

RESEARCH ARTICLE

Bioinformatics Analysis Reveals Significant Genes and Pathways to Target for Oral Squamous Cell Carcinoma

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Abstract

Purpose: The purpose of our study was to explore the molecular mechanisms in the process of oral squamous cells carcinoma (OSCC) development. **Method:** We downloaded the affymetrix microarray data GSE31853 and identified differentially expressed genes (DEGs) between OSCC and normal tissues. Then Gene Ontology (GO) and Protein-Protein interaction (PPI) networks analysis was conducted to investigate the DEGs at the function level. **Results:** A total 372 DEGs with $\log_{2}FC > 1$ and P value < 0.05 were obtained, including NNMT, BAX, MMP9 and VEGF. The enriched GO terms mainly were associated with the nucleoplasm, response to DNA damage stimuli and DNA repair. PPI network analysis indicated that GMNN and TSPO were significant hub proteins and steroid biosynthesis and synthesis and degradation of ketone bodies were significantly dysregulated pathways. **Conclusion:** It is concluded that the genes and pathways identified in our work may play critical roles in OSCC development. Our data provides a comprehensive perspective to understand mechanisms underlying OSCC and the significant genes (proteins) and pathways may be targets for therapy in the future.

Keywords: Bioinformatics - oral SCC - differentially expressed genes - protein-protein interactions - pathways

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Introduction

Oral squamous cell carcinoma (OSCC) is one of the common solid tumors originating from abnormal squamous cell in oral region. The incidence of OSCC is highest (80%) among head and neck squamous cell carcinomas (LandisMurray et al., 1999). OSCC is characterized by invasion of epithelial tumor cells to underlying tissues and more prevalent in old population (Deyhimi et al., 2013; Neville et al., 2002). OSCC is often asymptomatic in early stage until it is advanced to late stage (Scott et al., 2005). The diagnosis of OSCC is usually delayed and the five-year survival rates among the patients with advanced oral cancers were only 20% (Spiro, 1985; Mashberg and Samit, 1989). OSCC is a serious global health issue remained to be resolved.

OSCC with remarkable incidence and fairly poor prognosis have encouraged many studies to explore the underlying mechanism of OSCC development. Some genes were indicated to be aberrantly methylated in the progression of OSCC such as p16, p15, hMLH1 (Human Mut-L Homologue 1), MGMT (O 6-methylguanine-DNA methyltransferase) and E-cadherin (Viswanathan et al., 2003). Patients with these methylated genes were indicated to have high risk for OSCC. In addition, some cytokines and pathways were reported to play a key role in OSCC development. The expression of interleukin-8 (IL-8) was demonstrated to be elevated and speculated to contribute to the invasion of oral squamous tumor cell by regulating

matrix metalloproteinase pump-1 (MMP-7) expression (Watanabe et al., 2002). Other cytokines and pathways closely associated with OSCC include cyclooxygenase-2 (Chan et al., 1999), myeloid cell leukemia-1 (Shin et al., 2013), serine/threonine kinase signaling pathway (Gao et al., 2013) and PD-1/PD-L1 (the inhibitory receptor Programmed Death-1/Programmed cell death ligand 1) pathway (Lyford-Pike et al., 2013).

Although immeasurable contributions have been made, the molecular mechanism of OSCC seems to be less well clarified. In this work, we use bioinformatics methods to explore the differentially expressed genes (DEGs) between oral squamous carcinoma tissues and normal tissues. And the GO (Gene Ontology) analysis was constructed to investigate the critical genes in the progression of OSCC. Furthermore, protein-protein interaction (PPI) networks construction and pathway enrichment analysis was performed to estimate the significant pathways. This work provided a systematic perspective to understand the underlying mechanism in OSCC development.

