

## Synthesis and Hydroxyl Radicals Scavenging Activity of 2-Pyridine-acetyl-*N*-trimethyl Chitosan Chloride

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**ABSTRACT.** A novel chitosan derivative with double quaternary ammonium salt—2-pyridine-acetyl-*N*-trimethyl chitosan chloride (PATMCS) was synthesized and the antioxidant activity of PATMCS against hydroxyl radicals was assessed. The results indicated that PATMCS had potent hydroxyl scavenging activity. The IC<sub>50</sub> of PATMCS was 0.13 mg/mL. PATMCS showed 100% scavenging effect at a dose of 0.8 mg/mL which markedly better than that of *N*-trimethyl chitosan chloride (TMCS). It was confirmed that quaternary chitosan derivatives showed potent antioxidant activity. PATMCS has double quaternary ammonium salt structure in the molecules. Therefore, the antioxidant activity of PATMCS was better than TMCS. The above results are theoretically fundamental for further development and making use of chitosan resources to prepare new antioxidants.

**Key words:** Double quaternary chitosan, Synthesis, Antioxidant activity

### INTRODUCTION

Chitosan, the deacetylated derivative of chitin, is the second abundant natural polysaccharide found on the earth next to cellulose. As a natural renewable resource, it has attracted people's attention for a number of unique properties, such as antimicrobial activity, nontoxicity, biodegradability, as well as its antioxidant activity.<sup>1-3</sup> The quaternization of chitosan gave rise to an important derivative of chitosan and its bioactivities had been reported.<sup>4,5</sup> Guo's work had proved that the antioxidant activity of chitosan and its derivatives should be related to the different forms of nitrogen atom in the molecules of chitosan, and the QCSs had the best antioxidant activity. The increased activity should be attributed to the positive charge density of the nitrogen atom at C-2 in the molecules of QCSs was strengthened after being quaternized.<sup>6</sup> To further investigate the relationship between antioxidant activity and the charge density of the cation in QCSs, Liu et al. had introduced two more powerful electronegative groups, the -CBr<sub>3</sub> and -CCl<sub>3</sub> groups into the chitosan and synthesized two new kinds of QCSs (TBEDMCS and TCEDMCS). In the molecules of TBEDMCS and TCEDMCS, the nitrogen atoms at C-2 had the most positive charge density due to the strongest electronegative of their substituted electronegative groups. They exhibited the best antioxidant activity which further demonstrated the relationship between the antioxidant activity and the positive density of the chi-

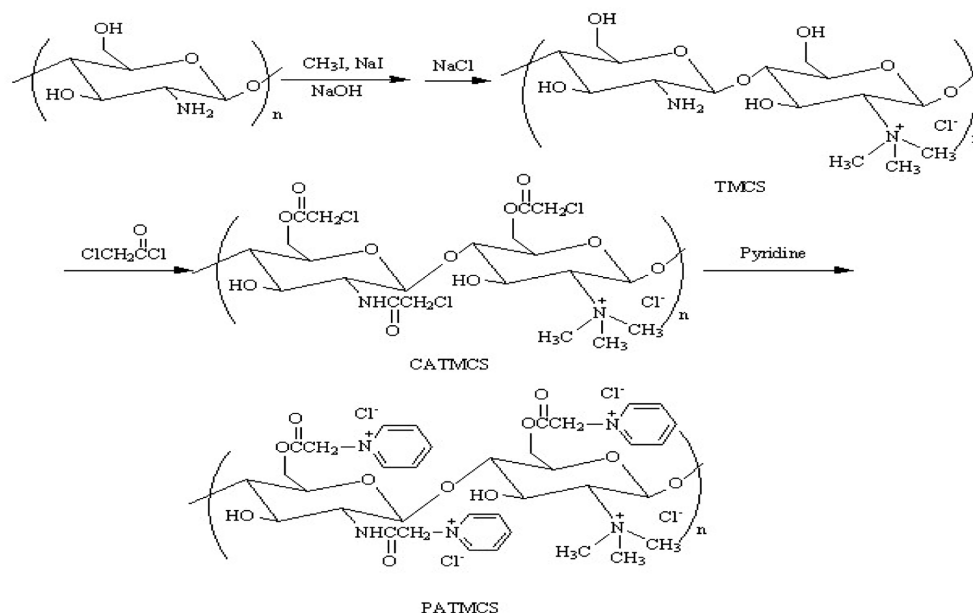
tosan derivative molecules.<sup>7</sup>

Based on above mentioned results, in this paper, a novel chitosan derivative with double quaternary ammonium salt—2-pyridine-acetyl-*N*-trimethyl chitosan chloride (PATMCS) was synthesized. Among various reactive oxygen species, hydroxyl radicals have the strongest chemical activity, which can damage a wide range of essential biomolecules such as amino acids, proteins, and DNA.<sup>8</sup> The antioxidant activity of PATMCS against hydroxyl radicals was assessed. PATMCS was expected to have potent activity due to the double quaternary ammonium salt structure in the molecules.

