

Asian-Aust. J. Anim. Sci. Vol. 26, No. 2 : 157-162 February 2013 http://dx.doi.org/10.5713/ajas.2012.12459

> www.ajas.info pISSN 1011-2367 eISSN 1976-5517

Phylogenetic Analysis of a Privately-owned Korean Native Chicken Population Using mtDNA D-loop Variations

M. R. Hoque, N. R. Choi, H. Sultana, B. S. Kang¹, K. N. Heo¹, S. K. Hong², C. Jo and J. H. Lee*

Department of Animal Science and Biotechnology, Chungnam National University, Daejeon 305-764, Korea

ABSTRACT: The use of Korean native chicken is increasing, and the discovery of new genetic resources is very important from both economic and genetic conservation points of view. In this study, mtDNA D-loop sequences from 272 privately-owned Korean native chickens from a Hyunin farm were investigated. Seventeen nucleotide substitutions were identified from the sequence analysis and they were classified as 6 haplotypes. Previously investigated haplotypes in five Korean native chicken populations have been compared with the Hyunin chicken population. The results indicated that two haplotypes, H10 and H15, in the Hyunin chicken population were not previously identified in other Korean native chicken populations, representing 33.09% (90/272) and 1.1% (3/272) of the Hyunin population, respectively. On the other hand, four other haplotypes were identical to those of a previous study of Korean native chicken populations. This result is indicative of conservation strategies of Hyunin chicken populations for expanding the genetic diversity in the Korean native chicken population. (**Key Words:** D-loop Variation, Haplotype, Korean Native Chicken, mtDNA)

INTRODUCTION

Increased research and genetic profiling of native species is an important component of modern perspectives of genetic diversity conservation. There is also an increasing demand for native chicken meat currently, even though 90% of chicken consumed in Korea is from imported breeds (MIFAFF, 2009). Five lines of Korean native chicken populations were developed in the 1990's, and this breeding was mainly controlled by a national institution called National Institute of Animal Science (NIAS) in Korea. However, there are still native chicken breeds available in the private farms, and one of them, the Hyunin population, is used in this study for comparison with the primary Korean native chicken populations for which we have previous genetic information.

Maternally-inherited mitochondrial DNA (mtDNA) sequences have been successfully used for genetic diversity studies of avian and domestic animals (Komiyama et al., 2003, 2004; Liu et al., 2006; Odahara et al., 2006; Sasazaki et al., 2006; Lei et al., 2007; Wang et al., 2007; Li et al.,

² Hyunin Farm, Paju 413-863, Korea.

Submitted Aug. 27, 2012; Accepted Oct. 6, 2012; Revised Oct. 29, 2012

2008). In the mitochondrial genome, coding genes are predominantly used for biodiversity studies of many different species, whereas, the non-coding control region is used for the inter-specific population studies (Baker and Marshall, 1997; Moore and Defilippis, 1997). MtDNA has the advantages of the absence of recombination and rapid nucleotide substitutions compared with genomic DNA sequences (Aquadro and Greenberg, 1983; Lansman et al., 1983; Cann et al., 1984). Chicken D-loop sequences were previously determined to investigate the origin of breeds or species, and various haplotypes were reported (Komiyama et al., 2003, 2004; Liu et al., 2006; Oka et al., 2007). Previously, three SNPs of D-loop region were found to have significant discrimination power for two commercial native chicken populations in Korea (Hoque et al., 2011). Also, the D-loop hypervariable region of mtDNA was used to determine the relationships between Korean native chickens and other chicken breeds (Hoque et al., 2009).

In this study, the chicken mtDNA D-loop region was used for the demarcation of haplotypes in the privatelyowned Hyunin native chicken population, and was compared with those of other Korean native chicken breeds, which were previously developed in NIAS. This Hyunin population has distinct feather colors and they were classified as eleven colors. Some of these (red, yellow, black, gray and white) are the same colors identified in Korean native chickens at NIAS. These results provide a

Copyright © 2013 by Asian-Australasian Journal of Animal Sciences

^{*} Corresponding Author: Jun-Heon Lee. Tel: +82-42-821-5779, Fax: +82-42-825-9754, E-mail: junheon@cnu.ac.kr

¹ Poultry Science Division, National Institute of Animal Science, RDA, Cheonan 331-801, Korea.

basis for an appropriate conservation breeding program for the Korean native chicken.

