

Laboratory Investigation

Analysis of Molecular Expression in Adipose Tissue-Derived Mesenchymal Stem Cells : Prospects for Use in the Treatment of Intervertebral Disc Degeneration

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Objective : Recent studies have shown encouraging progress toward the use of autogenic and allogenic mesenchymal stem cells (MSCs) to arrest, or even lead to partial regeneration in, intervertebral disc (IVD) degeneration. However, this technology is still in its infancy, and further development is required. The aim of this study was to analyze whether rat adipose-derived mesenchymal stem cells (ADMSC) can differentiate towards IVD-like cells after treatment with transforming growth factor β 3 (TGF- β 3) *in vitro*. We also performed quantitative analysis of gene expression for ADMSC only, ADMSCs treated with TGF- β 3, and co-cultured ADMSCs treated with TGF- β 3.

Methods : ADMSCs were sub-cultured to homogeneity and used in fluorocytometry assays for CD11, CD45, and CD90/Thy1. ADMSCs were differentiated in spheroid culture towards the chondrogenic lineage by the presence of TGF- β 3, dexamethasone, and ascorbate. We also co-cultured pure ADMSCs and nucleus pulposus cells in 24-well plates, and performed immunohistochemical staining, western blotting, and RT-PCR for quantitative analysis of gene expression.

Results : Results of fluorocytometry were positive for CD90/Thy1 and negative for CD11 and CD45. TGF- β 3-mediated induction of ADMSCs led to the expression of the differentiation markers of intervertebral disc-like cells, such as aggrecan, collagen II, and sox-9. Co-cultured ADMSCs treated with TGF- β 3 showed higher expression of differentiation markers and greater extracellular matrix production compared with ADMSCs treated with TGF- β 3 alone.

Conclusion : ADMSC treated with TGF- β 3 may be an attractive source for regeneration therapy in degenerative IVD. These findings may also help elucidate the pathologic mechanism of MSC therapy in the degeneration of IVD *in vivo*.

Key Words : Adipose tissue · Mesenchymal stem cells · Intervertebral disc · Cell and tissue engineering.

INTRODUCTION

Stem cell-mediated cell transplantation can reconstruct the normal structure and function of degenerated intervertebral discs (IVDs) by supplementing the loss of nucleus pulposus cells and promoting the formation of extracellular matrix⁷. Recent studies have shown encouraging progress toward the use of autogenic and allogenic mesenchymal stem cell (MSCs) to arrest, or even lead to partial regeneration of, IVD degeneration

in various animal models^{9,10}. However, it is clear that this technology is still in its infancy, and further developments are required while many issues await clarification⁵. One major issue is the identification of a suitable cell population with an expression profile resembling that of native IVD tissue and the capacity to regenerate this tissue. An additional concern is the acquisition of the cells in sufficient quantities for application in a clinical setting¹¹. Further, a greater understanding of the biologic mechanism of MSC transplantation, which remains uncertain, would

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be helpful in developing therapies. Therefore, we analyzed whether rat adipose-derived stem cells (ADMSCs) can differentiate towards IVD-like cells following transforming growth factor β 3 (TGF- β 3)-mediated induction *in vitro*. We also performed quantitative analysis of gene expression for ADMSC only, ADMSCs treated with TGF- β 3, and co-cultured ADMSCs treated with TGF- β 3, and evaluated the biologic mechanism of MSC therapy for regeneration of IVD tissue.