### EXPERIMENTAL

Chitosan was purchased from Qingdao Baicheng Biochemical Corp. (China). Its degree of deacetylation was 97%, and the viscosity-average molecular weight was  $2.0 \times 10^5$ . Sodium iodide (NaI) and iodomethane (CH<sub>3</sub>I) were purchased from the Sigma-Aldrich Chemical Co. The other reagents were all analytical grades and were used without further purification. The IR spectra were measured on a Jasco-4100 FT-IR spectrometer with KBr disks. The <sup>13</sup>C NMR spectra were recorded on a Bruker AVIII-500 NMR spectrometer in D<sub>2</sub>O solvent.

*N*-trimethyl chitosan chloride (TMCS) was the product of the fourth step prepared according to the method of Snyman via four steps.<sup>9</sup> PATMCS was synthesized as follows: TMCS (0.01 mol) was dispersed in NMP (20 mL)



**Scheme 1.** Synthetic pathway of PATMCS.

and stirred at room temperature for 2 h, then Chloroacetyl chloride (0.02 mol) was added. After stirring for 24 h at room temperature, the solution was precipitated in ether and the precipitate was washed with methanol and ether by turns, the Chloroacetyl-*N*-trimethyl chitosan (CATMCS) was obtained by drying at 40 °C in vacuum for 12 h. CATMCS (0.3 g) was dissolved in DMSO (20 mL), then pyridine (0.5 mL) was added. The solution was stirred for 24 h at 60 °C. The product was obtained by precipitation with excess acetone, and the PATMCS was obtained by drying at 40 °C in vacuum for 12 h (Scheme 1).

The antioxidant activity was carried out according to Guo.<sup>6</sup> The reaction mixture, total volume 4.5 mL, containing the samples of chitosan and its derivatives (chitosan, TMCS, CATMCS and PATMCS), was incubated with EDTA-Fe<sup>2+</sup> (220 μM), safranin O (0.23 μM), and H<sub>2</sub>O<sub>2</sub> (60 μM) in potassium phosphate buffer (150 mM, pH 7.4) for 30 min at 37 °C. The absorbance of the mixture was measured at 520 nm. Hydroxyl radicals bleached the safranin O, so increased absorbance of the reaction mixture indicated increased hydroxyl radicals' scavenging ability and the capability of scavenging hydroxyl radicals was calculated using the following equation:

$$\text{Scavenging effect (\%)} = \left[ \frac{(A_{\text{sample}520\text{nm}} - A_{\text{blank}520\text{nm}})}{(A_{\text{control}520\text{nm}} - A_{\text{blank}520\text{nm}})} \right] \times 100$$

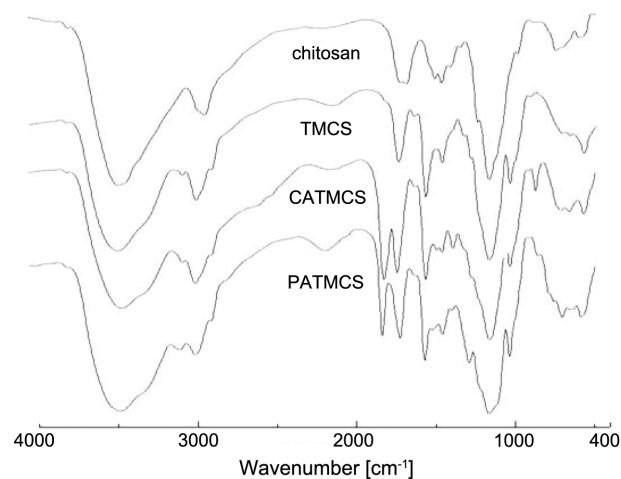
where  $A_{\text{blank}520\text{nm}}$  was the absorbance of the blank (distilled water instead of the samples),  $A_{\text{control}520\text{nm}}$  is the absor-

bance of the control (distilled water instead of H<sub>2</sub>O<sub>2</sub>).

All data are expressed as means ± SD. Data were analyzed by an analysis of variance ( $P < 0.05$ ) and the means were separated by Student-Newman-Keuls (SNK). The results were processed by the computer programs: Excel and Statistica software (SPSS).

## RESULTS AND DISCUSSION

In the FT-IR spectra of chitosan, TMCS, CATMCS and PATMCS (Fig. 1), chitosan was typically characterized by absorption regions as follows:<sup>10</sup> the major peaks of chitosan at about 898, 1079, 1600 cm<sup>-1</sup> belonging to pyranose



**Fig. 1.** IR spectra data of chitosan, TMCS, CATMCS and PATMCS.

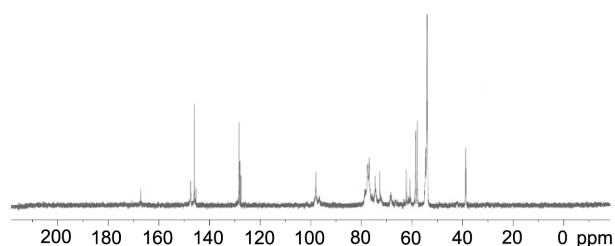


Fig. 2.  $^{13}\text{C}$  NMR spectra of PATMCS.

