However, at 0.46 T, when the NMR takes place, some other phenomena appear that should explain the measurement of extra thermal power as compared to the delivered by the coil in the cases of distilled water and glycerol, and the lack of it in the case of egg-albumin. Although the loss of energy as heat in case of NMR has been elsewhere theoretically predicted, it seems that the found amount of power, although very small, is higher than the predicted one. Moreover, the measured lack of power has not been shown in the literature. So, both phenomena seem to be anomalies that deserve further investigations.

Acknowledgements

This work was supported in part by a grant from CONACyT-Mexico to N. Felix-Gonzalez (180890); and a grant by the Technical University of Madrid to A. L. Urbano-Bojorge and N. Felix-Gonzalez.

References

- [1] C. M Weil, Biological effects of radiofrequency radiation, EPA-600/8-83-026A, (1984) pp. 3.2.
- [2] P. Bottomley, *Phys. Med. Biol.* **23**, 630 (2002).
- [3] F. G. Shellock, *Ann. N. Y. Acad. Sci.* **649**, 260 (1992).
- [4] F. G. Shellock, *J. Magn. Reson. Imaging* **12**, 30 (2000).
- [5] P. A. Bottomley, *Med. Phys.* **8**, 510 (1981).
- [6] M. J. Hagmann, *IEEE T. Microw. Theory* **27**, 804 (1979).
- [7] P. Röschmann, *Med. Phys.* **14**, 922 (1987).
- [8] T. F. Budinger, *IEEE T. Nucl. Sci.* **2**, 2821 (1979).
- [9] D. Sperber, *Naturwissenschaften* **71**, 100 (1984).
- [10] F. G. Shellock, *Magn. Reson. Med.* **11**, 371 (1989).
- [11] D. L. Parker, *IEEE T. Biomed. Eng.* **31**, 161 (1984).
- [12] H. Van Koningsveld, *Rcl. Trav. Chim. Pay-B* **87**, 243 (1968).
- [13] S. Sugio, *Protein Eng.* **12**, 439 (1999).
- [14] R. L. White, *IEEE T. Magn.* **28**, 2482 (1992).
- [15] P. Andris, *Meas. Sci. Rev.* **1**, 115 (2001).
- [16] F. Fickett, *Proceedings of the Fourth International Conference on Magnet Technology*, Brookhaven National Laboratory, Upton, NY, (1972) pp. 539-541.