Materials and Methods

Affymetrix microarray data

The transcription profile of GSE31853 was obtained from GEO (Gene Expression Omnibus) database (<http://www.ncbi.nlm.nih.gov/geo/>). Total 11 chips were deposited by YapJenei et al. (2009). In this paper, we only selected 9 samples for analysis including 8 samples of oral

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Table 1. GO Analysis for Up-regulated Genes

Category	Term		Count	P value
Cluster 1	Enrichment Score: 6.3396960052874975			
CC_FAT	GO:0005654	nucleoplasm	18	4.07E-09
CC_FAT	GO:0031981	nuclear lumen	21	3.33E-08
CC_FAT	GO:0031974	membrane-enclosed lumen	23	7.54E-08
CC_FAT	GO:0043233	organelle lumen	22	2.87E-07
CC_FAT	GO:0070013	intracellular organelle lumen	21	9.95E-07
CC_FAT	GO:0005730	nucleolus	9	0.003131
Cluster 2	Enrichment Score: 5.608042749152675			
BP_FAT	GO:0006259	DNA metabolic process	19	1.22E-12
BP_FAT	GO:0006974	response to DNA damage stimulus	11	3.06E-06
BP_FAT	GO:0033554	cellular response to stress	12	2.00E-05
BP_FAT	GO:0006281	DNA repair	9	2.31E-05
BP_FAT	GO:0006302	double-strand break repair	5	1.32E-04
BP_FAT	GO:0006310	DNA recombination	5	9.87E-04
Cluster 3	Enrichment Score: 5.001798060654299			
CC_FAT	GO:0005694	chromosome	14	4.93E-09
CC_FAT	GO:0044427	chromosomal part	12	8.52E-08
CC_FAT	GO:0000793	condensed chromosome	7	6.33E-06
CC_FAT	GO:0000228	nuclear chromosome	7	2.33E-05
CC_FAT	GO:0043228	non-membrane-bounded organelle	23	2.60E-05
CC_FAT	GO:0043232	intracellular non-membrane-bounded organelle	23	2.60E-05
BP_FAT	GO:0051276	chromosome organization	10	1.77E-04
BP_FAT	GO:0006323	DNA packaging	4	0.013146

squamous carcinoma tissues and 1 sample of normal tissue based on the platform of GPL96 [HG-U133A] Affymetrix Human Genome U133A Array. The raw data and the probe annotation files were downloaded for further analysis.

Differentially expressed genes (DEGs) analysis

The raw data in CEL files were normalized using RMA (Robust Multiarray Analysis) algorithm in R Affy package (IrizarryHobbs et al., 2003). Then 9 datasets were assigned into two groups: oral squamous carcinoma group (8 samples) and control group (1 sample). The differentially expressed genes between oral squamous carcinoma tissue and normal tissue were estimated by T-test of limma package (DibounWernisch et al., 2006). A mass of LogFC and P values were obtained and we set $\log_2FC > 1$ and $P < 0.05$ as the cutoff criterion.

Gene Ontology (GO) analysis

Gene Ontology (GO) is a freely available tool for gene or gene sequence annotation, which concerns domains of molecular and cellular biological (HarrisClark et al., 2004). DAVID bioinformatics resources possesses abundant tools for systematic and integrative analysis of large lists of genes (Da Wei Huangand Lempicki, 2008). The DEGs obtained in this paper were divided into two groups including up-regulated group and down-regulated group. To investigate the differentially expressed genes in a functional level, DEGs in two groups underwent GO analysis independently by using DAVID (Huang daSherman et al., 2007). Finally, we identified the over-represented GO categories with $P < 0.05$.

Protein-protein interaction (PPI) network

Protein-protein interactions can help to understand functions of proteins in molecule level and explore cell regulatory mechanism. And the interactions of proteins can be inferred from the respective interactions of genes

coded them (Von MeringHuynen et al., 2003a). STRING database is used for evaluating and pre-computing proteins interactions on global view (von MeringHuynen et al., 2003b) which contains dataset of 89 whole genomes sequencing including 261033 orthologous genes.

The DEGs in up-regulated and down-regulated group were mapped into PPI network by cytoscape software. The interactions between two proteins were scored by STRING 9.0 tool. The protein pairs with Required Confidence Score > 4 were defined to have close interactions.

Cluster analysis for PPI networks

We performed cluster analysis for PPI networks using ClusterONE of Cytoscape software and set P value $< 1.0E-5$ to select the enriched modules.

The Kyoto Encyclopedia of Genes and Genomes (KEGG) is a database consists of a large scale of genome information which can provide classifications of predicted genes to their respective pathways (Altermannand Klaenhammer, 2005). Then the KEGG pathway analysis was conducted to evaluate the modules in functional level by DAVID online tool. We defined the significant pathways with P value < 0.05 .

