

Ginger and Its Pungent Constituents Non-Competitively Inhibit Serotonin Currents on Visceral Afferent Neurons

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Nausea and emesis are a major side effect and obstacle for chemotherapy in cancer patients. Employ of antiemetic drugs help to suppress chemotherapy-induced emesis in some patients but not all patients. Ginger, an herbal medicine, has been traditionally used to treat various kinds of diseases including gastrointestinal symptoms. Ginger is effective in alleviating nausea and emesis, particularly, for cytotoxic chemotherapy drug-induced emesis. Ginger-mediated antiemetic effect has been attributed to its pungent constituents-mediated inhibition of serotonin (5-HT) receptor activity but its cellular mechanism of action is still unclear. Emetogenic chemotherapy drugs increase 5-HT concentration and activate visceral vagal afferent nerve activity. Thus, 5-HT mediated vagal afferent activation is essential to provoke emesis during chemotherapy. In this experiment, water extract of ginger and its three major pungent constituent's effect on 5-HT-evoked responses were tested on acutely dispersed visceral afferent neurons with patch-clamp methods. The ginger extract has similar effects to antiemetic drug ondansetron by blocking 5-HT-evoked responses. Pungent constituents of the ginger, [6]-shogaol, [6]-gingerol, and zingerone inhibited 5-HT responses in a dose dependent manner. The order of inhibitory potency for these compounds were [6]-shogaol > [6]-gingerol > zingerone. Unlike well-known competitive 5-HT₃ receptor antagonist ondansetron, all tested ginger constituents acted as non-competitive antagonist. Our results imply that ginger and its pungent constituents exert antiemetic effects by blocking 5-HT-induced emetic signal transmission in vagal afferent neurons.

Key Words: 5-HT receptor, Antiemetic, Chemotherapy, Shogaol, Vagal afferent nerves


INTRODUCTION

Over 70% of cancer patients, those who properly treated with antiemetic drugs, still experience emesis during chemotherapy [1]. Supplementary use of ginger further reduces chemotherapy-induced nausea in such patients [2]. The rhizome of ginger (*Zingiber officinale* Roscoe) has been traditionally used as herbal medicine to treat various kinds of diseases including gastrointestinal symptoms [3]. Ginger is especially effective to alleviate nausea and emesis due to various causes such as motion sickness, hyperemesis gravidarum, and chemotherapy with emetogenic drugs [4-6]. Among them antiemetic efficacy against chemotherapy-induced emesis has attracted much attention [1,2,5]. The cytotoxic chemotherapy drugs increases 5-HT release in the intestine and augments activity of visceral afferent vagus

nerves [7,8]. Abdominal vagotomy or administration of 5-HT type 3 (5-HT₃) receptors antagonists suppress cytotoxic drug-induced vomiting in an animal model [9]. Therefore, 5-HT₃ receptor mediated activation of visceral afferent nerve is integral for induced emesis during chemotherapy. Recently, Abdel-Aziz et al. [10] found that pungent constituents of ginger inhibited 5-HT₃ receptor function in neuroblastoma cells and suggest that the antiemetic efficacy of ginger may contribute to its 5-HT₃ receptor blocking activity. In spite of these advances, ginger and its pungent constituent's effects on visceral afferent neurons have not been tested. In this study patch-clamp methods were used to test the impact of ginger extract and its three major pungent constituents (Fig. 1) on 5-HT-induced responses in acutely dispersed visceral afferent neurons.

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METHODS

Nodose neuron dissociation

Visceral afferent neurons were dissociated from nodose ganglia from two to three week old Sprague-Dawley rats (Orient Bio Inc., Seongnam, Korea) of either gender as pre-

ABBREVIATIONS: 5-HT, serotonin; CPBG, 1-(3-Chlorophenyl) biguanide hydrochloride; Ondansetron, 1,2,3,9-Tetrahydro-9-methyl-3-[(2-methyl-1-imidazol-1-yl)methyl]-4-carbazol-4-onehydrochloride.