MATERIALS AND METHODS

Adipose-derived mesenchymal stem cell (ADMSC) isolation

Female Sprague-Dawley (SD) rats were used in this study. Approximately 10 g of fat tissue was harvested from the bilateral inguinal area. Harvested fat tissue was washed in saline and digested in 0.075% collagenase (filtered with a 0.20- μ m syringe filter) at 37°C for 1 h. The harvested fat tissue was treated with an equal volume of low-glucose Dulbecco's modified Eagle's medium (DMEM : Gibco, Invitrogen) containing 10% fetal bovine serum (Gibco, Invitrogen) and 1% penicillin/streptomycin (Gibco, Invitrogen). Cells were collected by centrifugation for 10 min at 600 \times g, and the supernatant was discarded. The pellet was resuspended in 160 mM ammonium chloride and incubated for 3 min at room temperature. ADMSCs were isolated by mash filtration (70- μ m nylon mash filter) and centrifuged at 600 \times g for 10 min. The pellet was suspended in a 75-cm² flask (Nunc, Invitrogen) as the primary culture and incubated at 37°C under 5% CO₂. Non-adherent cells were removed by replacing the medium. Cells were fed every 3 days with 10 mL of complete DMEM media. When this primary culture of ADMSCs reached 80% confluence, cells were harvested using 0.25% Trypsin-EDTA, and washed in 10 mL of phosphate-buffered saline (PBS). Cell viability was assessed by Trypan blue staining.

Identification of ADMSCs

Fluorescence-activated cell sorting (FACS) was used to analyze the surface markers of rat ADMSCs. After 3 passages, the cells were trypsinized and resuspended in DMEM containing 10% fetal bovine serum (FBS). Samples were counted, centri-

fuged, and resuspended in PBS. The cells were placed into eppendorf tubes at 1 \times 10⁶ cells per 1.5 mL, washed twice with PBS, and incubated for 1 hour at room temperature with the following fluorescein isothiocyanate-conjugated antibodies (Peprotech Inc., USA) : anti-rat CD90 (1 : 200), anti-rat CD45 (1 : 200), and anti-rat CD11b (1 : 200). In control groups, the cells were incubated in PBS without antibodies. The samples were then washed twice with PBS and analyzed by FACS.

Chondrogenic differentiation of ADMSCs

Chondrogenic differentiation of ADMSCs was achieved using the StemPro[®] Chondrogenesis Differentiation kit (Invitrogen, USA). About 1.6 \times 10⁷ ADMSCs were resuspended in 1 mL of pre-warmed complete chondrogenesis medium. Micromass cultures were generated by seeding 5- μ L droplets of cell solution in the center of multi-well plates. After cultivating micromass cultures for 2 hour under high humidity, warmed chondrogenesis medium was added to cultured vessels and incubated at 37°C and 5% CO₂ for 14 days.

Nucleus pulposus (NP) cells isolation for co-culture

We carefully extract intervertebral disc from SD rat tail and hold the annulus and nucleus, separated as much as possible. Harvested nucleus pulposus (NP) were washed in Hank's Balanced Salt Solution (HBSS : Gibco, Invitrogen) and diced the sample with sterilized tools and washed twice with HBSS. The diced tissues were digested in pronase solution [5% FBS (Gibco, Invitrogen), 0.2% pronase (Calbiochem, USA) in PBS (pH 7.4)] for 1 hour at 37°C. Pellet was collected by centrifugation for 10 min at 600 \times g, then the supernatant was discarded. After that the pellet was resuspended in collagenase-P solution [5% FBS, 0.02% collagenase-P (Roche, USA) in PBS (pH 7.4)] and digestion for 1 h at 37°C. Cells were collected by centrifugation for 5 min at 600 \times g, then the supernatant was discarded either. The pellet was resuspended in HBSS and washed for 5 min, and cells were collected by centrifugation for 10 min at 600 \times g, and the supernatant was discarded. NP cells placed in a 75-cm² flask (Nunc, Invitrogen) as the primary culture and incubated at 37°C under 5% CO₂. Non-adherent cells were removed by replacing the medium. Cells were fed every 3 days with 10 mL of DMEM media.

Table 1. Primers

	Primer	Product size
Aggrecan	5'-tcaacctcccactgaaggc-3' 5'-tttactatgacctcagggtg-3'	350 bp
Collagen II	5'-aggacctaaaggacctcc-3' 5'-agagacaccaggctgccagg-3'	364 bp
Sox-9	5'-tgtcggaggactcggctggtt-3' 5'-agggtctctctcgtctcgt-3'	384 bp
HIF-1 α	5'-actatgtcgtcttctgg-3' 5'-gtttctgctgcttgta-3'	195 bp
β -actin	5'-ggtatgggtcagaaggactcc-3' 5'-Cctgtcagcaatgctggta-3'	801 bp

Co-culture of chondrogenesis-differentiated ADMSCs and NP cells

We used a 6-well co-culture system (Nunc, USA). Due to the low-protein binding property of PET membranes, wide contact area and numerous micro pore cell, culture inserts are especially suited for co-culture. Chondrogenesis-differentiated ADMSCs were placed on the bottom layer, while the nucleus pulposus cells were placed on the upper

layer. The cells were incubated for 14 days and the medium was changed every 3 days (Fig. 1).

