Research Article Open Access

Leaf Rot and Leaf Ring Spot Caused by Rhizoctonia solani in Chinese Cabbage

Chang-Ki Shim, Min-Jeong Kim*, Yong-Ki Kim, Hyeong-Jin Jee, Sung-Jun Hong, Jong-Ho Park, Eun-Jung Han and Jong-Chul Yun

Organic Agriculture Division, National Academy of Agricultural Science, Rural Development Administration, Suwon 441-707, Korea

(Received on November 15, 2013; Revised on November 25, 2013; Accepted on November 26, 2013)

This study was conducted to determine the occurrence of leaf rot and leaf ring spot, caused by *Rhizoctonia solani* in Chinese cabbage under seedling nursery and cultivation greenhouses. Symptoms of leaf rot and leaf ring spot were found in three Chinese cabbage cultivars, *Brassica campestris* subsp. *pekinensis*, 'Ryeokgwang', 'Daetong', and 'CR mat'. In Hwacheon, the disease incidence was 73.8% in the seedling stage of the Chinese cabbage. In Icheon, the symptoms were observed on the upper leaves of the Chinese cabbage cultivar, 'Norangmini' with 20.5% of disease incidence. The symptoms appeared as primary lesions consisting of small, circular necrotic ring spots with gray color, 1.4–3.0 mm in diameter, accompanied by secondary rot lesions with large irregular borders of leaves. The color of mycelial mat of 20 isolates was dark brown and light brown. The average hyphal diameter of all the isolates was within 5.01–11.12 µm. Among the 20 strains isolated from Chinese cabbage, 16 isolates and four isolates anastomosed with the AG-1 (IB) and AG-1 (IC), respectively. Twenty isolates tested were only virulent on foliage parts of Chinese cabbage leaves but were avirulent on stem parts of the plants. Based on the mycological characteristics and pathogenicity test on host plants, the fungus was identified as *Rhizoctonia solani*.

Keywords : Anastomosis group, Bottom rot, Brassica campestris subsp. pekinensis, Rhizoctonia solani

Introduction

Chinese cabbage (*Brassica rapa* subsp. *pekinensis*) is one of the economically important vegetable crops in eastern Asia including Korea. It is usually harvested after completing the leaf heading period. Chinese cabbages as the major component in the preparation of Kimchi, a traditional fermented food and contain a majority of the components essential for human nutrition in Korea (Hong and Kim, 2011).

Bottom rot is a common disease caused by *Rhizoctonia* solani Kühn [telomorph: *Thanatephorus cucumeris* (Frank) Donk] in Chinese cabbage cultivation areas (Carling, 1996). Kim *et al.* (1995) reported that *R. solani* is one of the 65 diseases causing have to a wide range of plants including rice, potato, pepper, pine tree, etc. in Korea. Kang and Kim (1986) first reported 30% of overall bottom rot infection rate in Chinese cabbage under plastic film house cultivation in Gyeongnam province was caused by *R. solani*. The disease severely

*Corresponding author Phone) +82-31-290-0545, Fax) +82-31-290-0507 Email) kjs0308@korea.kr occurred in poly vinyl mulching fields during the wet and cold seasons. The symptoms were bottom rot of leaves, root rot and damping off of seedlings during cultivation. *R. solani* is one of the potent soilborne pathogens of economically important crops which develop both in cultured and non-cultured soils, causing diseases in different crops such as rice, bean and tomato, among others (Sneh and Akira, 1991). Numerous *Rhizoctonia* species have been reported to occur on many hosts in Korea (Kim *et al.*, 1993; Kim, 1996).

The pathogen shows considerable diversity in morphology, geographic location, host specificity and pathogenicity (Ogoshi, 1987). Isolates of *R. slolani* have been classified into fourteen anastomosis group (AG), AG-1 through AG-13 and AG-BI (bridging isolates) by different in pathogenicity (Carling, 1996; Ogoshi, 1987). Among the fourteen anastomosis group, AG-1 is subgrouped as AG-1 (1A), AG-1 (1B), and AG-1 (1C) and AG-2-2 (III B). The best known subsets of AG-2 are AG-2-1, AG-2-2 (IIIB), AG-2-2 (IV), AG-2-2 (LP), and AG-2-3 (Hyakumachi and Sumino, 1984; Watanabe and Matsuda, 1966).

