

# Changes in the Laboratory Data for Cancer Patients Treated with Korean-medicine-based Inpatient Care

Jeungwon Yoon<sup>1</sup>, Chong-Kwan Cho<sup>1</sup>, Ji-Eun Shin<sup>2</sup>, Hwa-Seung Yoo<sup>1\*</sup>

## **Key Words**

cancer coagulation, cancer immunity, fibrinogen, natural killer cells, Korean medicine, complementary alternative medicine

#### **Abstract**

**Objectives:** The study aimed to determine changes in laboratory data for cancer patients receiving Korean medicine (KM) care, with a focus on patients' functional status, cancer-coagulation factors and cancer immunity.

Methods: We conducted an observational study of various cancer patients in all stages admitted to the East-West Cancer Center (EWCC), Dunsan Korean Hospital of Daejeon University, from Mar. 2011 to Aug. 2011. All patients were under the center's multi-modality Korean-medicine-based inpatient cancer care program. The hospitalization stay at EWCC ranged from 9 to 34 days. A total of 80 patients were followed in their routine hematologic laboratory screenings performed before and after hospitalization. Patients were divided into three groups depending on the status of their treatment: prevention of recurrence and metastasis group, KM treatment only group, and combination of conventional and KM treatment group. The lab reports included natural killer cell count (CD16 + CD56), fibrinogen, white blood cell (WBC), lymphocytes, monocytes, neutrophil, red

blood cell (RBC), hemoglobin, platelet, Erythrocyte Sedimentation Rate (ESR), and Eastern Cooperative Oncology Group (ECOG) performance status.

Results: With a Focus on patients' functional status, cancer-coagulation factors and cancer immunity, emphasis was placed on the NK cell count, fibrinogen count, and ECOG scores. Data generally revealed decreased fibrinogen count, fluctuating NK cell count and decreased ECOG, meaning improved performance status in all groups. The KM treatment only group showed the largest decrease in mean fibrinogen count and the largest increase in mean NK cell count. However, the group's ECOG score showed the smallest decrease, which may be due to the concentration of late-cancer-stage patients in that particular group.

**Conclusions:** Multi-modality KM inpatient care may have positive effect on lowering the cancer coagulation factor fibrinogen, but its correlation with the change in the NK cell count is not clear.

#### 1. Introduction

Cancer is a hyperproliferative disorder that involves morphological cellular transformation, dysregulation of apoptosis, uncontrolled cellular proliferation, invasion, angiogenesis, and metastasis [1]. As the disease runs its course, the human body, which is made up of several components that together make a system, is in

<sup>&</sup>lt;sup>1</sup> East-West Cancer Center, Dunsan Korean Hospital of Daejeon University, Daejeon, Korea

<sup>&</sup>lt;sup>2</sup> Department of Statistics, Chungnam National University, Daejeon, Korea

 <sup>⊕</sup> This is an Open-Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

a constant cycle of attack, battle, and recovery. Numerous studies have recognized an association between the key immune, coagulation and inflammatory factors and the biological status of the body during the processes [1-3]. NK cells, which are innate immune system cells, are localized in peripheral blood, lymph nodes, the spleen, and bone marrow. NK cells constitutively express a lytic mechanism able to kill target cells independently from any previous activation. While the cytotoxic T-cells recognize infected cells by their peptides, the major histocompatibility complex (MHC) class I molecules present, NK cells look for the absence of MHC class I molecules. One proposal is that NK cells complement cytotoxic T-cells by taking charge in situations where MHC class I expression is hindered through the effects of virus infection [4]. Once Natural Killer (NK) cells are activated, they release small cytoplasmic granules of proteins called perforin and granzyme, which cause the target cell to go through apoptosis. These functional features have suggested a role for NK cells in the control of tumor growth and metastasis.

Fibrinogen, the major component of clots also has important roles in angiogenesis. Tumor-associated blood vessels are typically leaky; hence, fibrinogen and other clotting molecules extravasate from the defective vessels and form fibrin deposits around the tumor. These deposits provide a pro-angiogenic matrix that facilitates endothelial cell adhesion, migration, proliferation, and differentiation into tubules [5]. Formation of the fibrin matrix allows the malignant cells to adhere to and invade the tissues in metastatic sites and protects the cancer cells from immune surveillance [6-8]. The fibrin matrix also supports the migration of tumor cells and provides a scaffold for the formation of new blood vessels [9].