Immunohistochemical evaluation

After 2 weeks of co-culture, all cells were fixed with acetone for 10 min. The cells were washed 3 times with pH 7.6 Tris-buffered saline (TBS), and then blocked with TBS containing 5% bovine serum albumin and 0.1% Triton X-100 for 1 hour at room temperature. After incubating overnight with primary antibody at 48°C, the sections were washed 3 times with TBS and incubated with fluorescent-conjugated secondary antibody (1 : 1000; Invitrogen, USA). Polyclonal antibodies against anti-aggrecan (1 : 200; Abcam, UK) and collagenase type II (1 : 200; Abcam, UK) were used as primary antibodies. Stained cells were observed under a fluorescence microscope (Carl Zeiss Meditec, Germany). Appropriate controls, including omission of either primary or secondary antibodies, were included to confirm that the primary antibodies produced specific staining.

RT-PCR

For RNA isolation, media was aspirated from the dishes and the cells were washed with 1 mL of diethylpyrocarbonate-treated water. One milliliter of TRIzol® reagent was added to each dish. Homogenized samples were incubated for 5 min at room temperature, and 200 µL of chloroform was added per 1 mL of TRIzol® reagent by pipetting. The samples were shaken vigorously for 15 s and centrifuged at 12000× g at 4°C for 10 min. The aqueous phase was transferred to a fresh tube, and 500 µL of isopropyl alcohol was added per 1 mL of TRIzol® reagent. The sample was incubated at room temperature for 10 min to allow RNA to precipitate, and then centrifuged at 12000× g for 10 min at 4°C. The RNA was washed with 1 mL of 75% EtOH and centrifuged at 7500× g for 5 min at 4°C. The RNA pellet was briefly dried, resuspended in RNase-free water, and incubated for 10 min at 60°C. The cDNA synthesis reactions were incubated at 55°C for 50 min [cDNA mixture : 2 µL of 10× RT buffer, 4 µL of 25 mM MgCl₂, 2 µL of 0.1 M dithiothreitol, 1 µL of RNaseOUT™ (40 U/µL), 1 µL of Superscript™ III (200 u/200 µL)]. The mixture was then incubated at 85°C for 5 min and placed on ice. One microliter of RNase H was then added and incubated for 20 min at 37°C. For PCR, sense and anti-sense primers, template DNA, and distilled water were used. Primers were used for β-actin, aggrecan, collagen type II, SOX9, and HIF-1α (Table 1).

Western blotting

ADMSCs were harvested after 3 passages. Chondrogenesis-differentiated ADMSCs and co-cultured chondrogenesis-differentiated ADMSCs were harvested after 14 days of treatment. Proteins were extracted by homogenization in 100 µL of lysis buffer containing 20 mM Tris pH 7.4, 50 mM NaCl, 1% Triton X100, and protease inhibitors (Intron, Korea). Debris was cleared by centrifugation, and aliquots of the supernatants were stored

at -80°C. Samples were assayed for protein concentration using bicinchoninic acid assay (Bio-Rad, Richmond, CA, USA). After heating at 100°C for 5 min in 5× sample buffer, equal amounts of denatured protein (30 µg) were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis on 10% gels. Proteins were transferred onto polyvinylidene fluoride membranes at 20 V for 30 min by using a Semi-Dry Trans-Blot apparatus (Bio-Rad). After incubating for 1 hour at room temperature with blocking solution (0.4% Tween-20 and 5% dry milk in PBS), membranes were incubated overnight with primary antibodies against aggrecan (1 : 200; Abcam, UK) or collagen type II (1 : 200; Abcam, UK). An antibody against β-actin (1 : 10000; Sigma, St. Louis, MO, USA) was used as a control for equal loading. After washing with TBS/0.1% Tween-20, blots were incubated with horseradish peroxidase-conjugated secondary antibodies (Sigma, USA) and developed using enhanced chemiluminescence reagents (Pierce Biotechnology, USA).

