

## Model Systems in Radiation Biology; Implication for Preclinical Study of Radiotherapy

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In radiation biology, analysis of various mechanisms in response to radiation has been accomplished with the use of model organisms. These model organisms are powerful tools for providing a biologically intact *in vivo* environment to assess physiological and pathophysiological processes affected by radiation. Accumulated data using these models have been applied to human clinical studies (including the evaluation of radiotherapeutic efficacy) and discovery of radiotherapy reagents. However, there are few studies to provide overall integrated information about these useful model organisms. Thus, this review summarizes the results of radiation biology studies using four well-known model organisms: yeast, *Caenorhabditis elegans*, *Drosophila melanogaster*, and mice.

**Key words** : Radiation biology, yeast, *C. elegans*, *D. melanogaster*, mouse

### Introduction

The molecular, cellular and physiological analyses of various response pathways in humans have been accomplished by using of model organisms including yeast, fruit flies, and mice. These models allow us to understand other molecular mechanisms of cellular system, including cell cycle regulation, apoptosis, and tumorigenesis [73]. In addition, model organisms have been used to examine the pathogenesis of various diseases such as heart disease, neurodegenerative disorders, and cancers as well as discover novel targets for treating these conditions and enhancing the efficiency of targeted therapies [110].

In the same context, several model organisms have been adopted for conducting radiation biology studies [1,7,83]. Data from cell-based assays have helped to identify molecular targets in response to radiation and screen potent drugs. These attempts have resulted in effective outcomes in cellular conditions. Cell-based studies, however, are limited in that they overlook the potential roles of physiological conditions in tissues and organs. Experiments using model organisms could address this limitation because these models provide biologically intact systems available for evaluating the physiological effects of radiation. Moreover, data from

studies using yeast, worms, flies, and mice could be applied to human-based medical and medicinal investigations such as evaluating radiotherapeutic efficacy, developing radiotherapeutic adjuvants, and examining the mechanisms of radiation-induced injury in normal tissue.

Each model organism has unique properties, including life cycles, average lifespans, cellular pathways that respond to exogenous signals, and degrees of similarity to human cells in terms of genes and proteins. Based on these properties, each organism has advantages as well as limitations when using these models for different experimental methods. For example, yeast can be a good model for anti-cancer drug screening but it may be difficult to use these cells to directly explain various mechanisms of human diseases such as pulmonary fibrosis [24,110]. It is thus important for investigators to determine which organism is appropriate for their experiments. In this review, we provide an overview of four representative model organisms that have been used for radiation biology investigation: yeast, *Caenorhabditis (C.) elegans*, *Drosophila (D.) melanogaster*, and mice.

Yeast: an amenable and rapid research model eukaryotic organism

Yeasts, including *Saccharomyces (S.) cerevisiae* and *Schizosaccharomyces (S.) pombe*, have been widely employed

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as model systems for eukaryotic cell biology studies including genomics, cancer research, medicinal research, and radiation biology investigations [12,110]. Several reports studying the yeast genome have found that amino acid sequences and the functions of proteins in yeast and mammals, including ubiquitin, cytoskeletal elements, and a number of enzymes, are highly well conserved [12,40,110]. In particular, the analysis of yeast data has provided information about individual gene functions, protein-protein interactions, high-throughput screening, and genome-wide screening which can be applied to other eukaryotic organisms, including humans [11,55,70].

In radiation biology, yeasts have also been frequently used as experimental tools for investigating the DNA damage response following irradiation and mechanisms underlying radiosensitization and radioresistance. A variety of studies using yeasts have been conducted which focused on individual gene functions and protein interactions induced by irradiation [10,27,95]. Other yeast-based studies have demonstrated that an antioxidant defense mechanism is induced in response to ionizing radiation (IR) [69,123]. Several groups have shown that anti-oxidant enzymes, such as superoxide dismutase (SOD) and Yap1, are activated to protect against IR-associated oxidative stress [69,78]. Additionally, many checkpoint and DNA repair genes, including *RAD9*, *RAD17*, *RAD24* and *RAD53*, were identified during the response to DNA lesions induced by IR, leading to regulation of radio-sensitivity in *S. cerevisiae* [18,34,64]. These genes may play a role in survival signaling that acts against IR-induced damage. Studies of these genes have provided fundamental information for understanding the mechanisms underlying radioresistant properties of human cancer cells.

In the early 2000s, a genome-wide screening for radiation-affected genes in budding yeast *S. cerevisiae* was conducted. This study revealed that a large number of genes, involved in cell cycle control, nuclear pore formation, DNA damage repair, transcription regulation, Golgi/vacuolar activities, ubiquitin-mediated protein degradation, mitochondrial activities, and cell wall maintenance, were associated with resistance to IR. More than half of these genes shared a similar homology with human genes. Some of these, such as *SAC6* and *DHHL* (putative orthologs of human oncogene *LCP1* and *DDX6*, respectively), are involved in malignancies, indicating that they affect the radioresistant properties during radiotherapy for treating cancer [7]. Another study using the fission yeast *S. pombe* was reported. In this investigation,

global gene expression in response to conditions or reagents that induced DNA damage, including oxidative stress, heat shock, osmotic shock, heavy metal stress, DNA-damaging agents, and  $\gamma$ -irradiation was analyzed by DNA microarrays. Comparative analyses have demonstrated that genes involved in the DNA damage response, cell cycle regulators, signal transducers, stress-response genes, and genes involved in the metabolism of carbohydrates, lipids, and proteins are up-regulated by  $\gamma$ -irradiation. Among these, some subsets, such as *rhp51*, *rhp54*, and *dinB* which encode DNA repair proteins, might only respond to  $\gamma$ -irradiation [123].

Altogether, yeast is a simple and efficient model organism for providing fundamental information about IR-induced responses, such as antioxidant defense mechanism and DNA damage response, and investigating functionally related human genes. Such information obtained from yeast has been conveniently applied to the screening of anti-cancer drugs and radiotherapeutic adjuvants [110]. Several radiotherapeutic agents have been examined in yeast-based assay for their protective role against radiation-induced damage [2,3,92]. The data obtained from yeast cell-based studies, however, may have a limited ability to explain IR-induced clinical responses in human tissues or organs. To address this difficulty, data from higher-level eukaryotes, such as fruit flies and mice, are required.

#### *C. elegans*: an excellent organism for behavior study

The small nematode *C. elegans* has been an attractive model organism for biological research since it was first used as a model for developmental and behavior studies by Sydney Brenner over 40 years ago [15]. *C. elegans* has several advantages including its small size, rapid life cycle, short lifespan, genetic manipulability, and availability of its complete genome sequence [112,115]. Due to these properties, a number of studies have been performed, which provide the bulk of genotypic and phenotypic data available to researchers. Thus, *C. elegans* has been adopted as a good *in vivo* model organism in the field of radiation biology.

*C. elegans* was first used to evaluate the response to radiation in 1976 [49]. Subsequent studies have helped identify IR-response genes, including *rad-1* and *rad-2*, the mechanism of radiation-induced DNA damage repair, and radiation quality effect [46,47,84]. After the *C. elegans* genome was fully sequenced, analyses using high-throughput screening

and bioinformatics tools have found that about 53% of the genes are significantly similar to genes of other organisms, including yeasts and humans. Some of these genes are associated with factors that respond to DNA damage induced by oxidative stress, radiation, and chemical mutagens [13,83,114,126].

*C. elegans* has been used to examine cellular mechanisms involved in the response to radiation, including apoptosis, cell cycle arrest, and DNA damage repair. A variety of investigations have contributed to discovering the mechanism underlying IR-induced apoptosis in germ cells of *C. elegans* [39]. Several components, such as CED-3 (caspase homolog), CED-4 (apoptotic protease-activating factor 1 homolog), CED-9 (Bcl-2 homolog), and CEP-1 (p53 homolog) were playing critical roles in activation of apoptotic pathway after irradiation. [20,33,39,41,61,73,103,104,120,125]. In addition, *C. elegans* has been often used as a model to explain the aging mechanism regulated by oxidative stress which is possibly induced by radiation exposure. Several genetic studies have identified age-related genes, such as *age-1*, *clk-1*, *daf-2*, *daf-16*, and *mev-1* [36,52,59,67,121,127,129]. These genes are responsible for activation of survival signaling pathway such as DNA damage repair and antioxidant defense in response to oxidative stress, resulting in lifespan extension. Another report has presented that IR-induced oxidative damage was alleviated and consequently lifespan of *C. elegans* was extended by treatment of anti-oxidative agent resveratrol, indicating that *C. elegans* could be also an attractive candidate for investigation of pharmacological mechanism, like yeast [128].

