Original Research Article

# Evaluation of Chlorophyll Content, Antioxidant Activity and Phenolic Compounds in the Seedlings of Rice-type Tartary Buckwheat

### Pankaja Sharma, Kooyeon Lee and Cheol Ho Park\*

Department of Bio-Health Technology, Kangwon National University, Chuncheon 200-701, Korea

**Abstract** - Rice type tartary buckwheat is used as a substitute for rice in many Asian countries due to its easy dehulling character. The objective of the present study was to determine the chlorophyll, total polyphenol (TP), total flavonoid (TF), antioxidant activity and to quantify the bioactive compounds rutin, quercetin and chlorogenic acid in the seedlings of rice-type tartary buckwheat (RTTB). Young seedlings exhibited higher antioxidant (DPPH radical inhibition) activity in dose dependent manner. TP and TF content were highest (3017.46  $\pm$  201.84  $\mu$ g TAE /100 mg dw and 1916.0  $\pm$  102.95  $\mu$ g QE / 100 mg dw respectively) in 3 days after germination (DAG) seedlings compare to 6 and 9 DAG. The contents of rutin and quercetin increased with growing stage of seedlings. However, the chlorogenic acid decreased with increasing growth. Overall, RTTB seedlings can be regarded as a strong source of phenolics and have high possibility for food and nutraceutical application due to their efficient antioxidant properties, higher chlorophyll and phytochemical content.

Key words - Antioxidant, Chlorophyll, Seedlings, Rice-type tartary buckwheat

### Introduction

Buckwheat is an annual plant belonging to the family Polygonaceae and is cosmopolitan in nature. This plant is considered as a functional food as it is rich in phenolic compounds including rutin, quercetin, orientin, vitexin, isovitexein and isoorientin (Hagels et al., 1995, Li and Zhang, 2001). Among these compounds, rutin has been recognized as a major antioxidant component that accounts for about 85-90% of the total antioxidant activity (Morishita et al., 2007). Rutin is also known to have anti-inflammatory, anticarcinogenic effects (Liu et al., 2008) and is effective for preventing hemorrhagic disease and arteriosclerosis (Fabjan et al., 2003). Other bioactive compounds like quercetin and chlorogenic acid present in buckwheat have been identified as strong antioxidants, antimicrobial, antifungal (Bowels and Miller, 1994), anti-angiogenesis and anticancer (Jackson and Venema, 2006) agent.

Apart from phenolic compounds, chlorophylls are also a major component in green plants. Commonly two types of chlorophyll, a and b, are found in terrestrial plant. These chlorophylls absorb the light in the process of photosynthesis that converts light energy to chemical energy. These days, chlorophylls are used for the supplementary food because of its important health benefit role to human body. Researches have shown that the chlorophylls can be used for the treatment of different acute and chronic suppurative diseases (Benjamin, 1940). Chlorophyll derivatives have been considered as an antiproliferative effect (Chiu et al., 2003; Gomaa et al., 2012). Therefore, in this study it is worthy to analyze the chlorophyll content in the seedlings of the rice type tartary buckwheat.

Nowadays, buckwheat seedlings and sprout have gained popularity due to the functional compounds present in them and are considered as a new vegetable (Kim et al., 2001). So far, several researches have been reported regarding the phenolic content, antioxidant activities, and biological effects of the buckwheat sprouts (Kim et al., 2008, 2006; Liu et al., 2008; Chang et al., 2010). Compared to common buckwheat, tartary buckwheat is known to have higher flavonoid content, mainly rutin. Recently, another type of tartary buckwheat known as rice-type tartary buckwheat (RTTB) has gained much attention due to its unique character of being readily dehulled. This RTTB is used as a substitute for rice in some parts of Nepal, Bhutan, Inner Mongolia and southwest China

<sup>\*</sup>Corresponding author. E-mail: chpark@kangwon.ac.kr



Fig. 1. Seedlings of rice-type tartary buckwheat cultivated in greenhouse (A). B, C and D represent seedlings harvested in 3, 6 and 9 days after germination, respectively.

