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Analysis of Geographic and Pairwise Distances among Chinese Cashmere Goat Populations

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ABSTRACT: This study investigated the geographic and pairwise distances of nine Chinese local Cashmere goat populations through the analysis of 20 microsatellite DNA markers. Fluorescence PCR was used to identify the markers, which were selected based on their significance as identified by the Food and Agriculture Organization of the United Nations (FAO) and the International Society for Animal Genetics (ISAG). In total, 206 alleles were detected; the average allele number was 10.30; the polymorphism information content of loci ranged from 0.5213 to 0.7582; the number of effective alleles ranged from 4.0484 to 4.6178; the observed heterozygosity was from 0.5023 to 0.5602 for the practical sample; the expected heterozygosity ranged from 0.5783 to 0.6464; and Allelic richness ranged from 4.7551 to 8.0693. These results indicated that Chinese Cashmere goat populations exhibited rich genetic diversity. Further, the Wright's F-statistics of subpopulation within total (FST) was 0.1184; the genetic differentiation coefficient (GST) was 0.0940; and the average gene flow (Nm) was 2.0415. All pairwise FST values among the populations were highly significant (p<0.01 or p<0.001), suggesting that the populations into at least four clusters, with the Hexi and Yashan goat populations alone in one cluster. These results have provided useful, practical, and important information for the future of Chinese Cashmere goat breeding. (**Key Words:** Cashmere Goat, Geographic Distance, Pairwise Distance, Genetic Diversity)

INTRODUCTION

Goats occupy an extremely important position in human history as one of the first animal species to be domesticated (Luikart et al., 2001; Fernandez et al., 2006). Goats possess a special ability to adapt rapidly to different environmental conditions, and the species' special characteristics put it in a position of economic importance. Cashmere goats are found only in specific areas of Asia, with modern Cashmere goat breeds generally originating from the territorial plateau of southwestern China and adjacent areas of central Asia (Tu, 1989). The world's largest number of Cashmere goat populations is raised in China, and these populations are a significant resource: more than ten different populations have developed after long-term natural and human selection (Qi, 2004). Among these populations, the Liaoning and Inner Mongolian Cashmere goats (including three types of Alashan, Erlangshan, and Wuzhumuqin) are well known worldwide for their high cashmere yield and cashmere fineness, respectively. Cashmere goat breeding plays an important role in Chinese animal agriculture, and particularly so in underdeveloped, rural, and mountain areas, where production systems are based on local or native breeds. In the past three decades, a high cashmere yield has been the main focus in breeding. Importantly, genetic admixture among some breeds, as well as inbreeding within the breed/types, is possible. These activities may have led to a loss of genetic structure and diversity in some of these breed/types. In recent decades, an increasing focus on cashmere production within animal agriculture has resulted in changes in Cashmere goat management systems: there is an increasing emphasis on the breeding of a small number of very productive cosmopolitan breeds. In light of this, it is of great importance to conserve genetic resources in order

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to maintain more extensive breeding options. The effective assessment of genetic variability in domestic Cashmere goat populations constitutes the first step toward this genetic resource conservation.

Phylogenetic studies of population diversification have allowed researchers to identify the essential features in many species' evolutionary histories (Avise, 2000). Population differentiation through reduced gene flow, selection, and genetic drift can alter the geographic species shift scenarios that occur under regional warming (Davis and Shaw, 2001). The intensification of production systems and the successes observed in industrial breeding have led some farmers to abandon certain native breeds. Additionally, domesticated animals are presently experiencing a loss of genetic diversity as a result of several other factors. First, the intensive selection of highly productive breeds has placed inadequate emphasis on the preservation of overall genetic diversity. Second, native breeds in marginal areas are facing extinction, and little or no action is being taken to reverse this trend (Taberlet et al., 2008). Microsatellites provide accurate genetic information on individual genotypes and the genetic relationships between populations (Arranz et al., 1998; Ruane, 1999; Kantanen et al., 2000). However, for analysis of microsatellite data, the efficiencies of distance measures in phylogenetic reconstruction as compared to the traditional distance metrics remain unknown. In this paper, we will consider only one neighbor-joining (NJ) method (Saitou and Nei, 1987) that is assessed using genetic distances (D_s) and (D_A) , because this particular NJ method is considered to be robust across many situations (Nei, 1991). The calculation of genetic distances using microsatellite data can provide useful information for the overall monitoring and management of the genetic diversity of rare breeds.