Hematology lab testing follows the numbers of cells in blood, including red blood cells (RBC), white blood cells (WBC), platelets and others. Symptoms such as anemia, infection, and abnormal bleeding are looked for to determine how the body is dealing with the chemotherapy drugs or radiation treatments and to follow up on the general state of the patient's health. Studies report that Korean Medicine (KM) treatments have anti-coagulative effects under hyper-coagulative conditions [10-12], and correlations between KM treatment and the promotion of immune response have also been reported [13-15]. With the cross-link between inflammation, immunity, coagulation, and angiogenesis playing important roles in cancer progression and the treatment prognosis [1-3], the effect of KM inpatient care at the East-West Cancer Center (EWCC), Dunsan Korean Medical Hospital of Daejeon University, was investigated by following the cancer patients' hematological lab data and their quality-of-life reports.

## 2. Methods

The study population consisted of inpatients who were admitted to the East-West Cancer Center, Dunsan Korean Medical Hospital of Daejeon University, Korea, from March 1, 2011 to August 31, 2011. Patients were histologically or cytologically diagnosed with various cancers and received wheel balanced cancer therapy (WBCT), the center's inpatient multi-modality Complementary and Alternative Medicine (CAM) cancer program, for durations ranging from 9 to 34 days. A total of 80 patients, 57 females and 23 males with a mean age of 48.95 years (range: 27 - 82 years), were followed, and routine laboratory screenings were performed. All variables, including demographic, clinical and pathological findings, were recorded. The mean hospitalization period was 16.75 days. As a retrospective follow up, all data were collected as part of routine clinical practice. All treatments were conducted with prior patient consent and permission was obtained to use or disclose their health information for future research purposes.

All patients received EWCC's multi-modality CAM cancer care program, WBCT. WBCT incorporates various treatment modalities under four sub-programs: (1) anti-cancer nutrition; (2) metabolism activation: acupuncture, pharmacopuncture, thermotherapy, massage, physical therapy, and hydrotherapy; (3) herbal therapy: EWCC's signature anti-angiogenic and immune enhancing herbal prescriptions, which include Hang-am plus (ingredients: Panax Notoginseng Radix, Cordyceps Militaris, Cremastrae appendiculata Tuber, Panax ginseng Radix, Bovis Calculus, Margarita, Boswellia carteri and Commiphora myrrha), Myun-yuk plus (ingredients: Astragalus membranaceus, Salvia miltiorrhiza, Hericium erinaceum, and Panax ginseng) and other herbal formulas to suit each patients symptoms according to the Korean Medicine differential diagnosis; and (4) mind-body program: meditation, qi gong, hiking, etc.

Emphasis was placed on cancer-coagulation factors, immunity, and patients' functional status. The lab reports, including NK cell count (CD16 + CD56), fibrinogen, WBC, lymphocytes, monocytes, neutrophils, RBC, hemoglobin, platelets, ESR, and Eastern Cooperative Oncology Group (ECOG) performance status values [16], which were recorded before and after hospitalization, were closely monitored.

Patients were divided into 3 groups depending on the status of their treatment: prevention of recurrence and metastasis group (prevention group), Korean medicine treatment only group (OM Tx only group), and combination of conventional and Korean medicine treatment group

(combination group). The prevention group included cancer-free patients who were under surveillance for further disease progression. The KM Tx only group and the combination group included patients with cancer who were undergoing either Korean medicine treatment or combined treatment with both Korean medicine and conventional medicine.

Results are given as mean values  $\pm$  standard deviations (SDs). For continuous variables, all statistical analyses were performed using SPSS version 12.0 for Windows (SPSS, Chicago, Ill, USA). The paired t-test was used to compare the changes in the NK cell count, the fibrinogen count, and the ECOG performance status between assessment groups. P-values of less than 0.05 were considered statistically significant.