Results

Differentially expressed genes (DEGs)

To explore the DEGs between oral squamous carcinoma tissues and normal tissues, we applied limma package to analyze the GSE31853 dataset from GEO. Finally, we selected 372 DEGs with $\log_2FC > 1$ and P value < 0.05 including 67 up-regulated genes such as NNMT (Nicotinamide N-methyltransferase), VEGFA (Vascular endothelial growth factor) and 305 down-regulated genes such as MMP9 (Matrix Metalloproteinases-9), LAMC2 (gamma two chain gene), BAX (Bcl-2-associated X protein) and CD44 (a cell-surface glycoprotein). The

Table 2. GO Analysis for Down-regulated Genes

Category	Term		Count	P Value
Cluster 1	Enrichment Score: 3.961083343919374			
BP_FAT	GO:0016126	sterol biosynthetic process	11	4.94E-10
BP_FAT	GO:0006695	cholesterol biosynthetic process	8	3.39E-07
BP_FAT	GO:0016125	sterol metabolic process	13	3.54E-07
BP_FAT	GO:0006694	steroid biosynthetic process	12	4.58E-07
BP_FAT	GO:0008202	steroid metabolic process	16	5.46E-06
BP_FAT	GO:0008203	cholesterol metabolic process	11	7.67E-06
BP_FAT	GO:0008610	lipid biosynthetic process	19	3.38E-05
BP_FAT	GO:0006720	isoprenoid metabolic process	7	1.50E-04
BP_FAT	GO:0008299	isoprenoid biosynthetic process	5	4.44E-04
CC_FAT	GO:0005789	endoplasmic reticulum membrane	14	0.002148
CC_FAT	GO:0042175	nuclear envelope-endoplasmic reticulum network	14	0.003409
CC_FAT	GO:0031090	organelle membrane	35	0.003683
CC_FAT	GO:0005783	endoplasmic reticulum	31	0.005727
CC_FAT	GO:0012505	endomembrane system	26	0.008702
Cluster 2	Enrichment Score: 3.67837332129948			
CC_FAT	GO:0044421	extracellular region part	41	2.62E-06
CC_FAT	GO:0005615	extracellular space	26	0.001598
CC_FAT	GO:0005576	extracellular region	57	0.002205
Cluster 3	Enrichment Score: 3.6440749425044983			
MF_FAT	GO:0004866	endopeptidase inhibitor activity	12	9.11E-05
MF_FAT	GO:0030414	peptidase inhibitor activity	12	1.47E-04
MF_FAT	GO:0004867	serine-type endopeptidase inhibitor activity	9	3.32E-04
MF_FAT	GO:0004857	enzyme inhibitor activity	15	5.96E-04

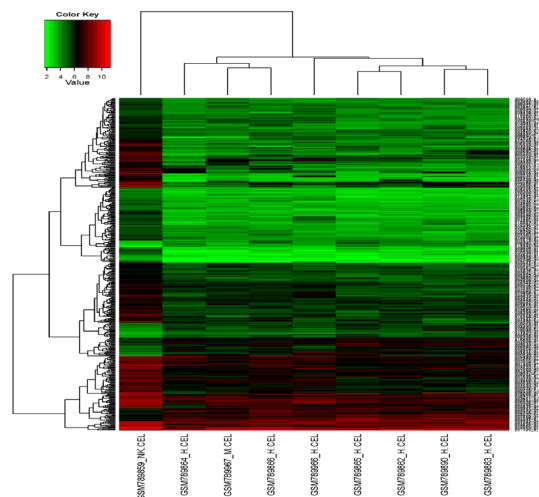


Figure 1. The Heat Map of DEGs. The x-coordinate represents sample symbols (from left to right: 1 sample of normal tissue and 8 samples of oral squamous carcinoma tissues), y-coordinate represents differentially expressed probes. Green: low expression and Red: high expression

DEGs expression heat map was shown in Figure 1.