Microsatellite DNA is currently the most useful and widely used marker that can be used to perform a wide range of molecular genetic studies, including establishing the population structure (Bruford, 1993), population differentiation, and reconstruction of phylogenetic relationships among populations (Bowcock, 1994; MacHugh, 1997; Wimmers, 2000). These microsatellites provide accurate genetic information on individual genotypes and the genetic relationships between populations (Arranz et al., 1998; Ruane, 1999; Kantanen et al., 2000). Such genetic diversity analyses can provide insight into domestication events, relationships among breeds, loss of within-breed genetic diversity, breed structure, etc., and are essential for establishing conservation priorities (Toro et al., 2009). In recent years, efforts have been made to carry out studies that are aimed to characterize the genetic diversity in goat breeds from several different countries and regions; the available information has been reviewed recently (Groeneveld et al.,

2010). The vast majority of these genetic diversity studies has used microsatellite markers to obtain information, and most of the studies cover a sample of breeds from a given country (Ganai and Yadav, 2001; Li et al., 2002; Tadlaoui-Ouafi et al., 2002; Visser et al., 2004; Xiang-Long and Valentini, 2004; Iamartino et al., 2005; Sechi et al., 2005; Araújo et al., 2006; Gour et al., 2006; Martínez et al., 2006; Oliveira et al., 2007; Fatima et al., 2008; Glowatzki-Mullis et al., 2008; Wang et al., 2008; Qi et al., 2009; Traoré et al., 2009; Bruno-de-Sousa et al., 2011; Ozgecan and Okan, 2012). Some studies, however, have considered a broader geographical scope, including larger samples of breeds from Asia (Barker et al., 2001), the Mediterranean region (Canon et al., 2006), Africa (Muema et al., 2009), and breeds from multiple continents (Martinez-Martinez et al., 2008; Oliveira et al., 2010; Di et al., 2011; Gamaa and Bressan, 2011). Furthermore, Di et al. revealed that Chinese Cashmere goat breeds/types exhibited a rich genetic diversity (2011), with expected heterozgosity and allelic richness values ranging from 0.54 to 0.65 and from 3.90 to 5.96 respectively. Further, Di et al. showed that the expected heterozgosity of Chinese Cashmere goats was similar to that of West African goats, but lower than that of Iranian goats. The aim of the present study was to assess how useful were microsatellite polymorphisms for analysis of the genetic relationships between nine Chinese local Cashmere goat populations, with the ultimate aim of maintaining and conserving the populations. Results of this analysis will help to further understand the morphology and geographical distribution among Chinese Cashmere goat populations.

MATERIAL AND METHODS

Animals and DNA samples

We selected nine Chinese local Cashmere goat populations for investigation in this study. For analysis, 10 ml blood samples were collected from the jugular vein of each investigated animal. From the 10 ml samples, 2 ml samples were quickly frozen in liquid nitrogen and stored at -80°C for genomic DNA extraction, as described previously (d'Angelo et al., 2006). The total DNA was extracted from whole blood using the saturated salt method (Sambrook et al., 1989), quantified spectrophotometrically, and adjusted to 50 ng/µl. Blood samples were collected from 809 goats from nine breeds/types, spanning the entire distribution range of Chinese Cashmere goats. The genetic characteristics for these Cashmere goats were analyzed in order to ascertain the historical relationships and relative genetic contributions for the populations. The investigated populations included the following numbers and corresponding population types: 84 Hexi Cashmere goat (HX), 108 Chaidamu Cashmere goat (CDM), 112 Xinjiang Cashmere goat (XJ), 116 Liaoning Cashmere goat (LN), 96 Erlangshan Cashmere goat (ELS), 72 Alashan Cashmere goat (ALS), 76 Hanshan Cashmere goat (HS), 86 Wuzhumuqin Cashmere goat (WZMQ), and 59 Yashan Cashmere goat (YS) raised in China (Table 1).