(Wang et al., 2007). In previous study, Li et al. (2012) quantified rutin and quercetin in mature flower, seed, leaf and stem of RTTB. However, to the best of our knowledge, there is no literature regarding the phenolic compounds (rutin, quercetin and chlorogenic acid), chlorophyll content and the antioxidant activity of seedlings of RTTB. Therefore, in this research, we made an effort to assess the chlorophyll content, antioxidant activity, as well as the content of phytochemicals, particularly rutin, quercetin and chlorogenic acid in the different growth stages of seedlings of RTTB.

### Materials and Methods

#### Chemicals

Analytical grade organic solvent (methanol and acetonitrile) used for the extraction of rice-type tartary buckwheat and detection in HPLC were purchased from Merck KGaA Darmstadt, Germany. Tannic acid, Quercitin and 1, 1-diphenyl-2-picrylhydrazyl (DPPH) purchased from Sigma Chemical Co. (St. Louis, Mo). Folin-Ciocalteu's reagent was purchased from Wako Pure Chemicals, Japan. The pure compounds rutin, quercetin and chlorogenic acid were purchased from Sigma Chemical Co. (St. Louis, MO). All other chemicals and reagents used were of analytical grade.

### Cultivation and sampling of plant materials

The seeds of RTTB were sown in a greenhouse at  $24 \pm 3$  °C and watered one time in a day. The young seedlings (Fig. 1) were harvested in 3, 6, and 9 days after germination (DAG). The harvested samples were washed and dried in an oven at a temperature of  $45 \pm 1$  °C. To estimate the fresh weight and

moisture content of seedlings another batch of 10 seedlings each in 3 replications were also harvested and dried in the oven.

### Preparation of plant extracts

The dried powdered samples  $(2\ g)$  of seedlings were taken and  $100\ ml$  of 100% ethanol was added to each and incubated overnight in a shaker followed by filtration using Advantech 5B filter paper (Tokyo Roshi Kaisha, Japan). The extract was dried using a rotatory evaporator (Eyela digital water bath SB-1000, Tokyo, Rikakikai Co., Ltd. Japan) at a temperature of  $40\ C$ . The extracts were vacuum freeze dried and the yield was measured and stored in the refrigerator for further experiment.

### **Extraction of chlorophylls**

The freshly harvested and pre-weighed seedlings were homogenized with 20 ml of 100% ethanol in an extraction tube for 10 min. The extract obtained was centrifuged at 5000 rpm for about 10 min. The supernatant was separated and the maximum absorbance of chlorophyll a and b were determined spectrophotometrically at 666 and 654 nm respectively by means of equations proposed by Lichtenthaler and Wellburn (1983):

Chlorophyll a =  $15.65 A_{666} - 7.340 A_{654}$ 

Chlorophyll b =  $27.05 A_{654} - 11.21 A_{666}$ 

### Estimation of total polyphenol and total flavonoid content

Total polyphenol (TP) content of samples was estimated using the Folin-Ciocalteu colorimetric method (Ghimeray et

al., 2009). The appropriate dilutions of the extracts were oxidized with 0.2N Folin-Ciocalteu's reagent and then the reaction was neutralized with 10% sodium carbonate. The absorbance of the resulting blue color was measured at 725 nm using spectrophotometer after incubation for 1 hr at room temperature. Quantification was done on the basis of the standard curve of Tannic acid. Results were expressed as  $\mu g$  of Tannic acid equivalent (TAE) per 100 mg of dry weight (dw).

Total flavonoid (TF) content was determined using the protocol of Eom *et al.* (2008). Briefly, an aliquot of 1 ml of the sample (1 mg/ml) was mixed with 0.1 ml of aluminum nitrate (10%) and 0.1 ml of potassium acetate (1 M). To the mixture, 3.8 ml of ethanol was added to make the total volume 5 ml. The mixture was vortexed and the absorbance was measured after 40 min at 415 nm using a spectrophotometer (UV, 1800 Shimadzu, Japan). The TF was calculated from a calibration curve ( $R^2 = 0.999$ ) using quercetin equivalents (QE) in  $\mu g$  per 100 mg dry weight.

### Measurement of antioxidant activity by DPPH free radical scavenging assay

DPPH radical scavenging activity of rice-type tartary buckwheat extracts were determined by using the method of Sharma *et al.* (2012). Briefly, 1 ml of each of extract at different concentration (500, 250, 125 and 62.5 ppm) was added to 3 ml of DPPH (0.15 mM) solution. The mixtures were shaken vigorously and left to stand at room temperature in the dark for 30 min. The absorbance was measured at 517 nm in a spectrophotometer and the percent inhibition activities of the extracts were calculated against a blank using the following expression: Inhibition (%) = (1-B/A) × 100, where, A is the absorbance of the mixture without extract and B is the absorbance with extracts.