R. solani species and anastomosis groups are reported to differ in sensitivity to common fungicides (Campion

et al., 2003). Anastomosis occurs between fungal isolates of the same AG but not between isolates of different AG's. The concept of anastomosis group is a widely accepted principle for identifying intraspecific groups in the R. solani complex and providing genetic relations for the breeding of new varieties (Carling et al., 1988). AG-1 isolates have been subdivided into four subgroups, AG-1 (IA), AG-1 (IB), AG-1 (IC) and AG-1 (ID) based on pathogenicity and culture morphology (Priyatojo et al., 2001). Currently, two intraspecific groups of R. solani AG-1 comprise the RFB (Rhizoctonia foliar blight) complex: intraspecific group A (IA), the causal agent of sheath blight of rice which causes aerial blight, intraspecific group B (IB), which causes web blight of soybean, and intraspecific group C (IC), which causes damping off (Joy et al., 1990; Sneh et al., 1991; Yang et al., 1990).

However, anastomosis group concept is not an ideal method for classification of *R. solani* as misidentification is caused from the varied frequency of hypal fusion in some AG (Liu and Sinclair, 1992). Recently, molecular biological techniques have been used in combination with morphological and physiological markers for the analysis of population (Guleria *et al.*, 2007; Jeon *et al.*, 2010) and the detection of genetic variability among the isolates of *R. solani* (Hong *et al.*, 1998; Lee *et al.*, 2005)

This paper will discuss the incidence and the causal organism of leaf rot and leaf ring spot diseases of Chinese cabbage under the organic Chinese cabbage seedling nursery and cultivation greenhouses in Hwacheon and Icheon.

Materials and Methods

Disease incidence. The study was conducted in 2011 at the three seedling nursery greenhouses in Hwacheon and ten cultivation greenhouses in Icheon. Chinese cabbage with leaf rot and leaf ring spot symptoms were visually determined and recorded based on the percentage of diseased plants in 20 replicates of each greenhouse one week before transplanting and harvesting

time, respectively, in the seedling nursery and cultivation greenhouses.

Isolation and purification. *R. solani* isolates were taken from symptomatic plants of Chinese cabbage collected from Icheon and Hwacheon farms in Korea. Eight pieces (1 cm^2) were excised from the margin of healthy and diseased tissues of Chinese cabbage that showed symptoms of *R. solani*. The excised species were surface-sterilized in 1.5% NaOCl for two minutes and extensively rinsed in sterile distilled water. Four pieces per plant were placed on potato dextrose agar (PDA, Difco), all plates were incubated at 25°C. Several isolates of *R. solani* were obtained by single hyphal tip isolation. The isolated *R. solani* were stored at 5°C in PDA slants.

Mycological characterization. The isolates were identified to genus level, based on microscopic and cultural characteristics. From the colony morphology of 20 isolates, color of mycelial mat was recorded after 10 days incubation on PDA. Three weeks after inoculation on PDA, sclerotia production was evaluated at 20°C. Vegetable growth of the 20 isolates of *R. solani* was observed at seven temperatures levels, 5, 10, 15, 20, 25, 30 and 35°C. Mycelial growth was determined in petri dish containing 15 ml of PDA inoculated with a 6 mm diameter of mycelial disk cut from a 5-day-old culture of 20 isolates on PDA. Mycelial growth in four replicated cultures at each temperature was measured at 24 hr intervals.

Determination of anastomosis groups. The standard isolates of the eight anastomosis groups, AG-1 to AG-4, and unidentified two strains, *R. cerealis*, and *R. edophytica* were obtained from the Korean Agricultural Culture Collection (KACC) (Table 1). The determination of the anastomosis group of the isolates was determined by using the methodology of Kim *et al.* (1993). Twenty isolates of *R. solani* were paired with the ten standard isolates of anastomosis groups of *R. solani*. Tester strains of multinucleate *Rhizoctonia* including AG-1 (IA), AG-1 (IB), AG-1 (IC), AG-2-I, AG-2-2 (IIIB), AG-2-2 (IV), AG-3, AG-4 and unidentified two strains, *R. cerealis*, and *R. edophytica* were used. Single 5-mm-diameter