ring, glucoside and amine groups, respectively, were identifiable. In the spectrum of TMCS, the characteristic absorption of  $\text{NH}_2$ - at  $1600\text{ cm}^{-1}$  disappeared and a new peak appeared at  $1650\text{ cm}^{-1}$ , which was assigned to the quaternary ammonium salt.<sup>11</sup> There is a strong new peak at about  $1481\text{ cm}^{-1}$ , which was assigned to the characteristic absorbance band of the methyl in  $-\text{N}^+(\text{CH}_3)_3$ .<sup>12</sup> In the spectrum of CATMCS, new strong peaks at about  $1743\text{ cm}^{-1}$  and  $1662\text{ cm}^{-1}$  appeared which attributed to carbonyl groups  $\text{C}=\text{O}$  of ester group and acylamide respectively.<sup>13</sup> The peak at about  $787\text{ cm}^{-1}$  was assigned to  $\text{C}-\text{Cl}$ . After pyridine grafted onto the CATMCS, the characteristic absorbance of  $\text{C}-\text{Cl}$  at about  $787\text{ cm}^{-1}$  disappeared and new peaks at about  $1438$ ,  $771$  and  $3027\text{ cm}^{-1}$  were due to the characteristic absorbance of pyridine appeared in the spectrum of PATMCS.<sup>14</sup>

The  $^{13}\text{C}$  NMR spectrum further confirmed the success preparation of PATMCS (Fig. 2). Except the chitosan backbone peaks, there was strong peak at about  $53.9\text{ ppm}$  which attributed to  $-\text{N}^+(\text{CH}_3)_3$ . The peak at about  $167.2\text{ ppm}$  was assigned to  $\text{C}=\text{O}$ .<sup>13</sup> The peaks at about  $145.5$ ,  $128.4$  and  $147.5\text{ ppm}$  were peaks of pyridine.<sup>15</sup> At about  $38.9$  and  $58.8\text{ ppm}$ , there were peaks of methylene in  $-\text{NHCOCH}_2-$  and  $-\text{CH}_2\text{OCOCH}_2$ .<sup>16</sup> The spectrum data indicated that PATMCS was obtained.

In this paper, the material chitosan was not soluble in water at pH 7. We measured the activity of chitosan by dissolving it in HOAc solution. All the prepared chitosan derivatives were water-soluble. In Fig. 3, we compared the antioxidant activity of chitosan and its derivatives, TMCS, CATMCS and PATMCS. The hydroxyl radicals, generated by the Fenton reaction in this system, were scavenged by chitosan and the derivatives. All the samples exhibited concentration-dependent  $\cdot\text{OH}$  elimination effect. All of the three derivatives had better  $\cdot\text{OH}$  scavenging activity than chitosan. CATMCS exhibited worse  $\cdot\text{OH}$  scavenging activity than that of TMCS due to the reduced contents of the active hydroxyl and amino groups in the molecule chains.<sup>17</sup> PATMCS had the best  $\cdot\text{OH}$  sca-

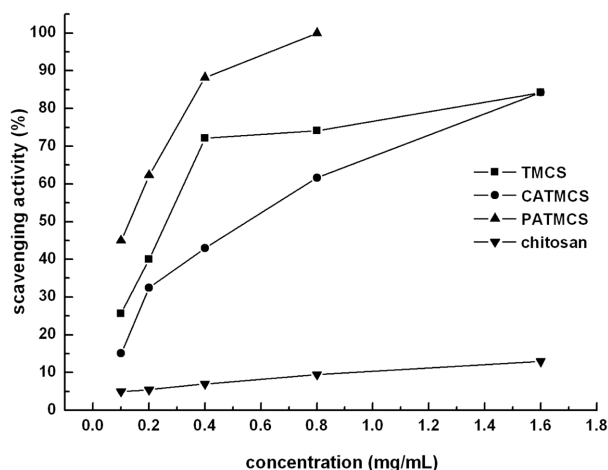


Fig. 3. Scavenging effect of chitosan, TMCS, CATMCS and PATMCS on hydroxyl radicals. Values are means  $\pm$  SD ( $n=3$ ).

vening activity. The  $\text{IC}_{50}$  value, which means the antioxidant concentration to reduce the  $\cdot\text{OH}$  by 50%, is a good parameter to evaluate the antioxidant ability. The  $\text{IC}_{50}$  of TMCS and PATMCS was  $0.26$  and  $0.13\text{ mg/mL}$ , respectively. The scavenging effect of PATMCS was 100% at the concentration of  $0.8\text{ mg/mL}$ . However, for TMCS, the scavenging effect was merely 84.5% at the concentration of  $1.6\text{ mg/mL}$ . Obviously, PATMCS improved the antioxidant activity of TMCS greatly.

## CONCLUSION

TMCS is a good substance for research of *N*-substituted quaternary chitosan with good water-solubility, easy-preparation and positive electricity of the molecules. In this paper, TMCS was further modified and a new double quaternary ammonium salt derivative of chitosan, PATMCS, was prepared. PATMCS exhibited potent antioxidant activity which may due to its double quaternary ammonium salt structure. The results are theoretical foundation for further development and making use of chitosan to prepare new antioxidants.

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