### Quantitation of Rutin, Quercetin and Chlorogenic acid by HPLC

The quantitative estimation of different compounds (rutin, quercetin, and chlorogenic acid) was performed by HPLC. The HPLC system (CBM-20A, Shimadzu Co, Ltd., Japan) with two gradient pumps (LC-20AT, Shimadzu), an auto sample injector (SIL-20A, Shimadzu), a UV-detector (SPD-

10A, Shimadzu) and a column oven (35°C CTO-20A, Shimadzu) were used for analysis. The separation was performed on a C18 column (Synergi 4  $\mu$  MAX-RY, 150 × 4.6 mm, 4 micron Phenomenex). Flow rate of mobile phage solution was 1.0 ml/min, and detection was at 355 nm. 10  $\mu$ l of each sample was injected in to the HPLC machine. HPLC conditions were as follows: Solvent A (water in 0.1% Trifloroacetic acid) and solvent B (acetonitrile). Gradient elution used was 0-10 min, 5-6% B; 10-15 min, 6-10% B; 15-45 min, 10-19% B; 45-65 min, 19-20% B. The compounds were identified in solvents by matching their retention times and spectra with that of the standards and the data were calculated on the basis of the peak area obtained.

#### Data analysis

All data were expressed as the mean value  $\pm$  standard deviation (SD) of each experimental group (n = 3). The results were processed using Excel 2010 (Microsoft, Redmond, WA, USA).

### Results and Discussion

## Estimation of fresh weight (FW), moisture and chlorophyll content in seedlings

Table 1 lists the FW, moisture and chlorophyll contents of seedlings harvested in different days after germination. The FW of seedlings increased with the growth of the seedlings which measured  $760.6 \pm 18.2$ ,  $1951.7 \pm 29.6$  and  $4426.4 \pm 17.1$  mg at 3, 6 and 9 DAG, respectively. The data of moisture content remain almost consistent (ranging from 91.83 - 92.58%) in all the seedlings harvested in different days. This moisture content of seedlings resembles with the previous report of common and tartary buckwheat seedlings shown by Kim *et al.* (2008).

The chlorophyll pigments are a major source applied in food and neutraceutical products (Mortensen, 2006). According to other findings the ethanol molecules penetrate lipid-protein bilayers more efficiently than methanol due to higher hydrophobicity (Patra *et al.*, 2006). Therefore, we also used ethanol to extract the chlorophyll from the seedlings. According to our data, the 3, 6 and 9 DAG seedlings showed higher chlb content ranging from 46.20-47.82 µg/ml compared to chl-a,

Table 1. Fresh weight (FW), moisture content (%) and chlorophyll content ( $\mu$ g/ml) in rice-type tartary buckwheat seedlings at different stages of germination<sup>z</sup>

Days after germination	FW (mg)	Moisture (%)	Chl-a (μg/ml) <sup>y</sup>	Chl-b (µgml) <sup>y</sup>
3	760.6 ±	91.83 ±	23.93 ±	47.82 ±
	18.2	0.53	2.61	2.71
6	$1951.7~\pm$	$92.58 \pm$	$24.34~\pm$	$47.75~\pm$
	29.6	0.46	1.96	2.54
9	$4426.4~\pm$	$92.39 \pm$	$23.76~\pm$	$46.20~\pm$
	17.1	0.75	2.14	3.04

<sup>&</sup>lt;sup>z</sup>Values are means ± standard error of 3 replications containing 10 seedlings in each replica.

which ranged from 23.76-24.34  $\mu$ g/ml. However, not much difference in chlorophyll content was observed in the different growing stages (3, 6 and 9 DAG) of seedlings.