 Table 1. Standard isolates of anastomosis groups (AG), Rhizoctonia solani AG-1 to AG-4, R. cerealis, and R. edophytica obtained from the Korean Agricultural Culture Collection (KACC)

KACC No.	Species	Host	KACC No.	Species	Host
40101	R. solani AG-1 (IA)	Oryza sativa	40131	R. solani AG-2-2 (IV)	Daucus carota
40108	R. solani AG-1 (IB)	Lactuca sativa	40136	R. solani AG-3	Solanum tubersoum
40113	R. solani AG-1 (IC)	Brassica campestris spp. pekinensis	40139	R. solani AG-4	Raphanus sativus
40119	R. solani AG-2-1	Brassica campestris spp. pekinensis	40154	R. cerealis	Agrostis palustris
40127	R. solani AG-2-2 (III-B)	Citrullus lanatus	40713	R. endophytica	Orchid symbiont

agar disc was cut from the perimeter of a 2- to 3-dayold colony of each isolate on PDA. Tester isolates were placed 3 to 4 cm away from each tested isolate and incubated at 25°C for 24 to 48 h in the dark. When the hyphae from the two disks were overlapping, they were stained with safranin O and 3% KOH and examined microscopically to determine anastomosis reaction (Carling, 1996).

Pathogenicity tests. Pathogenicity testing and leaf or root damage assessments of the isolates were carried out based on Kang and Kim (1986). The only modification was that the testing was conducted at 28°C instead of 21°C. Agar-disk assay was used in the pathogenicity tests. From stored cultures, isolates were transferred to the PDA plates and incubated at 25°C in the dark for seven days. Inoculations were made by placing a mycelial-agar disk (6 mm in diameter, taken from a 3-day-old culture on PDA) in the foliage part and stem of each leaf. Twenty isolates of R. solani were tested for pathogenic potential on Chinese cabbage, 'Daetong' with 15 seedlings for each isolate. After inoculations, leaves were covered with a plastic bag for five days. Leaves were similarly prepared, but only PDA disks served as an untreated control. All experiments were replicated twice. Five days after inoculation, the leaf or root damage assessment scale was 0 = no damage, 1 =minor discoloration of leaves, 2 = discoloration plus small necrotic lesions (< 1 mm in diameter) on leaves, 3 = discoloration with large necrotic lesions (1 mm or larger in diameter) on leaves, and 4 = death of the seedling.

Results

Disease incidence. In 2011, the occurrence of bottom rot diseases of Chinese cabbage caused by R. solani were investigated at the Chinese cabbage nursery greenhouses in Hwacheon and the cultivation greenhouses in Icheon. In Hwacheon Chinese cabbage nursery farm, only leaf ring spot symptom was observed on the upper leaves of three Chinese cabbage cultivars after 25 days of sowing with disease incidence of 22.6% for 'Ryeokgwang', 22.6% for 'Daetong' and 20.8% for 'CR mat', respectively (Table 2). At 35 days of seedlings, leaf ring spot and leaf rot symptoms occurred on leaves of three cultivars of Chinese cabbage with disease incidence of 60.1% both for 'Ryeokgwang' and 'Daetong' and 58.9% for 'CR mat' (Table 2). One week before transplanting, leaf rot and leaf ring spot symptoms of three Chinese cabbage cultivars were investigated having 73.8% of disease incidence (Fig. 1).

In Icheon organic Chinese cabbage farm, the leaf

Table 2. Occurrence of leaf rot and leaf ring spot caused by *Rhizoctonia solani* on three cultivars of Chinese cabbage seedlings grown in spring season of 2011

Cultivar	Disease incidence (Mean \pm SD, %)							
Cultival	25 days ^a	35 days	45 days					
Ryeokgwang	22.6 ± 0.05	60.1 ± 0.06	73.8 ± 0.13					
Daetong	19.6 ± 0.03	60.1 ± 0.07	75.4 ± 0.13					
CR mat	20.8 ± 0.08	58.9 ± 0.10	74.2 ± 0.11					

^aDays after sowing of Chinese cabbage seed.