## 3. Results

## 3.1. Laboratory changes in patients

The mean lab values checked before and after hospitalization of all cancer patients who received inpatient care at the EWCC showed minimal changes in most measures. The ranges of the lab values showed slight tendencies to move closer to the reference ranges, but showed no remarkable changes. The mean ECOG performance status of the patients dropped from 2.01  $\pm$  0.19 before to 1.08  $\pm$  0.63 after hospitalization. The bottom status for patients after hospitalization was 0, fully active and able to carry on all pre-disease activities without restriction, whereas the bottom status for patients before hospitalization was 1, restricted in physically-strenuous activity but ambulatory

Table 1 Changes in lab values and ECOG before and after inpatient care

	Before Hospitalization	After Hospitalization	Reference Range	
	(n = 80)	(n = 80)	(n = 80)	
NK Cell (CD16+CD56)	12.01 ± 8.53	14.77 ± 9.61	5 - 35	
	(0.10 - 51.0)	(3.30 - 52.60)		
Fibrinogen	$457.79 \pm 168.84$	$451.05 \pm 150.33$	254 - 510	
	(203 - 992)	(236 - 987)		
MDC	$5.28 \pm 2.04$	$5.11 \pm 1.87$	4.5 - 11	
WBC	(0.46 - 12.26)	(0.94 - 11.44)		
T 1	$39.66 \pm 12.01$	$40.68 \pm 9.12$	15 45	
Lymphocytes	(17.3 - 92.9)	(21.7 - 69)	15 - 45	
M	$8.34 \pm 2.69$	$9.74 \pm 4.66$		
Monocytes	(1.4 - 18.7)	(4.2 - 33.7)	2 - 10	
NT . 1.1	$48.57 \pm 12.81$	$45.98 \pm 10.82$	40 - 80	
Neutrophil	(2.8 - 78.1)	(11.6 - 67.3)		
PDC	$3.87 \pm 0.54$	$3.85\pm0.52$	4 - 7.2	
RBC	(2.13 - 5.27)	(2.4 - 5.3)		
*** 1.1.	$12.04 \pm 1.66$	11.95 ± 1.55	12 - 16	
Hemoglobin	(6.4 - 16.4)	(7.2 - 16.4)		
DI . I .	$24.84 \pm 9.01$	$24 \pm 8.25$	15 - 45	
Platelet	(7.1 - 55.4)	(3.6 - 46.6)		
FOR	$21.61 \pm 16.14$	$20.84 \pm 16.38$	0 - 20	
ESR	(0 - 62)	(0 - 60)		
TOO G	$2.01 \pm 0.19$	$1.08 \pm 0.63$	27/4	
ECOG	(1 - 3)	(0 - 3)	N/A	

Values in the table are shown as means  $\pm$  SD. The values in the parentheses represent the range of values from lowest to highest observed.

and able to carry out work of a light or sedentary nature (Table 1).

#### 3.2. Comparison of changes in assessment groups

Patients were divided in to three assessment groups depending on their treatment phases, prevention group, KM Tx only group, and combination group, and changes in the NK cell, fibrinogen count, and ECOG performance status values were investigated. The mean ages of the patients for all 3 groups were similar (prevention group:  $48.30 \pm 7.66$  years, KM Tx group:  $49.40 \pm 21.87$  years, combination group:  $49.02 \pm 10.16$  years) as were the mean hospitalization days (prevention group:  $16.40 \pm 3.75$  days, KM Tx group:  $17.60 \pm 9.34$  days, combination group:  $16.74 \pm 5.01$  days). The mean stages of the patients differed by groups, with a higher mean stage being found in the KM Tx only group  $(3.40 \pm 1.34)$  than in either the prevention group  $(2.60 \pm 1.07)$  or the combination group  $(2.23 \pm 1.20)$  (Table 2).