*C. elegans* is also an efficient model for analyzing changes in behavior patterns in response to radiation. This organism has about 300 neurons connected by synapses and gap junctions and is able to display dynamic patterns of behavior, including learning, locomotion, and socialization in response to environmental conditions [23,50,98,102,124]. *C. elegans* shows a decreased locomotory rate in the presence of food; this has been defined as the "basal slowing response" [102]. A previous report showed that IR-induced locomotion of *C. elegans* was reduced in the absence of food, indicating that IR might affect the nervous system and regulate motor-behavior [99,113]. In addition, another study demonstrated that IR-induced responses might act as a modulator that influences food-NaCl associative learning by regulating specific sensory neurons related to GPC-1 (one of the two  $\gamma$  subunits of the heterotrimeric G-protein) in the nervous system, con-

sequently leading to altered learning behavior [100]. Such IR-induced behavior data derived from the *C. elegans* model has helped identify potential risks associated with radiotherapy for treating brain tumors, including decreased learning and memory.

All together, *C. elegans* as an *in vivo* radiation biology model can render significant clues to elucidate the several cellular mechanisms of radiation-induced responses. In addition, *C. elegans* studies can provide fundamental information to describe the aging mechanism and neurological response to IR in different biological systems. These outcomes can help apply for understanding of IR-induced human response, although, like yeast as a model organism, there are significant differences in physiological characteristics between *C. elegans* and mammals [101].

#### *D. melanogaster*: a landmark model organism for phenotype analysis

*D. melanogaster* (fruit fly) has been a preeminent animal model used in modern biological sciences since the heredity study conducted by Morgan in the early 20<sup>th</sup> century. The concept that chromosomes encode heritable traits was first developed using the fly along with many other outstanding discoveries in the field of genetics and development [96]. In 1927, Muller showed that IR causes chromosomal damage using male fruit fly germ cells. For this finding, Muller received a Nobel Prize in 1946 [82]. *D. melanogaster* has many attractive features that make it a landmark model for studying radiation biology. The fly genome has been completely sequenced and annotated, and contains more than 14,000 genes which have a nearly 75% homology to human genes associated with various diseases including cancer as well as immune and neurodegenerative diseases. Although overall identity between flies and mammals at the nucleotide or protein sequence is approximately 40%, the levels of similarity can be 80 to 90% in conserved functional domains [76,91]. The fly has unique characteristics traditional rodent models lack that makes flies ideal for studying radiation biology. These include as a short lifespan of about 60 days, excellent fertility rates (hundreds of identical offspring can be produced by a single fertile mating pair), and distinct developmental stages.

The fly may be considered as a versatile model organism due to its defined developmental stages: embryo, larva, pupa, and adult. Each stage has its own specific advantages

for investigating the effects of radiation. Embryos are often irradiated in fundamental developmental studies including ones evaluating cell fate determination, gene expression and organogenesis [31,62,106]. Larvae, particularly 3<sup>rd</sup> instar larvae, are most commonly used for irradiation experiments to study developmental and physiological processes since tissues, known as imaginal discs, grow inside the larva during this developmental stage [5,9,37,58,79,88,107,108,119]. Beginning in the late third instar larval phase and proceeding through the pupal phase, the imaginal disc develops into most structures of the adult body, such as the head, legs, wings, thorax, and genitalia. Exposure to IR during the 3<sup>rd</sup> larva stage may be used to observe the detrimental effects of radiation on development. Adult flies are suitable for studying radiation resistance owing to the composition of post-mitotic tissues. This makes it possible to determine the effects of radiation without the confounding effects from mitotic tissues [48,89].

Above all, the most invaluable attribute of *D. melanogaster* as an animal model is the establishment of advanced genetic techniques for the production of transgenic flies. Since the initial development of a GAL4/UAS system for specific gene expression in 1993, many modifications and enhancements of this basic system have been created to further refine spatial and temporal expression in the fly body [94]. Moreover, a collection of RNAi (RNA interference) knockdown strains targeting about 90% of the entire fly genome has been constructed and is now available from the Vienna Drosophila RNAi Center (<http://www.vdrc.at/>) [25]. A great number of genes involved in responses to radiation have been investigated using these transgenic flies, including ones affecting apoptosis, DNA repair, stress responses, cell cycle regulation, and longevity [21,68,80,81,87].

Since the study conducted by Muller mentioned above, many groups have examined the dose-dependent effects of radiation on germ cell damage. Initial experiments involving thousands of *Drosophila* demonstrated that radiation induces mutation rates up to 150 times greater than spontaneous mutation rate [82]. This group hypothesized that radiation can cause severe infertility. These and similar experimental results using mouse sperm indicated that there is no safe threshold dose no matter how low. This serves as the basis for guide-lines to protect human from radiation exposure in the world [118].

Developments in transgenic fly technology have contributed to the characterization of gene functions related to

specific conditions such as neurodegenerative diseases, cancer, and diabetes [8,44]. Tumor development is a very complicated process involving various cellular responses including genome integrity, DNA damage response, apoptosis, and cell cycle regulation. These responses are highly conserved among different species, and many genes of these responses identified in *Drosophila* are altered in human cancers [16,19,86,111]. Recently, fruit flies were also used to evaluate the efficacy of cancer therapeutic approaches involving combinations of radiation and chemical reagents [32].

*Drosophila* have been usually used for studying innate immunity since key elements of the innate immune system are highly conserved between flies and humans [51]. Identification of the Toll and Imd signaling pathways, well-known for mediators of bacterial and fungal infections in the fruit fly, led to the identification of mammalian counterparts of the Toll-like receptor and tumor necrosis factor pathways [72]. Reactive oxygen species (ROS) and IR were also reported to be involved in the *Drosophila* immune response [97,106]. In addition, experiments conducted to study the effect of radiation on innate immunity showed that low dose radiation specifically activates a particular immune pathway in *Drosophila* [105]. Taken together, the findings of these studies illustrate how radiation affects innate immunity at the molecular level.

Considering the results of previous experiments, it is clear that *Drosophila* is a useful animal model in both the discovery for the physiological mechanism of radiation effect and the validation for the efficacy of reagents related to radiation response. Although there are many differences between flies and humans, the degree of biological and physiological conservation enables the use of *D. melanogaster* as an extremely valuable tool for radiation biology studies.

#### *Mus musculus* (mouse): a premier mammalian model

Among the higher eukaryotic organisms, mice are one of the most suitable models for investigating IR-mediated human physiological responses, since about 99% of the genes in mice and humans are shared. Additionally, common pathophysiological processes of various diseases, particularly cancer, obesity, diabetes, neurological disorders, and heart disease, are also shared [122]. From the early 1900s to now, many studies, involved in basic biology, ge-

nomics, cancer research, and transplantation biology, have provided much information which has systematically accumulated to form the infrastructure of mouse-based databases and resources (*i.e.*, mouse genome informatics; <http://www.informatics.jax.org>) [90]. In the same context, a large number of genetically modified mice, including inbred mouse strains, transgenic mice, and knock-out mice, have been established and are commercially available [63]. Mice have a relatively short development period (about 20 days), so researchers can obtain their desired data easily and quickly. For these reasons, the mouse is considered to be a premier model organism for understanding the genetic basis and complicated mechanisms of various human diseases and normal physiological processes such as radio-resistance in cancer cells and inflammatory response in the normal tissue during radiotherapy.

Many groups have employed the mouse as an experimental model for protein profiling analysis in the context of the radiation response [4,43,74,93]. One study using C57BL/6 mice found that IR induces differences in protein expression patterns between brain and intestine (organs known to be resistant and sensitive to radiation, respectively) by comparative 2-DE and gel imaging analyses. Based on these results, several proteins, including transaldolase 1 and phosphoglycerate kinase 1, were identified as potential markers of IR responses in brain and intestine, respectively [74]. Other groups demonstrated IR-induced changes of serum protein content in the skin of BABL/c mice and performed IR-induced protein expression profiling in blood-plasma of CBA/CaJ mice primarily using 2-DE proteomic analysis [43,93]. In addition, cardiac mitochondrial proteome and function using C57BL/6N mice was investigated by analyses of several proteomic approaches and interactome network [6]. These data have been used to identify tissue-specific protein biomarkers to help prognose and diagnose the IR-induced response of normal and cancerous tissues.