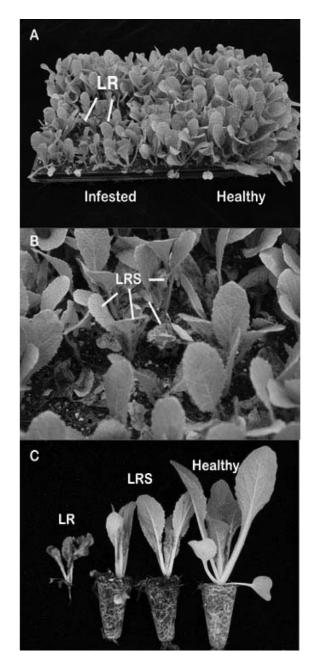


Fig. 1. Symptoms of leaf rot (LR; A, C) and leaf ring spot (LRS; B, C) caused by *Rhizoctonia solani* in Chinese cabbage seedlings in Hwacheon in 2011.

ring spot incidence of 14.1% was observed 15 days after transplanting and leaf rot incidence 3.2% was observed on the upper leaves of 'Norangmini' Chinese cabbage cultivar. At 25 days after transplanting, disease incidence of the leaf ring spot was 19.5% and the leaf rot was 11.5% observed only on the upper leaves. At 35 days after transplanting, the bottom rot symptoms were observed on the stem of the Chinese cabbage with 1.2% disease incidence, counted as 20.5% for leaf ring spot and 19.8% for leaf rot symptoms (Table 3, Fig. 2).

Fungal isolates and characterization. In spring season of 2011, *Rhizoctonia* samples we collected from the main organic Chinese cabbage cultivation areas of

Table 3. Occurrence of leaf rot and leaf ring spot caused by *Rhizoctonia solani* on Chinese cabbage cultivar 'Norangmini' grown in Icheon in 2011

Symptom	Disease incidence (Mean ± SD, %)						
Symptom	Feb. 25	Mar. 7	Mar. 17				
Leaf rot	3.2 ± 0.55	11.5 ± 1.56	19.8 ± 2.03				
Leaf ring spot	14.1 ± 2.23	19.5 ± 2.70	20.5 ± 1.53				
Bottom rot	0.0 ± 0.00	0.0 ± 0.00	1.2 ± 0.15				

Hwacheon and Icheon. Twenty isolates collected and were grown on PDA for 6–7 days.

The colony morphology of these isolates was recorded after 10 days incubation on PDA. The color of mycelial mat of 20 isolates was dark brown except for four isolates, ICC15, ICC16, ICC17 and ICC18 collected from Icheon which was light brown. The average hyphal diameter of all the isolates was within $5.01-11.12 \ \mu m$ which concurs with the previous reported data for *R. solani*.

After four days of growth on PDA, white sclerotial initials were visible and after at eight days, all sclerotia of 20 isolates were dark brown and showed small (0.38–1.28 mm) and uniform of sclerotia formation on PDA. Formation stopped after 9 days, even if the colony had reached profusely at the edge of the petri dish.

The temperature for the twenty tested isolates ranged from 5 to 35°C, with optimum of 25 to 30°C. All of the twenty isolates grew relatively well at 15°C, where all of them did not grow in less than 10°C. Most isolates showed optimum growth at 30°C. No significant (p =0.05) variations were observed in temperature optimum requirements among twenty isolates (Table 5).



Fig. 2. Symptoms of leaf rot (A) and leaf ring spot (B, C) caused by *Rhizoctonia solani* on Chinese cabbage in Icheon in 2011. Microscopic observation shows the mycelia stained with 0.02% cotton blue staining solution on the surface of lesions (D, ×60; E, ×200).

Chang-Ki Shim et al.