GO enrichment analysis

To identify the functions of the DEGs, we performed GO analysis and set $P < 0.05$ as the cutoff value. The results were shown in Table 1 and 2. The DEGs mainly enriched in 3 GO categories including Biological Process (BP), Molecular Function (MF) and Cellular Component (CC). The up-regulated DEGs mainly enriched in CC and BP. The CC involved GO terms contained nucleoplasm, nuclear lumen, membrane-enclosed lumen, organelle lumen, intracellular organelle lumen and nucleolus. And the BP associated GO terms included DNA metabolic process, response to DNA damage stimulus, cellular response to stress, DNA repair, double-strand break repair,

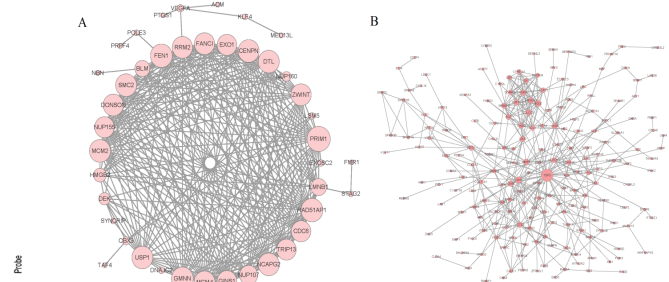


Figure 2. PPI Networks of DEGs. A: PPI network of up-regulated genes. B: PPI network of down-regulated genes. Nodes represent proteins, edges represent interactions between two proteins. The higher the node shape, the greater degree of connection

DNA recombination, chromosome, chromosomal part, condensed chromosome and nuclear chromosome.

The most enriched GO terms of down-regulated DEGs were relevant with BP (sterol metabolism, cholesterol metabolism, steroid metabolism and isoprenoids metabolism), endoplasmic reticulum related CC (endoplasmic reticulum membrane, nuclear envelope-endoplasmic reticulum network, endoplasmic reticulum) and MF associated with enzymic inhibition (endopeptidase inhibitor activity, peptidase inhibitor activity, serine-type endopeptidase inhibitor activity and enzyme inhibitor activity).

PPI network construction

After demonstrated the protein interacting pairs with required confidence score > 4 , we established the PPI networks of up-regulated and down-regulated genes by cytoscape software. As shown in figure 2 A, the up-regulated network included 42 nodes, 282 edges and the significant hub proteins containing PRIM1 (DNA primase 1, Degree=26), RAD51AP1 (RAD51 associating protein-1, Degree=24), FEN1 (DNase IV, Degree=24),

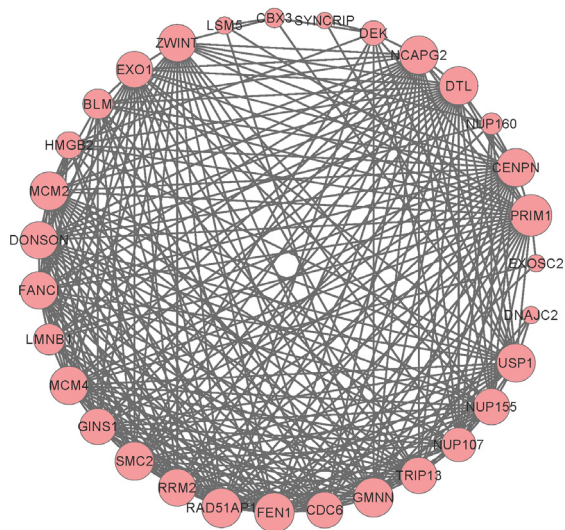


Figure 3. Cluster Analysis for Up-regulated Network (P value=0). Nodes represent proteins, edges represent interactions between two proteins. The higher the node shape, the greater degree of connection

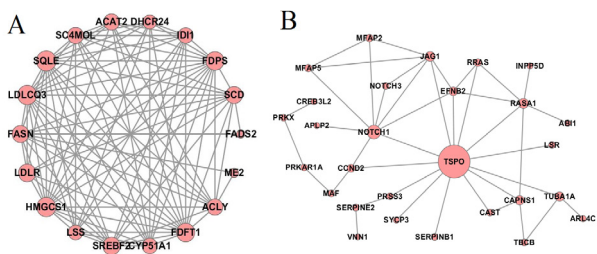


Figure 4. Cluster Analysis for Down-regulated Network. A: P value=5.914E-7. B: P value=1.488E-6. Nodes represent proteins, edges represent interactions between two proteins. The higher the node shape, the greater degree of connection

GMNN (Geminin, Degree=24). The most significant hub protein in the down-regulated PPI network (195 nodes and 439 edges) was TSPO (Translocator Protein) with Degree=43 (Figure 2 B).