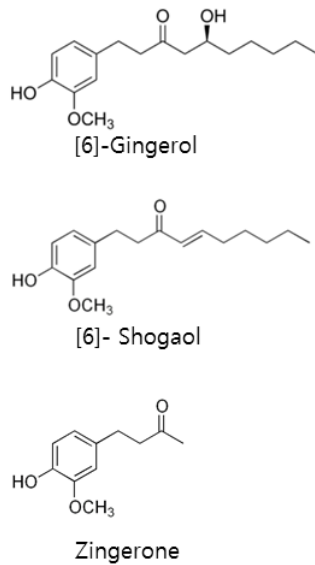


Fig. 1. Molecular structure of [6]-shogaol, [6]-gingerol, and zingerone.

viously described [11]. All animal procedures were conducted with the approval of the institutional Animal Care and Use Committee in Kyung Hee University, Seoul, ROK, study protocol #KHUASP(SE)-13-002 (approval date: April 2, 2013). Briefly, the rats were deeply anesthetized (Zolazepam 0.3 ml/kg, Virbac Laboratories., Carros, France), and the vagus nerve from the jugular foramen was exposed for a length of 0.5 cm caudally. After the vagus nerve was carefully freed from surrounding tissue, the attached ganglion was immediately placed into a Petri dish containing chilled (4°C) artificial cerebrospinal fluid (ACSF) solution, which was composed of the following (in mM): 125 NaCl, 3 KCl, 1.2 KH₂PO₄, 1.2 MgSO₄, 25 NaHCO₃, 10 glucose, and 2 CaCl₂, and bubbled with 95% O₂-5% CO₂. After, gently removing excess connective tissue, the bulb of the nodose ganglion was incubated for 28~40 min at 31°C in ACSF solution containing type II-S trypsin (4~5 mg/ml at 1,310 units/mg; Sigma-Aldrich., St. Louis, MO). Following enzyme treatment, the ganglion was placed in enzyme-free ACSF for 1~3 hours at room temperature. The ganglia were then mechanically dispersed using fire-polished Pasteur pipettes in a glass bottom perfusion chamber filled with standard external recording solution containing (in mM): 150 NaCl, 5 KCl, 1 MgCl₂, 2 CaCl₂, 10 HEPES, and 10 glucose (pH was adjusted to 7.4 with Tris-base).

Electrical measurements

For voltage-clamp recording, isolated neurons were visualized using a phase contrast microscope (IX70; Olympus, Tokyo, Japan) with a 40× objective and 10× ocular lens. Recordings utilized an EPC 9 (HEKA, Lambrecht, Germany) and pClamp 9 software (Molecular Devices, Sunnyvale, California). Electrical recordings used a modified perforated patch method using a nystatin concentration of 450 μg/ml [12]. The ionic composition of the internal patch-pipette solution for nystatin perforated patch recordings was (mM): 50 KCl, 100 K gluconate, and 10 HEPES. The pH

of this solution was adjusted to 7.2 with Tris-OH. The nystatin-internal solution filled patch-pipettes has 5~7 MΩ resistance. Junction potential for these internal and standard external solutions was 11.9 mV and compensated throughout experiments. The access resistance, measured at approximately 15 min after formation of a gigaohm seal, was 8.5±1.2 MΩ. Neurons were voltage clamped to -50 mV and currents were sampled at 10 KHz with 3 KHz filtering and saved to computer. All experiments were conducted at room temperature (21~22°C).