	Local region	Anastomosis group									
Isolates		AG-1 (IA)	AG-1 (IB)	AG-1 (IC)	AG-2-1	AG-2-2 (IIIB)	AG-2-2 (IV)	AG-3	AG-4	R. cerealis	R. edophytica
ICC01	Hwacheon	-	_	+	-	-	-	-	-	_	-
ICC02	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC03	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC04	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC05	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC06	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC07	Hwacheon	-	_	+	-	-	-	-	-	_	-
ICC08	Hwacheon	-	_	+	-	-	-	-	-	_	-
ICC09	Hwacheon	-	_	+	-	-	-	-	-	_	-
ICC10	Hwacheon	-	-	+	-	-	-	-	-	-	-
ICC11	Icheon	-	_	+	-	-	-	-	-	_	-
ICC12	Icheon	-	-	+	-	-	-	-	-	-	-
ICC13	Icheon	-	_	+	-	-	-	-	-	_	-
ICC14	Icheon	-	_	+	-	-	-	-	-	_	-
ICC15	Icheon	-	+	-	-	-	-	-	-	_	-
ICC16	Icheon	-	+	-	-	-	-	-	-	_	-
ICC17	Icheon	-	+	-	-	-	-	-	-	-	-
ICC18	Icheon	-	+	-	-	-	-	-	-	-	-
ICC19	Icheon	-	-	+	-	-	-	-	-	-	-
ICC20	Icheon	-	-	+	-	-	-	-	-	-	-

Table 4. Anastomosis group of 20 isolates of Rhizoctonia slolani isolated from spring season Chinese cabbage in Icheon in 2011

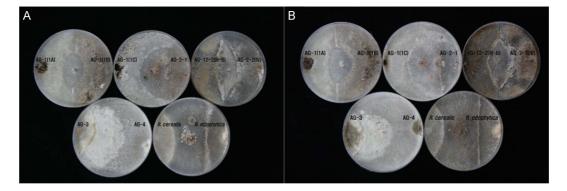


Fig. 3. Determination of anastomosis group (AG) of Hwacheon isolates (A, ICC01) and Icheon (B, ICC11) isolates of *Rhizoctonia solani* from lesions of leaf rot and leaf ring spot symptoms of Chinese cabbage.

Anastomosis Grouping and Pathogenicity Tests. Twenty isolates of *R. solani* obtained from Chinese cabbage leaves found in Hwacheon and Icheon belonged to two anastomosis groups, AG-1 (IB) and AG-1 (IC) (Fig. 3). Ten isolates collected from Hwacheon anastomosed with AG-1 (IB). From Icheon, four isolates were anastomosed with AG-1 (IB) and six isolates anastomosed with AG-1 (IC) (Table 4, Fig. 3).

The pathogenicity of twenty isolates of R. solani was

also investigated on 35-day-old Chinese cabbage seedlings through the biological assay method. All isolates tested were only highly virulent on foliage parts of Chinese cabbage leaves but were avirulent on stem parts. None of the control seedlings developed symptoms. *R. solani* was reisolated from infected seedlings and compared with the original isolate by morphological characteristics. All the isolates had characteristics similar to the original fungus.

Inclator	Colony diameter (mm) of Rhizoctonia solani on potato dextrose agara									
Isolates -	5°C	10°C	15°C	20°C	25°C	30°C	35°C	Mycelial color		
ICC01	0	0	4.3	16.9	28.5	24.4	12.4	dark		
ICC02	0	0	5.1	18	28.9	24.3	13.1	dark		
ICC03	0	0	4.5	16.9	28.8	25	13.1	dark		
ICC04	0	0	4.6	16.9	28.9	24.3	12.9	dark		
ICC05	0	0	4.6	16.9	28.9	25.2	13.1	dark		
ICC06	0	0	5.0	18.2	29.0	25.2	13.3	dark		
ICC07	0	0	4.6	16.9	29.0	25.0	12.4	dark		
ICC08	0	0	4.9	18.1	29.1	24.7	12.4	dark		
ICC09	0	0	4.9	18	28.9	25.1	11.9	dark		
ICC10	0	0	4.9	18.2	28.3	24.6	13.4	dark		
ICC11	0	0	4.8	16.9	28.5	23	12.9	dark		
ICC12	0	0	4.8	17.1	28.2	24.3	12.8	dark		
ICC13	0	0	5.0	18.1	30.0	26.4	14.2	dark		
ICC14	0	0	4.7	16.9	28.5	24.8	12.5	dark		
ICC15	0	0	4.7	16.4	28.5	25	12.5	light		
ICC16	0	0	4.8	18.2	28.4	24.2	13.3	light		
ICC17	0	0	4.5	16.8	28.3	24.4	12.5	light		
ICC18	0	0	4.6	16.5	28.3	25.1	12.5	light		
ICC19	0	0	5.2	17.8	30.1	25.7	13.4	dark		
ICC20	0	0	4.5	16.4	28.7	24.6	12.0	dark		

 Table 5. Morphological characteristics and optimum temperature for mycelia growth of 20 isolates of *Rhizoctonia slolani* isolated from Chinese cabbage in Icheon in 2011

^aValues are the mean mycelial growth of four replicates measured at 7 days.