Statistical analyses

The SPSS Version 12.0 (SPSS Inc., Chicago, IL, USA) statistical package was utilized for statistical analyses. Data was represented as mean±standard deviation and repeated measure analysis of variance with post-hoc analysis was used to analyze the differences in radiological measurements at 2, 4, and 6 weeks after cell injection in NC, IN, and TS segments. *p* values less than 0.05 were considered statistically significant.

RESULTS

Culture of isolated ADMSCs in monolayer and identification of ADMSCs

ADMSCs cultured in monolayer had a fibroblast-like shape under an inverted phase-contrast microscope. FACS analysis showed that rat ADMSCs expressed a number of specific markers of stem cells : CD90, CD45, and CD11. Therefore, we concluded that rat ADMSCs were positive for CD90 and negative for CD45 and CD11b (Fig. 2).

Chondrogenic differentiation and immunohistochemical evaluation of ADMSCs

We determined the chondrogenic differentiation status of AD-

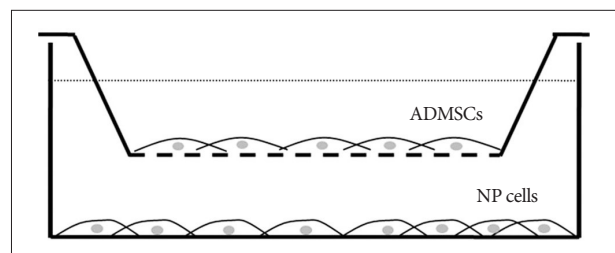


Fig. 1. Schematic diagram shows co-culture system. Insert has low-protein binding property of PET membranes, wide contact area and numerous micro pore. ADMSC : adipose-derived mesenchymal stem cells, NP : nucleus pulposus.

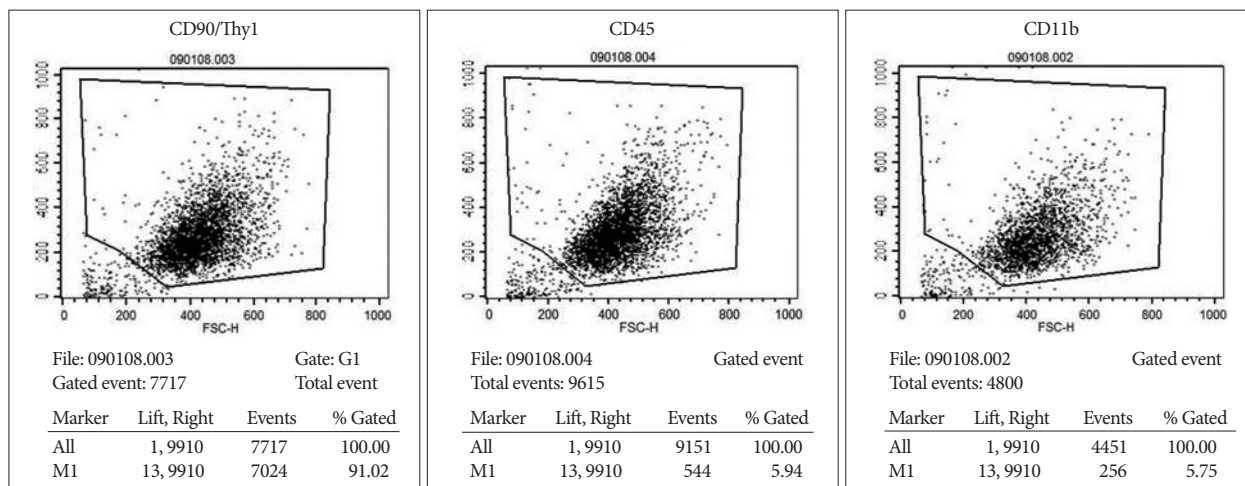


Fig. 2. FACS analysis shows that rat ADMSCs express high levels of CD90/thy1 and low levels of CD45 and CD11b. FACS : fluorescence-activated cell sorting, ADMSCs : adipose-derived stem cells.