## Total Polyphenol (TP) and Total Flavonoid (TF) content in seedlings

The TP contents in the extracts of RTTB seedlings were determined from regression equation of calibration curve and expressed in Tannic acid equivalent (µg TAE /100 mg) of dry weight (dw) of plant material (Fig. 2). The results revealed that the TP content was highest (3017.46  $\pm$  201.84  $\mu$ g TAE / 100 mg dw) in 3DAG seedlings. However, 6 and 9 DAG seedlings showed almost similar TP content which was found to be 2257.94  $\pm$  180.43 and 2163.72  $\pm$  154.02  $\mu g$  TAE /100 mgdw respectively. Similarly, the TF contents of the seedlings at different stages of growth of RTTB were expressed in quercetin equivalent ( $\mu g$  QE /100 mg) of dry weight (dw) of plant material (Fig. 2). The TF content followed the similar trend as the TP content. The content was found to be slightly higher (1916.0  $\pm$  102.95  $\mu$ g QE/100 mg dw) in 3DAG seedlings compared to the 6 and 9DAG seedlings (1762.41  $\pm$  179.16 and  $1621.50 \pm 107.68 \,\mu g$  QE /100 mg dw respectively). This variation in the TP and TF content among the growing stages of the seedlings could be due to the fact that exposure to natural light and the age of the buckwheat seedlings affects the phenolic and flavonoid composition of buckwheat (Kim et al., 2008).

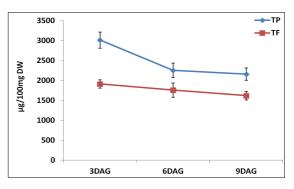


Fig. 2. Total polyphenol (TP) and total flavonoid (TF) content in rice-type tartary buckwheat seedlings at 3, 6 and 9 days after germination (DAG). TP content is expressed in  $\mu g$  TAE/100 mgdw and TF content is expressed in  $\mu g$  QE/100 mg dw. Bar means standard deviation.

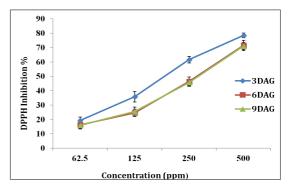


Fig. 3. DPPH free radical scavenging activity (%) of rice-type tartary buckwheat seedlings at 3, 6 and 9 days after germination (DAG).

#### DPPH free radical scavenging activity

DPPH is a stable radical with a deep purple color whose reaction with other radicals or compounds leads to loss of color at  $517 \, \text{nm}$ . The results are presented in percent inhibition in dose dependent manner (Fig. 3). At the concentration of  $500 \, \text{ppm}$ , the 3DAG seedling of RTTB showed a higher inhibition of  $78.49 \pm 1.64\%$ . However, 6 and 9DAG seedling showed  $71.55 \pm 3.58$  and  $71.09 \pm 2.51\%$  inhibitions respectively. With similar trend, the 3DAG seedlings showed higher free radical scavenging activity compared to 6 and 9DAG at all the concentrations used in the experiment. This increased antioxidant activity of 3DAG seedlings compared to others could be due to the higher polyphenol and flavonoid content or higher amount of chlorogenic acid or synergistic effect of some other compounds (including anthocyanins) present in the early stage of growth.

<sup>&</sup>lt;sup>y</sup>Chl-a and Chl-b represent chlorophyll a and chlorophyll b, respectively.

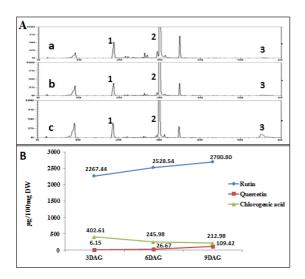


Fig. 4. HPLC chromatogram (A) and quantitation of rutin, quercetin and chlorogenic acid (B) of rice type tartary buckwheat seedlings harvested in different days. Chromatogram a, b and c denotes seedlings of rice type tartary buckwheat at 3, 6 and 9 days respectively. The compounds were expressed in  $\mu g/100 \, \mathrm{mg}$  dw.