Discussion

This study identified twenty isolates of *R. solani* obtained from organic Chinese cabbage fields by morphology and anastomosis grouping. *R. solani* Kühn [teleomoph: *T. cucumeris* (Frank) Donk] is a destructive and widely spread fungal pathogen with wide host range including rice, turfgrass, Chinese cabbage, and others vegetable crops. Bottom rot results from infection by *R. solani*, which lives in garden soil. When bottom rot occurs, dark brown, possibly soft and watery, lesions are found at the cabbage base (Kang and Kim, 1986; Keinath, 1995).

The results indicated that only leaf rot and leaf ring spot symptoms were observed on the foliage part of Chinese cabbage of seedlings and plants in Hwacheon seedling nursery and Icheon cultivation greenhouse. Earlier researches showed that most bottom rot pathogen, *R. solani* causal agent is known to infect moist soil. Infection in young plants into the environment due to improper plant growth will severely slow its progress (Kang and Kim 1986; Kim *et al.*, 1993). The other

hand, the twenty tested isolates showed leaf rot and leaf ring spot symptoms and did not infect stem of Chinese cabbage except for bottom rot symptoms at Icheon Chinese cabbage cultivation field with 1.2% disease incidence one week before harvesting.

Unlike many other sclerotia forming plant pathogens, *R. solani* sclerotia only undergo direct myceliogenic germination, whereby vegetative hyphae capable of infecting the host grow directly out of the sclerotium. Mycelia and sclerotia can grow and develop on plant debris, allowing inoculum to survive in the soil as well as on seed from season to season (Dijst, 1988). Genetic variability among the isolates of *R. solani* was reported by Nelson *et al.* (1996). Kaminski and Verma (1985) found a variable response of different *R. solani* isolates at different levels of temperatures. It was found that there were uniforms among the isolates regarding colony diameter and sclerotia formation on PDA with same temperature. It also suggested that there was an existence of homogenous among the 20 isolates of *R. solani*.

Anastomosis groups appear to be fairly host plant specific. The classification of *R. solani* isolates into anastomosis group (AG) is widely accepted as the first way of subgrouping this heterogeneous species into more homogeneous subspecific groups. Although this determination needs experience and probably timeconsuming observations, AG remains a reliable and universal classification within *R. solani* (Carling, 1996).

The present study revealed that twenty isolates of *R. solani* were mostly obtained from foliage part of Chinese cabbage leaves in Hwacheon and Icheon and belonged to two anastomosis groups, AG-1 (IB) and AG-1 (IC). AG-1 (IB) is most commonly associated with species in Fabaceae and Poacea (Tsai, 1970). O'Neill *et al.* (1977) showed that *R. solani* AG-I spreads through the soybean canopy by mycelial growth from infected to adjacent healthy leaves. In Germany, *R. solani* AG-1 (IB) is the causal agent of bottom rot on lettuce (Grosch *et al.*, 2004).

It has been reported that cultural types AG-1 (IA) and (IB) inhabit on aerial parts of plant and AG 2-1 inhabits on ground surface of crops (Watanabe and Matsuda, 1966). In general, AG2-1 was commonly isolated from Chinese cabbage in the field and greenhouse, and showed damping-off and bottom rot symptom on the petiole of the outmost leaves (Kang and Kim, 1986). One important subgroup within the R. solani is AG-1, the causal agent of sheath blight of rice and reported from most rice-growing regions of the world (Dagupta, 1992). In Germany, R. solani AG 1-IB is the causal agent of bottom rot on lettuce, and disease affecting field-grown lettuce (Grosch et al., 2004). AG-1 (IC) is mildly virulent or causes damping-off on variety of hosts (Hyakumachi and Sumino, 1984; Sneh et al., 1991). Some post emergence damping-off occurred and was especially serious for AG-1 and AG2-2, where over 50% of the plants that emerged died by three weeks after planting (Grisham and Anderson, 1983).