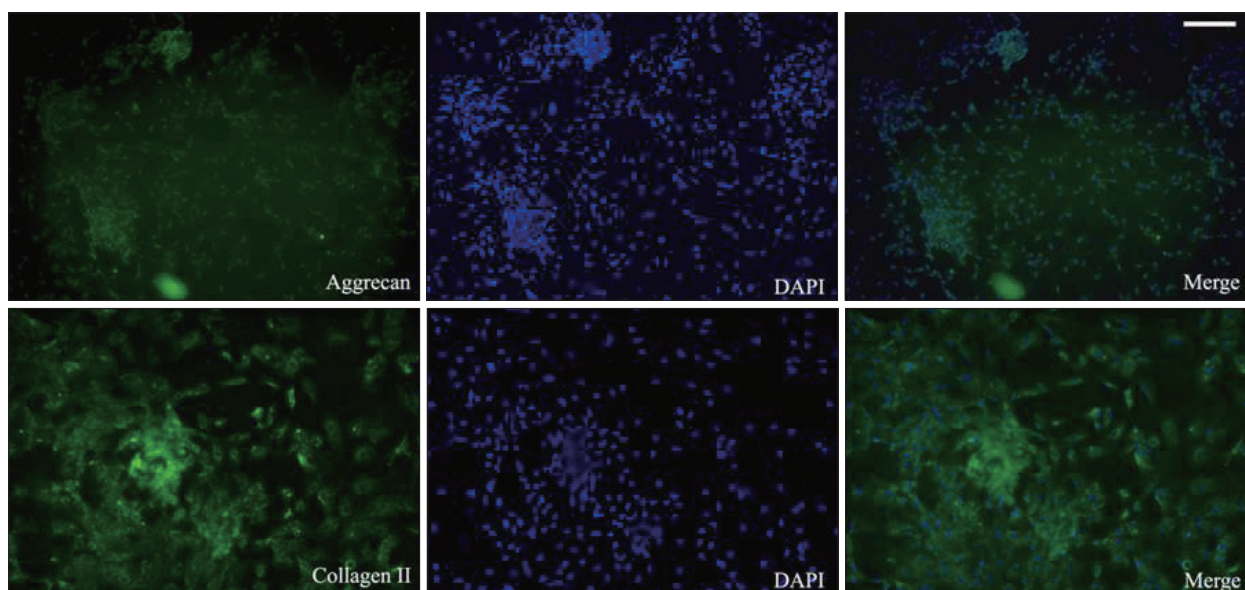


Fig. 3. Immunohistochemical analysis shows positive staining for aggrecan and collagen II (scale bar=100 μm). ADMSCs : adipose-derived stem cells, TGF-β3 : transforming growth factor β3.

MSCs by inverted phase-contrast microscopy. Immunohistochemical analysis revealed positive staining for aggrecan and collagen II (Fig. 3).

RT-PCR

The results of RT-PCR indicated that ADMSCs treated with TGF-β3 showed a significant increase in the gene expression of aggrecan, collagen II, and Sox-9 over untreated ADMSCs. Furthermore, co-cultured ADMSCs treated with TGF-β3 had further elevated expression of these genes compared to the ADMSCs treated with TGF-β3 alone (Fig. 4). Therefore, co-cultured ADMSCs treated with TGF-β3 showed the maximum expression of differentiation markers among these samples.

Western blotting

Western blotting showed results similar to RT-PCR. Moreover,

co-cultured ADMSCs treated with TGF-β3 expressed more extracellular matrix compared with ADMSCs alone or ADMSCs treated with TGF-β3 (Fig. 5).

DISCUSSION

Adipose tissue is an abundant, accessible, and replenishable source of adult stem cells. The ADMSCs are multipotent and differentiate along the adipocyte, chondrocyte, myocyte, neuronal, and osteoblast lineages. ADMSCs have potential applications for the repair and regeneration of acute and chronically damaged tissues. Most importantly, a comparative analysis of the MSCs obtained from the bone marrow, adipose tissue, and umbilical cord clearly showed that adipose-derived cells were not different from the MSCs derived from other tissues with regard to morphology, immune phenotype, success rate of MSC

isolation, colony frequency, and differentiation capacity³). Therefore, this suggests that adipose tissue is the most attractive source for MSCs for researchers and clinicians in nearly all medicinal subspecializations²). In our study, we used three-dimensional cultured ADMSCs which was confirmed their identity by FACS analysis.