MATERIALS AND METHODS

Sampling and DNA extraction

A total of 272 native chicken samples were collected from a single flock in a Hyunin chicken farm. Genomic DNAs were extracted from blood samples using a manual extraction method (Miller et al., 1998) applied to a large volume, which was stored at -20°C until use. For the comparison of haplotypes, previous D-loop sequence data from two commercial chicken populations Hanhyup (40 samples) and Yellim (37 samples) and three Korean native chicken breeds (88 samples) were included in the analysis (Hoque et al., 2011).

PCR amplification and sequencing

PCR was used to amplify a 591 bp fragment in the D-loop hypervariable region in mtDNA by the following primer pair (Forward: 5'- AGGACTACGGCTTGAAAAGC -3' and Reverse: 5'-ATGTGCCTGACCGAGGAACCAG -3'). The PCR reactions included approximately 100 ng of genomic DNA, 2.5 µl of 10× buffer (Tris-HCl (pH 9.0), PCR enhancers, (NH4)₂SO₄, 20 mM MgCl₂), 2.0 µl of 10 mM dNTPs mixture (2.5 mM each of dATP, dCTP, dGTP and dTTP), 1 µl of 10 pM of each primer and 1 U HS Prime Taq (GeNet Bio, Korea) in a 25 µl reaction volume. PCR was performed in a My-Genie96 Thermal Block (Bioneer) with an initial denaturation step at 94°C for 10 min, followed by 35 cycles of 30 s at 94°C, 30 s at 61°C, 40 s at 72°C and a final extension step at 72°C for 10 min. The PCR products for mtDNA were electrophoresed on 1.5% agarose gels with ethidium bromide, and DNA bands were visualized under ultraviolet light. Purification of PCR products was performed using an Accuprep[®]PCR purification kit (Bioneer) according to the manufacturer's instructions. Purified PCR products were also confirmed using agarose gels for sequencing. All the purified PCR products were sequenced by Genotech (www.genotech.co.kr) commercial company.

Analysis

The chicken mtDNA D-loop nucleotide sequence data were aligned using the ClustalW program (Thompson et al., 1994). Nucleotide replacement export data from mtDNA were carried out in haplotype sequences by using MEGA5 (Tamura et al., 2011). The number of haplotypes, nucleotide variable site, haplotype diversity and nucleotide diversity (Nei, 1982) were calculated using DnaSP V.5.10 (Rozaset al., 2003). A rectangular Kimura 2-parameter model neighbor-joining (NJ) phylogenetic tree with 1000 bootstrap replications was constructed using MEGA5. Also, an outline for median-joining network profiles with positional SNPs for the differentiation of haplotypes by using NETWORK4610 software (Bandelt et al., 1999).

RESULTS AND DISCUSSION

Genetic diversity and haplotype analysis

Mitochondrial D-loop sequence variations were used to calculate the number of variable sites (S), number of haplotypes (h), haplotype diversity (Hd) and nucleotide diversity (Pi) among the chicken populations (Table 1). Analysis of 272 mtDNA D-loop sequence data from the Hyunin population revealed six haplotypes, which were comprised of seventeen nucleotide substitutions. All of the six mtDNA haplotypes for the Hyunin chicken population that were discovered are shown in Table 2. Based on the statistical analysis, the identified Hd and Pi were 0.768 and 0.0083, respectively, in Hyunin chickens. Based on the previous analysis of 40 sequences from Hanhyup commercial chicken population, 15 variable sites were identified which have 5 haplotypes with 0.778 of Hd and 0.0068 of Pi. Interestingly, 37 mtDNA D-loop sequences were conserved in the Yellim commercial chicken population (Hoque et al., 2011). Moreover, three Korean native chicken breeds have been compared with the three Korean native chicken populations, dominated by black, red, and yellow, respectively. The black Korean native chicken (KB) breed was observed to be highly variable in terms of S, h, Hd and Pi for 22, 9, 0.823 and 0.0113, respectively. Red Korean native chicken (KR) breeds also had a high nucleotide substitution of 20, and 6 haplotypes which used