It was concluded that on the basis of mycological characteristics and pathogenicity test on host plants, the twenty isolates were identified as *R. solani* AG-1 (IB) and AG-1 (IC).

Acknowledgement

This study was carried out with the support from "Cooperative Research Program for Agricultural Science & Technology Development (Project No. PJ00912702), Rural Development Administration, the Republic of Korea.

References

Band, R. E., Caning, D. E. and Mullinix, B. G. 1966. Charac-

terization and comparison of isolates of *Rhizoctonia solani* AG-7 from Arkansas, Indiana and Japan and select AG-4 isolates. *Plant Dis.* 80: 1421–1424.

- Bolkan, H. A. and Ribeiro, W. R. C. 1985. Anastomosis groups and pathogenicity of *Rhizoctonia solani* isolates from Brazil. *Plant Dis.* 69: 599–601.
- Carling, D. E. 1996. Grouping in *Rhizoctonia solani* by hyphal anastomosis reaction. In: *Rhizoctonia solani*: Taxonomy, Molecular Biology, Ecology, Pathology and Disease Control. ed. by B. Sneh, S. Jabaji-Hare, S. Neate and G. Dijst, pp. 37– 47. Springer Netherlands, Alaska.
- Carling, D. E., Kuninaga, S. and Leiner, R. H. 1988. Relatedness within and among intraspecific groups of *Rhizoctonia solani*: A comparison of grouping by anastomosis and by DNA hybridization. *Phytoparasitica* 16: 209–210.
- Carling, D. E., Pope, E. J., Brainard, K. A. and Carter, D. A. 1999. Characterization of mycorrhizal isolates of *Rhizoctonia solani* from an orchid, including AG-12, a new anastomosis group. *Phytopathology* 89: 942–946.
- Dagupta, M. K. 1992. Rice sheath blight: The challenge continues. In: Plant Disease of International Importance. Disease of Cereal and Pulses. Vol. 1. ed. by B. Sneh, S. Jabaji-Hare, S. Neate and G. Dijst. pp. 37–47. Kluwer Academic, Dordrecht.
- Dijst, G. 1988. Formation of sclerotia by *Rhizoctonia solani* on artificial media and potato tubers. *Neth. J. Plant Pathol.* 94: 233–242.
- Grisham, M. P. and Anderson, N. A. 1983. Pathogenicity and host specificity of Rhizoctonia solani isolated from carrots. *Phytopathology* 73: 1564–1569.
- Grosch, R., Schneider, J. H. M. and Kofoet, A. 2004. Characterization of *Rhizoctonia solani* anastomosis groups causing bottom rot in field grown lettuce in Germany. *Eur. J. Plant Pathol.* 110: 53–62.
- Guleria, S., Aggarwal, R., Thind, T. S. and Sharma, T. R. 2007. Morphological and pathological variability in rice isolates of *Rhizoctonia solani* and molecular analysis of their genetic variability. *Phytopathology* 155: 654–661.
- Hong, S. J. and Kim, Y. G. 2011. Study on the Chinese cabbage producder's using patterns about a new variety of seed. *CNU J. Agri. Sci.* 38: 549–557. (In Korean)
- Hong, S. B., Go, S. J., Ryu, J. C., Kim, W. K. and Kim, I. S. 1998. Differentiation of intraspecific groups within Korean isolates of *Rhizoctonia solani* using PCR-RFLP of ribosomal DNA. *Korean J. Plant Pathol.* 14: 157–163.
- Hyakumachi, M. and Sumino, A. 1984. New morphological type (IC) in *Rhizoctonia solani* AG-1 isolated from the sugar beetmanufactory-waste soils ad some of its characteristics. *Ann. Phytopath. Soc. Jpn* 50: 507–514.
- Jeon, Y. A., Kim, W. G., Kim, D. H., Kwon, S. W. and Hong, S. B. 2010. Taxonomic position of Korean isolates of *Rhizoctonia solani* based on RAPD and ITS sequencing of Ribosomal DNA. *Plant Pathology J.* 26: 83–89.