Cluster analysis for PPI networks

In order to explore the functions of the modules of networks, we implemented the cluster analysis. As shown in Figure 3, 1 module (P value <1.0E-5) of up-regulated PPI network was obtained with 31 nodes and 271 edges. In down-regulated PPI network, there were 2 enrichment modules: A with 18 nodes and 97 edges, B with 28 nodes and 41 edges (Figure 4).

After KEGG pathway analysis for the enriched modules, we obtained significant pathways with P value<0.05 (Figure 5). In the up-regulated module, the significant pathways were DNA replication and Cell cycle, while in the down-regulated modules, the significant pathways in A and B were Steroid biosynthesis, Terpenoid backbone biosynthesis, synthesis and degradation of ketone bodies; Notch signaling pathway, respectively.

Discussion

Oral squamous cell carcinoma (OSCC) with ranked mortality rate has been a health problem hotly discussed

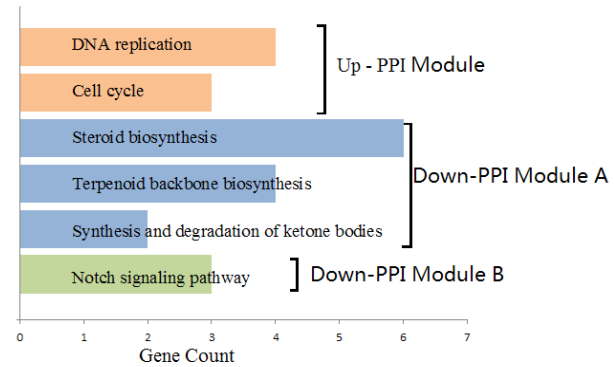


Figure 5. KEGG Pathway Analysis for PPI Networks. x-coordinate represents gene count; y-coordinate represents pathway term. Orange: pathways in up-regulated PPI network; Blue: pathways in down-regulated network A; Green: pathways in down-regulated network B

all over the world (Lo et al., 2007). A comprehensive analysis of the mechanism underlying OSCC development is of crucial importance for management policy. In this paper, we provided a systematic perspective to understand the mechanism using bioinformatics method. We first analyzed the differentially expressed genes (DEGs) between oral squamous carcinoma tissues and normal tissues. Besides we performed GO analysis to evaluate the DEGs in function level and construct the protein-protein interaction (PPI) networks for significant pathways.

Results showed that there were total 372 DEGs detected in our study. The differentially expressed genes including NNMT, BAX, MMP9 and VEGF have been reported to be tightly associated with OSCC disease. NNMT is indicated to be significantly up-regulated in tumor tissues compared with normal tissue in our study. NNMT coded for Nicotinamide N-methyltransferase play a key role in biotransformation of drug or other xenobiotics. A previous study suggested that NNMT, a novel gene, regulated cell migration which was necessary for cancer cell invasion and tumor stage (Wu et al., 2008). The expression of NNMT was extremely elevated in the progression and development of several tumor cells. It has been reported that the NNMT expression is related with differentiation of oral cancer cells and may be a biomarker for OSCC treatment (Emanuelli et al., 2010). A recent study revealed that the abnormal expression of NNMT was closely associated with the metabolism of oral squamous carcinoma cells (Pozzi et al., 2013). The decreased expression of NNMT was able to suppress cancer cell proliferation and tumor development. The inhibition of NNMT showed potential capacity for oral cancer treatment by molecular approach (Pozzi et al., 2013). Besides, MMP9 (Matrix Metalloproteinases-9) is reported to be significantly activated in malignant tissues of patients with oral cancer (Patel et al., 2007). The expressions of BAX and VEGF are demonstrated to be the prognosis biomarker for OSCC (Bose et al., 2012; Kämmereret al., 2012).