Primer design and PCR amplification

Primers flanking 20 microsatellite loci located at several different chromosomes were designed by an available genome sequence (Table 2) (Crawford et al., 1995) and synthesized by the Shanhai Shenggong Biological Engineering Company. The 20 selected microsatellite DNA markers were recommended by the Food and Agriculture Organization of the United Nations (FAO) and the International Society for Animal Genetics (ISAG). PCR reactions took place in a 25 µl reaction system containing 2 μl genomic DNA template; 1.5 μl MgCl₂ (25 mmol/L); 2 μl dNTP (10 mmol/L); 0.25 µl Taq DNA polymerase (5 µl/U) (TaKaRa, China); 2 µl each forward and reverse primers (8 pmol/ μ l); 2.5 μ l 10×PCR Buffer; and ddH₂O (up to 25 μ l). The PCR conditions were as follows: initial denaturation for 5 min at 94°C, 35 cycles at 94°C for 1 min, annealing temperatures at 52 to 58°C for 40 s, and extension at 72°C for 40 s. The final extension step was followed by a 5 min extension at 72°C.

DNA sequencing and agarose electrophoresis analysis

For analysis, 2 μ l PCR product was mixed with 6 μ l denaturing solution (95% formamide, 25 mM EDTA, 0.025% xylenecyanole, and 0.025% bromophenol blue), heated for 10 min at 98°C, and then chilled on ice. The samples were electrophoresed with 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The electrophoresis ran at 250 V and 40 mA (preelectrophoresis) for 10 min, followed by 150 V and 24 mA (Kucharczyk Techniki Elektroforetyczne) for 8 h (silver staining). A refrigerated circulator was used to control the gel temperature (4°C). Then, each locus of the number of alleles was computed using Excel Microsatellite Toolkit (Version 3.1) software. The fragment length of PCR products was determined using GeneMapper software (Version 3.7).

Statistical analysis

The gene and genotypic frequencies were estimated using direct gene counting. The N_a , Ne (Kimura and Crow, 1974), Pa, He (Nei et al., 1983), Ho, PIC (Botstein et al., 1980), and AR values were estimated using POPGENE software (version 1.31) (Yeh et al., 1997). The significance of the F-statistics was determined from permutation tests, in which the sequential Bonferroni procedure was applied over loci (Hochberg, 1988). The F-statistics indices (Wright, 1978) were computed using the FSTAT program (Goudet, 2002), and the estimate was based on the following relationship: $N_m = 0.25(1-F_{ST})/F_{ST}$, where F_{ST} was the mean F_{ST} value across all loci (Slatkin and Barton, 1989). Reynolds' genetic distance between populations was subsequently calculated on the basis of the F_{ST} values (Reynolds et al., 1983). A consensus neighbour-joining (NJ) dendrogram of the nine Chinese local Cashmere goat populations, based on Nei's genetic distances (D_s) (Nei, 1972) and Nei's genetic distances (D_A) (Nei et al., 1983), was reconstructed using DISPAN software (version 1.1); this was the most effective way to obtain accurate phylogenetic trees (Saitou and Nei, 1987; Takezaki and Nei, 1996). Finally, G_{ST} was calculated on the basis of the data from all loci (Takezaki et al., 2010).

RESULTS

Genetic diversity

Table 2 and 3 display the following data concerning the 20 microsatellite loci used from the nine Chinese local Cashmere goat populations: a description of the markers, including the chromosomal localization, the number of alleles per marker, and the fragment sizes; observed and expected heterozygosities; the heterozygote deficiency within population (F_{IS}); and the deviation from the Hard-Weinberg equilibrium. The 20 microsatellites included a total of 206 alleles, and the mean number of alleles per locus in the population was 10.30. Most of the markers used had allele number values that were higher than nine, and the

Table 1. Name, code, sample size, and source region of nine Chinese cashmere goat populations

Population	Population code	Sample number	Sampling location
Hexi cashmere goat	HX	84	Mongolian and Yugu Autonomous and Tianzhu County, Gansu
Chaidamu cashmere goat	CDM	108	Delingha City, Qinghai
Xinjiang cashmere goat	XJ	112	Wulumuqi City, Xinjiang
Liaoning cashmere goat	LN	116	Kuandian County and Gaizhou City, Liaoning
Erlangshan cashmere goat	ELS	96	Bayannaor League, Inner Mongolia
Alashan cashmere goat	ALS	72	Alashan League, Inner Mongolia
Hanshan cashmere goat	HS	76	Chifeng City, Iner Mongolia
Wuzhumuqin cashmere goat	WZMQ	86	Xilingol League, Inner Mongolia
Yashan cashmere goat	YS	59	Xixia County, Shandong

values ranged from seven (SRCRSP3, OarFCB48, BMS1494, and BM1818) to 16 (DRBP1) alleles per marker. The fragment sizes ranged from 107 bp to 123 bp, from 138 bp to 178 bp, from 227 to 237 bp, from 200 to 284 bp, and from 106 to 154 bp, respectively (Table 2).