Table 1. The calculated number of variable sites (S), number of haplotype (h), haplotype diversity (Hd) and nucleotide diversity (Pi) using mitochondrial D-loop sequence polymorphisms for the Korean native chicken populations

	· 1		1 1		
Breed/population	Sample no.	S	h	Hd	Pi
Hyunin (Hn)	272	17	6	0.768	0.0083
Hanhyup (Hf)	40	15	5	0.778	0.0068
Yellim (Yf)	37	0	1	0	0
Black Korean native chicken (KB)	28	22	9	0.823	0.0113
Red Korean native chicken (KR)	30	20	6	0.662	0.0093
Yellow Korean native chicken (KY)	30	10	4	0.591	0.0077

S = Number of variable sites, h = Number of haplotype, Hd = Haplotype diversity, Pi = Nucleotide diversity.

Except for the sequences from Hyunin, the sequence data were obtained from Hoque et al. (2011).

Hoque et al. (2013) Asian-Aust. J. Anim. Sci. 26:157-162

		Nucleotide substitution ²																	
Haplotype	1	1	2	2	2	2	2	2	2	2	2	3	3	3	3	3	4	- 61	
	6	9	1	1	1	2	4	4	5	6	9	1	1	1	6	7	4	Sample no.	
	7	9	0	2	7	5	3	6	6	1	1	0	3	5	7	0	6		
Hnhap1	С	Т	С	G	Т	Т	Т	С	Т	С	G	С	Т	С	С	С	С	29	
Hnhap2		С																51	
Hnhap3			Т								А		С		Т	Т		90	
Hnhap4											А		С		Т	Т		72	
Hnhap5	Т				С	С	С		С	Т	А	Т	С		Т	Т	Т	27	
Hnhap6	Т			А		С	С	Т			А		С	Т	Т	Т		3	
Total																		272	
D ()			1			1 .	** 1												

Table 2. The identified haplotypes with the mitochondrial D-loop sequence polymorphisms in Hyunin chicken

Dot (.) represents the identical nucleotide with the haplotype Hnhap1.

to calculate a Hd value of 0.662 and a Pi value of 0.0093. On the other hand, the yellow Korean native chicken (KY) breed had a comparatively low number of variable sites of 10 and 4 haplotypes, from which a Hd and Pi of 0.591 and 0.0077 were calculated, respectively.

A total of 28 nucleotide substitutions were observed among the chicken populations which represent 15 haplotypes (Table 3). Among these, 6 haplotypes were identified in the Hyunin population. Of these, two haplotypes were unique to the Hyunin population. On the

Table 3. The identified haplotypes with the mitochondrial D-loop sequence polymorphisms among the Korean native chicken populations