Some studies showed that TGF- β -mediated induction protocols for MSCs in 3D culture resulted in a gene expression profile highly similar to that of native IVDs tissue. Furthermore its molecular and histological appearance closer to fibrocartilage than hyaline articular cartilage⁶). However, these common protocols for TGF- β -mediated chondrogenesis seem to be most suitable for generation of a collagen type I-rich fibrocartilaginous tissue and has tendency towards a molecular phenotype and morphological features of hyaline cartilage. Therefore, these methods may need to be adapted to reach transcription levels close to IVD tissue. In our study, to elucidate the effect of TGF- β -mediated chondrogenesis and co-culture ADMSC with NP cells, we performed comparative study for cultured ADMSCs, ADMSCs treated with TGF- β and co-cultured ADMSCs treated with TGF- β and accomplished immunohistochemical staining and extracellular matrix expression by PCR and western blotting. We found that all of them have nucleus pulposus-like activity and the expression of differentiation markers and extracellular matrix. Co-culture methods NP cell with ADMSCs treated with TGF- β was one of the modification methods to reach transcription levels close to IVD tissue.

The previous studies have been reported the suitable cell population with the capacity to regenerate IVD tissue and acquisition methods of an unlimited cell source with an expression profile resembling that of native IVD tissue⁶). Although promising, stem cell-based regenerative medicine remains a developing field. For disc regeneration treatment, further advances are required, and many issues await clarification. The one major issue is the biologic mechanism of MSC transplantation. A previous study reported that MSCs directly penetrate into damaged tissue and secrete or indirectly generate various cytokines that are useful for tissue regeneration, including nerve growth fac-

tor, neurotrophic factor, and vascular endothelial cell growth factor. Other studies have revealed that MSCs stimulate intrinsic stem cells in the tissue and differentiate target tissues⁴). In this study, we revealed that the rat ADMSCs and rat ADMSCs treated with TGF- β could be differentiated into functional NP-like cells. The rat ADMSCs and rat ADMSCs treated with TGF- β expressed differentiation markers and extracellular matrix. Therefore, we expected that all of them were suitable for regeneration therapy following IVD degeneration. Furthermore, co-cultured ADMSCs treated with TGF- β was more effective to the regeneration of intervertebral disc than the other cells.

The other major issue is clarification of the suitable cell popu-

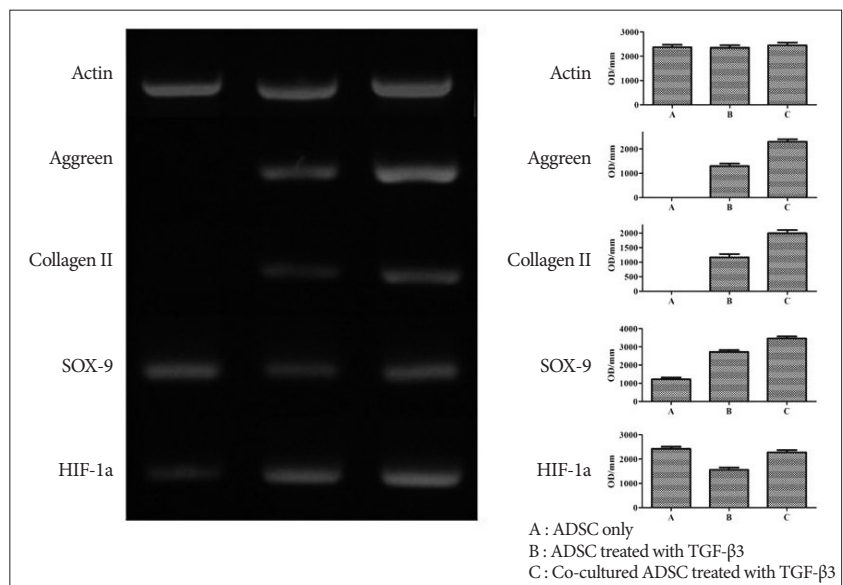


Fig. 4. Results of RT-PCR show that ADMSCs treated with TGF- β 3 displayed a significant increase in the gene expression of aggrecan, collagen II, and Sox-9 over untreated ADMSCs. Co-cultured ADMSCs treated with TGF- β 3 was further elevated compared with the ADMSCs treated only with TGF- β 3.