Drugs & ginger extraction

Serotonin, CPBG (1-(3-Chlorophenyl)biguanide hydrochloride), zingerone, [6]-gingerol, and [6]-shogaol were purchased from Sigma-Aldrich (St. Louis, MO). All three pungent constituents of ginger were dissolved in DMSO and then diluted to the standard external recording solution. The final concentration of DMSO was less than 0.3%, a concentration which had no effect on the cells. 5-HT₃ receptor selective antagonist ondansetron (1,2,3,9-Tetrahydro-9-methyl-3-[(2-methyl-1*H*-imidazol-1-yl)methyl]-4*H*-carbazol-4-one hydrochloride) was purchased from Tocris Cookson (Ballwin, MO). Ginger extract were prepared from freeze-dried *Zingiberis officinalis* roots. Finely grounded 10 g ginger was extracted with 400 mL water. The extract was filtered, freeze-dried and kept -20°C until use. Each 10 g of dried ginger yielded 1.61 g dried power. All of the drugs were applied via a rapid application Y-tube microperfusion system that provided complete solution changes surrounding the recorded neurons within 0.1 seconds [13].

Data analysis

Data were analyzed off-line using Clampfit 9 software. All data are represented as mean±standard error of the mean (SEM). For statistical comparisons, one-way ANOVA with post-hoc multiple comparisons (Bonferroni/Dunn's correction, StatView, SAS Institute Inc., Cary, NC) was used. Differences were considered statistically significant for *p* values < 0.05. The dose response curves for concentration-inhibition relationships were constructed according to the following equation (1) using a least-square fitting routine:

$$I=1 - [C^{nH}/(C^{nH} + IC_{50}^{nH})] \quad (1)$$

Where *I* is the current amplitude normalized to the control response, *C* is the corresponding ginger constituents or ondansetron concentrations. IC₅₀ and *nH* denote the half-maximum inhibition concentration and the Hill coefficient, respectively. The continuous theoretical curves for the concentration response relationships were constructed according to a Hill equation (2) using a least-square fitting routine as described elsewhere [14] :

$$I=I_{max} [C^{nH}/(C^{nH} + EC_{50}^{nH})] \quad (2)$$

Where *I* is the current amplitude normalized to the control response and *C* is the corresponding 5-HT concentration. EC₅₀ and *nH* denote the half-maximum concentration and Hill slope, respectively.

RESULTS

Inhibition of 5-HT-induced current by ginger extract

To characterize the base serotonin (5-HT) responses, whole-cell current recordings were made at a holding potential (V_H) of -50 mV. Serotonin evoked inward currents in 29% ($n=9/31$) of assayed nodose neurons in a concentration dependent manner (0.001 to 1 mM). Three micromole 5-HT was used for subsequent control responses, at which approximately 25% of the maximal current amplitude was obtained. As shown in Fig. 2A, 4 min repeated application of the $3 \mu\text{M}$ 5-HT produced constant 5-HT responses. Co-application of 5-HT₃ receptor selective antiemetic drug ondansetron (10 nM) reversibly blocked the 5-HT-evoked currents by an average of $99.5 \pm 0.4\%$ ($p=0.0006$, $n=7$). Serotonin type 3 receptor selective agonist, CPBG (1 μM), mimicked the 5-HT effect by inducing inward current responses and ondansetron effectively inhibited CPBG responses ($n=5$, Fig. 2B). These results demonstrate that 5-HT-evoked current responses by activating 5-HT₃ receptors. In the continuous experiments, inhibitory effects of the ginger extract