Mice were also used for investigating IR-induced pathogenesis of various conditions such as thymic lymphomas, hepatic injury, and lung fibrosis [4,38,56,65,66]. One study investigated radiation-induced DNA damage in C57BL/6J mice. This damage might result in the accumulation of mutations in tumor suppressor genes, such as *p53* and *CTIP2*, thus leading to the development of mouse thymic lymphomas [14,65]. In addition to DNA damage, irradiated liver tissues from C3H/HeJ mice were shown to have up-regu-

lated levels of antioxidant enzymes, including glutathione *S*-transferase Pi, peroxiredoxin VI, and cytochrome *c* which could help protect against radiation injury mediated by IR-generated ROS [1,71]. Another study was performed to investigate different responses to IR in the lungs of C3H/HeJ (a lung fibrosis-resistant strain) and C57BL/6J (a lung fibrosis-sensitive strain) mice. Comparative analysis using a proteomic approach has shown that the expression of several antioxidant proteins, including SOD1, cytochrome *c* oxidase, and glutamate dehydrogenase, were decreased in the irradiated C57BL/6J mice compared to C3H/HeJ mice [4]. This finding suggests that IR leads to the dysregulation of cellular defense systems which protect against oxidative stress and could be responsible for lung toxicity, possibly leading to the development of lung fibrosis. Together, such studies have provided insights into physiological mechanism underlying normal tissue injury and tumorigenesis in humans receiving radiotherapy.

In addition to the use of mouse model for studying normal tissue damage, mice are good model for understanding the enhancement of radiotherapeutic efficiency for treating cancer. When anti-cancer drugs as well as radioprotective agents that prevent normal tissue damage were identified by wide-range screening, they were further assessed by cell-based *in vitro* and *ex vivo* experiments, such as cytotoxicity screens and biochemical assays, to further reveal their functions. Based on data produced by these studies, mice were employed as *in vivo* models to test various chemicals administered in combination with radiotherapy. This was done to confirm for their activities in physiologically relevant environments through the use of tumor xenografts, transplantation techniques and genetically engineered mice [17,60,85].

Tumor xenografts in mice have been used as models for preclinical testing of radiotherapy strategies. An investigation using tumor xenografts in mice to establish an IR-guided drug delivery system for specifically treating lung cancer has been presented. In this study, an anti-cancer drug with specific peptides targeting the radiation-inducible membrane protein, tax-interacting protein-1, was evaluated in a tumor model (lung cancer cells grafted in C57 and athymic nude mice). The results showed that the drug specifically targeted tumor mass, leading to the reduction of tumor volume [45]. In addition to tumor xenograft models, genetically engineered mice, including knockout mice, have been also developed to explore the *in vivo* functions of many

genes in response to IR. Finding from studies involved these animals could support the *in vitro* data, and consequently to render the clues for discovering novel target-specific drugs [22,35,75].

Many groups have adopted and/or (if necessary) modified a specific mouse strain as a disease model based on the characteristics of each strain or using the genetic engineering and tumor xenograft methods. These mouse models have been recently used for obtaining the clinical implication to apply appropriate radiotherapy strategy to many diseases. Several investigations have been conducted to understand radiation-induced lung injury (fibrosis and pneumonitis) using various mouse strains including C3H, CBA, C57BL/6, C57L, BALB/c, C57BR/J, and A/J mice [26,28-30,57,109,117]. Some of these strains have been controversial to be proper models for lung injury model because they have shown the unrelated phenomena to lung fibrosis or pneumonitis in response to radiation. Thus, one study using comparative analysis of several strains have reported that, based on histopathological analyses, CBA, C3H, and, especially, C57L strains are more desirable for investigating radiation-induced lung fibrosis and pneumonitis and developing appropriate radioprotective therapies [56,57]. In addition to lung model, there are more models designed for specific disease. A study using a specifically designed DBA/2J mouse as a glaucoma (a leading cause of visual loss) disease model has reported that the development of glaucoma might be resulted from neuroinflammatory response, which could be prevented effectively by localizing tissue radiation [53,54]. The mouse prostate reconstitution model system have been generated using RM-9 cells (the specific intent to mimic the phenotypic and genetic characteristics of human prostate cancer) to investigate the intratumoral mechanism (regulation of immune and hypoxia response) induced by co-treatment of chemotherapy and radiotherapy [42,77,116]. Many groups have made a great effort to establish the desirable model mice for specific disease although the outcome was still insufficient. These efforts could definitely help to investigate the mechanism of radiation-induced disease development and to enhance the efficacy of radiotherapy with high accuracy in a relatively short time. Moreover, such a disease model may satisfy the preclinical requirements for approval of radiation adjuvants for enhanced radiotherapy or radiation protectors against radiation-induced normal tissue injury for human use.

Altogether, the mice have several important advantages

for studying radiation biology compared to other organisms such as yeast, worms, and flies. Unlike lower organisms, the mouse has a highly conserved genome and proteome compared to humans. Genetically engineered mice and disease model mice can directly provide clinical information about the response to radiation. In addition, these mice are also useful for validating the physiological efficacy of candidate radiotherapeutic drugs discovered by cell-based screening.

## Conclusions

This review discussed the use of model organisms to conduct the radiation biology investigations. All models can produce valuable information important for understanding the response to radiation (Table 1). This accumulated databases and resources of each organism are generally accessible as well (Table 2). Basic studies in yeast have contributed to elucidating conserved biological responses, promoted the development of genomic methodologies, and provided functional information about IR-responsive eukaryotic proteins through genome-wide high-throughput approaches. Moreover, yeast-based assays may be used for the discovery of radiotherapeutic agents and measuring the enhanced efficacy of radiotherapy for treating cancer. *C. elegans* may be an appropriate model for studying IR-induced basic cellular responses including apoptosis, cell cycle arrest, and DNA damage repair. In addition, *C. elegans* can be used as a basic model to investigate the effect IR on the nervous system. Data from these two organisms, yeast and *C. elegans*, may have a limited ability to apply to directly IR-induced clinical response in human, because they are so simple compared to human physiology. *D. melanogaster* is often used for basic and applied studies of IR-induced responses based on phenotype analysis. Transgenic fly models allowed us to investigate mechanisms of physiological changes induced by IR in humans. However, data from *D. melanogaster* may also be insufficient to explain the IR-mediated physiological response in human owing to different body system between fly and human. Since the mouse is a higher organism, it is the most suitable among the four models for clinical studies to understand human physiological and pathological responses mediated by IR. Mice can be also used to evaluate the effects of drugs administered in combined with radiation. On the other hand, mouse has relatively more expensive and strict care condition and need more time to obtain experimental results than others. Based on these proper-

Table 1. Comparison of four model organisms



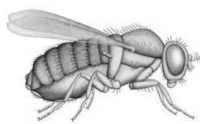

	Yeast	<i>C. elegans</i>	<i>D. melanogaster</i>	Mouse
<b>Feature</b>				
<b>Advantage of experiments</b>	Simple growth requirements, Rapid cell growth, Ease of genetic manipulation, Genome-wide screening	Short lifespan, Rapid life cycle, Small body size, Transparent body, Ease of genetic manipulation, Knockout mutant libraries, Behavior pattern	Excellent fertility (identical offsprings), Distinct developmental stages, Transgenic flies	Higher functional genetic and proteomic conservation to human homolog, Transplantation, Gene-knockout or -knockin mice, Proteomics (tissue- or organ-based), Construction of disease model
<b>Clinical meanings</b>	Determination of candidate genes and proteins in response to radiation Cell-based drug screening for radiotherapy (basic tool)	Cellular response to radiation, IR-induced aging mechanisms, IR-mediated neuronal pathway	Analysis of IR-induced phenotype changes, IR-affected innate immunity Examination of heritable effects	Disease model in radiation biology, Drug screening for radiotherapy (physiological application), Drug delivery system