Overall, NK cell count increased in all groups. The highest

mean increase was seen in the KM Tx only group (from 9.82  $\pm$  2.99 to 12.98  $\pm$  5.11, with -3.16  $\pm$  6.84 paired difference). However, the difference was not measurably greater than it was in the other two groups. The NK cell count change in the combination Tx group was statistically significant (P = 0.003). Fibrinogen showed a general tendency to decrease in all groups except for the combination treatment group. The highest mean decrease of fibrinogen was seen in the KM Tx only group (from  $671.40 \pm 218.91$  to  $569.20 \pm$ 194.59, with  $102.20 \pm 117.65$  paired difference). The paired differences for the prevention group and the combination Tx group were  $28.30 \pm 87.24$  and  $-3.92 \pm 113.49$ , respectively. ECOG performance status improved in all groups, with the most improvement being seen in the prevention group (from  $2.00 \pm 0.00$  to  $0.90 \pm 0.57$ , with  $1.10 \pm 0.57$  paired difference). The paired differences for the KM Tx only group and the combination Tx group were  $0.60 \pm 0.89$  and 0.94± 0.56, respectively. The ECOG performance status changes in the prevention group and the combination Tx group were statistically significant (P = 0.000) (Table 3).

Table 2 Comparison of patient characteristics, lab values, and ECOG between assessment groups

		Prevention Group (n = 10)	KM Tx only Group (n = 5)	Combination Group (n = 65)
A ( )		$48.30 \pm 7.66$	49.40 ± 21.87	49.02 ± 10.16
Age (yrs)		(35 - 63)	(27 - 82)	(27 - 76)
O.		$2.60\pm1.07$	$3.40 \pm 1.34$	$2.23 \pm 1.20$
Stage		(1 - 4)	(1 - 4)	(1 - 4)
Hospitalization		16.40 - 3.75	$17.60 \pm 9.34$	$16.74 \pm 5.01$
		(9 - 21)	(11 - 34)	(9 - 31)
NK cell (CD16+CD56)	Before	$9.86 \pm 5.00$	$9.82 \pm 2.99$	$12.51 \pm 9.19$
		(4.4 - 17.8)	(7 - 13.10)	(0.10 - 51.0)
	After	$12.39 \pm 4.33$	$12.98 \pm 5.11$	$15.28 \pm 10.41$
		(7.10 - 19.90)	(5.60 - 18.50)	(3.30 - 52.60)
	Before	$425.90 \pm 116.06$	$671.40 \pm 218.91$	$446.26 \pm 162.35$
		(280 - 646)	(422 - 992)	(203 - 972)
Fibrinogen	After	$397.60 \pm 83.07$	$569.20 \pm 194.59$	$450.18 \pm 151.73$
		(236 - 509)	(350 - 785)	(250 - 987)
W000	Before	$2 \pm 0.00$	$2.20 \pm 0.45$	$2 \pm 0.18$
		(2 - 2)	(2 - 3)	(1 - 3)
ECOG	After	$0.90 \pm 0.57$	$1.60 \pm 1.14$	$1.06 \pm 0.58$
		(0 - 2)	(0 - 3)	(0 - 3)

Values in the table are shown as means  $\pm$  SD. The values in the parentheses represent the range of values from lowest to highest observed.

Table 3 Comparison of NK cell count, fibrinogen, and ECOG changes between groups

Groups		Prevention Group (n = 10)	t	P
Prevention group (n = 10)	NK cell	$-2.53 \pm 5.07$	-1.577	0.149
	Fibrinogen	$28.30 \pm 87.24$	1.026	0.332
	ECOG	$1.10\pm0.57$	6.128	0.000**
KM Tx only group (n = 5)	NK cell	$-3.16 \pm 6.84$	-1.033	0.360
	Fibrinogen	$102.20 \pm 117.65$	1.942	0.124
	ECOG	$0.60 \pm 0.89$	1.500	0.208
Combination Tx group (n = 65)	NK cell	$-2.77 \pm 7.10$	-3.143	0.003*
	Fibrinogen	$-3.92 \pm 113.49$	-0.279	0.781
	ECOG	$0.94 \pm 0.56$	13.619	0.000**

Values in the table are shown as means  $\pm$  SD. The values in the parentheses represent the range of values from lowest to highest observed. \*P < 0.01, \*\*P < 0.001

## 4. Discussion

The relationship between NK cell count and cancer progression is controversial, and the results of research have been conflicting [17-19], even though more research tends to show a positive correlation between low NK cell count and cancer. Plonquet *et al.* found the peripheral blood natural killer cell count to be associated with clinical outcome in patients with aaIPI 2–3 diffuse large B-cell lymphomas [20]. Stewart *et al.* found the CD3-16+56+ cell count to be an independent predictive factor for autologous blood stem cell mobilization (BSCM) in the heterogeneous group of cancer patients [21]. Studies on NK cell activities in mediation of cancer immunity also support the correlation between the NK cell count and cancer progression [22-24].