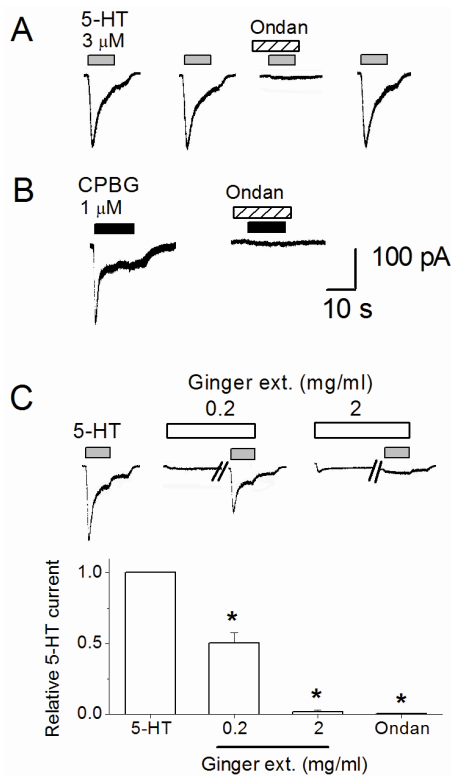


Fig. 2. Inhibition of 5-HT-evoked current by ginger dry extract (Ginger ext.) and ondansetron (Ondan). (A) Serotonin ($3 \mu\text{M}$) was repeatedly applied with 4 min intervals with or without 5-HT₃ receptor antagonist ondansetron (10 nM). (B) CPBG-evoked current was blocked by 10 nM ondansetron. (C) The traces show that 5-HT-evoked currents were suppressed by 0.2 and 2 mg/ml ginger dry extract. Ginger extract was pre-applied 2 min before co-application with $3 \mu\text{M}$ 5-HT. The diagram shows summary of average 5-HT current change by 0.2 and 2 mg/ml ginger extract and 10 nM ondansetron treatments. *Significant difference from control ($p \leq 0.01$). Each column and vertical lines represent the mean \pm SEM.

on the $3 \mu\text{M}$ 5-HT-induced control responses were tested. Before the simultaneous application with 5-HT, an external solution containing 0.2 or 2 mg/ml ginger extract was pre-treated for 2 min (Fig. 2C). The 5-HT current was suppressed by $49.5 \pm 7.2\%$ and $98.2 \pm 1.2\%$ by 0.2 and 2 mg/ml ginger extract, respectively. As shown in Fig. 2C traces, ginger extract alone evoked 37.5 ± 17.1 pA ($n=8$) and 38.8 ± 8.9 pA ($n=6$) currents at 0.2 mg/ml and 2 mg/ml, respectively. Those responses are not inhibited by ondansetron ($p > 0.05$, $n=6$).

In summary, the above result shows that ginger extract and ondansetron have similar modes of action to inhibit 5-HT-evoked response in nodose ganglia neurons.

Inhibition of 5-HT current by ginger constituents

To determine which of the pungent ginger constituents, [6]-gingerol, [6]-shogaol and zingerone, block 5-HT-evoked responses in nodose neurons we tested them independently. The neurons were perfused with an external solution containing each compound from 30 seconds prior until simultaneous application with 5-HT. All constituents concentra-

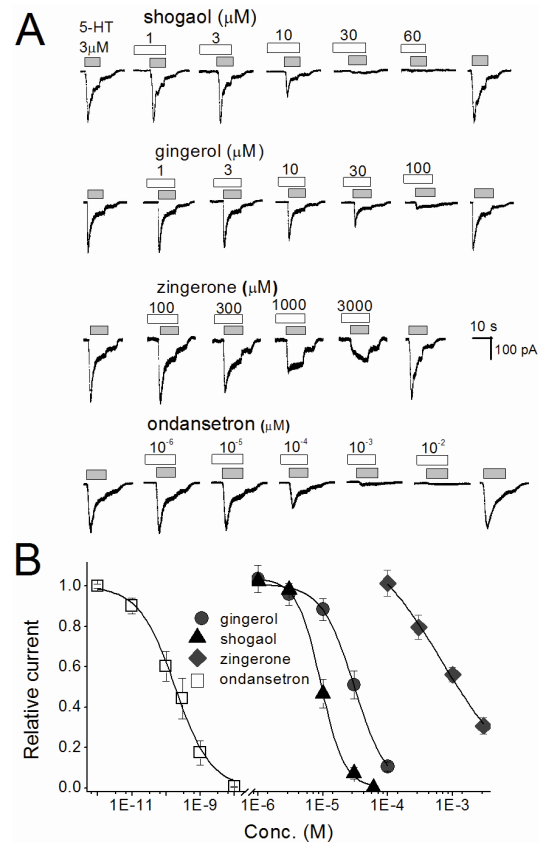


Fig. 3. Concentration-inhibition relationships for single compounds of ginger and ondansetron in nodose ganglion neurons. (A) The traces shown are inhibition of $3 \mu\text{M}$ 5-HT-evoked current by [6]-shogaol, [6]-gingerol, zingerone and ondansetron. The traces were taken from different set of neurons. Various concentrations of reagents were pretreated 30 sec, and then co-application with 5-HT. (B) Represents the concentration-inhibition relationship of [6]-shogaol, [6]-gingerol, zingerone and ondansetron. Each point is the average of 4~7 neurons.