Within												N	ucleo	tide s	subst	itutio	ns												sample no.	Haplotype
population	1	1	1	2	2	2	2	2	2	2	2	2	2	2	2	2	2	3	3	3	3	3	3	3	3	3	3	4		
napiotype	6	7	9	1	1	1	2	2	3	4	4	4	5	6	6	8	9	0	1	1	1	3	4	6	6	7	9	4		
	7	7	9	0	2	7	2	5	9	2	3	6	6	1	5	1	1	6	0	3	5	0	2	3	7	0	1	6		
KBhap1	Т	Α	С	С	G	С	Α	С	Α	G	С	С	С	Т	С	Α	Α	Т	Т	С	С	С	Α	С	Т	Т	С	Т	11	H1
KYhap1																													7	
Hfhap1																													13	
Hnhap5			Т																										27	H2
KYhap3			Т																										3	
KRhap1			Т																										2	
KBhap8			Т																										1	
Hfhap2			Т																								А		10	H3
KBhap5			Т																			Т							2	H4
Hfhap4			Т																			Т							8	
KBhap9			Т				G															Т							1	H5
KBhap3		Т	Т			Т									Т	G		С					G	Т				С	4	H6
KRhap5		Т	Т			Т									Т	G		С					G	Т				С	3	
KBhap6			Т			Т				Α						G								Т	С			C	2	H7
KRhap6			Т			Т				А						G								Т	С			C	2	
Hfhap5			Т			Т				А						G								Т	С			C	2	
KBhap7			Т			Т				А				С		G							G	т	C			С	2	H8
KBhap2	Ċ		Т			Т		Т			Т		Т	Č					Ċ									Č	3	H9
KRhan3	Č		Т			Т		Т			Т		Т	Č					Č									Č	17	
Hnhap4	Č		Т			Т		Т			Т		Т	Č					Č									Č	72	
Hfhap3	Č		Т			Т		Т			Т		Т	Č					Č	ż					÷			Ĉ	7	
Yfhap1	C		Т			Т		Т			т		Т	C					C									С	37	
Hnhap3	C		Т	Ť		Т		Т			т		Т	č					c									C	90	H10
KBhap4	Č					Т		Т			Т		Т	Č			G		Č	т					Ċ	Ċ		Č	2	H11
KRhan2	Ċ	-	-		-	т	-	т	-		т	-	т	Ċ	-		G	-	Ċ	т	-	-	-	-	Ċ	Ĉ	-	Ĉ	3	
Hnhan2	Ċ	-	-		-	т	-	т	-		т	-	т	Ċ	-		G	-	Ċ	т	-	-	-	-	Ċ	Ĉ	-	Ĉ	51	
KRhan4	C	•	Т	•	•	т	•	т	•	•	т		т	c		•	G		c	т	•	•	•		c	C	•	C	3	H12
Hnhan1	c	•	т	•	•	т	•	т	•	•	т	•	т	c	•	•	G	•	C	т	•	•	•	•	c	c	•	c	29	1112
KYhan?	c	•	т	•	•	т	•	т	Ġ	•	т	•	т	c	•	•	0	•	c	1	•	•	•	•	C	C	•	c	18	H13
KVhan4	c	•	т	•	•	т	•	т	G	•	т	•	т	c	•	•	•	•	C	•	•	•	•	•	•	•	•	c	2	H14
Hnhan6	C	•	т	•	Δ	т	•	1	U	•	1	Т	т	C	•	•	•	•	C	•	Т	•	•	•	•	•	•	c	2	H15
Total	•	•	1	•	11	1	•	•	•	•	•	1	1	2	. 8	•	•	•	C	•	1	•	•	•	•	•	•	C	437	15

KBhap = Black Korean native chicken haplotype; KRhap = Red Korean native chicken haplotype; KYhap = Yellow Korean native chicken haplotype; Hfhap = Hanhyup chicken haplotype; Yfhap = Yellim chicken haplotype; Hnhap = Hyunin chicken haplotype.

Dot (.) represents the identical nucleotide with the upper haplotype.

other hand, four haplotypes were mixed with other Korean chicken populations. Of these, four haplotypes (Hnhap5, Hnhap4, Hnhap2 and Hnhap1) were highly related with the KR breed (25 samples) which represents H2, H9, H11 and H12, respectively. In the case of the KB breed, only six chickens shared three Hyunin haplotypes (Hnhap5, Hnhap4 and Hnhap2) which are the H2, H9 and H11, respectively. Only one haplotype (Hnhap5) was shared with the KY breed (3 samples). Based on the haplotype sharing among the Korean native chicken breeds, the Hyunin chicken population is very closely related to the KY chicken breed. Two chicken populations (Hanhyup and Yellim) had Hnhap4 representing the H9 haplotype. On the other hand, specific haplotypes for Hyunin population have contained 33.09% of Hnhap3 (90 samples) and 1.1% of Hnhap6 (3 samples) which represent H10 and H15 haplotypes, respectively. The H10 haplotype, containing the D-loop nucleotide substitution C210T, is unique to the Hyunin chicken breed. Also, three SNP positions, G212A, C246T and C315T, in the D-loop region are specific for H15 discrimination of the Hyunin breed. Previously, we successfully differentiated two commercial chicken populations by the selected three D-loop SNPs at positions C225T, A239G and C243T (Hoque et al., 2011). Also, it was demonstrated that allele-specific SNP typing in mtDNA D-loop region at the five SNP sites (C310T, A342G, C446T, A686G and C1213T) were SNP haplotypes that can be used for identification of the origins of chicken meat (Harumi et al., 2011). The D-loop region in mtDNA may also play a vital role in the measurement of diversity, and in evaluating the population structure of native Korean chickens.