Recent evidence from several laboratories has linked malignant transformation, tumor angiogenesis, and metastasis to thrombus formation, mediated perhaps by signaling cascades that can be triggered in a clotting-dependent and/or clotting-independent manner [25-27]. The precise role of the cytoplasmic domain of tissue factor (TF), which has been targeted as the likely signaling region of the molecule, remains controversial [28, 29]. TF is a 47-kDa transmembrane protein that initiates the coagulation cascade. Similar functions have been suggested for TF and vascular endothelial growth factor (VEGF). TF and VEGF have been found to be co-localized on malignant cells from human lung cancer and breast cancer samples [30]. The increased expression of TF in tumors results in an angiogenic phenotype by up regulation of the pro-angiogenic factor VEGF and down regulation of the anti-angiogenic factor thrombospondin [31]. An aberrant increased expression of TF is known to occur in a variety of tumor types, including breast cancer, leukemia, glioma, non-small-cell lung cancer, and colon cancer [32-36]. Increased levels of TF have been associated with increased angiogenesis, advances stages of disease, and poorer outcome [36].

In our study, a decrease of ECOG was seen in all groups, implying general improvement of performance status in all the patients. The largest decrease was seen in the prevention group, and the smallest decrease was seen in the KM Tx group. This result was presumably related to the different concentrations of late-stage-cancer patients between the groups. The data from the present study revealed a general decrease in the fibringen count, fluctuating NK cell counts, and enhanced ECOG performance status in all groups. The NK cell count of CD16+56 increased in all assessment groups after the CAM inpatient care, though not by much. The KM Tx only group showed the largest increase, but the differences between groups were not enough to say that the treatment received had had any effect on the overall NK cell count. On a closer look, the NK cell count fluctuated with treatment in individual patients and did not seem to show any consistent correlation with the treatment or the functional improvement; this was presumably due to tumor escape by way of alteration of NK cell function and to the resistance to killing associated with tumor progression and chronic inflammation. The level of the clotting factor fibrinogen decreased most in the KM Tx only group, despite that group's having higher mean cancer stages and worse progression. That increased by a minuscule amount in the combination group, which is thought to have been caused by the multiple mechanisms

of concurrent cytotoxic conventional therapies causing clotting response and of Korean medicine treatment fighting off the coagulation at the same time.

#### 5. Conclusion

Despite the short follow-up time, the results support the positive role of the multi-modality KM-based inpatient care program in complementing the disease treatment course by lowering the cancer coagulation factor fibrinogen and improving the patients functional status. However, its correlation with changes in the NK cell count is not clear. In the future, further studies that have longer treatment durations and use more diverse follow up of the cancer immune, coagulation and inflammation factors as measures to accurately evaluate the efficacy of the treatment are needed.

## **Acknowledgement**

This study was supported by the Korean Institute of Oriental Medicine (grant #20110004). The authors have no conflict of interest to declare.