Table 1. Half-maximum inhibition concentration (IC_{50}) for zingerone, [6]-shogaol, [6]-gingerol, and ondansetron on 3 μ M 5-HT evoked currents in nodose neurons

Compound	Molecular weight	IC_{50} (M)
Zingerone	194.22	1.19×10^{-3}
[6]-Shogaol	276.37	9.27×10^{-6}
[6]-Gingerol	294.38	3.03×10^{-5}
Ondansetron	293.4	1.84×10^{-10}

The IC_{50} was determined by fitting the data obtained from 4~7 cells to concentration-inhibition equation in the method using a least square fitting routine.

tion-dependently inhibited 5-HT-evoked currents (Fig. 3). Where control 5-HT responses were significantly ($p < 0.05$) attenuated by $\geq 10 \mu$ M gingerol, $\geq 10 \mu$ M shogaol, and $\geq 300 \mu$ M zingerone. For comparison, ondansetron was tested in same conditions and the 5-HT responses were significantly ($p < 0.0001$) inhibited by $\geq 10^{-4} \mu$ M concentrations. All ginger constituents reversibly inhibited the control response in a concentration-dependent manner. The control responses were recovered by 4 to 5 min washing with ACSF. The order of inhibitory potency for these compounds were [6]-shogaol > [6]-gingerol > zingerone. The IC_{50} of [6]-shogaol and [6]-gingerol were 128 and 39 times lower than zingerone, respectively (Table 1). Thus, [6]-shogaol and [6]-gingerol are more effective than zingerone.

Pungent ginger constituents act as non-competitive antagonists

The effects of three ginger constituents were examined at different 5-HT concentrations. Fig. 4 shows the concentration-response curves of 5-HT in the absence and presence of [6]-shogaol (10 μ M), [6]-gingerol (30 μ M), or zingerone (1 mM). The EC_{50} of the 5-HT was $7.81 \pm 0.63 \mu$ M. In the presences of [6]-shogaol, [6]-gingerol, or zingerone, respective values for EC_{50} were 7.2 ± 1.32 , 7.9 ± 1.7 and $6.59 \pm 2.24 \mu$ M. The EC_{50} value for 5-HT was not changed in the presence of three tested constituents ($p > 0.05$, $n = 4$). In contrast that the maximal response induced by 1 mM 5-HT was significantly decreased ($p < 0.026$, $n = 4$) to 89 ± 3.4 , 85 ± 3.6 , and $79 \pm 2.7\%$ of control values in the presence of [6]-shogaol (10 μ M), [6]-gingerol (30 μ M), and zingerone (1 mM), respectively. These results support a non-competitive interaction by ginger constituents at the 5-HT site.

DISCUSSION

Nausea and emesis are inherent defense mechanisms for avoid ingestion or expel toxic food, however, these symptoms are also a major side effect and obstacle for chemotherapy. Combined use of 5-HT₃ antagonists and dexamethasone help to suppress chemotherapy-induced moderate to severe emesis in some ($\leq 23.3\%$) but not all patients [1]. Ginger was found to be effective in animal experiments by suppressing cytotoxic chemotherapy drug-induced emesis [5,15]. In clinical trials, for the ethical reason, antiemetic efficacy of the ginger was only tested in the presence of antiemetic drugs. Ginger supplementation effectively reduced severity of chemotherapy-induced nausea in cancer