Network profiling and phylogenetic analysis

The estimation of network profiles was traced using 15

haplotypes in the D-loop nucleotide positions of native chicken populations (Figure 1). In our studies, haplotype H10 was found to specifically represent the Hyunin population that differentiated H9 by the C210T SNP. Also, the H15 haplotype for the Hyunin population is distinguished from other haplotypes by the three nucleotide positions at 212, 246, and 315. Moreover, the H9 Hyunin Haplotype is differentiated with H12 by the four nucleotide positions at 291, 313, 367 and 370. Besides, the H12 haplotype is differed from the H11 haplotype by 199 nucleotide position for the Hyunin haplotype. Only haplotype H2 is markedly different from other Hyunin haplotypes. These specific SNPs observed in the D-loop region will provide valuable markers for the discrimination of Korean native chicken breeds.

The phylogenetic analysis among the chicken populations was conducted for identifying relationships between Hyunin chicken populations and previously identified Korean native chicken breeds (Figure 2). In our phylogenetic haplotypes were analysis, clearly differentiated among the two groups. Five haplotypes for the Hyunin population were clustered together in one clade (lower clade in Figure 2) and only one haplotype was contained in another clade (upper clade in Figure 2), while, three haplotypes in the lower clade were mostly related with the KR breed. The internal Hyunin haplotype Hnhap4 is also related with two commercial chicken populations and the KB haplotype. In addition, Hnhap2 contains the KB haplotype. Interestingly, Hnhap3 and Hnhap6 are completely separated from other haplotypes, which was determined through network profile analysis. On the other hand, Hnhap5 in the upper clade is closely related with KR and KB haplotypes. Our phylogenetic analyses also gives some idea as to the relationship among Korean native



Figure 1. Network profiles of the 15 haplotypes from the six Korean native chicken populations. The links are labeled by the nucleotide positions. The order of the mutations on the branch is arbitrary. The median vectors (mv) are nucleotide junctions.



Figure 2. The neighbor-joining phylogenetic tree using Kimura 2-parameter model with 1000 bootstrap replications in six Korean native chicken populations.

chicken populations.

In this study, we investigated the D-loop SNPs and haplotypes for the identification of Korean native chicken populations in a Hyunin private farm. Two previously unidentified haplotypes were investigated, and the phylogenetic relationships were observed by comparing them with haplotypes of other chicken breeds. The results indicate that the private farms still hold valuable genetic resources which had not been previously identified. To maintain this valuable native chicken population, an appropriate conservation breeding program is ultimately required, and should be conducted with the aid of molecular genetic techniques.

ACKNOWLEDGEMENTS

This study was carried out with the support of "FTA

Agriculture Research Project (Project No. PJ9070112011)", RDA, Republic of Korea.

REFERENCES

- Aquadro, C. F. and B. D. Greenberg. 1983. Human mitochondrial DNA variation and evolution: Analysis of nucleotide sequences from seven individuals. Genetics 103:287-312.
- Baker, A. J. and H. D. Marshall. 1997. Mitochondrial control region sequences as tools for understanding evolution. In: Avian molecular evolution and systematic (Ed. D. P. Mindell). Academic press, San Diego. 51-82.
- Bandelt, H. J., P. Forster and A. Rohl. 1999. Median-joining networks for inferring intraspecific phylogenies. Mol. Biol. Evol. 16:37-48.
- Cann, R. L., W. M. Brown and A. C. Wilson. 1984. Polymorphic sites and the mechanism of evolution in human mitochondrial DNA. Genetics 106:479-499.