#### References

- 1. Lin WW, Karin M. A cytokine-mediated link between innate immunity, inflammation, and cancer. J Clin Invest. 2007;117(5):1175-83.
- 2. Reynés G, Vila V, Martín M, Parada A, Fleitas T, Reganon E, *et al.* Circulating markers of angiogenesis, inflammation, and coagulation in patients with glioblastoma. J Neurooncol. 2011;102(1):35-41.
- 3. Lwaleed BA, Cooper AJ, Voegeli D, Getliffe K. Tissue factor: a critical role in inflammation and cancer. Biol Res Nurs. 2007;9(2):97-107.
- Stadnisky MD, Xie X, Coats ER, Bullock TN, Brown MG. Self MHC class I-licensed NK cells enhance adaptive CD8 T-cell viral immunity. Blood. 2011;117(19):5133-41.
- 5. Fernandez PM, Patierno SR, Rickles FR. Tissue factor and fibrin in tumor angiogenesis. Semin Thromb Hemost. 2004;30(1):31-44.
- Buller HR, van Doormaal FF, van Sluis GL, Kamphuisen PW. Cancer and thrombosis: from molecular mechanisms to clinical presentations. J Thromb Haemost. 2007;5:246-54.
- Palumbo JS. Mechanisms linking tumor cell-associated procoagulant function to tumor dissemination. Semin

- Thromb Hemost. 2008;34(2):154-60.
- 8. Rickles FR, Patierno S, Fernandez PM. Tissue factor, thrombin, and cancer. Chest. 2003;124(3 Suppl):58S-68S.
- 9. Khorana AA, Fine RL. Pancreatic cancer and thromboembolic disease. Lancet Oncol. 2004;5(11):655-63.
- 10. Liu L, Duan JA, Tang Y, Guo J, Yang N, Ma H, *et al.* Taoren-Honghua herb pair and its main components promoting blood circulation through influencing on hemorheology, plasma coagulation and platelet aggregation. J Ethnopharmacol. 2012;139(2):381-7.
- 11. Liang JQ, Mi SQ, Wang NS. [Anticoagulative effect and antiplatelet aggregation effect of combination of Hirudo and Tabanus on rat model of blood stasis syndrome]. Zhong Yao Cai. 2009;32(9):1347-50. Chinese.
- 12. Ma H, Yang J, Nan SL, Gong JN, Liu XF. [Effects of Yang-yin Shengjin Decoction on hemorheological parameters and coagulation factors in model rabbits with syndrome of excessive heat consuming body fluid and blood stasis]. Zhong Xi Yi Jie He Xue Bao. 2005;3(1):39-42. Chinese.
- 13. Meng MB, Wen QL, Cui YL, She B, Zhang RM. Meta-analysis: traditional Chinese medicine for improving immune response in patients with unresectable hepatocellular carcinoma after transcatheter arterial chemoembolization. Explore (NY). 2011;7(1):37-43.
- 14. Liu Z, Shen Y, Cui N, Yang J. Clinical observation of immunity for severe acute pancreatitis. Inflammation. 2011;34(5):426-31.
- 15. Zhao CL, Peng LJ, Zhang ZL, Zhang T, Li HM. [Effect of acupuncture on the activity of the peripheral blood T lymphocyte subsets and NK cells in patients with colorectal cancer liver metastasis]. Zhongguo Zhen Jiu. 2010;30(1):10-2. Chinese.
- 16. Oken MM, Creech RH, Tormey DC, Horton J, Davis TE, McFadden ET, *et al.* Toxicity and response criteria of the Eastern Cooperative Oncology Group. Am J Clin Oncol. 1982;5(6):649-55.
- 17. Luo J, Ling Z, Mao W. [Circulating lymphocyte subsets in patients with lung cancer and their prognostic value]. Zhongguo Fei Ai Za Zhi. 2011;14(8):669-73. Chinese
- 18. Soygür T, Bedük Y, Baltaci S, Yaman O, Tokgöz G. The prognostic value of peripheral blood lymphocyte subsets in patients with bladder carcinoma treated using neoadjuvant M-VEC chemotherapy. BJU Int. 1999;84(9):1069-72.
- 19. Hus I, Staroslawska E, Bojarska-Junak A, Dobrzyńska-Rutkowska A, Surdacka A, Wdowiak P, *et al.* CD3+/CD16+CD56+ cell numbers in peripheral blood are correlated with higher tumor burden in patients with diffuse large B-cell lymphoma. Folia Histochem Cyto-