Table 3 contains genetic variation metrics for the nine Chinese local Cashmere populations. The highest mean numbers of effective alleles were found in LN (8.4025) and XJ (8.3608), followed by CDM (7.9613) and ELS (7.8812). Meanwhile, YS had the lowest mean number of effective alleles (6.7683). The highest average numbers of effective alleles were found in XJ (4.6178) and LN (4.5649), followed by ELS (4.4936) and CDM (4.3350); meanwhile, the lowest average number of effective alleles was found in YS (4.0484). The genetic diversity difference index (PIC value) for the whole dataset was high (0.6500), and most of the populations had PIC values above 0.6198. The lower PIC values were present in YS (0.5213) and WZMQ (0.5468), while higher PIC values were observed for XJ and LN (0.7582 and 0.7187, respectively). The observed

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Locus	Primer sequence (5'- 3')	Chromosome	Allele number	Fluorescent marker	Annealing temperature (°C)	Fragment size
ILSTS029	TGTTTTGATGGAACACAGCC	1	9	NED	53	152-180
	TGGATTTAGACCAGGGTTGG					
OarFCB20	AAATGTGTTTAAGATTCCATACAGTG	2	11	6FAM	55	82-104
	GGAAAACCCCCATATATACCTATAC					
MAF70	CACGGAGTCACAAAGAGTCAGACC	4	10	PET	60	142-184
	GCAGGACTCTACGGGGGCCTTTGC					
MCM527	GTCCATTGCCTCAAATCAATTC	5	11	NED	58	151-173
	AAACCACTTGACTACTCCCCAA					
ILSTS087	AGCAGACATGATGACTCAGC	6	11	PET	52	138-158
	CTGCCTCTTTTCTTGAGAGC					
SPS113	CCTCCACACAGGCTTCTCTGACTT	7	9	PET	58	130-152
	CCTAACTTGCTTGAGTTATTGCCC					
ILSTS005	GGAAGCAATGAAATCTATAGCC	7	8	6FAM	55	181-216
	TGTTCTGTGAGTTTGTAAGC					
ILSTS011	GCTTGCTACATGGAAAGTGC	9	11	6FAM	58	268-282
	CTAAAATGCAGAGCCCTACC					
SRCRSP3	CGGGGATCTGTTCTATGAAC	10	7	NED	55	107-123
	TGATTAGCTGGCTGAATGTCC					
CSRD247	GGACTTGCCAGAACTCTGCAAT	14	10	VIC	58	211-246
	CACTGTGGTTTGTATTAGTCAGG					
INRA063	ATTTGCACAAGCTAAATCTAACC	14	9	NED	58	165-199
	AAACCACAGAAATGCTTGGAAG					
MAF065	AAAGGCCAGAGTATGCAATTAGGAG	15	14	VIC	55	113-147
	CCACTCCTCCTGAGAATATAACATG					
OarFCB48	GACTCTAGAGGATCGCAAAGAACCAG	17	7	6FAM	57	138-178
	GAGTTAGTACAAGGATGACAAGAGGCAC					
MAF209	TCATGCACTTAAGTATGTAGGATGCTG	17	8	VIC	55	87-103
	GATCACAAAAAGTTGGATACAACCGTGG					
BMS1494	TCTGGAGCTTGCAAAAGACC	18	7	6FAM	55	227-237
	AATGGATGACTCCTGGATGG					
SRCRSP5	GGACTCTACCAACTGAGCTACAAG	18	12	VIC	58	152-182
	TGAAATGAAGCTAAAGCAATGC					
BM1818	AGCTGGGAATATAACCAAAGG	20	7	VIC	50	200-284
	AGTGCTTTCAAGGTCCATGC					
OarFCB304	CCCTAGGAGCTTTCAATAAAGAATCGG	22	15	VIC	55	150-188
	CGCTGCTGTCAACTGGGTCAGGG					
DRBP1	ATGGTGCAGCAGCAAGGTGAGCA	23	16	PET	53	106-154
	GGGACTCAGTCTCTCTATCTCTTTG					
OarAE54	TACTAAAGAAACATGCTCCCAC	25	14	6FAM	56	112-134
	AGAAACATTTATTCTTATCCTCAGTG					

NED: Black fluorescent; 6FAM: Blue fluorescent; PET: Red fluorescent; VIC: Green fluorescent.