After GO analysis, the results showed that the over-represented GO terms were related with nucleoplasm, response to DNA damage stimulus and DNA repair. Nucleoplasm plays a fundamental role in the proliferation of cancer cells (Tsai and McKay, 2002). Nucleophosmin

(NPM1) has multiple functions in transcription, centrosome duplication, ribosome biogenesis and genomic stability. The overexpression of acetylated NPM1 is found to be existing in high grade of OSCC (Shandilya et al., 2009). Acetylated NPM1 primarily located in nucleoplasm and regulated cell survival and proliferation of oral cancer cells (Shandilya et al., 2009). Endogenous DNA damage increased the risk of cancer development (Loft and Poulsen, 1996). Cdc7-Dbf4 kinase (Dbf4-dependent kinase, DDK) is a critical factor regulating DNA replication and DNA damage (Costanzo et al., 2003). Cdc7 was found to affect the outcome of OSCC patients with chemotherapy. The up-regulated Cdc7 enhanced drug resistance of patients by suppressing the apoptosis and promoting DNA repair of oral squamous cancer cells (Cheng et al., 2013).

Furthermore the results from PPI network analysis indicated that the significant hub proteins (GMNN and TSPO) and the obviously dysregulated pathways (Steroid biosynthesis and Synthesis and degradation of ketone bodies) played key roles in OSCC development. GMNN is a critical factor in cell cycle, especially in S phase to M phase transition (Ma et al., 2012). Recently, GMNN was revealed to be a common cancer gene and amplified in tumor cells (Kim et al., 2012). It was also found that the expression of GMNN was increased in oral epithelial dysplasia (OED) cases, likewise the GMNN/Ki67 ratios were also significantly increased compared with premalignant and malignant tumours (Torres-Rendon et al., 2009). GMNN was considered to be a prognostic biomarker for OSCC progression. TSPO (translocator protein) was found to be involved in cell proliferation, tumor invasion, and metastasis (Batarseh and Papadopoulos, 2010). There was a significant increase of TSPO expression in oral cancer tumors compared with adjacent normal tissues. And patients with high levels of TSPO showed low five-year survival rate (Nagler et al., 2010). Although the detailed mechanism of TSPO affecting on OSCC is far from being clear, TSPO is of critical importance in OSCC development and progression.

Steroid biosynthesis pathway was found to be dysregulated compared with Oral squamous cancer cells and normal cells. The reports concerning the role of steroid in OSCC are relatively rare. A recent study suggested that the steroid hormones (estrogen beta, ER β) was abundantly expressed in oral squamous cancer cells from both female and male patients (Marocchio et al., 2013). The detection of steroid expression (ER β) may helpful to understand the role of these proteins in OSCC progression. Another significant pathway in this paper was found to be synthesis and degradation of ketone bodies. The production of ketone bodies has been found in the systemic metabolic response to early stage oral cancer. And the accumulation of ketone bodies is more pronounced in later stage cancer (Tiziani et al., 2009). The synthesis of ketone bodies was considered to be a marker for therapeutic implication of cancers (Veech, 2004). Recently, a novel compound BBSKE (1, 2-[bis (1, 2-Benzisoseleazolone-3 (2H)-ketone)]ethane) was reported to possess anti-cancer properties and have potential therapeutic effect on oral squamous cell carcinoma (OSCC) (Xing et al., 2008).

BBSKE showed significant effect on inhibiting cancer cell proliferation and inducing cancer cell apoptosis. The synthesis and degradation of ketone bodies was of crucial importance throughout the process of OSCC development.

In summary, the differentially expressed genes, hub proteins and pathways identified in our work showed close association with the progression of OSCC. The bioinformatics analysis provided a comprehensive perspective to understand the mechanism underlying OSCC development. The significant gene and pathways may be targets of treatment management for OSCC. However, further investigations are still necessary for unraveling the mechanism in the process of OSCC development.