Table 2. Websites for database of four model organisms

<b>Yeast</b>	
<i>Saccharomyces</i> Genome Database	<a href="http://www.yeastgenome.org/">http://www.yeastgenome.org/</a>
Mammalian homology to yeast	<a href="http://www.yeastgenome.org/mammal/">http://www.yeastgenome.org/mammal/</a>
Yeast homologs of human disease-associated genes	<a href="http://mips.gsf.de/proj/yeast/reviews/human_diseases.htm">http://mips.gsf.de/proj/yeast/reviews/human_diseases.htm</a>
<b><i>C. elegans</i></b>	
WormBase	<a href="http://www.wormbase.org/">http://www.wormbase.org/</a>
<i>Caenorhabditis</i> Genetics Center	<a href="http://www.cbs.umn.edu/CGC/">http://www.cbs.umn.edu/CGC/</a>
<b><i>D. melanogaster</i></b>	
<i>Drosophila</i> Genomics Resource Center	<a href="https://dgrc.cgb.indiana.edu/">https://dgrc.cgb.indiana.edu/</a>
<i>Drosophila</i> functional genomics and proteomics	<a href="http://www.gen.cam.ac.uk/~flychip/">http://www.gen.cam.ac.uk/~flychip/</a>
Database of human disease gene homologs in <i>Drosophila</i>	<a href="http://homophila.sdsc.edu/">http://homophila.sdsc.edu/</a>
Vienna <i>Drosophila</i> RNAi Center	<a href="http://www.vdrc.at/">http://www.vdrc.at/</a>
<b>Mouse</b>	
Mouse Genome Informatics	<a href="http://www.informatics.jax.org">http://www.informatics.jax.org</a>
International Mouse Strain Resources	<a href="http://www.informatics.jax.org/imsr/index.jsp">http://www.informatics.jax.org/imsr/index.jsp</a>
Mouse Mutant Resource	<a href="http://www.jax.org/mmr/index.html">http://www.jax.org/mmr/index.html</a>

ties, yeast, *C. elegans*, and *D. melanogaster* can be appropriate models for initial step in investigation of radiation biology, such as identifying IR-induced novel biomarker protein, investigating cellular mechanism of the protein, screening novel radiotherapeutic drug, and testing its cytotoxicity. In addi-

tion, *D. melanogaster* might be a good model for testing IR-induced simple physiological response and heritable effects as well. Among four organisms, mouse, especially as a disease model, is the best-suited for preclinical study of radiation response, including validation of the physiological efficacy

of novel radiotherapeutic drugs and determination of the appropriate dose for administration.

In radiation biology, model organisms are considered mandatory for confirming *in vitro* data because the responses to radiation might vary in different tissues or cells. The underlying biological mechanisms in humans and these model organisms can be quite different as well. Nevertheless, if investigators comprehend the properties of each model organism and choose the proper one for studying of radiation mechanisms, these models can make valuable contributions to basic radiation biology investigations as well as the development of clinical applications for humans.

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#### Abbreviations

*C. elegans*, *Caenorhabditis elegans*; *D. melanogaster*, *Drosophila melanogaster*; IR, ionizing radiation; RNAi, RNA interference; ROS, reactive oxygen species; *S. cerevisiae*, *Saccharomyces cerevisiae*; *S. pombe*, *Schizosaccharomyces pombe*; SOD, superoxide dismutase

#### References

- An, J. H., Kim, J. and Seong, J. 2004. Redox signaling by ionizing radiation in mouse liver. *Ann. N. Y. Acad. Sci.* **1030**, 86-94.
- Anjaria, K. B., Bhat, N. N., Shirsath, K. B. and Sreedevi, B. 2011. Differential modifying effects of food additive butylated hydroxytoluene toward radiation and 4-nitro-quinoline 1-oxide-induced genotoxicity in yeast. *J. Environ. Pathol. Toxicol. Oncol.* **30**, 189-197.
- Anjaria, K. B. and Rao, B. S. 2001. Effect of caffeine on the genotoxic effects of gamma radiation and 4-NQO in diploid yeast. *J. Environ. Pathol. Toxicol. Oncol.* **20**, 39-45.
- Ao, X., Lubman, D. M., Davis, M. A., Xing, X., Kong, F. M., Lawrence, T. S. and Zhang, M. 2008. Comparative proteomic analysis of radiation-induced changes in mouse lung: fibrosis-sensitive and -resistant strains. *Radiat. Res.* **169**, 417-425.
- Arias, A. M. 2008. *Drosophila melanogaster* and the development of biology in the 20th century. *Methods Mol. Biol.* **420**, 1-25.
- Barjaktarovic, Z., Schmaltz, D., Shyla, A., Azimzadeh, O., Schulz, S., Haagen, J., Dorr, W., Sarioglu, H., Schafer, A., Atkinson, M. J., Zischka, H. and Tapio, S. 2011. Radiation-induced signaling results in mitochondrial impairment in mouse heart at 4 weeks after exposure to X-rays. *PLoS One* **6**, e27811.
- Bennett, C. B., Lewis, L. K., Karthikeyan, G., Lobachev, K. S., Jin, Y. H., Sterling, J. F., Snipe, J. R. and Resnick, M. A. 2001. Genes required for ionizing radiation resistance in yeast. *Nat. Genet.* **29**, 426-434.
- Bilen, J. and Bonini, N. M. 2005. *Drosophila* as a model for human neurodegenerative disease. *Annu. Rev. Genet.* **39**, 153-171.
- Bol'shakov, M. A., Librikht, O. K., Kniazeva, I. R., El'chaninov, A. A., Klimov, A. I. and Rostov, V. V. 2007. Life expectancy and fertility of postembryo stage *drosophila* after the pulse-periodic X-ray irradiation. *Radiat. Biol. Radioecol.* **47**, 22-27.
- Boreham, D. R., Trivedi, A., Weinberger, P. and Mitchel, R. E. 1990. The involvement of topoisomerases and DNA polymerase I in the mechanism of induced thermal and radiation resistance in yeast. *Radiat. Res.* **123**, 203-212.
- Botstein, D. and Fink, G. R. 2011. Yeast: an experimental organism for 21st Century biology. *Genetics* **189**, 695-704.
- Botstein, D. and Fink, G. R. 1988. Yeast: an experimental organism for modern biology. *Science* **240**, 1439-1443.
- Boulton, S. J., Gartner, A., Reboul, J., Vaglio, P., Dyson, N., Hill, D. E. and Vidal, M. 2002. Combined functional genomic maps of the *C. elegans* DNA damage response. *Science* **295**, 127-131.
- Brathwaite, O., Bayona, W. and Newcomb, E. W. 1992. p53 mutations in C57BL/6J murine thymic lymphomas induced by gamma-irradiation and N-methylnitrosourea. *Cancer Res.* **52**, 3791-3795.
- Brenner, S. 1974. The genetics of *Caenorhabditis elegans*. *Genetics* **77**, 71-94.
- Brodsky, M. H., Nordstrom, W., Tsang, G., Kwan, E., Rubin, G. M. and Abrams, J. M. 2000. *Drosophila* p53 binds a damage response element at the reaper locus. *Cell* **101**, 103-113.
- Cao, C., Albert, J. M., Geng, L., Ivy, P. S., Sandler, A., Johnson, D. H. and Lu, B. 2006. Vascular endothelial growth factor tyrosine kinase inhibitor AZD2171 and fractionated radiotherapy in mouse models of lung cancer. *Cancer Res.* **66**, 11409-11415.
- Carr, A. M. 1994. Radiation checkpoints in model systems. *Int. J. Radiat. Biol.* **66**, S133-139.
- Chang, H. C. and Rubin, G. M. 1997. 14-3-3 epsilon positively regulates Ras-mediated signaling in *Drosophila*. *Genes Dev.* **11**, 1132-1139.
- Chen, F., Hersh, B. M., Conradt, B., Zhou, Z., Riemer, D., Gruenbaum, Y. and Horvitz, H. R. 2000. Translocation of *C. elegans* CED-4 to nuclear membranes during programmed cell death. *Science* **287**, 1485-1489.
- Chen, J., Xie, C., Tian, L., Hong, L., Wu, X. and Han, J. 2010. Participation of the p38 pathway in *Drosophila* host defense against pathogenic bacteria and fungi. *Proc. Natl. Acad. Sci. USA* **107**, 20774-20779.
- Day, T. K., Hooker, A. M., Zeng, G. and Sykes, P. J. 2007. Low dose X-radiation adaptive response in spleen and pro-