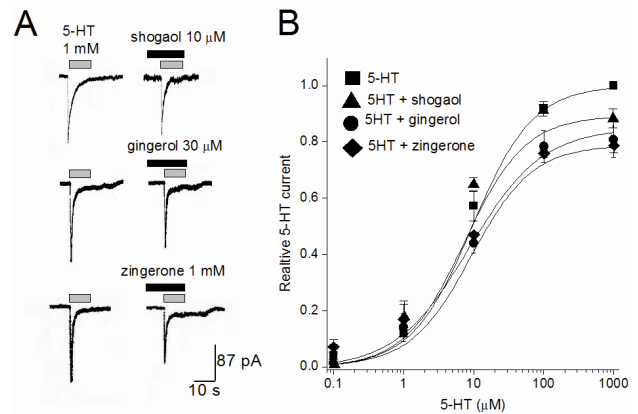


Fig. 4. The effect of [6]-shogaol, [6]-gingerol, and zingerone on 5-HT concentration-response curves. (A) Representative currents traces activated by 1 mM 5-HT in the presence or absence of [6]-shogaol (10 μ M), [6]-gingerol (30 μ M), and zingerone (1 mM). Various concentrations of reagents were pretreated 30 sec, and then co-application with 5-HT. (B) Concentration-response relationship for 5-HT in the absence and presence of 10 μ M [6]-shogaol, 30 μ M [6]-gingerol, and 1 mM zingerone. The current amplitude is normalized to the current activated by 1 mM 5-HT. Data points are the mean \pm S.E.M. ($n = 4 \sim 6$). The curve is the best fit of the data to the Hill equation described in methods.

patients [1,2]. Several studies have been conducted to determine the mechanism of gingers' antiemetic action. Based on anti-serotonergic effects of ginger extract on isolated guinea-pig ileum, Yamahara et al. [15] suggested that the antiemetic actions of ginger may be attributed to 5-HT₃ receptor antagonism. In recent studies, Abdel-Aziz et al. [10] reported that ginger extract and its constituents inhibit 5-HT₃ receptor function in neuroblastoma cells and rat ileum. Emetogenic chemotherapy drugs increase 5-HT concentration in the plasma and intestine [8,16] and activate vagal afferent nerves via 5-HT₃ receptors [7]. Resection of afferent vagus nerve or administration of 5-HT₃ receptors antagonists inhibits cisplatin-induced emesis [9]. Thus, 5-HT₃ mediated activation of visceral afferent nerves underlies the mechanism for induction of emesis during chemotherapy. For the first time, we tested ginger extract and its major pungent ingredients on 5-HT-evoked currents in acutely isolated visceral afferent nodose neurons and found that all tested substances inhibited the 5-HT response in a concentration-dependent manner. In spite of the results of current experiments, the following question remains. If active ginger constituents block 5-HT₃ receptors at much higher IC_{50} concentration than antiemetic drug ondansetron, how ginger extract exert additive antiemetic efficacy in cancer patients, who treated with antiemetic drugs? Interestingly, previous reports suggest that ginger constituents have distinct binding sites from that of 5-HT₃ receptor antagonists. Unlike, 5-HT₃ receptor selective antiemetic drugs which bind at the 5-HT binding site [17], the major active constituents of ginger inhibit the 5-HT₃ receptor ion channel by binding to a modulatory binding site distinct from the serotonin recognition site [18]. In addition that Walstab et al. [19] showed that ginger extract and [6]-gingerol non-competitively inhibited 5-HT induced Ca^{2+} influx through human recombinant 5-HT₃ receptors. In this experiment, we showed that [6]-shogaol, [6]-gingerol, and zingerone act as non-competitive antagonists for

5-HT₃ receptors in visceral afferent neurons. Thus, overlapping inhibition to 5-HT₃ receptors via combination therapy of pharmaceutical 5-HT₃ antagonists and ginger may increase antiemetic efficacy for more patients. In conclusion, our results suggest that ginger and its pungent constituents may suppress chemotherapy-induced emesis by blocking 5-HT-induced emetic signal transmission in vagal afferent nerves.

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