### Quantitation of Rutin, Quercetin and Chlorogenic acid

The quantitative estimation of phenolic compounds (rutin, quercetin and chlorogenic acid) in RTTB seedlings are shown in Fig. 4 (A and B). The data revealed that rutin was present as a major compound in RTTB seedlings. The average content of rutin in the growing stages of seedlings of 3, 6 and 9DAG were 2267.44, 2528.54 and 2700.80  $\mu$ g/100 mg dw respectively. The results showed that the rutin content increased with the growth of the seedlings. This increase in the level of rutin content could be due to longer growth in the presence of sun light which increases the rutin contents in buckwheat (Yao et al., 2004). Likewise, the quercetin content also increased with the increase in growth. At 3DAG, the quercetin content was  $6.15 \,\mu\text{g}/100 \,\text{mg}$  dw, which further increased to 26.67 and  $109.4 \,\mu\text{g}/100 \,\text{mg}$  dw at 6 and 9 DAG respectively. Beside the above compounds, chlorogenic acid was also present in higher amount. But the content was decreased with the growing stages of seedlings. In 3DAG seedlings the chlorogenic acid content was 402.6  $\mu$ g/100 mg dw, however, the content decreased to 245.98 and 213.98  $\mu$ g/100 mg dw in 6 and 9 DAG seedlings respectively. Our findings are in agreement with the previous findings by Sharma et al. (2012), showing similar trends in which the quercetin content increased with the growth of the seedlings and the chlorogenic acid content decreased with the maturation of the seedlings of common and tartary buckwheat. However, comparing the RTTB seedlings with 7 days old common and tartary seedlings used in the previous studies (Sharma *et al.*, 2012), the contents of chlorogenic acid in 3DAG RTTB seedlings is 2.58 and 1.37 fold higher respectively and the quercetin content is similar to tartary buckwheat but 1.30 fold higher than common buckwheat. In 9DAG RTTB seedlings, the quercetin content is 15.28 and 22.98 fold higher than common and tartary buckwheat (7 days old) seedlings respectively.

Phenolic or flavonoid composition in buckwheat depends upon cultivar, location, growing season, soil type, harvesting times and other environmental conditions (Oomah and Mazza, 1996; Kitabayashi *et al.*, 1995; Hagel *et al.*, 1995). In our study, we analyzed seedlings of RTTB in different growing stages and observed higher value of chlorophyll (a and b), TP and TF contents. RTTB seedlings also showed a higher antioxidant activity in the DPPH free radical scavenging assay in a dose dependent manner. Likewise, RTTB seedlings contain considerable amount of rutin, chlorogenic acid and quercetin. Therefore, RTTB seedlings which are more potent than common and tartary buckwheat seedlings can be regarded as a strong source of phenolics and has high possibility for food and nutraceutical application due to their efficient antioxidant properties, higher chlorophyll and phytochemical content.

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### References

Bowels, B.L. and A.J. Miller. 1994. Caffeic acid activity against *Clostridium botulinum* spores. J. Food Sci. 59:905-908.

Benjamin, G. 1940. Chlorophyll – Its therapeutic place in acute and suppurative disease. American J. Surgery. New Series Vol. XLIX, No. 1. 49-55.

Chang, K.J., G.S. Seo, Y.S. Kim, D.S. Huang, J.I. Park, J.J. Park, Y.S. Lim, B.J. Park, C.H. Park and M.H. Lee. 2010. Components and biological effects of fermented extract from tartary buckwheat sprouts. Kor. J. Plant Res. 23(2):

- 131-137.
- Chiu, L.C., C.K. Kong and V.E. Ooi. 2003. Antiproliferative effect of chlorophyllin derived from a traditional Chinese medicine *Bombyx mori* excreta on human breast cancer MCF-7 cells. International J. Oncology 23:729-735.
- Eom, S.H., H.J. Park, C.W. Jin, D.O. Kim, D.W. Seo, Y.H. Jeong and D.H. Cho. 2008. Changes in antioxidant activity with temperature and time in *Chrysanthemum indicum* L (Gamguk) teas during elution processes in hot water. Food Sci. Biotech. 17:408-412.
- Fabjan, N., J. Rode, I.J. Kosir, Z. Zang and I. Kreft. 2003. Tartary buckwheat (*Fagopyrum tartaricum* Gaertn.) as a source of dietary rutin and quercetrin. J. Agric. Food Chem. 51:6452-6455.
- Gomaa, I., S.E. Ali, T.A. El-Tayeb and M.H. Abdel-kader. 2012. Chlorophyll derivative mediated PDT *versus* methotrexate: An *in vitro* study using MCF-7 cells. Photodiagnosis and Photodynamic Therapy 9:362-368.
- Ghimeray, A.K., C.W. Jin, B.K. Ghimire and D.H. Cho. 2009. Antioxidant activity and quantitative estimation of azadirachtin and nimbin in *Azadirachta indica* A. Juss grown in foothills of Nepal. African J. Biotechnol. 8:3084-3091.
- Hagels, H.D., Wagenbreth and H. Schilcher. 1995. Phenolic compounds of buckwheat herb and influence of plant and agricultural factors (*Fagopyrum esculentum* Moench and *Fagopyrum tataricum* Gartner): *In* Matano, T. and A. Ujihara (eds.), Current Advances in Buckwheat Research, Vol. I, Shinshu University Press, Matsumoto, Japan. pp. 801-809.
- Jackson, S.J. and R.C. Venema. 2006. Quercetin inhibits eNOS, microtubule polymerization, and mitotic progression in bovine aortic endothelial cells. J. Nutr. 136:1178-1184.
- Kim, S.J., I.S.M. Zaidul, T. Suzuki, Y. Mukasa, N. Hashimoto, S. Takigawa, T. Noda, M.E. Chie and H. Yamauchi. 2008. Comparison of phenolic compositions between common and Tartary buckwheat (*Fagopyrum*) sprouts. Food Chem. 110: 814-820.
- Kim, S.J., C. Kawaharada, T. Suzuki, K. Saito, N. Hashimoto and S. Takigawa. 2006. Effect of natural light periods on rutin, free amino acid and vitamin C contents in the sprouts of common (*Fagopyrum esculentum M*) and tartary (*F. tataricum G*) buckwheats. Food Sci. Technol. Res.12:199-205.