- tate of Atm knockout heterozygous mice. *Int. J. Radiat. Biol.* **83**, 523-534.
23. de Bono, M. and Bargmann, C. I. 1998. Natural variation in a neuropeptide Y receptor homolog modifies social behavior and food response in *C. elegans*. *Cell* **94**, 679-689.
  24. Degryse, A. L. and Lawson, W. E. 2011. Progress toward improving animal models for idiopathic pulmonary fibrosis. *Am. J. Med. Sci.* **341**, 444-449.
  25. Dietzl, G., Chen, D., Schnorrer, F., Su, K. C., Barinova, Y., Fellner, M., Gasser, B., Kinsey, K., Oppel, S., Scheiblauer, S., Couto, A., Marra, V., Keleman, K. and Dickson, B. J. 2007. A genome-wide transgenic RNAi library for conditional gene inactivation in *Drosophila*. *Nature* **448**, 151-156.
  26. Dileto, C. L. and Travis, E. L. 1996. Fibroblast radiosensitivity in vitro and lung fibrosis in vivo: comparison between a fibrosis-prone and fibrosis-resistant mouse strain. *Radiat. Res.* **146**, 61-67.
  27. Dolling, J. A., Boreham, D. R., Bahen, M. E. and Mitchel, R. E. 2000. Role of RAD9-dependent cell-cycle checkpoints in the adaptive response to ionizing radiation in yeast, *Saccharomyces cerevisiae*. *Int. J. Radiat. Biol.* **76**, 1273-1279.
  28. Down, J. D. and Steel, G. G. 1983. The expression of early and late damage after thoracic irradiation: a comparison between CBA and C57B1 mice. *Radiat. Res.* **96**, 603-610.
  29. Down, J. D., Tarbell, N. J., Warhol, M. and Mauch, P. 1988. Dose-limiting complications from upper half body irradiation in C3H mice. *Int. J. Radiat. Oncol. Biol. Phys.* **14**, 483-489.
  30. Down, J. D. and Yanch, J. C. 2010. Identifying the high radiosensitivity of the lungs of C57L mice in a model of total-body irradiation and bone marrow transplantation. *Radiation Res.* **174**, 258-263.
  31. Ducau, J., Bregliano, J. C. and de La Roche Saint-Andre, C. 2000. Gamma-irradiation stimulates homology-directed DNA double-strand break repair in *Drosophila* embryo. *Mutat. Res.* **460**, 69-80.
  32. Edwards, A., Gladstone, M., Yoon, P., Raben, D., Frederick, B. and Su, T. T. 2011. Combinatorial effect of maytansinol and radiation in *Drosophila* and human cancer cells. *Dis. Model. Mech.* **4**, 496-503.
  33. Fairlie, W. D., Perugini, M. A., Kvensakul, M., Chen, L., Huang, D. C. and Colman, P. M. 2006. CED-4 forms a 2 : 2 heterotetrameric complex with CED-9 until specifically displaced by EGL-1 or CED-13. *Cell Death Differ.* **13**, 426-434.
  34. Fasullo, M., Bennett, T., AhChing, P. and Koudelik, J. 1998. The *Saccharomyces cerevisiae* RAD9 checkpoint reduces the DNA damage-associated stimulation of directed translocations. *Mol. Cell. Biol.* **18**, 1190-1200.
  35. Finnberg, N., Gruber, J. J., Fei, P., Rudolph, D., Bric, A., Kim, S. H., Burns, T. F., Ajuha, H., Page, R., Wu, G. S., Chen, Y., McKenna, W. G., Bernhard, E., Lowe, S., Mak, T. and El-Deiry, W. S. 2005. DR5 knockout mice are compromised in radiation-induced apoptosis. *Mol. Cell. Biol.* **25**, 2000-2013.
  36. Friedman, D. B. and Johnson, T. E. 1988. A mutation in the age-1 gene in *Caenorhabditis elegans* lengthens life and reduces hermaphrodite fertility. *Genetics* **118**, 75-86.
  37. Fryxell, K. J. and Kumar, J. P. 1993. Characterization of the radiation-sensitive stage in the development of the compound eye of *Drosophila*. *Mutat. Res.* **285**, 181-189.
  38. Fu, Q., Berbee, M., Wang, W., Boerma, M., Wang, J., Schmid, H. A. and Hauer-Jensen, M. 2011. Preclinical evaluation of Som230 as a radiation mitigator in a mouse model: post-exposure time window and mechanisms of action. *Radiat. Res.* **175**, 728-735.
  39. Gartner, A., Milstein, S., Ahmed, S., Hodgkin, J. and Hengartner, M. O. 2000. A conserved checkpoint pathway mediates DNA damage-induced apoptosis and cell cycle arrest in *C. elegans*. *Mol. Cell* **5**, 435-443.
  40. Goffeau, A., Barrell, B. G., Bussey, H., Davis, R. W., Dujon, B., Feldmann, H., Galibert, F., Hoheisel, J. D., Jacq, C., Johnston, M., Louis, E. J., Mewes, H. W., Murakami, Y., Philippsen, P., Tettelin, H. and Oliver, S. G. 1996. Life with 6000 genes. *Science* **274**, 546, 563-547.
  41. Greiss, S., Schumacher, B., Grandien, K., Rothblatt, J. and Gartner, A. 2008. Transcriptional profiling in *C. elegans* suggests DNA damage dependent apoptosis as an ancient function of the p53 family. *BMC Genomics* **9**, 334.
  42. Gridley, D. S., Makinde, A. Y., Luo, X., Rizvi, A., Crapo, J. D., Dewhirst, M. W., Moeller, B. J., Pearlstein, R. D. and Slater, J. M. 2007. Radiation and a metalloporphyrin radioprotectant in a mouse prostate tumor model. *Anticancer Res.* **27**, 3101-3109.
  43. Guipaud, O., Holler, V., Buard, V., Tarlet, G., Royer, N., Vinh, J. and Benderitter, M. 2007. Time-course analysis of mouse serum proteome changes following exposure of the skin to ionizing radiation. *Proteomics* **7**, 3992-4002.
  44. Hafen, E. 2004. Cancer, type 2 diabetes, and ageing: news from flies and worms. *Swiss Med Wkly.* **134**, 711-719.
  45. Hariri, G., Yan, H., Wang, H., Han, Z. and Hallahan, D. E. 2010. Radiation-guided drug delivery to mouse models of lung cancer. *Clin. Cancer Res.* **16**, 4968-4977.
  46. Hartman, P. S. 1985. Epistatic interactions of radiation-sensitive (rad) mutants of *Caenorhabditis elegans*. *Genetics* **109**, 81-93.
  47. Hartman, P. S., Simpson, V. J., Johnson, T. and Mitchell, D. 1988. Radiation sensitivity and DNA repair in *Caenorhabditis elegans* strains with different mean life spans. *Mutat. Res.* **208**, 77-82.
  48. Helfand, S. L. and Rogina, B. 2003. Genetics of aging in the fruit fly, *Drosophila melanogaster*. *Annu. Rev. Genet.* **37**, 329-348.
  49. Herman, R. K., Albertson, D. G. and Brenner, S. 1976. Chromosome rearrangements in *Caenorhabditis elegans*. *Genetics* **83**, 91-105.
  50. Hobert, O. 2003. Behavioral plasticity in *C. elegans*: paradigms, circuits, genes. *J. Neurobiol.* **54**, 203-223.
  51. Hoffmann, J. A. and Reichhart, J. M. 2002. *Drosophila* innate immunity: an evolutionary perspective. *Nat. Immunol.* **3**, 121-126.
  52. Houthoofd, K., Braeckman, B. P., Lenaerts, I., Brys, K., De Vreese, A., Van Eygen, S. and Vanfleteren, J. R. 2002. Axenic