 Table 3. Basic genetic variation information of the nine Chinese
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 Cashmere goat populations
 diagonal

Population	Allelic	diversity	Genetic diversity				
	Na	Ne	He	Ho	PIC	AR	
HX	7.4823	4.1671	0.6452	0.5197	0.7128	7.0816	
CDM	7.9613	4.3350	0.6331	0.5061	0.6883	6.8639	
XJ	8.3608	4.6178	0.6464	0.5602	0.7582	8.0693	
LN	8.4025	4.5649	0.5884	0.5371	0.7187	6.9698	
ELS	7.8812	4.4936	0.5896	0.5049	0.6469	6.4853	
ALS	7.7659	4.3271	0.5932	0.5119	0.6328	6.2616	
HS	7.7264	4.4207	0.6169	0.5402	0.6198	6.2951	
WZMQ	7.3627	4.4092	0.5846	0.5413	0.5468	6.4539	
YS	6.7683	4.0484	0.5783	0.5023	0.5213	4.7551	

 N_a : Mean number of effective alleles; N_e : Number of effective alleles; H_e : Expected heterozygosity; H_o : Observed heterozygosity; *PIC*: polymorphism information content; *AR*: Allelic richness.

heterozygosity (Ho) averaged over all loci was 0.5249, while the expected heterozygosity (He) was 0.6084. The allelic richness in the nine populations ranged from 4.7551 to 8.0693; it was higher in XJ and LN populations than in YS Cashmere goat populations (Table 3). Therefore, the 20 microsatellite loci could be used to evaluate the genetic diversity in Cashmere goat populations. Furthermore, results demonstrated that the investigated Chinese Cashmere goat populations exhibited a rich genetic diversity.

Wright's F-statistics

The genetic structure and genetic variation of the Cashmere goat populations were analyzed using the 20 microsatellite loci along with Wright's *F*-statistics. The mean F_{IT} was 0.1374, the mean F_{ST} was 0.1184, and the mean F_{IS} was 0.0229. Meanwhile, the G_{ST} values ranged from 0.0273 to 0.1738; N_m values for the markers varied from 1.0171 to 3.7246; the mean G_{ST} was 0.0940; and the mean N_m was 2.0415. The variation results both among and within populations for the small number of individuals can be characterized as inbred. Meanwhile, the distribution of the nine Chinese Cashmere goat populations differed in various microsatellite loci (p<0.05, p<0.01 or p<0.001) (Table 4).

Phylogenetic analysis

The genetic phylogeny was analyzed using three methods described by Peter et al. (2007). We determined an optimum K value of 4, following Evanno et al. (2005). Evanno et al. found that an *ad hoc* quantity, which was based on the second order rate of change of the likelihood function with respect to K, did show a clear peak at the true value of K. Meanwhile, Pritchard and Falush have

Table 4. F-statistics of nine Chinese cashmere goat populations at 20 microsatellite loci

Locus ——		All studied Chinese Cashmere goat populations							
	F_{IT}	F_{ST}	F_{IS}	G_{ST}	N_m				
OarFCB20	0.0785	0.0866**	0.0088	0.0762	2.6368				
ILSTS029	0.1738**	0.1029***	0.0790**	0.1103	2.1795				
MAF70	0.0676	0.0758**	-0.0089	0.0608	3.0482				
MCM527	0.0738	0.0629**	0.0116	0.0331	3.7246				
ILSTS087	0.1679**	0.1215***	0.0528**	0.1118	1.8076				
SPS113	0.1193*	0.1054***	0.0155	0.0836	2.1219				
ILSTS005	0.1036*	0.1372***	-0.0389	0.0273	1.5722				
ILSTS011	0.1151*	0.1385***	-0.0272	0.1482	1.5551				
SRCRSP3	0.2421***	0.1973***	0.0558**	0.1738	1.0171				
CSRD247	0.0309	0.0716**	-0.0438	0.0573	3.2416				
INRA063	0.1280*	0.1469***	-0.0222	0.1461	1.4518				
MAF065	0.0627	0.0915**	-0.0317	0.0690	2.4822				
OarFCB48	0.0806	0.0981**	-0.0194	0.0673	2.2984				
MAF209	0.2534***	0.1632***	0.1078***	0.1554	1.2819				
BMS1494	0.1986***	0.1358***	0.0727**	0.1412	1.5909				
SRCRSP5	0.1782**	0.1083***	0.0779**	0.0427	2.0584				
BM1818	0.1548**	0.1278***	0.0310*	0.0538	1.7062				
DRBP1	0.2186***	0.1064***	0.1256***	0.0862	2.0996				
OarFCB304	0.1028*	0.1369***	-0.0395	0.0956	1.5762				
OarAE54	0.1968***	0.1535***	0.0512**	0.1406	1.3787				
Total	0.1374**	0.1184***	0.0229*	0.0940	2.0415				