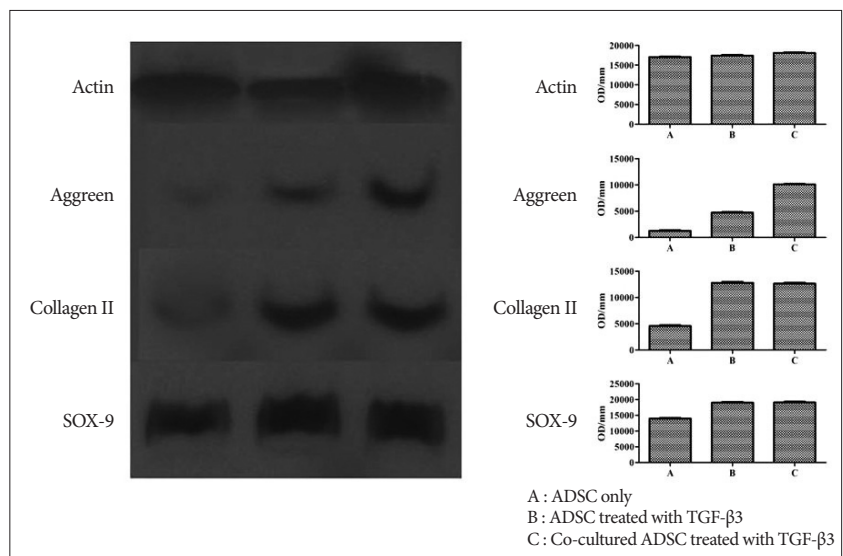


Fig. 5. Western blotting showed that co-cultured ADMSCs treated with TGF- β 3 had greater expression of extracellular matrix compared with ADMSCs alone or ADMSCs treated only with TGF- β 3.

lation and environment. The pre-loading of biological signals to MSCs prior to implantation in order to “direct” MSC or host cell differentiation, such as using cytokines or genes, or co-culture, has been tested as a means to enhance stem cell therapy⁵⁾. Cells in the NP are heterogeneous, comprised of notochordal cells and chondrocyte-like nucleus pulposus cells. Notochordal cells in the nucleus pulposus have been proposed to act as “organizers” or behave as stem cells for IVD maintenance and repair¹⁾. The origin, function, and characteristics of NP cells, and the signals required for NP cell differentiation, remain unclear. Some previous studies have attempted to stimulate MSCs to differentiate into chondrocyte-like or NP-like cells using cytokines or genes. For example, MSCs carrying a SOX9-expressing adenovirus were seeded into a synthetic poly-lactic acid polymer scaffold prior to implantation⁸⁾. Other studies have used co-culturing methods with differentiated cells that can determine the fate of MSCs. They revealed that adipose-derived rabbit MSCs can activate chondrocytic markers such as collagen II and aggrecan by NP tissues through soluble mediators⁶⁾. This study shows that ADMSCs treated with TGF- β 3 expressed more than ADMSCs alone. Furthermore, ADMSCs treated with TGF- β 3 and co-cultured with NP cell expressed more differentiation markers and extracellular matrix than ADMSCs treated with TGF- β 3. Therefore, the cellular microenvironment (including the co-culture status), was found to be the most important factor, as chondrocyte-specific gene expression and extracellular matrix production were the maximum greatest in our co-culture system.

The limitation of this study is that the design of study was *in vitro* type. Furthermore, evaluation parameter of this study has not totally representative for intervertebral disc regeneration effect. Therefore, further study should be combined *in vitro* and *in vivo* study with detailed design.

CONCLUSION

ADMSC treated with TGF- β 3 maybe an attractive source for regeneration therapy in degenerative IVD. These findings may also provide evidence into the physiologic mechanism of MSC

therapy for the degeneration of IVD *in vivo*.

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