- Harumi, T., A. Sano, T. Minematsu and M. Naito. 2011. Allelespecific PCR typing and sequencing of the mitochondrial Dloop region in four layer breeds. Anim. Sci. J. 82:223-226.
- Hoque, M. R., K. C. Jung, B. K. Park, K. D. Choi and J. H. Lee. 2009. Genetic variability of mtDNA D-loop region in Korean native chickens. Korean J. Poult. Sci. 37:323-328.
- Hoque, M. R., S. H. Lee, K. C. Jung, B. S. Kang, M. N. Park, H. K. Lim, K. D. Choi and J. H. Lee. 2011. Discrimination of Korean native chicken populations using SNPs from mtDNA and MHC polymorphisms. Asian-Aust. J. Anim. Sci. 24:1637-1643.
- Komiyama, T., K. Ikeo and T. Gojobori. 2003. Where is the origin of the Japanese gamecock? Gene 317:195-202.
- Komiyama, T., K. Ikeo and T. Gojobori. 2004. The evolutionary origin of long-crowing chicken: its evolutionary relationship with fighting cocks disclosed by the mtDNA sequence analysis. Gene 333:91-99.
- Lansman, R. A., J. C. Avise and M. D. Huettel. 1983. Critical experimental test of the possibility of 'paternal leakage' of mitochondrial DNA. Proc. Natl. Acad. Sci. USA. 80:1969-1971.
- Lei, C. Z., W. Zhang, H. Chen, F. Lu, Q. L. Ge, R. Y. Liu, R. H. Dang, Y. Y. Yao, L. B. Yao, Z. F. Lu and Z. L. Zhao. 2007. Two maternal lineages revealed by mitochondrial DNA dloop sequences in Chinese native water buffaloes (*Bubalus bubalis*). Asian-Aust. J. Anim. Sci. 20:471-476.
- Li, J. L., Y. Shi, C. Fan and D. Manglai. 2008. mtDNA diversity and origin of Chinese Mongolian horses. Asian-Aust. J. Anim. Sci. 21:1696-1702.
- Liu, Y. P., G S. Wu, Y. G. Yao, Y. W. Miao, G. Luikart, M. Baig, A. B. Pereira, Z. L. Ding, M. G. Palanichamy and Y. P. Zhang. 2006. Multiple maternal origins of chickens: Out of the Asian jungles. Mol. Phylogenet. Evol. 38:12-19.
- MIFAFF. 2009. Primary statistics of food. Agriculture, Forestry and Fisheries, Korea.

- Miller, S. A., D. D. Dykes and H. F. Polesky. 1988. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res. 16:1215.
- Moore, W. S. and V. R. Defilippis. 1997. The window of taxonomic resolution for phylogenies based on mitochondrial cytochrome b. In: Avian molecular evolution and systematic (Ed. D. P. Mindell). Academic press, San Diego. 84-119.
- Nei, M. 1982. Evolution of human races at the gene level. Prog. Clin. Biol. Res. 167-181.
- Odahara, S., H. J. Chung, S. H. Choi, S. L. Yu, S. Sasazaki, H. Mannan, C. S. Park and J. H. Lee. 2006. Mitochondrial DNA diversity of Korean native goats. Asian-Aust. J. Anim. Sci. 19: 482-485.
- Oka, T., Y. Ino, K. Nomura, S. Kawashima, T. Kuwayama, H. Hanada, T. Amano, M. Takada, N. Takahata, Y. Hayashi and F. Akishinonomiya. 2007. Analysis of mtDNA sequences shows Japanese native chickens have multiple origins. Anim. Genet. 38:287-293.
- Rozas, J., J. C. Sanchez-DelBarrio and X. Messeguer. 2003. DnaSP, DNA polymorphism analyses by the coalescent and other methods. Bioinformatics 19:2496-2497.
- Sasazaki, S., S. Odahara, C. Hiura, F. Mukai and H. Mannen. 2006. Mitochondrial DNA variation and genetic relationships in Japanese and Korean cattle. Asian-Aust. J. Anim. Sci. 19: 1394-1398.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei and S. Kumar. 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol. Biol. Evol. 28:2731-2739.
- Thompson, J. D., D. G. Higgins, T. J. Gibson and W. Clustal. 1994. Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res. 22: 4673-4680.
- Wang, X., H. Chen and C. Z. Lei. 2007. Genetic diversity and phylogenetic analysis of the mtDNA D-loop region in Tibetan sheep. Asian-Aust. J. Anim. Sci. 20:313-315.