 F_{TT} : Wright's *F*-statistics of inbreeding within total; F_{ST} : Wright's *F*-statistics of subpopulation within total; F_{IS} : Wright's *F*-statistics of inbreeding within subpopulation; G_{ST} : Genetic differentiation coefficient; N_m : Gene flow; $N_m = 0.25(1-F_{ST})/F_{ST}$. Significance levels of deficit in heterozygotes: * p<0.05; ** p<0.01; *** p<0.001.

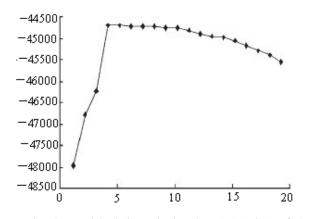


Figure 1. The model choice criterion $\ln P(D)$ (\pm SD) of the Structure analysis for *K* values ranging from 2 to 20 for the admixture model.

developed methods that allow for linkage between loci, which can produce highly accurate assignments using modest numbers of loci, (Pritchard et al., 2000; Falush et al., 2003). A clear clustering (K = 4) was observed, which was in accordance with population types (Figure 1). Further, the genetic distance between populations was analyzed using Nei's genetic distance (D_A) and Nei's standard genetic distance (D_S), and neighbour-joining phylogenetic trees for both methods were similar (Figures 2 and 3). More specifically, the neighbour-joining dendrogram that was based on Nei's genetic distances effectively divided the nine Chinese local Cashmere goat populations into four groups. YS, which produces very little cashmere, was genetically distinct and was the first to separate. The other Chinese goats were then divided into three sub-clusters. The first cluster included LN, CDM, and XJ, which belonged to the high-yield-cashmere type. The second cluster included ALS, ELS, WZMQ, and HS, which belonged to the finecashmere type. The third cluster included HX, and was farther from all of the other Chinese goats, with the exception of YS. The three-factor correspondence analysis for frequency distributions of the 20 microsatellite alleles in

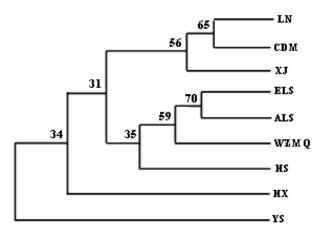


Figure 2. A neighbour-joining dendrogram of nine Cashmere goat populations based on Nei's genetic distances (D_A).

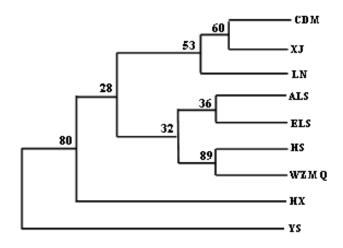


Figure 3. A neighbour-joining dendrogram of nine Cashmere goat populations based on Nei's standard genetic distances (D_S).

the nine Chinese local Cashmere goat populations is shown in Figure 4. The first two factors accounted for 31.42% and 25.52% of the total variation, respectively, and clearly distinguished the following principal components: Axis I (ELS, ALS, WZMQ, YS, and HS), Axis II (XJ, CDM, and LN), and Axis III (HX). The first two factors fit well with the geography, while the third factor, contributing 11.77% of the total variation, played an important role in discriminating the Hexi Cashmere goat population (Figure 4).

Geographic and pairwise distances

Table 5 presents the genetic distance and pairwise distance among the nine investigated populations, as shown by D_A and D_S values. The D_A values ranged from 0.0673 to 0.2873, and the D_S values from 0.0428 to 0.2745. The genetic distance between LN and CDM was the smallest ($D_A = 0.0673$, $D_S = 0.0428$), and the genetic distance between CDM and YS was the largest ($D_A = 0.2873$, $D_S = 0.2745$).