- growth up-regulates mass-specific metabolic rate, stress resistance, and extends life span in *Caenorhabditis elegans*. *Exp. Gerontol.* **37**, 1371-1378.
53. Howell, G. R., Libby, R. T., Jakobs, T. C., Smith, R. S., Phalan, F. C., Barter, J. W., Barbay, J. M., Marchant, J. K., Mahesh, N., Porciatti, V., Whitmore, A. V., Masland, R. H. and John, S. W. 2007. Axons of retinal ganglion cells are insulted in the optic nerve early in DBA/2J glaucoma. *J. Cell Biol.* **179**, 1523-1537.
54. Howell, G. R., Soto, I., Zhu, X., Ryan, M., Macalinao, D. G., Sousa, G. L., Caddle, L. B., MacNicoll, K. H., Barbay, J. M., Porciatti, V., Anderson, M. G., Smith, R. S., Clark, A. F., Libby, R. T. and John, S. W. 2012. Radiation treatment inhibits monocyte entry into the optic nerve head and prevents neuronal damage in a mouse model of glaucoma. *J. Clin. Invest.* **122**, 1246-1261.
55. Huttenhower, C., Hibbs, M. A., Myers, C. L., Caudy, A. A., Hess, D. C. and Troyanskaya, O. G. 2009. The impact of incomplete knowledge on evaluation: an experimental benchmark for protein function prediction. *Bioinformatics* **25**, 2404-2410.
56. Jackson, I. L., Vujaskovic, Z. and Down, J. D. 2011. A further comparison of pathologies after thoracic irradiation among different mouse strains: finding the best preclinical model for evaluating therapies directed against radiation-induced lung damage. *Radiat. Res.* **175**, 510-518.
57. Jackson, I. L., Vujaskovic, Z. and Down, J. D. 2010. Revisiting strain-related differences in radiation sensitivity of the mouse lung: recognizing and avoiding the confounding effects of pleural effusions. *Radiat. Res.* **173**, 10-20.
58. Kanao, T. and Miyachi, Y. 2006. Exposure to low-dose X-rays promotes peculiar autophagic cell death in *Drosophila melanogaster*, an effect that can be regulated by the inducible expression of Hml dsRNA. *Mutat. Res.* **595**, 60-68.
59. Kenyon, C., Chang, J., Gensch, E., Rudner, A. and Tabtiang, R. 1993. A *C. elegans* mutant that lives twice as long as wild type. *Nature* **366**, 461-464.
60. Kim, K. W., Hwang, M., Moretti, L., Jaboin, J. J., Cha, Y. I. and Lu, B. 2008. Autophagy upregulation by inhibitors of caspase-3 and mTOR enhances radiotherapy in a mouse model of lung cancer. *Autophagy* **4**, 659-668.
61. Kimura, T., Takahashi, K., Suzuki, Y., Konishi, Y., Ota, Y., Mori, C., Ikenaga, T., Takanami, T., Saito, R., Ichiishi, E., Awaji, S., Watanabe, K. and Higashitani, A. 2008. The effect of high strength static magnetic fields and ionizing radiation on gene expression and DNA damage in *Caenorhabditis elegans*. *Bioelectromagnetics* **29**, 605-614.
62. King, B. H. and Bryant, P. J. 1982. Developmental responses of the *Drosophila melanogaster* embryo to localized X irradiation. *Radiat. Res.* **89**, 590-606.
63. Kislinger, T. and Gramolini, A. O. 2010. Proteome analysis of mouse model systems: A tool to model human disease and for the investigation of tissue-specific biology. *J. Proteomics* **73**, 2205-2218.
64. Koltovaia, N. A., Nikulushkina Iu, V., Poshchina, M. P. and Devin, A. B. 2008. [Interaction between checkpoint genes RAD9, RAD17, RAD24, and RAD53 involved in the determination of yeast *Saccharomyces cerevisiae* sensitivity to ionizing radiation]. *Genetika* **44**, 761-770.
65. Kominami, R. and Niwa, O. 2006. Radiation carcinogenesis in mouse thymic lymphomas. *Cancer Sci.* **97**, 575-581.
66. Kuperwasser, C., Pinkas, J., Hurlbut, G. D., Naber, S. P. and Jerry, D. J. 2000. Cytoplasmic sequestration and functional repression of p53 in the mammary epithelium is reversed by hormonal treatment. *Cancer Res.* **60**, 2723-2729.
67. Larsen, P. L. 1993. Aging and resistance to oxidative damage in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. USA* **90**, 8905-8909.
68. Le Bourg, E. 2007. Hormetic effects on longevity of hydrogen peroxide in *Drosophila melanogaster* flies living on a poorly nutritious medium. *Biogerontology* **8**, 327-344.
69. Lee, J. H., Choi, I. Y., Kil, I. S., Kim, S. Y., Yang, E. S. and Park, J. W. 2001. Protective role of superoxide dismutases against ionizing radiation in yeast. *Biochim. Biophys. Acta* **1526**, 191-198.
70. Lee, J. H., Terzaghi, W. and Deng, X. W. 2011. DWA3, an Arabidopsis DWD protein, acts as a negative regulator in ABA signal transduction. *Plant Sci.* **180**, 352-357.
71. Lee, K., Park, J. S., Kim, Y. J., Lee, Y. S., Hwang, T. S., Kim, D. J., Park, E. M. and Park, Y. M. 2002. Differential expression of Prx I and II in mouse testis and their up-regulation by radiation. *Biochem. Biophys. Res. Commun.* **296**, 337-342.
72. Lemaitre, B. and Hoffmann, J. 2007. The host defense of *Drosophila melanogaster*. *Annu. Rev. Immunol.* **25**, 697-743.
73. Lettre, G. and Hengartner, M. O. 2006. Developmental apoptosis in *C. elegans*: a complex CEDnario. *Nat. Rev. Mol. Cell Biol.* **7**, 97-108.
74. Lim, Y. B., Pyun, B. J., Lee, H. J., Jeon, S. R., Jin, Y. B. and Lee, Y. S. 2011. Proteomic identification of radiation response markers in mouse intestine and brain. *Proteomics* **11**, 1254-1263.
75. Liu, C., Gao, F., Li, B., Mitchel, R. E., Liu, X., Lin, J., Zhao, L. and Cai, J. 2011. TLR4 knockout protects mice from radiation-induced thymic lymphoma by downregulation of IL6 and miR-21. *Leukemia* **25**, 1516-1519.
76. Lloyd, T. E. and Taylor, J. P. 2010. Flightless flies: *Drosophila* models of neuromuscular disease. *Ann. NY Acad. Sci.* **1184**, e1-20.
77. Makinde, A. Y., Luo-Owen, X., Rizvi, A., Crapo, J. D., Pearlstein, R. D., Slater, J. M. and Gridley, D. S. 2009. Effect of a metalloporphyrin antioxidant (MnTE-2-PyP) on the response of a mouse prostate cancer model to radiation. *Anticancer Res.* **29**, 107-118.
78. Molin, M., Renault, J. P., Lagniel, G., Pin, S., Toledano, M. and Labarre, J. 2007. Ionizing radiation induces a Yap1-dependent peroxide stress response in yeast. *Free Radic. Biol. Med.* **43**, 136-144.
79. Montero, L., Muller, N. and Gallant, P. 2008. Induction of apoptosis by *Drosophila Myc*. *Genesis* **46**, 104-111.
80. Moskalev, A. 2007. Radiation-induced life span alteration