- Joye, G. F., Berggren, G. T., Jr. and Bemer, D. K. 1990. Effects of row spacing and within-row plant population of Rhizoctonia aerial blight of soybean and soybean yield. *Plant Dis.* 74: 158–160.
- Kaminski, D. A. and Verma, P. R. 1985. Cultural characteristics, virulence and in vitro temperature effect on mycelial growth of *Rhizoctonia* isolates from rape seed. *Can. J. Plant Pathol.* 7: 256–261.
- Kang, S. W. and Kim, H. K. 1986. Incidence and control of bottom rot of Chinese cabbage caused by *Rhizoctonia solani* Kühn. *Korean J. Plant Pathol.* 2: 193–198. (In Korean)
- Keijer, J., Houterman, P. M., Dullemans, A. M. and Korsman, M. G. 1996. Heterogeneity in electrophoretic karyotype within and between anastomosis groups of *Rhizoctonia solani*. *Mycol. Res.* 100: 789–797.
- Keinath, A. P. 1995. Relationships between inoculum density of *Rhizoctonia solani*, wirestem incidence and severity, and growth of cabbage. *Phytopathology* 85: 1487–1492.
- Kim, W. G. 1996. Pathogenicity of anastomosis groups and cultural types of *Rhizoctonia solani* on crops. *Korean J. Plant Pathol.* 12: 21–32. (In Korean)
- Kim, W. G., Cho, W. D. and Lee, Y. H. 1993. Anastomosis groups and pathogenicity of *Rhizoctonia solani* isolates from Chinese cabbage. *Korean J. Plant Pathol.* 9: 200–205. (In Korean)
- Kim, W. G, Cho, W. D. and Ryu, H. Y. 1995. Dignosis and control of Rhizoctonia diseases on crops. *Res. Report RDA*. (In Korean)
- Lee, Y. S., Choi, H. S., Kim, K. S., Woo, S. J., Kang, W. H., Kim, M. J., Shim, J. O. and Lee, M. W. 1998. Analysis of genetic relationships of *Rhizoctonia solani* isolates from various crop species and rapid identification of anastomosis groups with

RAPD method. Korean J. Mycol. 26: 373-379.

- Liu, Z. L. and Sinclair, J. B. 1992. Genetic diversity of *Rhizoctonia solani* anastomosis group 2. *Phytopathology* 82: 778–787.
- Nelson, B., Helms, T., Christianson T. and Rural, I. 1996. Characterization and pathogenicity of *Rhizoctonia* from soybean. *Plant Dis.* 80: 74–80.
- O'Neill, N. R., Rush, M. C., Horn, N. L. and Carver, R. B. 1977. Aerial blight of soybean caused by *Rhizoctonia solani*. *Plant Dis. Rep.* 61: 713–717.
- Priyamojo, A., Escopalao, V. E., Tangonan, N. G., Pascual, C. B., Kageyama, K. and Hyakumachi, M. 2001. Characterization of a new subgroup ID of *Rhizoctonia solani* anastomosis group 1 (AG1-ID), causal agent of anecrotic leaf spot on coffee. *Phytopathology* 91: 1054–1061.
- Sharma M., Gupta, S. K. and Sharma, T. R. 2005. Characterization of variability in *Rhizoctonia solani* by using morphological and molecular markers. *Phytopathology* 153: 449–456.
- Sneh, B., Burpee, L. and Ogoshi, A. 1991. Identification of *Rhizoctonia* Species. APS Press, St. Paul, MN., USA. 133 pp.
- Tsai, W. H. 1970. Studies on the relations between weeds and rice diseases. J. Taiwan Agric. Res. 19: 48–51.
- Yang, X. B., Berggren, G. T., Jr. and Snow, J. P. 1990. Types of Rhizoctonia foliar blight on soybean in Louisiana. *Plant Dis.* 74: 501–504.
- Watanabe, B. and Matsuda, A. 1966. Studies on the grouping of *Rhizoctonia solani* Kühn pathogenic to upland crops. Appointed Experiment (Plant diseases and insects). *Agric. For. Fish. Res. Counc. Ibaraki Agric. Exp. Stn. Jpn. Bull.* 7: 137.