- biol. 2011;49(1):183-7.
- 20. Plonquet A, Haioun C, Jais JP, Debard AL, Salles G, Bene MC, *et al.* Peripheral blood natural killer cell count is associated with clinical outcome in patients with aaIPI 2-3 diffuse large B-cell lymphoma. Ann Oncol. 2007;18(7):1209-15.
- 21. Stewart DA, Guo D, Luider J, Auer I, Klassen J, Morris D, *et al.* The CD3- 16+ 56+ NK cell count independently predicts autologous blood stem cell mobilization. Bone Marrow Transplant. 2001;27(12):1237-43.
- 22. Gillgrass A, Ashkar A. Stimulating natural killer cells to protect against cancer: recent developments. Expert Rev Clin Immunol. 2011;7(3):367-82.
- 23. Shen Y, Lu C, Tian W, Wang L, Cui B, Jiao Y, *et al.* Possible association of decreased NKG2D expression levels and suppression of the activity of natural killer cells in patients with colorectal cancer. Int J Oncol. 2012;40(4):1285-90.
- 24. Konjevic G, Radenkovic S, Srdic T, Jurisic V, Stamatovic Lj, Milovic M. Association of decreased NK cell activity and IFN $\gamma$  expression with pSTAT dysregulation in breast cancer patients. J BUON. 2011;16(2):219-26.
- 25. Boccaccio C, Sabatino G, Medico E, Girolami F, Follenzi A, Reato G, *et al.* The MET oncogene drives a genetic programme linking cancer to haemostasis. Nature. 2005;434(7031):396-400.
- 26. Rong Y, Post DE, Pieper RO, Durden DL, Van Meir EG, Brat DJ. PTEN and hypoxia regulate tissue factor expression and plasma coagulation by glioblastoma. Cancer Res. 2005;65(4):1406-13.
- 27. Yu JL, May L, Lhotak V, Shahrzad S, Shirasawa S, Weitz JI, et al. Oncogenic events regulate tissue factor expression in colorectal cancer cells: implications for tumor progression and angiogenesis. Blood. 2005;105(4):1734-41.
- 28. Abe K, Shoji M, Chen J, Bierhaus A, Danave I, Micko C, *et al.* Regulation of vascular endothelial growth factor production and angiogenesis by the cytoplasmic tail of tissue factor. Proc Natl Acad Sci U S A. 1999;96(15):8663-8.
- 29. Belting M, Dorrell MI, Sandgren S, Aguilar E, Ahamed J, Dorfleutner A, *et al.* Regulation of angiogenesis by tissue factor cytoplasmic domain signaling. Nat Med. 2004;10(5):502-9.
- 30. Shoji M, Hancock WW, Abe K, Micko C, Casper KA, Baine RM, *et al.* Activation of coagulation and angiogenesis in cancer: immunohistochemical localization in situ of clotting proteins and vascular endothelial growth factor in human cancer. Am J Pathol. 1998;152(2):399-411.
- 31. Zhang Y, Deng Y, Luther T, Müller M, Ziegler R, Waldherr R, *et al.* Tissue factor controls the balance of angi-

- ogenic and antiangiogenic properties of tumor cells in mice. J Clin Invest. 1994;94(3):1320-7.
- 32. Contrino J, Hair G, Kreutzer DL, Rickles FR. In situ detection of tissue factor in vascular endothelial cells: correlation with the malignant phenotype of human breast disease. Nat Med. 1996;2(2):209-15.
- 33. Hair GA, Padula S, Zeff R, Schmeizl M, Contrino J, Kreutzer DL, *et al.* Tissue factor expression in human leukemic cells. Leuk Res. 1996;20(1):1-11.
- 34. Guan M, Jin J, Su B, Liu WW, Lu Y. Tissue factor expression and angiogenesis in human glioma. Clin Biochem. 2002;35(4):321-5.
- 35. Sawada M, Miyake S, Ohdama S, Matsubara O, Masuda S, Yakumaru K, *et al.* Expression of tissue factor in non-small-cell lung cancers and its relationship to metastasis. Br J Cancer. 1999;79(3-4):472-7.
- 36. Nakasaki T, Wada H, Shigemori C, Miki C, Gabazza EC, Nobori T, *et al.* Expression of tissue factor and vascular endothelial growth factor is associated with angiogenesis in colorectal cancer. Am J Hematol. 2002;69(4):247-