- Kim, S.L., K.S. Young, J.J. Hwang, S.K. Kim, H.S. Hur and C.H. Park. 2001. Development and utilization of buckwheat sprouts as functional vegetables. Fagopyrum 18:49-54.
- Kitabayashi, H., A. Ujihara, T. Hirose and M. Minami. 1995.
  Varietal differences and heritability for rutin content in common buckwheat, *Fagopyrum esculentum* Moench. Jpn. J. Breed. 45:75-79.
- Li, X., N.I. Park, Y.B. Kim, H.H. Kim, C.H. Park, Q. Wu and S.U. Park. 2012. Accumulation of flavonoids and expression of flavonoid biosynthesis genes in tartary buckwheat and rice-tartary buckwheat. Process Chemistry 47:2306-2310.
- Li, S.Q. and Q.H. Zhang. 2001. Advances in the development of functional foods from buckwheat. Crit. Rev. Food Sci. Nutr. 41:451-464.
- Lichtenthaler, H.K. and A.R. Wellburn. 1983. Determination of total carotenoids and chlorophyll a and b of leaf extract in different solvents. Biochem. Soc. Trans. 11:591-592.
- Liu, C.L., Y.S. Chen, J.H. Yang and B.H. Chiang. 2008. Antioxidant activity of tartary (*Fagopyrum tataricum* (L.) Gaertn.) and common (*Fagopyrum tataricum* Gaertn). buck-wheat sprouts. J. Agric. Food Chem. 56:173-178.
- Morishita, T., H. Yamaguchi and K. Degi. 2007. The contribution of polyphenols to antioxidative activity in common buckwheat and tartary buckwheat grain. Plant Prod. Sci. 10:99-104.
- Mortensen, A. 2006. Carotenoids and other pigments as natural colorants. Pure and Applied Chemistry 78:1477-1491.
- Oomah, B.D. and G. Mazza. 1996. Flavonoids and antioxidative activities in buckwheat. J. Agric. Food Chem. 44:1746-1750.
- Patra, M., E. Salonen, E. Terama, I. Vattulainen, R. Faller and B.W. Lee. 2006. Under the influence of alcohol: The effect of ethanol and methanol on lipid bilayers. Biophysical J. 90:1121-1135.
- Sharma, P., A.K. Ghimeray, A. Gurung, C.W. Jin, H.S. Rho and D.H. Cho. 2012. Phenolic contents, antioxidant and αglucosidase inhibition properties of Nepalese strain buckwheat vegetables. African J. Biotechnol. 11:184-190.
- Wang, Y. and C. Campbell. 2007. Tartary buckwheat breeding through hybridization with its rice-tartary type. Euphytica 156:399-405.
- Yao, L.H., Y.M. Jiang, J. Shi, F.A. Tomas-Barberan, N. Datta and R. Singanusong. 2004. Flavonoids in food and their health benefits. Plant Foods Human Nutr. 59:113-122.