- of *Drosophila* lines with genotype differences. *Biogerontology* **8**, 499-504.
81. Moskalev, A. A., Plusnina, E. N. and Zainullin, V. G. 2007. The influence of low dose gamma-irradiation on life span of *Drosophila* mutants with defects of DNA damage sensation and repair. *Radiat. Biol. Radioecol.* **47**, 571-573.
  82. Muller, H. J. 1927. Artificial transmutation of the gene. *Science* **66**, 84-87.
  83. Nelson, G. A., Jones, T. A., Chesnut, A. and Smith, A. L. 2002. Radiation-induced gene expression in the nematode *Caenorhabditis elegans*. *J. Radiat. Res.* **43 Suppl**, S199-203.
  84. Nelson, G. A., Schubert, W. W., Kazarians, G. A., Richards, G. F., Benton, E. V., Benton, E. R. and Henke, R. 1994. Radiation effects in nematodes: results from IML-1 experiments. *Adv. Space Res.* **14**, 87-91.
  85. Newcomb, E. W., Lukyanov, Y., Kawashima, N., Alonso-Basanta, M., Wang, S. C., Liu, M., Jure-Kunkel, M., Zagzag, D., Demaria, S. and Formenti, S. C. 2010. Radiotherapy enhances antitumor effect of anti-CD137 therapy in a mouse Glioma model. *Radiat. Res.* **173**, 426-432.
  86. Ollmann, M., Young, L. M., Di Como, C. J., Karim, F., Belvin, M., Robertson, S., Whittaker, K., Demsky, M., Fisher, W. W., Buchman, A., Duyk, G., Friedman, L., Prives, C. and Kopczynski, C. 2000. *Drosophila* p53 is a structural and functional homolog of the tumor suppressor p53. *Cell* **101**, 91-101.
  87. Orme, M. and Meier, P. 2009. Inhibitor of apoptosis proteins in *Drosophila*: gatekeepers of death. *Apoptosis* **14**, 950-960.
  88. Palermo, A. M., Rey, M. and Munoz, E. R. 1994. Protective effect of ethanol on X-ray-induced mitotic recombination in *Drosophila melanogaster*. *Environ. Mol. Mutagen.* **24**, 137-142.
  89. Parashar, V., Frankel, S., Lurie, A. G. and Rogina, B. 2008. The effects of age on radiation resistance and oxidative stress in adult *Drosophila melanogaster*. *Radiat. Res.* **169**, 707-711.
  90. Peters, L. L., Robledo, R. F., Bult, C. J., Churchill, G. A., Paigen, B. J. and Svenson, K. L. 2007. The mouse as a model for human biology: a resource guide for complex trait analysis. *Nat. Rev. Genet.* **8**, 58-69.
  91. Reiter, L. T., Potocki, L., Chien, S., Gribskov, M. and Bier, E. 2001. A systematic analysis of human disease-associated gene sequences in *Drosophila melanogaster*. *Genome Res.* **11**, 1114-1125.
  92. Reliene, R., Pollard, J. M., Sobol, Z., Trouiller, B., Gatti, R. A. and Schiestl, R. H. 2009. N-acetyl cysteine protects against ionizing radiation-induced DNA damage but not against cell killing in yeast and mammals. *Mutat. Res.* **665**, 37-43.
  93. Rithidech, K. N., Honikel, L., Rieger, R., Xie, W., Fischer, T. and Simon, S. R. 2009. Protein-expression profiles in mouse blood-plasma following acute whole-body exposure to (137)Cs gamma rays. *Int. J. Radiat. Biol.* **85**, 432-447.
  94. Roman, G., Endo, K., Zong, L. and Davis, R. L. 2001. P[Switch], a system for spatial and temporal control of gene expression in *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. USA* **98**, 12602-12607.
  95. Rowley, R. and Zhang, J. 1996. Effect of B-type cyclin over-expression on radiation-induced mitotic delay in the fission yeast. *Int. J. Radiat. Biol.* **69**, 565-573.
  96. Rubin, G. M. and Lewis, E. B. 2000. A brief history of *Drosophila's* contributions to genome research. *Science* **287**, 2216-2218.
  97. Ryu, J. H., Ha, E. M., Oh, C. T., Seol, J. H., Brey, P. T., Jin, I., Lee, D. G., Kim, J., Lee, D. and Lee, W. J. 2006. An essential complementary role of NF-kappaB pathway to microbicidal oxidants in *Drosophila* gut immunity. *EMBO J.* **25**, 3693-3701.
  98. Saeki, S., Yamamoto, M. and Iino, Y. 2001. Plasticity of chemotaxis revealed by paired presentation of a chemo-attractant and starvation in the nematode *Caenorhabditis elegans*. *J. Exp. Biol.* **204**, 1757-1764.
  99. Sakashita, T., Hamada, N., Ikeda, D. D., Suzuki, M., Yanase, S., Ishii, N. and Kobayashi, Y. 2008. Locomotion-learning behavior relationship in *Caenorhabditis elegans* following gamma-ray irradiation. *J. Radiat. Res.* **49**, 285-291.
  100. Sakashita, T., Hamada, N., Ikeda, D. D., Yanase, S., Suzuki, M., Ishii, N. and Kobayashi, Y. 2008. Modulatory effect of ionizing radiation on food-NaCl associative learning: the role of gamma subunit of G protein in *Caenorhabditis elegans*. *FASEB J.* **22**, 713-720.
  101. Sakashita, T., Takanami, T., Yanase, S., Hamada, N., Suzuki, M., Kimura, T., Kobayashi, Y., Ishii, N. and Higashitani, A. 2010. Radiation biology of *Caenorhabditis elegans*: germ cell response, aging and behavior. *J. Radiat. Res.* **51**, 107-121.
  102. Sawin, E. R., Ranganathan, R. and Horvitz, H. R. 2000. *C. elegans* locomotory rate is modulated by the environment through a dopaminergic pathway and by experience through a serotonergic pathway. *Neuron* **26**, 619-631.
  103. Schumacher, B., Hofmann, K., Boulton, S. and Gartner, A. 2001. The *C. elegans* homolog of the p53 tumor suppressor is required for DNA damage-induced apoptosis. *Curr. Biol.* **11**, 1722-1727.
  104. Schumacher, B., Schertel, C., Wittenburg, N., Tuck, S., Mitani, S., Gartner, A., Conradt, B. and Shaham, S. 2005. *C. elegans* ced-13 can promote apoptosis and is induced in response to DNA damage. *Cell Death Differ.* **12**, 153-161.
  105. Seong, K. M., Kim, C. S., Lee, B. S., Nam, S. Y., Yang, K. H., Kim, J. Y., Park, J. J., Min, K. J. and Jin, Y. W. 2012. Low-dose Radiation Induces *Drosophila* Innate Immunity through Toll Pathway Activation. *J. Radiat. Res.* **53**, 242-249.
  106. Seong, K. M., Kim, C. S., Seo, S. W., Jeon, H. Y., Lee, B. S., Nam, S. Y., Yang, K. H., Kim, J. Y., Min, K. J. and Jin, Y. W. 2011. Genome-wide analysis of low-dose irradiated male *Drosophila melanogaster* with extended longevity. *Biogerontology* **12**, 93-107.
  107. Shaposhnikov, M. V. and Moskalev, A. A. 2010. FOXO transcription factor role in radiation adaptive response and hormesis in *Drosophila melanogaster*. *Radiat. Biol. Radioecol.* **50**, 312-317.
  108. Shaposhnikov, M. V., Turysheva, E. V. and Moskalev, A.