References

- Altermann E, Klaenhammer TR (2005). PathwayVoyager: pathway mapping using the Kyoto Encyclopedia of Genes and Genomes (KEGG) database. *BMC Genomics*, **6**, 60.
- Batarseh A, Papadopoulos V (2010). Regulation of translocator protein 18kDa (TSPO) expression in health and disease states. *Mol Cell Endocrinol*, **327**, 1-12.
- Bose P, Klimowicz A, Kornaga E, et al (2012). Bax expression measured by AQUAnalysis is an independent prognostic marker in oral squamous cell carcinoma. *BMC Cancer*, **12**, 332.
- Chan G, Boyle JO, Yang EK, et al (1999). Cyclooxygenase-2 expression is up-regulated in squamous cell carcinoma of the head and neck. *Cancer Res*, **59**, 991-4.
- Cheng AN, Jiang SS, Fan CC, et al (2013). Increased Cdc7 expression is a marker of oral squamous cell carcinoma and overexpression of Cdc7 contributes to the resistance to DNA-damaging agents. *Cancer Lett*, **337**, 218-25.
- Costanzo V, Shechter D, Lupardus PJ, et al (2003). An ATR- and Cdc7-dependent DNA damage checkpoint that inhibits initiation of DNA replication. *Mol Cell*, **11**, 203-13.
- Deyhimi P, Torabinia N, Torabinia A (2013). A comparative study of histological grade and expression of Ki67 protein in oral squamous cell carcinoma in young and old patients. *Dental Res J*, **10**, 514.
- Diboun I, Wernisch L, Orengo C A, Koltzenburg M (2006). Microarray analysis after RNA amplification can detect pronounced differences in gene expression using limma. *BMC Genomics*, **7**, 252.
- Emanuelli M, Santarelli A, Sartini D, et al (2010). Nicotinamide N-Methyltransferase upregulation correlates with tumour differentiation in oral squamous cell carcinoma. *Histol Histopathol*, **25**, 15.
- Gao J, Li X, Jiang J (2013). [The study of serine/threonine kinase signaling pathway-mediated inhibition of proliferation and invasion of oral squamous cell carcinoma transfected with p53 gene]. *Hua Xi Kou Qiang Yi Xue Za Zhi*, **31**, 145-9.
- Harris M, Clark J, Ireland A, et al (2004). The Gene Ontology (GO) database and informatics resource. *Nucleic Acids Res*, **32**, D258-61.
- Huang da W, Sherman BT, Lempicki RA (2008). Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc*, **4**, 44-57.
- Huang da W, Sherman BT, Tan Q, et al (2007). The DAVID Gene Functional Classification Tool: a novel biological module-centric algorithm to functionally analyze large gene lists. *Genome Biol*, **8**, R183.
- Irizarry RA, Hobbs B, Collin F, et al (2003). Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics*, **4**, 249-64.