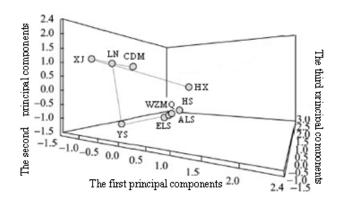


Figure 4. Scatter plot showing the first, second, and third principal components of genetic differentiation among nine Cashmere goat populations.

Population	HX	CDM	XJ	LN	ELS	ALS	HS	WZMQ	YS
HX	-	0.1738	0.1682	0.1636	0.1416	0.1521	0.1326	0.1165	0.2238
CDM	0.2295	-	0.0689	0.0673	0.0980	0.0942	0.1185	0.1372	0.2873
XJ	0.1687	0.0896	-	0.0783	0.1063	0.0980	0.1502	0.1192	0.2246
LN	0.2326	0.0428	0.0593	-	0.9841	0.1062	0.1266	0.1478	0.2224
ELS	0.1839	0.0985	0.0653	0.1068	-	0.1106	0.1123	0.1538	0.1918
ALS	0.1769	0.1063	0.0791	0.1596	0.0595	-	0.1165	0.1266	0.1937
HS	0.2239	0.1421	0.1439	0.1769	0.0663	0.0593	-	0.1196	0.1927
WZMQ	0.2298	0.1282	0.1399	0.1754	0.6526	0.0635	0.1095	-	0.2108
YS	0.2306	0.2745	0.1768	0.1893	0.1986	0.2262	0.1314	0.1380	-

Table 5. Nei's genetic distances (shaded area above the diagonal) and Nei's standard genetic distances (below the diagonal) among nine

 Chinese Cashmere goat populations

DISCUSSION

The polymorphism information content (PIC) for a population, and the heterozygosity of the individuals within, can reflect the extent of genetic variation, which shows high value for large. (Botstein et al., 1980). Information on the genetic variation within the populations investigated in this study is reported in Table 3. This study was expected to provide the genetic evidence for the classification of these breeds/types. The results showed that for the nine Chinese Cashmere goat populations investigated here, He and AR values ranged from 0.5783 to 0.6464 and from 4.7551 to 8.0693, respectively. The lowest value was found in YS and the highest in XJ and HX. The He observed for Chinese goats was similar to that which was reported in previous studies on Chinese goats (Li and Alessio, 2004; Chen et al., 2006; Kang et al., 2011) and West African goats, but lower than that observed in Iranian goats. Further, the allelic richness of the Chinese Cashmere goat populations studied here was higher than the allelic richness reported in other studies. And, the present study included more individuals (809) and more microsatellite markers (20) than the number of individuals (376) and microsatellite markers (14) included in the study by Di et al. Although we conducted similar analyses and used some of the same local populations (6 populations are the same, 3 populations are not the same), the allelic diversity and genetic diversity that resulted here was not consistent with that reported in the prior study (Di et al., 2011). In addition, the present study included PIC, F_{IT} , F_{IS} , G_{ST} , and N_m , while previous studies included a more limited set of parameters (Di et al., 2011). Regarding the population cluster produced in this study, Cashmere goat populations from Inner Mongolia were classified into one branch, and other Cashmere goats from Northern China were grouped into a second branch. This clustering further illustrated that in China, there were differences between Cashmere goats that were consistent with genetic distance and geographic distance.

The PIC value was originally introduced by Botstein et

al. (1980). Based on the number of detectable alleles and the distribution of their frequency, PIC can indicate how useful a marker can be for detecting polymorphism within a population. Additionally, the PIC value has proven to be a general measure of how informative is a marker (Guo and Elston, 1999): the higher the PIC value, the more the informative the marker. In some cases, populations that have similar frequencies at certain microsatellite loci may still have adaptively important differences that have been maintained by natural selection (Hedrick, 1999). In other cases, populations that have different allele frequencies at certain microsatellite loci may share adaptively important traits.

In general, differentiation at microsatellite loci should reflect the potential for adaptive differences among populations (Gutierrez-Espeleta et al., 2000). In this study, estimates of heterozygosity within the populations were based on a set of microsatellites that indicated that XJ had the largest genetic variability, while YS had the lowest. At the same time, the mean number of alleles and the mean observed and expected heterozygosities of XJ and YS were similar (Table 3). This may be because XJ included a large number of individuals and a broad distributing area, while YS existed in a remote area and had a small population size. Thus, there could have been less gene exchange between YS and other populations. YS ultimately showed an allelic diversity and genetic diversity that was lower than corresponding values for the other eight local Chinese Cashmere goat populations.