- A. 2009. Low-dose rate irradiation induced hormesis, hypersensitivity and adaptive response in *Drosophila melanogaster* of radiosensitive strains. *Radiat. Biol. Radioecol.* **49**, 46-54.
109. Siemann, D. W., Hill, R. P. and Penney, D. P. 1982. Early and late pulmonary toxicity in mice evaluated 180 and 420 days following localized lung irradiation. *Radiat. Res.* **89**, 396-407.
110. Simon, J. A. and Bedalov, A. 2004. Yeast as a model system for anticancer drug discovery. *Nat. Rev. Cancer* **4**, 481-492.
111. Song, Y. H. 2005. *Drosophila melanogaster*: a model for the study of DNA damage checkpoint response. *Mol. Cells* **19**, 167-179.
112. Sulston, J. E. and Horvitz, H. R. 1977. Post-embryonic cell lineages of the nematode, *Caenorhabditis elegans*. *Dev. Biol.* **56**, 110-156.
113. Suzuki, M., Sakashita, T., Yanase, S., Kikuchi, M., Ohba, H., Higashitani, A., Hamada, N., Funayama, T., Fukamoto, K., Tsuji, T. and Kobayashi, Y. 2009. Effects of ionizing radiation on locomotory behavior and mechanosensation in *Caenorhabditis elegans*. *J. Radiat. Res.* **50**, 119-125.
114. Tawe, W. N., Eschbach, M. L., Walter, R. D. and Henkle-Duhrsen, K. 1998. Identification of stress-responsive genes in *Caenorhabditis elegans* using RT-PCR differential display. *Nucleic Acids Res.* **26**, 1621-1627.
115. The *C. elegans* Sequencing Consortium. 1998. Genome sequence of the nematode *C. elegans*: a platform for investigating biology. *Science* **282**, 2012-2018.
116. Thompson, T. C., Timme, T. L., Park, S. H., Yang, G. and Ren, C. 2000. Mouse prostate reconstitution model system: A series of in vivo and in vitro models for benign and malignant prostatic disease. *Prostate* **43**, 248-254.
117. Travis, E. L., Down, J. D., Holmes, S. J. and Hobson, B. 1980. Radiation pneumonitis and fibrosis in mouse lung assayed by respiratory frequency and histology. *Radiat. Res.* **84**, 133-143.
118. Vaiserman, A. M. 2010. Radiation hormesis: historical perspective and implications for low-dose cancer risk assessment. *Dose Response* **8**, 172-191.
119. Vaiserman, A. M., Koshel, N. M. and Voitenko, V. P. 2004. Effect of X-irradiation at larval stage on adult lifespan in *Drosophila melanogaster*. *Biogerontology* **5**, 49-54.
120. van Haften, G., Romeijn, R., Pothof, J., Koole, W., Mullenders, L. H., Pastink, A., Plasterk, R. H. and Tijsterman, M. 2006. Identification of conserved pathways of DNA-damage response and radiation protection by genome-wide RNAi. *Curr. Biol.* **16**, 1344-1350.
121. Vanfleteren, J. R. and Braeckman, B. P. 1999. Mechanisms of life span determination in *Caenorhabditis elegans*. *Neurobiol. Aging* **20**, 487-502.
122. Waterston, R. H., Lindblad-Toh, K., Birney, E., Rogers, J., Abril, J. F., Agarwal, P., Agarwala, R., Ainscough, R., Alexandersson, M., An, P., Antonarakis, S. E., Attwood, J., Baertsch, R., Bailey, J., Barlow, K., Beck, S., Berry, E., Birren, B., Bloom, T., Bork, P., Botcherby, M., Bray, N., Brent, M. R., Brown, D. G., Brown, S. D., Bult, C., Burton, J., Butler, J., Campbell, R. D., Carninci, P., Cawley, S., Chiaromonte, F., Chinwalla, A. T., Church, D. M., Clamp, M., Clee, C., Collins, F. S., Cook, L. L., Copley, R. R., Coulson, A., Couronne, O., Cuff, J., Curwen, V., Cutts, T., Daly, M., David, R., Davies, J., Delehaunty, K. D., Deri, J., Dermitzakis, E. T., Dewey, C., Dickens, N. J., Diekhans, M., Dodge, S., Dubchak, I., Dunn, D. M., Eddy, S. R., Elnitski, L., Emes, R. D., Esvara, P., Eyras, E., Felsenfeld, A., Fewell, G. A., Flicek, P., Foley, K., Frankel, W. N., Fulton, L. A., Fulton, R. S., Furey, T. S., Gage, D., Gibbs, R. A., Glusman, G., Gnerre, S., Goldman, N., Goodstadt, L., Grafham, D., Graves, T. A., Green, E. D., Gregory, S., Guigo, R., Guyer, M., Hardison, R. C., Haussler, D., Hayashizaki, Y., Hillier, L. W., Hinrichs, A., Hlavina, W., Holzer, T., Hsu, F., Hua, A., Hubbard, T., Hunt, A., Jackson, I., Jaffe, D. B., Johnson, L. S., Jones, M., Jones, T. A., Joy, A., Kamal, M., Karlsson, E. K., Karolchik, D., Kasprzyk, A., Kawai, J., Keibler, E., Kells, C., Kent, W. J., Kirby, A., Kolbe, D. L., Korf, I., Kucherlapati, R. S., Kulbokas, E. J., Kulp, D., Landers, T., Leger, J. P., Leonard, S., Letunic, I., Levine, R., Li, J., Li, M., Lloyd, C., Lucas, S., Ma, B., Maglott, D. R., Mardis, E. R., Matthews, L., Mauceli, E., Mayer, J. H., McCarthy, M., McCombie, W. R., McLaren, S., McLay, K., McPherson, J. D., Meldrim, J., Meredith, B., Mesirov, J. P., Miller, W., Miner, T. L., Mongin, E., Montgomery, K. T., Morgan, M., Mott, R., Mullikin, J. C., Muzny, D. M., Nash, W. E., Nelson, J. O., Nhan, M. N., Nicol, R., Ning, Z., Nusbaum, C., O'Connor, M. J., Okazaki, Y., Oliver, K., Overton-Larty, E., Pachter, L., Parra, G., Pepin, K. H., Peterson, J., Pevzner, P., Plumb, R., Pohl, C. S., Poliakov, A., Ponce, T. C., Ponting, C. P., Potter, S., Quail, M., Reymond, A., Roe, B. A., Roskin, K. M., Rubin, E. M., Rust, A. G., Santos, R., Sapojnikov, V., Schultz, B., Schultz, J., Schwartz, M. S., Schwartz, S., Scott, C., Seaman, S., Searle, S., Sharpe, T., Sheridan, A., Shownkeen, R., Sims, S., Singer, J. B., Slater, G., Smit, A., Smith, D. R., Spencer, B., Stabenau, A., Stange-Thomann, N., Sugnet, C., Suyama, M., Tesler, G., Thompson, J., Torrents, D., Trevaskis, E., Tromp, J., Ucla, C., Ureta-Vidal, A., Vinson, J. P., Von Niederhausern, A. C., Wade, C. M., Wall, M., Weber, R. J., Weiss, R. B., Wendl, M. C., West, A. P., Wetterstrand, K., Wheeler, R., Whelan, S., Wierzbowski, J., Willey, D., Williams, S., Wilson, R. K., Winter, E., Worley, K. C., Wyman, D., Yang, S., Yang, S. P., Zdobnov, E. M., Zody, M. C. and Lander, E. S. 2002. Initial sequencing and comparative analysis of the mouse genome. *Nature* **420**, 520-562.
123. Watson, A., Mata, J., Bahler, J., Carr, A. and Humphrey, T. 2004. Global gene expression responses of fission yeast to ionizing radiation. *Mol. Biol. Cell* **15**, 851-860.
124. White, J. G., Southgate, E., Thomson, J. N. and Brenner, S. 1986. The Structure of the Nervous System of the Nematode *Caenorhabditis elegans*. *Philos. Trans. R. Soc. Lond., B, Biol. Sci.* **314**, 1-340.
125. Xue, D. and Horvitz, H. R. 1997. *Caenorhabditis elegans* CED-9 protein is a bifunctional cell-death inhibitor. *Nature* **390**, 305-308.

126. Yanase, S., Hartman, P. S., Ito, A. and Ishii, N. 1999. Oxidative stress pretreatment increases the X-radiation resistance of the nematode *Caenorhabditis elegans*. *Mutat. Res.* **426**, 31-39.
127. Yanase, S. and Ishii, N. 2008. Hyperoxia exposure induced hormesis decreases mitochondrial superoxide radical levels via Ins/IGF-1 signaling pathway in a long-lived age-1 mutant of *Caenorhabditis elegans*. *J. Radiat. Res.* **49**, 211-218.
128. Ye, K., Ji, C. B., Lu, X. W., Ni, Y. H., Gao, C. L., Chen, X. H., Zhao, Y. P., Gu, G. X. and Guo, X. R. 2010. Resveratrol attenuates radiation damage in *Caenorhabditis elegans* by preventing oxidative stress. *J. Radiat. Res.* **51**, 473-479.
129. Yokoyama, K., Fukumoto, K., Murakami, T., Harada, S., Hosono, R., Wadhwa, R., Mitsui, Y. and Ohkuma, S. 2002. Extended longevity of *Caenorhabditis elegans* by knocking in extra copies of hsp70F, a homolog of mot-2 (mortalin)/mthsp70/Grp75. *FEBS Lett.* **516**, 53-57.

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초록 : 방사선 생물학을 위한 모델 시스템; 방사선치료의 전임상 연구

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방사선 생물학에서 방사선에 대한 반응으로 매개되는 다양한 기작에 대한 분석을 위해 여러 종류의 모델 생물체를 사용해 왔다. 모델 생물체는 생물학적으로 온전한 *in vivo* 환경을 제공할 수 있기 때문에 방사선에 의해 발생하는 세포 내 현상은 물론 생리적인 현상이나 병리적인 현상을 규명하는 데 있어서 모델 생물체를 사용하는 것은 효과적인 방법이 될 수 있다. 지금까지 축적된, 모델 생물체를 이용한 방사선 생물학적 연구결과들은 새로운 방사선치료 보조제의 개발, 방사선치료 효율 증진 등에 적용되어 여러 질병에 대한 임상연구의 기초가 되어왔다. 이렇게 유용하게 사용된 여러 모델 생물체에 있어서, 각각의 모델에 대한 개별적인 정보에 대한 연구는 다양한 방면에서 이루어지고 있지만, 통합적인 비교, 분석 및 정리를 한 경우는 부족한 실정이다. 따라서, 본 논문에서는 방사선 생물학에서 지금까지 많이 사용된 모델 생물체 4종(효모, 예쁜꼬마선충, 초파리, 생쥐)에 대해 각 생물체가 갖는 모델로서의 특징과 장단점 그리고 방사선 생물학 연구에 이용된 사례 등을 서술하고자 한다.