- Kämmerer P, Koch F, Schiegnitz E, et al (2012). Associations between single-nucleotide polymorphisms of the VEGF gene and long-term prognosis of oral squamous cell carcinoma. *J Oral Pathol Med*, **42**, 374-81.
- Kim HE, Kim DG, Lee KJ, et al (2012). Frequent amplification of CENPF, GMNN and CDK13 genes in hepatocellular carcinomas. *PLoS One*, **7**, e43223.
- Landis SH, Murray T, Bolden S, Wingo PA (1999). Cancer statistics, 1999. *CA Cancer J Clin*, **49**, 8-31.
- Lo WY, Tsai MH, Tsai Y, et al (2007). Identification of over-expressed proteins in oral squamous cell carcinoma (OSCC) patients by clinical proteomic analysis. *Clin Chim Acta*, **376**, 101-7.
- Loft S, Poulsen H (1996). Cancer risk and oxidative DNA damage in man. *J Mol Med*, **74**, 297-312.
- Lyford-Pike S, Peng S, Young GD, et al (2013). Evidence for a role of the PD-1: PD-L1 pathway in immune resistance of HPV-associated head and neck squamous cell carcinoma. *Cancer Res*, **73**, 1733-41.
- Ma JY, Li M, Ge ZJ, et al (2012). Whole transcriptome analysis of the effects of type I diabetes on mouse oocytes. *PLoS One*, **7**, e41981.
- Marocchio LS, Giudice F, Corrêa L, et al (2013). Oestrogens and androgen receptors in oral squamous cell carcinoma. *Acta Odontol Scand*, **71**, 1513-9.
- Mashberg A, Samit AM (1989). Early detection, diagnosis, and management of oral and oropharyngeal cancer. *CA Cancer J Clin*, **39**, 67-88.
- Nagler R, Ben-Izhak O, Savulescu D, et al (2010). Oral cancer, cigarette smoke and mitochondrial 18kDa translocator protein (TSPO) - In vitro, in vivo, salivary analysis. *Biochim Biophys Acta*, **1802**, 454-61.
- Neville B, Damm D, Allen C, Bouquot J (2002). Oral and Maxillofacial Pathology (ed 2) Saunders. Philadelphia, PA, 582-3.
- Patel BP, Shah SV, Shukla SN, et al (2007). Clinical significance of MMP-2 and MMP-9 in patients with oral cancer. *Head Neck*, **29**, 564-72.
- Pozzi V, Sartini D, Morganti S, et al (2013). RNA-mediated gene silencing of nicotinamide N-Methyltransferase is associated with decreased tumorigenicity in human oral carcinoma cells. *PLoS One*, **8**, e71272.
- Scott S, Grunfeld E, McGurk M (2005). The idiosyncratic relationship between diagnostic delay and stage of oral squamous cell carcinoma. *Oral Oncol*, **41**, 396-403.
- Shandilya J, Swaminathan V, Gadad SS, et al (2009). Acetylated NPM1 localizes in the nucleoplasm and regulates transcriptional activation of genes implicated in oral cancer manifestation. *Mol Cell Biol*, **29**, 5115-27.
- Shin JA, Jung JY, Ryu MH, et al (2013). Mithramycin A inhibits myeloid cell leukemia-1 to induce apoptosis in oral squamous cell carcinomas and tumor xenograft through activation of Bax and oligomerization. *Mol Pharmacol*, **83**, 33-41.
- Spiro RH (1985). Squamous cancer of the tongue. *CA Cancer J Clin*, **35**, 252-6.
- Tiziani S, Lopes V, Günther UL (2009). Early stage diagnosis of oral cancer using 1H NMR-based metabolomics. *Neoplasia*, **11**, 269.
- Torres-Rendon A, Roy S, Craig G, Speight P (2009). Expression of Mcm2, geminin and Ki67 in normal oral mucosa, oral epithelial dysplasias and their corresponding squamous-cell carcinomas. *Br J Cancer*, **100**, 1128-34.
- Tsai RY, McKay RD (2002). A nucleolar mechanism controlling cell proliferation in stem cells and cancer cells. *Genes Dev*, **16**, 2991-3003.
- Veech RL (2004). The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism. *Prostaglandins Leukot Essent Fatty Acids*, **70**, 309-19.
- Viswanathan M, Tsuchida N, Shanmugam G (2003). Promoter hypermethylation profile of tumor-associated genes p16, p15, hMLH1, MGMT and E-cadherin in oral squamous cell carcinoma. *Int J Cancer*, **105**, 41-6.
- Von Mering C, Huynen M, Jaeggi D, et al (2003a). STRING: a database of predicted functional associations between proteins. *Nucleic Acids Res*, **31**, 258-61.
- von Mering C, Huynen M, Jaeggi D, et al (2003b). STRING: a database of predicted functional associations between proteins. *Nucleic Acids Res*, **31**, 258-61.
- Watanabe H, Iwase M, Ohashi M, Nagumo M (2002). Role of interleukin-8 secreted from human oral squamous cell carcinoma cell lines. *Oral Oncol*, **38**, 670-9.
- Wu Y, Siadaty M, Berens M, et al (2008). Overlapping gene expression profiles of cell migration and tumor invasion in human bladder cancer identify metallothionein 1E and nicotinamide N-methyltransferase as novel regulators of cell migration. *Oncogene*, **27**, 6679-89.
- Xing F, Li S, Ge X, et al (2008). The inhibitory effect of a novel organoselenium compound BBSKE on the tongue cancer Tca8113 in vitro and in vivo. *Oral Oncol*, **44**, 963-9.
- Yap LF, Jenei V, Robinson CM, et al (2009). Upregulation of Eps8 in oral squamous cell carcinoma promotes cell migration and invasion through integrin-dependent Rac1 activation. *Oncogene*, **28**, 2524-34.