In general, Inner Mongolian Cashmere goats have been considered to be one breed. However, all of the obtained pairwise F_{ST} values were highly significant (p<0.001 or p<0.01) at 20 microsatellite loci. This indicated that, for breeding purposes, the ALS, ELS, and WZ breeds from Inner Mongolia could be considered independent populations. According to the F_{ST} values and the structure plot, the Shandong YS was the most distinct of the Chinese Cashmere goats. Notably, a marked deviation of the Hardy-Weinberg proportions was observed for the marker, which may be explained by the following. The observed null alleles that led to a high within-breed global F_{ST} value of 0.1184 for Chinese Cashmere goats in this study was similar to that which was observed for Chinese goats (0.0629 to 0.1632) (Li et al., 2004; Di et al., 2011), higher than that observed for European and Middle Eastern goats (0.069) (Canon et al., 2006), and lower than that observed for Southeast Asian goats (0.143) (Barker et al., 2001). The information offered by G_{ST} in the present study allowed for a better differentiation among populations, and this was especially true for the Cashmere goat populations. The observed G_{ST} for Cashmere goats showed a high within breed genetic variability characterized by the lowest within breed. To determine whether the degree of scatter shown in the scatter plots increased with geographic distance and whether the populations were in drift-gene flow equilibrium, we correlated the residuals from the F_{ST} linear regression with geographic distance. Ultimately, these parameters do not allow gene flow patterns among populations to be ascertained. It has been suggested that the typical high within population variability of microsatellites may result in a low magnitude of differentiation measures (Hedrick, 1999; Balloux and Lugon-Moulin, 2002). This provides a useful context for the F_{IT} vales from this study: 0.0627 to 0.2534, as well as those of F_{IS} : 0.088 to 0.1078. In addition, the most commonly used estimators of gene flow, such as $N_m = 0.25(1-F_{ST})/F_{ST}$, (1.0171 to 3.7246 for this study), are derived on the basis of simplified models of population structure that assume constant population sizes, symmetrical migration at constant rates, and population persistence for periods long enough for achievement of genetic equilibrium (Wright, 1969). These shortcomings highlight the need for the application of new, more informative methodologies in an effort to effectively ascertain the evolutionary history of present-day populations, both for long term gene flow and for the recent migration patterns (Wilson and Rannala, 2003). Here, we further emphasize the need to apply more informative methodologies to ascertain the evolutionary history of current populations. In addition, we present methodologies for the estimation of recent migration patterns to furnish complementary information, thus allowing recent introgression processes to be ascertained.

The observed genetic relationships among the investigated populations have been illustrated using the NJ topology tree derived from the Nei (1978) standard genetic distance. Although the NJ topology tree is not well supported by the nodes, the dendrogram (Figures 2 and 3) shows a clear separation of the Chinese local Cashmere goat populations from different geographic locations. Similar results were observed in the phylogenetic trees of neighbour-joining dendrogram analysis using Nei's genetic

distances (D_A) and Nei's standard genetic distances (D_S), which indicated that the phylogenetic relationships among the nine Cashmere goat populations were not closely linearly correlated with their geographic distribution. This result was in agreement with the finding of previous studies, which showed that these populations were originally different, but shared the same genetic material because of natural and artificial selection, as well as their ecological habitat (Li et al., 2002; Chen et al., 2006; Kang et al., 2011). From the perspective of conservation genetics, Hedrick and Miller (1992) have indicated that populations should be managed in a way that retains adequate genetic variability, so as to provide for future adaptation and the successful expansion of native and reintroduced free-ranging populations. Because we cannot directly evaluate the biological significance of the genetic differences between locations, and because genetic differences are roughly proportional to geographic distances, the most conservative method of selecting stock for translocation would be to choose the closest available population to preserve local variation and/or potential adaptation (Gutierrez-Espeleta et al., 2000). According to our investigation, the individual exchanges among Cashmere goat breeds with the aim of increasing cashmere quantity (Li et al., 2004; Wang et al., 2008) might have led to a closer relationship among Cashmere goat populations. This study showed significant levels of genetic divergence between the nine Chinese local Cashmere goat populations, and these findings indicate that the genetic process was associated with historical ecogeographic barriers. To maintain the present genetic diversity and structure of these populations, genetic exchanges between populations must be carefully controlled on existing conservation farms.

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